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Co-exposure of C60 fullerene with benzo[a]pyrene results in enhanced biological effects in cells as determined by Fourier-transform infrared spectroscopy

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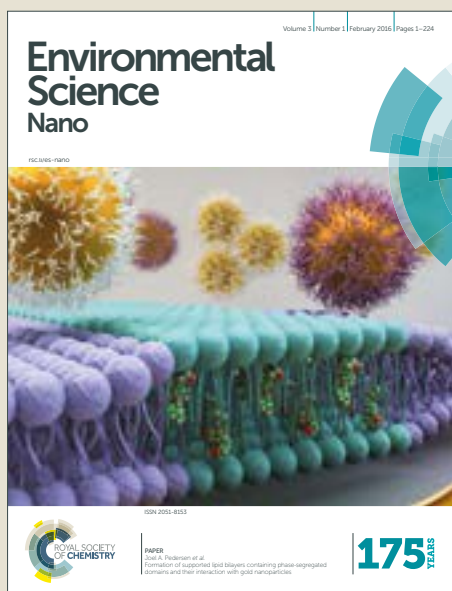
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Environmental Significance Statement

There is an urgent need for approaches capable of lending insights into nanoparticle (NP)-induced effects in biological cells. Conventional assays such as those employing genotoxicity endpoints remain inconsistent. With increasing usage, carbon-based NPs are entering the environment and their effects either directly or in combination with other environmental contaminants remain to be understood. This study primarily exploits FTIR spectroscopy to derive signature fingerprints of cellular material based on chemical composition. Using computational algorithms to process spectral datasets, alterations post-exposure to C₆₀ fullerene (C₆₀) with or without benzo[*a*]pyrene (B[*a*]P) were investigated. Exposure-induced spectral data points to gene expression and oxidative damage alterations; this is subsequently shown using more conventional assays. Low-dose C₆₀ increased B[*a*]P-induced alterations, while alterations at high C₆₀ concentrations appeared absent. This suggests that the interactions between NPs with toxic chemical contaminants are complex and remain to be fully understood.

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Co-exposure of C₆₀ fullerene with benzo[*a*]pyrene results in enhanced biological effects in cells as determined by Fourier-transform infrared spectroscopy

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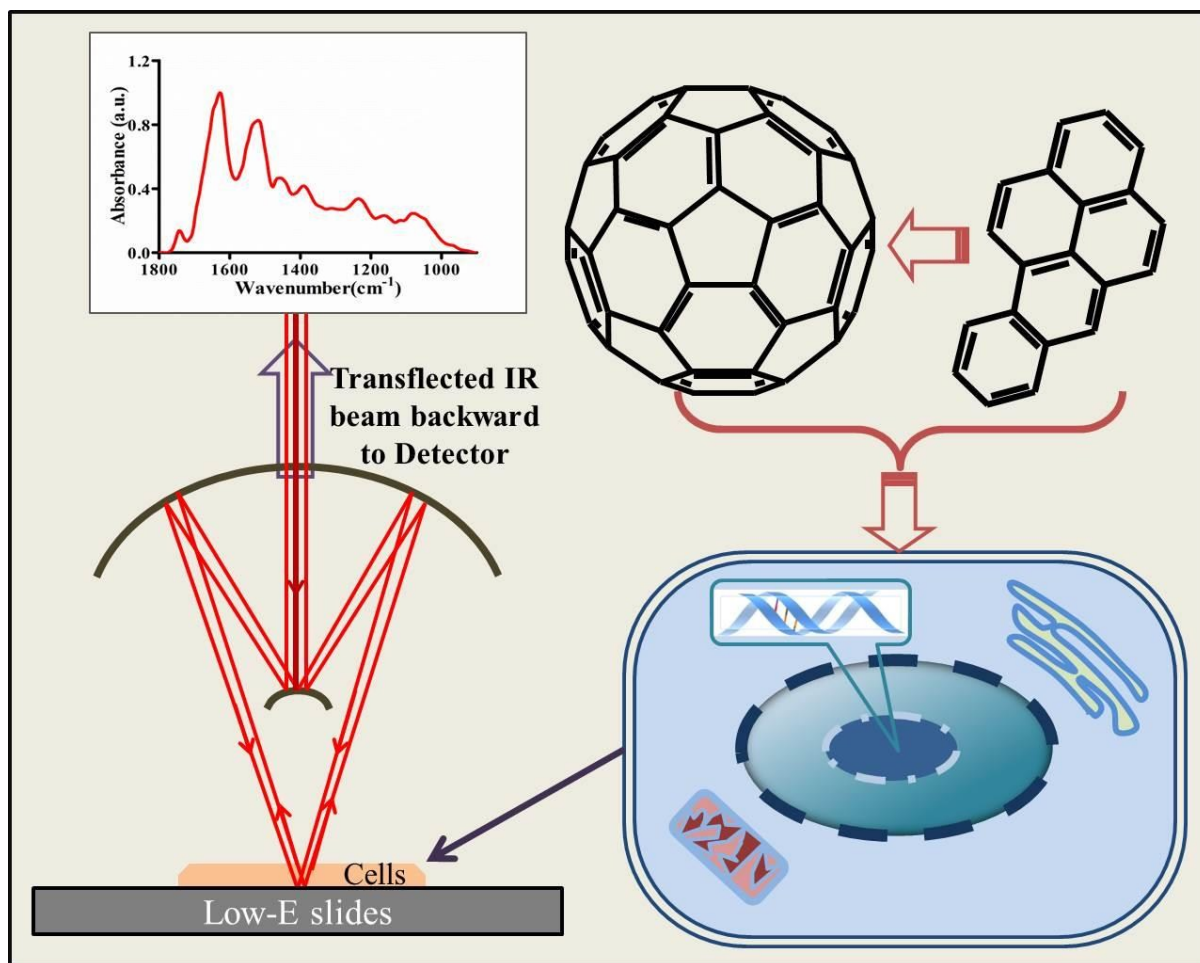
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TOC graphic

