

1 ***Toxoplasma gondii* infection in farmed wild boars (*Sus scrofa*) in three cities of**

2 **Northeast China**

3

4 **Meng-Jie Bai ¹, Yang Zou ^{1,2}, Hany M. Elsheikha³, Jian-Gang Ma ^{1,2}, Wen-Bin Zheng ^{1,2},**

5 **Quan Zhao ², Xiao-Xuan Zhang ^{1,2,*}, Xing-Quan Zhu^{1,4,*}**

6

7

8

9

10

11

12 ¹ State Key Laboratory of Veterinary Etiological Biology, Key Laboratory of Veterinary
13 Parasitology of Gansu Province, Lanzhou Veterinary Research Institute, Chinese Academy of
14 Agricultural Sciences, Lanzhou, Gansu Province 730046, PR China

15 ² College of Animal Science and Technology, Jilin Agricultural University, Changchun, Jilin
16 Province 130118, PR China

17 ³ Faculty of Medicine and Health Sciences, School of Veterinary Medicine and Science,
18 University of Nottingham, Sutton Bonington Campus, Loughborough, LE12 5RD, UK

19 ⁴ Jiangsu Co-innovation Center for Prevention and Control of Important Animal Infectious
20 Diseases and Zoonoses, Yangzhou, Jiangsu Province 225009, PR China

21 **Abstract**

22 The apicomplexan protozoan parasite *Toxoplasma gondii* is a widely distributed etiological
23 agent of food-borne illness. This parasite can cause production losses in livestock and serious
24 disease in humans through consumption of contaminated meat. Pig's meat is the most likely
25 source of human infection and wild boars may play a role in the transmission of *T. gondii* by
26 serving as a reservoir host. This study aimed to investigate the seroprevalence of antibodies to
27 *T. gondii* among farmed wild boars in China. In an 11-month survey, a total of 882 serum
28 samples were obtained from farmed wild boars from 3 cities (Jilin City, Siping City and
29 Fusong City) in Jilin province, Northeast China and were tested for antibodies specific for *T.*
30 *gondii*. Using modified agglutination test (MAT) and a cut-off titer of 1:25, the prevalence of
31 *T. gondii* infection in the examined samples was 9.9% (88 of 882). The highest
32 seroprevalence was observed in animals from Jilin city (15.3%, 43/281), followed by Siping
33 (11.4%, 30/263) and Fusong (4.4%, 15/338). Logistic regression analysis revealed a
34 significant correlation between the investigated geographic region and *T. gondii* infection.
35 Also, prevalence was higher in females compared to males and the highest prevalence was
36 detected in piglets. These findings indicate that farmed wild boars may become a source of
37 food-borne toxoplasmosis, posing a food safety threat to the public health in the investigated
38 areas. Implementation of effective measures to control *T. gondii* infection in farmed wild
39 boars in China may be warranted.

40

41

42 **Key words:** *Toxoplasma gondii* — Food safety — Seroprevalence — Risk factors —

43 Wild boars — China

44

45 **Introduction**

46 *Toxoplasma gondii*, the causative agent of the disease toxoplasmosis, is a protozoan parasite
47 that can infect virtually all warm-blooded animals and humans (Dubey, 2010; Calero-Bernal
48 *et al.*, 2016; de Souza *et al.*, 2016; Gennari *et al.*, 2016; Jiang *et al.*, 2016). *T. gondii* has a
49 complex lifecycle that encompasses sexual and asexual phases of reproduction. Sexual
50 development occurs in the intestine of the felid, definitive hosts (Frenkel, 1973) and ends up
51 with the formation and shedding of oocysts in the cat feces (Dubey, 2010). Asexual
52 reproduction and formation of bradyzoites-containing tissue cysts occur in the tissues of the
53 intermediate vertebrate host. Animals and humans acquire infection if they ingest food or water
54 contaminated with oocysts (Dubey, 2004). Also, infection can be acquired congenitally
55 through transplacental transmission of tachyzoites or postnatally via consumption of
56 undercooked or raw meat, containing tissue cysts (Elsheikha, 2008; Dubey, 2009). In general,
57 immunocompetent individual infected with *T. gondii* may develop flu-like symptoms.
58 However, infection can cause abortion and stillbirth during pregnancy and fatal consequences
59 in immunocompromised individuals (Montoya and Liesenfeld, 2004; Elsheikha, 2008; Wu *et*
60 *al.*, 2012a).

61 Toxoplasmosis is one of the most prevalent zoonotic parasitic diseases and is reported to
62 be the third-leading cause of foodborne related deaths in the USA (Mead *et al.*, 1999). Global
63 seroprevalence of *T. gondii* varies from 1% to 100% depending on lifestyle, dietary habits
64 (e.g. consumption of undercooked meat), environmental, socioeconomic, and hygienic
65 conditions (Elsheikha, 2008; Furtado *et al.*, 2011). In China, about 7.9% of the population is
66 chronically infected with *T. gondii* and the number of infected people is projected to increase
67 (Jiang *et al.*, 2014). Unfortunately, there is no effective vaccine and anti-*T. gondii*
68 chemotherapeutic drugs have limitations (Montoya and Liesenfeld, 2004; Serranti *et al.*,
69 2011). The extremely broad host range, and the various routes and means by which *T. gondii*

70 can transmit among its hosts add more challenges to effective control of this ubiquitous
71 parasite. Thus, while vaccination in humans or cats might be useful it would not completely
72 solve the problem of *T. gondii* infection as long as parasite transmission between hosts is not
73 prevented. Therefore, control strategies should also aim to minimize the parasite transmission
74 from animals to humans, and this requires more understanding of *T. gondii* epidemiology by
75 obtaining quantitative data on its prevalence.

76 Several studies on the prevalence of *T. gondii* in wild boars have been reported
77 worldwide (Table 1) and *T. gondii* prevalence in pigs has been investigated in some regions of
78 China (Table 2). Wild boar's meat is preferred in China because of its high nutritional value
79 and palatable taste. In the meantime, wild pigs can serve as a vehicle for dissemination of *T.*
80 *gondii* to humans (Choi *et al.*, 1997). These facts suggest that wild boars can potentially serve
81 as a source for human toxoplasmosis in China. There is a need to study the prevalence of *T.*
82 *gondii* in farmed wild boars in China due to the potential public health impact. In an effort to
83 improve the understanding of the epidemiology of toxoplasmosis in wild boars the present
84 study was carried out to determine the seroprevalence and risk factors associated with *T.*
85 *gondii* infection in farmed wild boars in three major cities (Jilin, Siping and Fusong) in
86 Northeast China. Data about the occurrence of *T. gondii* in wild boars in China was
87 discussed with respect to its implications for humans via the food chain and should help in
88 any future parasite risk management strategies.

89

90 **Materials and Methods**

91

92 *Ethics statement*

93 This study was approved by the Animal Ethics Committee of Lanzhou Veterinary
94 Research Institute, Chinese Academy of Agricultural Sciences. The wild boars from which

95 blood samples were collected were handled in strict accordance with good animal practices as
96 stipulated in the Animal Ethics Procedures and Guidelines of the People's Republic of China.

97

98 *Study population and sample collection*

99 This research took place between April 2015 and February 2016, and aimed to assess the
100 seroprevalence of *T. gondii* in free-ranged farmed wild boars foraging on mountain pasture
101 in three Northeastern Chinese cities in Jilin province (Fig. 1), where about 500,000 farmed
102 wild boars were raised. Farms based in Fusong city had a better farming and environmental
103 conditions and the farm's location was far from residential areas. Sampling strategy was
104 optimized in order to obtain a reliable estimate of the prevalence of *T. gondii* in the
105 investigated regions. Therefore, based on prevalence (P) of *T. gondii* in farmed wild boars in
106 Jilin Province of 19.2% (Xu *et al.*, 2015) with an accepted deviation of the true prevalence of
107 5% (d) and a confidence level of 95% ($z = 1.96$) the sample size was calculated as 247
108 [according to $n = P(1 - P)z^2/d^2$]. However, in the present study we aimed to examine more
109 samples to maximize the reliability of the results. We randomly selected 882 wild boars from
110 six piggeries in three cities: Jilin ($n = 281$), Siping ($n = 263$) and Fusong ($n = 338$). Blood
111 samples were collected from the precaval vein of wild boars using 18 gauge, 2.5" needles into
112 5 ml Vacutainer® blood collection tubes (Medical Equipment Factory, Liuyang City, Hubei
113 Province). Sera were separated from blood samples in local veterinary stations and were
114 transported to the laboratory and stored at -20°C until analysis. Information regarding
115 geographic origin, gender, age, and sampling time was collected for each sample.

116

117

118 *Detection of anti-T. gondii antibodies*

119 The modified agglutination test (MAT) was used to detect the antibodies against *T.*
120 *gondii*, and the test was carried out as described previously (Dubey, 2010; Wang *et al.*, 2016).
121 MAT has an acceptable level of sensitivity and specificity for the detection of *T. gondii*
122 antibodies and has been widely used in serological investigations in a wide range of animals
123 worldwide (Gauss *et al.*, 2005; Richomme *et al.*, 2009; Richomme *et al.*, 2010; Wu *et al.*,
124 2012b; Coelho *et al.*, 2014; Qin *et al.*, 2015; Gennari *et al.*, 2016; Wang *et al.*, 2016;). The
125 MAT was performed at serum dilutions of 1:10, 1:25, 1:100 and 1:500. Results are considered
126 positive when obtained at dilution of $\geq 1:25$. Antibody titers $< 1:25$ were considered “suspect”
127 and were retested (Gauss *et al.*, 2005; Wang *et al.*, 2016; Wu *et al.*, 2016b). Positive and
128 negative control sera were included in each test. In the present study, the cut-off of 1:25 was
129 used in accordance with previous investigations (Dubey, 1997; Forbes *et al.*, 20012) and
130 because using more conservative cut-off may risk underestimating the actual prevalence.

131

132 *Statistical analysis*

133 The variation in *T. gondii* seroprevalence (y) of wild boars by gender (x1), sampling time
134 (x2), age (x3) and region (x4) were analyzed by χ^2 test using SAS version 9.3 (SAS Institute
135 Inc., USA). In the multivariable regression analysis, each of these variables was included in
136 the binary Logit model as an independent variable. The best model was validated by Fisher’s
137 scoring algorithm. All tests were two-sided, and when the probability (*P*) value < 0.05 the
138 results were considered statistically significant. Odds ratios (ORs) and their 95% confidence
139 intervals (95% CIs) were obtained to explore the strength of the association between *T.*
140 *gondii*-seropositivity and the risk factors considered above.