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Biospectroscopy signatures effects of binary mixture of C₆₀ fullerene and benzo[*a*]pyrene in cells



AbstractView Article Online
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C₆₀ fullerene (C₆₀) is a promising manufactured carbon-based nanoparticles (NPs). With an increasing number of applications, it is being found in the environment. In addition, C₆₀ is likely to associate with other environmental toxic contaminants. How such interactions with C₆₀ can impact on the environmental fate, transport and bioavailability of toxicants remains unknown. Benzo[*a*]pyrene (B[*a*]P) is a polycyclic aromatic hydrocarbon (PAH). Herein, two cell lines (fish gill or MCF-7 cells) were employed to explore the biological impacts of co-exposure to C₆₀ and B[*a*]P. Post-exposure cells were interrogated using Fourier-transformation infrared (FTIR) microspectroscopy. By inputting spectral data into principal component analysis and linear discriminant analysis, data reduction allowed for visualisation of cell categorization and identification of wavenumber-related biomarkers corresponding to cellular alterations. Our results indicate that low-dose C₆₀ increases B[*a*]P-induced alterations, while C₆₀ at high concentrations reduces these effects. We also found that although C₆₀ co-exposure increases B[*a*]P-induced *CYP1A1* induction, co-exposure seemingly attenuates the levels of oxidative damage induced by either agent singly. This suggests that interactions between environmental NPs and contaminants are complex and unpredictable.

Introduction

With the rise of nanotechnology, there has been a rapid increase in the commercial use of nanoparticles (NPs). However, little is known regarding the fate and behaviour of engineered NPs in the environment, and concerns have emerged regarding their potential impact on human health.^{1,2} Furthermore, quantitative analytical methods are required to determine environmental concentrations and, enable both effect and exposure assessments. Many methods still need optimization and development, especially for new types of NPs.³⁻⁶ There is an urgent need for analytical methods to adequately assess the risk of NPs.