141

142 **Results and Discussion**

143 The lack of both *T. gondii* surveillance analysis in wild boars and etiological diagnosis in
144 meat-borne *T. gondii* infection has limited our knowledge about the transmission links of the
145 wild boar-borne *T. gondii* infection in China. Therefore, we conducted a cross-sectional
146 survey from April 2015 to February 2016 to evaluate the sero-prevalence of *T. gondii*
147 infection in 882 farmed wild boars from three cities in Northeast China using the MAT
148 assay.

149

150 *Sero-prevalence data*

151 The study showed that 88 (9.9%, 95% CI 8.00-11.96) of the 882 tested serum samples are
152 seropositive to *T. gondii* tested with titers of 1:25 (n=69), 1:50 (n=15) and 1:100 (n=4)
153 animals (Table 2), suggesting that wild boars are considered a potential source of exposure for
154 humans in China. *T. gondii* seroprevalence reported in our study was lower than that reported
155 in Finland 32.99% (65/197) (Jokelainen *et al.*, 2012) and in Estonia 23.99% (113/471)
156 (Jokelainen *et al.*, 2015) by the direct agglutination test (DAT), and in Poland 37.60%
157 (138/367) (Witkowski *et al.*, 2015) by the multi-species ID Screen (MIDS). It is also lower
158 than the 14-43.13% seroprevalence reported in wild boars in central Italy (Ranucci *et al.*,
159 2013), Romania (Paștiu *et al.*, 2013), Czech Republic (Bártová *et al.*, 2006; Račka *et al.*,
160 2015), Spain (Calero-Bernal *et al.*, 2016), Sweden (Wallander *et al.*, 2015), Korea (Kang *et*
161 *al.*, 2013), Latvia (Deksne *et al.*, 2013), Spain (Gauss *et al.*, 2005), Portugal (Coelho *et al.*,
162 2014), Mediterranean island (Richomme *et al.*, 2010), France (Richomme *et al.*, 2009), and
163 USA (Diderrich *et al.*, 1996). On the other hand, sero-prevalence of *T. gondii* in our study was
164 higher than that reported in Switzerland (6.67%, 10/150) (Berger-Schoch *et al.*, 2011), but

165 similar to that reported in Slovak Republic (8.13%, 26/320) (Antolová *et al.*, 2007) tested by
166 ELISA, and in Japan (6.29%, 11/175) (Matsumoto *et al.*, 2011) by the latex agglutination test
167 (LAT) (Table 1). Interestingly, the wild boars investigated in our study had lower *T. gondii*
168 seroprevalence compared with the remarkably high 11.26-70.00% *T. gondii* seroprevalence
169 reported in intensive pig farms in China (Table 1). Interestingly, the relatively higher
170 prevalence reported in some studies in Europe compared to China (Table 1) could be due to
171 the fact that wild boars investigated in Europe were free ranging and thus have more
172 opportunities to encounter the infection than the investigated animals in our study.

173

174 *Risk factors*

175 The effect of multiple variables on *T. gondii*-seropositivity was assessed by stepwise
176 logistic regression analysis using Fisher's test. In the final model, only the geographic locality
177 from which sera were collected had a significant effect on the risk of *T. gondii* infection
178 (OR=1.876, 95% CI 1.418-2.482). Wild boars tested from Fusong City (4.4%, 15/338; 95%
179 CI 2.24-6.63) had the lowest *T. gondii* seroprevalence compared to animals tested from Jilin
180 City (15.3%, 43/281; 95% CI 11.09-19.51) and Siping City (11.4%, 30/263; 95% CI
181 7.57-15.25); this difference was statistically significant ($P<0.0001$) (Table 2). *T. gondii*
182 transmission cycle in a piggery may be perpetuated by multiple factors, including (i) the
183 contamination of the environment with oocyst from cat feces and (ii) increased susceptibility
184 to infection in piglets due to immature immune defenses. Hence, large numbers of stray cats
185 live in Jilin and Siping may be one of the reasons why seroprevalence of wild boars tested
186 from Jilin and Siping was higher than that of animals from Fusong. This is not surprising, as
187 cats are definitive hosts of *T. gondii* and oocysts secreted by cats represent a major source of
188 *T. gondii* infection to wild boars. It is known that the prevalence of *T. gondii* in pigs can be

189 affected by management systems where in poorly managed systems, seroprevalence in pigs
190 can reach 68% (Gamble *et al.*, 1999). Therefore, the better farming and environmental
191 conditions (i.e. less contamination with oocysts) of wild boars in Fusong may have
192 contributed to the reduction in the frequency by which intermediate host (boars) have access to
193 oocysts of the definitive host, ultimately leading to a lower seroprevalence in wild boars from
194 Fusong.