C₆₀ fullerene (C₆₀), the first manufactured NP, possesses unique physical and chemical properties, which makes it a candidate agent for many nanotechnological applications in industrial and medical fields.⁷⁻¹⁰ However, its extremely small size, unique conformation, large surface area, and propensity for surface modification raise the possibility that C₆₀ could pose a hazard to humans and other living organisms.¹¹ It seems that the cytotoxicity of C₆₀ differs depending on the type of cells exposed and how test suspensions are prepared.^{12,13}

Included in the debate regarding NP-induced acute toxicity, there are emerging concerns about their release into the environment in that NPs may not only just interfere with biological systems, but also may interact with other contaminants such as polycyclic aromatic hydrocarbons (PAHs). Consequently, NPs could affect the fate, transportation and bioavailability of pollutants in binary mixtures. In aquatic environments, contaminants can accumulate in aqueous NPs and this accumulation appears to affect the physicochemical property of both NP and the co-contaminant.¹⁴ Investigations show that NPs seem to be highly reactive in their interactions with other contaminants. Furthermore, it has been noted that nano-silica could facilitate the cellular uptake of metals, and induce higher levels of

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3 damage than that induced by metal or nano-silica alone.¹⁵ This type of delivery mechanism is
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5 the so called ‘Trojan horse’ effect. In another example, it was found that nano-TiO₂ enhanced
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7 the bioaccumulation and toxicity of copper in *Daphnia magna*.¹⁶ However, studies
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9 investigating the effects of co-contamination with carbon-based NPs seem to be less
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11 conclusive, although carbon nanomaterials appear to be highly interacting with chemicals in
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13 the environment.¹⁷⁻¹⁹ Single-walled carbon nanotubes were found to act as a contaminant
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15 carrier and enhance the accumulation of phenanthrene in the digestive track of fish.²⁰ Another
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17 study suggests that co-exposure with carboxyl-functionalized single-walled carbon nanotubes
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19 significantly inhibits the bioactivity of adsorbed 17 α -ethinylestradiol (EE₂) in cultured cells.²¹
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Polycyclic aromatic hydrocarbons (PAHs) are a class of widespread organic compounds with two or more fused aromatic rings; they have a relatively low solubility in water, but are highly lipophilic.²² Benzo[*a*]pyrene (B[*a*]P) is a PAH that is pro-carcinogenic. It is a potent ligand for the cytosolic aryl hydrocarbon receptor (AhR), which may mediate teratogenic and carcinogenic effects of certain environmental pollutants.²³ In cells, B[*a*]P can bind to AhR and activate it, or it is effectively metabolised by several xenobiotic metabolizing enzymes to B[*a*]P-7,8-diol-9,10-epoxide (BPDE), which is the ultimate carcinogenic form, and generates bulky chemical-DNA adducts.²⁴⁻²⁶

It is highly possible that carbon-based nanomaterials would interact with PAHs and be a co-contaminating influence in the environment. In order to understand how this kind of co-exposure will impact on toxicity, cells were co-exposed to C₆₀ with B[*a*]P. Effects in cells were then assessed using Fourier-transform infrared (FTIR) microspectroscopy. Such biospectroscopy provides a rapid, reagent-free and non-destructive method for biological analysis.²⁷ Therefore, IR spectroscopy has been widely applied in biological research, for disease diagnosis²⁸, stem cell characterisation²⁹ and toxicity assessment³⁰. The mid-IR region

(4000-400 cm^{-1}) is specific for biological application and it provides abundant chemical bond information in the 'biochemical-fingerprint' region (1800-900 cm^{-1}), through identifiable peaks at specific absorption frequencies: Amide I (~1650 cm^{-1}), Amide II (~1550 cm^{-1}), Amide III (~1260 cm^{-1}), carbohydrates (~1155 cm^{-1}), glycogen (~1030 cm^{-1}), lipids (~1750 cm^{-1}), asymmetric phosphate stretching vibrations ($\nu_{as}\text{PO}_2^-$; ~1225 cm^{-1}), symmetric phosphate stretching vibrations ($\nu_s\text{PO}_2^-$; ~1225 cm^{-1}) and protein phosphorylation (~970 cm^{-1}).³¹ Coupled with computational multivariate analyses, FTIR spectroscopy is a sensitive bioanalytical tool.