195 Also, in this study we determined the prevalence of *T. gondii* in males and females,
196 which was found to be 5.34% and 10.79%, respectively (Table 2). This finding is similar to
197 that of previous studies in mice where female mice appeared to be more susceptible to
198 infection with *T. gondii* than male mice (Roberts *et al.*, 1995). However, other studies have
199 not reported any correlation between the gender of pigs and anti-*T. gondii* seropositivity
200 (Alvarado-Esquivel *et al.*, 2015; Gebremedhin *et al.*, 2015). In addition to gender-related
201 differences, we studied *T. gondii* seroprevalence in different age groups, which was found to
202 range from 5.38% to 30.30%. The highest seroprevalence tend to be in young piglets (<22
203 days) and with increasing age this gradually declined by as much as 6-fold for the prevalence
204 in pigs >66 days old. (Table 2). This pattern of early acquisition followed by a gradual decline
205 in prevalence is probably due to different husbandry practice, outdoor piggery density or
206 increased susceptibility in young pigs ((Dubey, 1986).

207 Some limitations should be highlighted. First, the study design did not include collection
208 of meat samples and hence we were not able to isolate any *T. gondii* strains for subsequent
209 genotyping characterization. Second, we were unable to conclusively confirm the associations
210 between geographic region, gender, age, year of study and anti-*T. gondii* seropositivity in the
211 multivariate analysis, probably due to the small ($n = 882$) and unbalanced sample size (619 and

212 263 in the year 2015 and 2016, respectively). Testing 882 animals from three cities is not
213 representative of the prevalence of *T. gondii* in farmed wild boar population in China. The exact
214 basis for association between geographic region, age, gender, year of study remains to be
215 elucidated. It is noteworthy that none of the previous prevalence studies or the present study
216 have conducted direct statistical comparisons between pig populations from different regions
217 and countries. This is probably due to the difficulty associated with controlling the confounding
218 effects caused by differences in pig populations, designs and methodologies of the different
219 studies, such as sampling site, sampling strategy, the type and age of pig included in the survey,
220 specificity and sensitivity of the detection methods, and ecological and geographical factors.

221 In conclusion, we have estimated the prevalence of *T. gondii* in wild farmed boars in
222 Northeast China, which provides new information for assessment of human exposure to *T.*
223 *gondii* through consumption of wild boar's meat. To our knowledge, our study is the first to
224 demonstrate the presence of *T. gondii* infection in (9.9%) farmed wild boars in China, with
225 the highest seroprevalence in Jilin (15.3%), followed by Siping (11.4%) and Fusong (4.4%).
226 Logistic regression analysis indicated that region is a risk factor associated with *T. gondii*
227 infection in the investigated wild boar populations. These results highlight the importance of
228 *T. gondii* in farmed wild boars as a potential pathogen to be considered in the control of
229 food-borne infection risks. The isolation of multiple strains of *T. gondii* known to cause
230 disease in humans would suggest that wild boars are a source/reservoir for *T. gondii* infection
231 in humans in China, although this requires further study. In the future, more research is
232 needed to isolate and characterize the genotype of *T. gondii* strains present in farmed wild
233 boars in order to dissociate the role of wild boars in food-borne *T. gondii* infection and to

234 differentiate foodborne illnesses related to consumption of wild boars from those related to
235 consumption of different types of meat.

236

237 **Acknowledgments**

238 Project support was kindly provided by the National Natural Science Foundation of
239 China (Grant No. 31230073), the Fundamental Research Funds of Chinese Academy of
240 Agricultural Sciences (Grant No. Y2016JC05), and the Agricultural Science and Technology
241 Innovation Program (ASTIP) (Grant No. CAAS-ASTIP-2014-LVRI-03). Laboratoire de
242 Parasitologie-Mycologie, Centre National de Référence de la Toxoplasmose, Centre de
243 Ressources Biologiques Toxoplasma, Hôpital Maison Blanche, Reims Cédex, France, is
244 thanked for providing the Toxoplasma MAT antigen.

245

246 **Author Disclosure Statement**

247 No competing financial interests exist. The findings and conclusions in this report are
248 those of the authors and do not necessarily represent the views of the funding agencies.

249

250 **References**

- 251 Alvarado-Esquivel C, Vazquez-Morales RF, Colado-Romero EE, Guzmán-Sánchez R,
252 Liesenfeld O, Dubey JP. Prevalence of infection with *Toxoplasma gondii* in landrace and
253 mixed breed pigs slaughtered in Baja California Sur State, Mexico. Eur J Microbiol
254 Immunol (Bp) 2015;5(1):112–115.
- 255 Antolová D, Reiterová K, Dubinský P. Seroprevalence of *Toxoplasma gondii* in wild boars
256 (*Sus scrofa*) in the Slovak Republic. Ann Agric Environ Med 2007;14:71–73.
- 257 Bártová E, Sedlák K, Literák I. Prevalence of *Toxoplasma gondii* and *Neospora caninum*
258 antibodies in wild boars in the Czech Republic. Vet Parasitol 2006;142:150–153.

259 Berger-Schoch AE, Bernet D, Doherr MG, Gottstein B, Frey CF. *Toxoplasma gondii* in
260 Switzerland: a serosurvey based on meat juice analysis of slaughtered pigs, wild boar,
261 sheep and cattle. *Zoonoses Public Health* 2011;58:472–478.