In this study, two cell lines, including a fish gill cell line and a mammalian cell line were used to examine the *in vitro* biological effects following co-exposure to C₆₀ and B[a]P; the cellular response was determined using FTIR spectroscopy. In line with previous investigations, three relatively low doses were employed (B[a]P exposure concentrations at 10⁻⁶ M, 10⁻⁷ M and 10⁻⁸ M; C₆₀ at 0.1 mg/L, 0.01 mg/L and 0.001 mg/L).^{32, 33} Spectral alterations were associated with effects on AhR-inducible *CYP1A1*, DNA damage inferred by *P21^{WAF1/CIP1}* and oxidative stress [measured by thiobarbituric acid reactive substances (TBARS) and levels of intracellular reactive oxygen species (ROS)]. This study aimed to gain insights into a binary exposure including a carbon-based NP and chemical contaminant.

Materials and Methods

Chemicals and carbon nanoparticles (NPs)

All test agents were purchased from Sigma. B[a]P was HPLC-grade (>96%) in powder-form, while dimethyl sulfoxide (DMSO) used as solvent was GC-grade (≥99.5%). Bovine serum albumin (BSA), also obtained from Sigma, was ≥98% pure. C₆₀ (from Sigma) had a purity >99.5% and particle size of 1 nm. It was analysed by Raman spectroscopy (Renishaw PLC, Gloucestershire, UK) with a 785 nm laser, and determined to be of high

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purity. Additionally, images of C₆₀ were taken using a scanning electron microscope (SEM) [JSM 5600 (JEOL)] [see Electronic Supporting Information (ESI) Figure S1].

B[a]P was dissolved in DMSO, and stock solutions were made at concentrations of 10⁻³ M, 10⁻⁴ M and 10⁻⁵ M. C₆₀ were dispersed in 1% BSA solution following a 15-min ultrasonication in an ice-water bath and stock solutions were made at concentrations of 100 mg/L, 10 mg/L and 1 mg/L. Agglomeration could be observed in the solution due to its lipophilic properties. To ensure a homogeneous mixture of chemical agents, solvent or exposure medium, and to avoid any solvent-specific effects, stock solutions and exposure medium were mixed prior to application to the cells. Accordingly, as NPs and B[a]P were dispersed in 1% BSA solution and DMSO respectively, each experimental medium contained a final level of 0.1% (vol/vol) 1% BSA solution and DMSO. An experimental medium containing 0.1% (vol/vol) 1% BSA solution and DMSO without test agent was used as vehicle control. All experimental media were prepared 72 h prior to cell exposure and stored at 4°C, which allowed absorption equilibration of B[a]P onto C₆₀. Mixture treatment compositions were as follows: Mix 1, 10⁻⁶ M B[a]P + 0.001 mg/L C₆₀; Mix 2, 10⁻⁶ M B[a]P + 0.01 mg/L C₆₀; Mix 3, 10⁻⁶ M B[a]P + 0.1 mg/L C₆₀; Mix 4, 10⁻⁷ M B[a]P + 0.1 mg/L C₆₀; and, Mix 5, 10⁻⁸ M B[a]P + 0.1 mg/L C₆₀.

Cell culture

Human breast cancer MCF-7 cells were cultured in Dulbecco's modified essential medium (DMEM) supplemented with 10% heat-inactivated foetal bovine serum, penicillin (100 U/mL), and streptomycin (100 µg/mL). MCF-7 cells were maintained in a humidified atmosphere with 5% CO₂ in air at 37°C. Gill cells for primary cultures were derived from gills of rainbow trout (*Oncorhynchus mykiss*)³⁴. The cells were cultured with Leibovitz's L-15 culture media supplemented with 10% heat-inactivated foetal bovine serum, penicillin

(100 U/mL), and streptomycin (100 µg/mL). These gill cells were maintained without CO₂ incubation in free gas exchange with air at an optimal temperature (~18°C). These cell lines were routinely cultured in 75 cm² cell culture flasks. MCF-7 cells were split twice a week, while gill cells were split once a week.

Exposure protocol

For MCF-7 cells, the same medium was used for incubation before and during exposure (namely exposure medium), while L15 medium was only used for gill cells prior to exposure, but a different medium, L15/ex, was then introduced as the experimental medium during exposures. L-15/ex medium was initially validated in the RTgill-w1 cell line. L15/ex medium contains only salts, galactose and pyruvate to provide an isotonic environment, and a source of energy; as such, it is fully defined^{35, 36}. Bioavailability of test chemicals in this *in vitro* system is not influenced by a serum component. Binding of hydrophobic test chemicals to constituents of serum was suggested to contribute to the systematic deviation of mammalian cell viability *versus* fish acute toxicity depending on the chemicals' octanol-water partition coefficient (Kow) and is also a likely cause of under-estimation of fish acute toxicity using fish cell lines³⁶.