262 Calero-Bernal R, Pérez-Martín JE, Reina D, Serrano FJ, Frontera E, Fuentes I, Dubey JP.
263 Detection of zoonotic protozoa *Toxoplasma gondii* and *Sarcocystis suis hominis* in wild
264 boars from Spain. *Zoonoses Public Health* 2016;63:346–350.

265 Coelho C, Vieira-Pinto M, Faria AS, Vale-Gonçalves H, Veloso O, Paiva-Cardoso Md,
266 Mesquita JR, Lopes AP. Serological evidence of *Toxoplasma gondii* in hunted wild boar
267 from Portugal. *Vet Parasitol* 2014;202:310–312.

268 Chang QC, Zheng X, Qiu JH, Wang CR, Zhu XQ. Seroprevalence of *Toxoplasma gondii*
269 infection in fattening pigs in Northeast China. *J Parasitol* 2013;99:544–545.

270 Choi WY, Nam HW, Kwak NH, Huh W, Kim YR, Kang MW, Cho SY, Dubey JP. Foodborne
271 outbreaks of human toxoplasmosis. *J Infect Dis* 1997;175:1280–1282.

272 de Souza JB, Soares VE, Maia MO, Pereira CM, Ferraudo AS, Cruz BC, Pires Teixeira WF,
273 Felippelli G, Maciel WG, Gonçalves WA, Junior, da Costa AJ, Zanetti Lopes WD.
274 Spatial distribution and risk factors for *Toxoplasma gondii* seropositivity in cattle
275 slaughtered for human consumption in Rondônia, North region, Brazil. *Vet Parasitol*
276 2016;226:145–149.

277 Deksne G, Kirjušina M. Seroprevalence of *Toxoplasma gondii* in domestic pigs (*Sus scrofa*
278 domestica) and wild boars (*Sus scrofa*) in Latvia. *J Parasitol* 2013;99:44–47.

279 Diderrich V, New JC, Noblet GP, Patton S. Serologic survey of *Toxoplasma gondii* antibodies
280 in free-ranging wild hogs (*Sus scrofa*) from the Great Smoky Mountains National Park
281 and from sites in South Carolina. *J Eukaryot Microbiol* 1996;43:122S.

282 Dubey J P. A review of toxoplasmosis in pigs. *Vet Parasitol* 1986;19:181–223.

283 Dubey JP. Validation of the specificity of the modified agglutination test for toxoplasmosis in
284 pigs. *Vet Parasitol* 1997;7(4):307–310

285 Dubey JP. *Toxoplasmosis of animals and humans*, 2nd ed. CRC Press, Boca Raton,
286 2010;pp.1–313.

287 Dubey JP. Toxoplasmosis in pigs--the last 20 years. *Vet Parasitol* 2009;164:89–103.

288 Dubey JP. Toxoplasmosis—A waterborne zoonosis. *Vet Parasitol* 2004;126:57–72.

289 Elsheikha HM. Congenital toxoplasmosis: priorities for further health promotion action.
290 *Public Health* 2008;122:335–353.

291 Forbes LB, Parker SE, Gajadhar AA. Performance of commercial ELISA and agglutination
292 test kits for the detection of anti-*Toxoplasma gondii* antibodies in serum and muscle fluid
293 of swine infected with 100, 300, 500 or 1000 oocysts. *Vet Parasitol*
294 2012;190(3-4):362–367

295 Frenkel JK. *Toxoplasmosis: parasite life cycle pathology and immunology*. Hammond, D.M.,
296 Long, P.L. (Eds.), *The Coccidia*, University Park Press, Baltimore 1973; pp.343–410.

297 Furtado JM, Smith JR, Belfort RJr, Gattey D, Winthrop K. Toxoplasmosis: a global threat. *J*
298 *Glob Infect Dis* 2011;3:281–284.

299 Gauss CB, Dubey JP, Vidal D, Ruiz F, Vicente J, Marco I, Lavin S, Gortazar C, Almería S.
300 Seroprevalence of *Toxoplasma gondii* in wild pigs (*Sus scrofa*) from Spain. *Vet Parasitol*
301 2005;131:151–156.

302 Gamble HR, Brady RC, Dubey JP. Prevalence of *Toxoplasma gondii* infection in domestic
303 pigs in the New England states. *Vet Parasitol* 1999;82:129–136.

304 Gebremedhin EZ, Kebeta MM, Asaye M, Ashenafi H, Di Marco V, Vitale M. First report on
305 seroepidemiology of *Toxoplasma gondii* infection in pigs in Central Ethiopia. *BMC Vet*
306 *Res* 2015;11:59.

307 Gennari SM, Niemeyer C, Soares HS, Musso CM, Siqueira GC, Catão-Dias JL, Dias RA,
308 Dubey JP. Seroprevalence of *Toxoplasma gondii* in seabirds from Abrolhos Archipelago,
309 Brazil. *Vet Parasitol* 2016;226:50–52.

310 Huang CQ, Lin YY, Dai AL, Li XH, Yang XY, Yuan ZG, Zhu XQ. Seroprevalence of
311 *Toxoplasma gondii* infection in breeding sows in Western Fujian Province, China. *Trop*
312 *Anim Health Prod* 2010; 42:115–118.

313 Jiang HH, Wang SC, Huang SY, Zhao L, Wang ZD, Zhu XQ, Liu Q. Genetic characterization
314 of *Toxoplasma gondii* isolates from pigs in Jilin Province, Northeastern China.
315 *Foodborne Pathog Dis* 2016;13:88–92.

316 Jiang HH, Zhang WB, Zhao L, Zhou DH, Song HQ, Xu CM, Deng SZ, Zhu XQ.
317 Seroprevalence of *Toxoplasma gondii* infection in pigs in Jiangxi Province, Southeastern
318 China. *Foodborne Pathog Dis* 2014;11:362–365.

319 Jokelainen P, Velström K, Lassen B. Seroprevalence of *Toxoplasma gondii* in free-ranging
320 wild boars hunted for human consumption in Estonia. *Acta Vet Scand* 2015;57:42.

321 Jokelainen P, Näreaho A, Hälli O, Heinonen M, Sukura A. Farmed wild boars exposed to
322 *Toxoplasma gondii* and *Trichinella spp.* *Vet Parasitol* 2012;187:323–327.

323 Kang SW, Doan HT, Noh JH, Choe SE, Yoo MS, Kim YH, Reddy KE, Nguyen TT, Van
324 Quyen D, Nguyen LT, Kweon CH, Jung SC. Seroprevalence of *Toxoplasma gondii* and
325 *Trichinella spiralis* infections in wild boars (*Sus scrofa*) in Korea. *Parasitol Int*
326 2013;62:583–585.

327 Li YN, Nie X, Peng QY, Mu XQ, Zhang M, Tian MY, Min SJ. Seroprevalence and genotype
328 of *Toxoplasma gondii* in pigs, dogs and cats from Guizhou province, Southwest China.
329 *Parasit Vectors* 2015;8:214.

330 Liu X, Liu C, Liu Y, Jin H, Zhao Y, Chen J, Yang M, Liu Q. Seroprevalence of *Toxoplasma*
331 *gondii* infection in slaughtered pigs and cattle in Liaoning Province, northeastern China.
332 J Parasitol 2012;98:440–441.

333 Matsumoto J, Kako Y, Morita Y, Kabeya H, Sakano C, Nagai A, Maruyama S, Nogami S.
334 Seroprevalence of *Toxoplasma gondii* in wild boars (*Sus scrofa leucomystax*) and wild
335 sika deer (*Cervus nippon*) in Gunma Prefecture, Japan. Parasitol Int 2011;60:331–332.

336 Mead PS, Slutsker L, Dietz V, McCaig LF, Bresee JS, Shapiro C, Griffin PM, Tauxe RV.
337 Food-related illness and death in the United States. Emerg Infect Dis 1999;5,pp.607–25.

338 Montoya JG, Liesenfeld O. *Toxoplasmosis*. Lancet 2004;363:1965–1975.

339 Paștiu AI, Györke A, Blaga R, Mircean V, Rosenthal BM, Cozma V. In Romania, exposure to
340 *Toxoplasma gondii* occurs twice as often in swine raised for familial consumption as in
341 hunted wild boar, but occurs rarely, if ever, among fattening pigs raised in confinement.
342 Parasitol Res 2013;112: 2403–2407.

343 Qin SY, Zhou DH, Cong W, Zhang XX, Lou ZL, Yin MY, Tan QD, Zhu XQ.
344 Seroprevalence, risk factors and genetic characterization of *Toxoplasma gondii* in
345 free-range white yaks (*Bos grunniens*) in China. Vet Parasitol 2015;211:300–302.

346 Račka K, Bártová E, Budíková M, Vodrážka P. Survey of *Toxoplasma gondii* antibodies in
347 meat juice of wild boar (*Sus scrofa*) in several districts of the Czech Republic. Ann Agric
348 Environ Med 2015;22:231–235.

349 Ranucci D, Veronesi F, Moretti A, Branciarri R, Miraglia D, Manfredi MT, PiergiliFioretti D.
350 Seroprevalence of *Toxoplasma gondii* in wild boars (*Sus scrofa*) from Central Italy.
351 Parasite 2013;20:48.

352 Richomme C, Afonso E, Tolon V, Ducrot C, Halos L, Alliot A, Perret C, Thomas M, Boireau
353 P, Gilot-Fromont E. Seroprevalence and factors associated with *Toxoplasma gondii*

354 infection in wild boar (*Sus scrofa*) in a Mediterranean island. *Epidemiol Infect*
355 2010;138:1257–1266.

356 Richomme C, Aubert D, Gilot-Fromont E, Ajzenberg D, Mercier A, Ducrot C, Ferté H,
357 Delorme D, Villena I. Genetic characterization of *Toxoplasma gondii* from wild boar
358 (*Sus scrofa*) in France. *Vet Parasitol* 2009;164:296–300.

359 Roberts CW, Cruickshank SM, Alexander J. Sex-determined resistance to *Toxoplasma gondii*
360 is associated with temporal differences in cytokine production. *Infect Immun*
361 1995;63:2549–2555.

362 Serranti D, Buonsenso D, Valentini P. Congenital toxoplasmosis treatment *Eur Rev Med*
363 *Pharmacol Sci* 2011;15:193–198.