For exposure, cells were directly grown on Low-E slides (Kevley Technologies, OH, USA) in 45-mm culture dishes. Prior to cell seeding, Low-E slides were immersed in ethanol for 30 min. Following rinsing in sterile water, Low-E slides were then stored in empty culture dishes and dried in an incubator. Confluent cells in 75 cm² flasks were disaggregated with trypsin (0.05%)/EDTA (0.02%) solution, and were immediately re-suspended in complete medium. Cells were then seeded to the culture dishes and allowed to attach on the slides and to form a cell layer on the Low-E slides. After 72 h, the original medium was removed and experimental medium containing test agents (or not) was added. After 24-h incubation, the

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cells on Low-E slides were rinsed in PBS and fixed in 4% formalin in PBS for 30 min. Once fixed, the slides were rinsed in PBS and given a quick wash (~3 sec) in distilled water. Following air-drying overnight, slides were stored in a desiccator until analysis.

Spectrochemical analysis

All cell samples on Low-E slides were interrogated using a Bruker TENSOR 70 FTIR spectrometer (Bruker Optics Ltd., Germany) equipped with a HYPERION 1000 microscope containing a liquid nitrogen-cooled detector. Instrument parameters were set at 32 scans and 8 cm⁻¹ resolution. For each slide, some 20 IR spectra were acquired at different points across the sample. Prior to starting a new slide or after each ten spectra, a background was taken.

Spectral data acquired from FTIR spectroscopy were processed using IRootLab toolbox (<http://trevisanj.github.io/irootlab/>) running on MATLAB r2010a (The MathWorks, Inc., US). IR spectra were pre-processed as follows: cut to 1800-900 cm⁻¹ (the biochemical fingerprint range), rubberband baseline corrected, and normalisation to Amide I peak. Computational analysis using multivariate techniques included principal component analysis (PCA) and linear discriminant analysis (LDA), which can efficiently analyse large spectral datasets. Following pre-processing, PCA was applied to the spectral dataset. PCA is an unsupervised technique employed to reduce the dimensions of the data. Undoubtedly, PCA is capable of identifying some important biochemical information in the spectral data. However, it has less discriminating power due to the fact that it is an unsupervised procedure. In order to interpret such complex biochemical information, further data analysis by using supervised procedures such as LDA is often applied. Thus, the output derived from PCA was inputted into LDA³⁷. The first ten PC factors from PCA were used for LDA since that accounted for >99% variance³¹. Multivariate analysis results were visualized either as scores plots and/or cluster vectors plot. In scores plots, nearness between two groups implies similarity,

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3 while distance indicates dissimilarity^{38, 39}. Cluster vectors plots from PCA-LDA help to View Article Online
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6 reveal the biochemical alterations associated with each category in the dataset. To simplify
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8 the identification of the main biochemical alterations of each group, cluster vectors peak plots
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10 were used to indicate the top seven peaks in the cluster vectors plots.
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13 14 **Quantitative real-time reverse transcriptase (RT)-PCR**