364 Tao Q, Wang Z, Feng H, Fang R, Nie H, Hu M, Zhou Y, Zhao J. Seroprevalence and risk
365 factors for *Toxoplasma gondii* infection on pig farms in central China. *J Parasitol*
366 2011;97:262–264.

367 Tenter AM, Heckeroth AR, Weiss LM. *Toxoplasma gondii*: from animals to humans. *Int J*
368 *Parasitol* 2000;30:1217–58.

369 Touloudi A, Valliakos G, Athanasiou LV, Birtsas P, Giannakopoulos A, Papaspyropoulos K,
370 Kalaitzis C, Sokos C, Tsokana CN, Spyrou V, Petrovska L, Billins C. A serosurvey for
371 selected pathogens in Greek European wild boar. *Vet Rec Open* 2015;2:e000077

372 Wang D, Liu Y, Jiang T, Zhang G, Yuan G, He J, Su C, Yang N. Seroprevalence and
373 genotypes of *Toxoplasma gondii* isolated from pigs intended for human consumption in
374 Liaoning province, northeastern China. *Parasit Vectors* 2016;9:248.

375 Wallander C, Frössling J, Vågsholm I, Ugglå A, Lundén A. *Toxoplasma gondii*
376 seroprevalence in wild boars (*Sus scrofa*) in Sweden and evaluation of ELISA test
377 performance. *Epidemiol Infect* 2015;143:1913–1921.

378 Witkowski L, Czopowicz M, Nagy DA, Potarniche AV, Aoanei MA, Imomov N, Mickiewicz
379 M, Welz M, Szaluś-Jordanow O, Kaba J. Seroprevalence of *Toxoplasma gondii* in wild
380 boars, red deer and roe deer in Poland. Parasite 2015;22:17.

381 Wu D, Lv R, Sun X, Shu F, Zhou Z, Nie K, Duan G, Zou F. Seroprevalence of *Toxoplasma*
382 *gondii* antibodies from slaughter pigs in Chongqing, China. Trop Anim Health Prod
383 2012a;44:685–687.

384 Wu SM, Ciren D, Huang SY, Xu MJ, Ga G, Yan C, Mahmoud MS, Zou FC, Zhu XQ. First
385 report of *Toxoplasma gondii* prevalence in Tibetan pigs in Tibet, China. Vector Borne
386 Zoonotic Dis 2012b;12: 654–656.

387 Xu P, Cai YN, Leng X, Wang J, Ma W, Mu GD, Jiang J, Liu XY, Wang ZD, Zhao Q, Yang
388 GL. Seroprevalence of *Toxoplasma gondii* infection in pigs in Jilin Province,
389 Northeastern China. Trop Biomed 2015;32:116–120.

390 Xu Y, Li RC, Liu GH, Cong W, Zhang XX, Yu XL, Zhu XQ. Seroprevalence of *Toxoplasma*
391 *gondii* infection in sows in Hunan province, China. ScientificWorldJournal
392 2014;2014:347908.

393 Yu HJ, Zhang Z, Liu Z, Qu DF, Zhang DF, Zhang HL, Zhou QJ, Du AF. Seroprevalence of
394 *Toxoplasma gondii* infection in pigs, in Zhejiang Province, China. J Parasitol
395 2011;97:748–749.

396 Zhou DH, Liang R, Yin CC, Zhao FR, Yuan ZG, Lin RQ, Song HQ, Zhu XQ. Seroprevalence
397 of *Toxoplasma gondii* in pigs from southern China. J Parasitol 2010;96:673–674.

398 Zou FC, Sun XT, Xie YJ, Li B, Zhao GH, Duan G, Zhu XQ. Seroprevalence of *Toxoplasma*
399 *gondii* in pigs in southwestern China. Parasitol Int 2009;58:306–307.

400
401
402

403 TABLE 1. GLOBAL PREVALENCE OF *TOXOPLASMA GONDII* INFECTION IN WILD BOARS.

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

	<i>Region</i>	<i>Test^a</i>	<i>No. positive/ no. tested</i>	<i>Prevalence (%)</i>	<i>Year</i>	<i>Reference</i>
Europe	Spain	ELISA	688/2881	23.8	2003-2011	Calero-Bernal <i>et al.</i> , 2016
	Spain	MAT	185/517	38.4	1993-2004	Gauss <i>et al.</i> , 2005
	Portugal	MAT	20/97	20.6	2011-2012	Coelho <i>et al.</i> , 2014
	Estonia	DAT	113/471	23.9	2012-2013	Jokelainen <i>et al.</i> , 2015
	Poland	MIDS	138/367	37.6	2009-2011	Witkowski <i>et al.</i> , 2015
	Sweden	ELISA	207/480	43.1	2005-2011	Wallander <i>et al.</i> , 2015
	Central Italy	IFAT	56/400	14	2009-2011	Ranucci <i>et al.</i> , 2013
	USA	MAT	33/108	30.5	1990	Diderrich <i>et al.</i> , 1996
	Romania	IFAT	24/150	16	2008-2010	Paștiu <i>et al.</i> , 2013
	Latvia	ELISA	201/606	33.1	2010-2011	Deksne <i>et al.</i> , 2013
	Finland	DAT	65/197	32.9	2007-2008	Jokelainen <i>et al.</i> , 2012
	Switzerland	ELISA	10/150	6.6	2006-2008	Berger-Schoch <i>et al.</i> , 2011
	Slovak Republic	ELISA	26/320	8.1	2003	Antolová <i>et al.</i> , 2007
	Mediterranean island	MAT	566/1399	40.4	2006-2007	Richomme <i>et al.</i> , 2010
	France	MAT	26/148	17.5	2002–2008	Richomme <i>et al.</i> , 2009
	The Czech Republic	ELISA	148/565	26.1	1999-2005	Bártová <i>et al.</i> , 2006
	The Czech Republic	ELISA	260/656	39.6	2008-2010	Račka <i>et al.</i> , 2015
	Greek	ELISA	5/94	5.2	2006-2010	Touloudi <i>et al.</i> , 2015
Asia	Korea	ELISA	131/521	25.1	2009-2011	Kang <i>et al.</i> , 2013
	Japan	LAT	11/175	6.2	2004-2007	Matsumoto <i>et al.</i> , 2011
	Guizhou, China	ELISA	49/70	70	2011-2012	Li <i>et al.</i> , 2015

Liaoning, China	MAT	233/2063	11.2	2013-2014	Wang et al., 2016
Jilin, China	IHA	236/1235	19.1	2013	Xu et al., 2015
Hunan, China	IHA	373/1191	31.3	2010-2012	Xu et al., 2014
Jiangxi, China	IHA	282/1232	22.8	2012	Jiang et al., 2014
Heilongjiang, China	IHA	47/1,014	4.6	2011-2012	Chang et al., 2013
Chongqing, China	IHA	278/908	30.6	UN ^b	Wu et al., 2012a
Liaoning, China	IHA	140/1,164	12.03	2011	Liu et al., 2012
Tibet, China	MAT	97/427	22.7	2010	Wu et al., 2012b
Zhejiang, China	ELISA	434/813	53.3	2009-2010	Yu et al., 2011
Central China	IHA	873/3,558	24.5	2008-2009	Tao et al., 2011
Guangdong, China	ELISA	276/1,022	27.01	2008-2009	Zhou et al., 2010
Fujian, China	IHA	87/605	14.3	2006-2007	Huang et al., 2010
Yunnan, China	IHA	141/831	16.9	2008-2009	Zou et al., 2009

422

423 ^aELISA, enzyme-linked immunosorbent assay; IHA, indirect hemagglutination test; MAT, modified agglutination test; DAT, direct agglutination
424 test; LAT, latex agglutination test; IFAT, immunofluorescence antibody test; MIDS, multi-species ID Screen.

425 ^bUN, unknown.

426 TABLE 2. SEROPREVALENCE OF *TOXOPLASMA GONDII* INFECTION IN WILD BOARS IN THREE CITIES IN NORTHEAST CHINA
 427 BY REGION, GENDER, AGE AND COLLECTION YEAR.
 428

Variable	Category	Sera with different MAT titers			No. tested	No. positive	Prevalence (%) (95%CI)	P-value	OR (95% CI)
		1:25	1:50	1:100					
Region	Fusong	11	2	2	338	15	4.4 (2.24-6.63)	<0.0001	1.0 (Reference)
	Siping	25	5	0	263	30	11.4 (7.57-15.25)		2.77 (1.46-5.27)
	Jilin	33	8	2	281	43	15.3 (11.09-19.51)		3.89 (2.11-7.17)
Gender	Male	6	1	0	131	7	5.3 (1.49-9.20)	0.0551	1.0 (Reference)
	Female	63	14	4	751	81	10.7 (8.57-13.00)		2.14 (0.97-4.75)
Age	<22 days ^a	8	2	0	33	10	30.3 (14.62-45.98)	<0.0001	1.0 (Reference)
	22-66 days	6	4	2	233	12	5.3(2.42_8.34)		7.65 (2.98-19.63)
	>66 days	55	9	2	626	66	10.5 (8.14-12.95)		2.07 (1.10-3.91)
Collection year	2015	441	10	4	619	58	9.3 (7.07-11.67)	0.3558	1.0 (Reference)
	2016	25	5	0	263	30	11.4 (7.57-15.25)		1.25 (0.78-1.99)
Total		69	15	4	882	88	9.98 (8.00-11.96)		

429 ^a pre-weaned.
 430
 431
 432
 433
 434
 435
 436

437 **Figure Legend:**

438

439 Fig. 1. Map showing three cities, Jilin, Siping and Fusong, in Jilin province, Northeast China where the serum samples have been collected.

440

441

442

443

444

445

446

447

448

449

450

451

452

Address correspondence to:

Xiao-Xuan Zhang PhD and Xing-Quan Zhu PhD

State Key Laboratory of Veterinary Etiological Biology,

Key Laboratory of Veterinary Parasitology of Gansu Province,

Lanzhou Veterinary Research Institute, Chinese Academy of Agricultural Sciences,

Lanzhou, Gansu Province 730046, PR China

E-mail: zhangxiaoxuan1988@126.com (X.X. Zhang);

xingquanzhu1@hotmail.com (X.Q. Zhu)