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16 Routinely-cultured MCF-7 cells were disaggregated and re-suspended in complete
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18 medium (DMEM, 10% FCS) prior to seeding aliquots (5 ml, $\approx 1 \times 10^5$ cells) into 60-mm petri
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20 dishes⁴⁰. Following 24-h incubation, cells were treated (using exposure medium as described
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22 above) for a further 24 h with 10^{-8} M B[a]P, 0.1 mg/L C₆₀ fullerene or a combination of both;
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24 a vehicle control was also included. Cells were then washed twice with PBS prior to lysis and
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26 total RNA extraction using the Qiagen RNeasy® Kit in combination with the Qiagen RNase-
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28 free DNase kit (QIAGEN Ltd, Crawley, UK). DNase was incorporated into the extraction
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30 procedure in order to remove residual DNA, *e.g.*, pseudogene. RNA quality was routinely
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32 assessed in a 1.2% formaldehyde agarose gel; yield and purity were checked using a
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34 spectrophotometer. RNA (0.4 μ g) was reverse transcribed in a final volume of 20 μ l
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36 containing Taqman® reverse transcription reagents (Applied Biosystems, Warrington, UK):
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38 1 \times Taqman reverse transcriptase (RT) buffer; MgCl₂ (5.5 mM); oligo d(T)₁₆ (2.5 μ M); dNTP
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40 mix (dGTP, dCTP, dATP and dTTP; each at a concentration of 500 μ M); RNase inhibitor
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42 (0.4 U/ μ l); RT (MultiScribe™) (1.25 U/ μ l) and RNase-free water. Reaction mixtures were
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44 then incubated at 25°C (10 min), 48°C (30 min) and 95°C (5 min).
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52 cDNA samples were stored at -20°C prior to use. Primers (Table 1) for cyclin-
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54 dependent kinase inhibitor 1A [*CDKN1A* (*P21*^{WAF1/CIP1}, GenBank accession no. NM_078467)]
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56 and *CYP1A1* (GenBank accession no. BC023019) and endogenous control β -*ACTIN*
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58 (GenBank accession no. AK222925) were chosen using Primer Express software 2.0
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60 (Applied Biosystems) and designed so that one primer spanned an exon boundary. Specificity

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3 was confirmed using the NCBI BLAST search tool. Quantitative real-time PCR was View Article Online
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5 performed using an ABI Prism 7000 Sequence Detection System (Applied Biosystems).
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7 Reaction mixtures contained 1 × SYBR® Green PCR master mix (Applied Biosystems);
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9 forward and reverse primers (Invitrogen Life Technologies) at a concentration of 300 nM
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11 (*P21^{WAF1/CIP1}*, *CYP1A1* or *β-ACTIN*); for *P21^{WAF1/CIP1}* or *CYP1A1* amplification 20 ng cDNA
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13 template or for *β-ACTIN* amplification 5 ng cDNA template; made to a total volume of 25 μl
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15 with sterile H₂O. Thermal cycling parameters included activation at 95°C (10 min) followed
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17 by 40 cycles each of denaturation at 95°C (15 sec) and annealing/extending at 60°C (1 min).
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19 Each reaction was performed in triplicate and ‘no-template’ controls were included in each
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21 experiment. Dissociation curves were run to eliminate non-specific amplification, including
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23 primer dimers. In control cell populations, averaged threshold cycle values of amplified
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25 cDNA were in the 25-30 range for *CYP1A1*.
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33 **The thiobarbituric acid (TBA) assay for lipid peroxidation**

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35 Lipid peroxidation was measured as a function of TBA reactive substances (TBARS),
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37 including malondialdehyde-TBA adduct production^{41, 42}. Routinely cultured MCF-7 cells
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39 were disaggregated and re-suspended in complete medium (DMEM, 10% FCS) prior to
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41 seeding aliquots (10 ml, ≈1 × 10⁶ cells) into 75 cm² flasks. Following 24-h incubation, cells
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43 were treated (using exposure medium as described above) for a further 24 h with 10⁻⁸ M
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45 B[a]P, 0.1 mg/L C₆₀ fullerene or a combination of both; a vehicle control was also included.
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47 Cells were harvested by scraping, washing with PBS and re-suspension in deionized water
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49 containing 8.1% SDS, 20% acetic acid and 0.8% thiobarbituric acid. Resultant mixtures were
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51 incubated in a boiling water bath for 1 h. After cooling, *n*-butanol:pyridine mixture (15:1,
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53 v/v) was added and the reaction mixtures were centrifuged at 1600 g for 15 min. In decanted
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55 supernatants, malondialdehyde was assayed at 532 nm. TBARS concentrations were
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expressed as nmol/mg protein, as determined using the Bradford method, using 1,1,3,3-tetraethoxypropane as a reference standard. Results are presented as the Mean \pm SD of five independent experiments, each performed in duplicate.

Determination of intracellular reactive oxygen species (ROS) levels

The fluorescent probe to oxidative damage, 2',7'-dichlorofluorescein diacetate (DCFH-DA), in combination with flow cytometry was employed. Routinely cultured MCF-7 cells were disaggregated and re-suspended in complete medium (DMEM, 10% FCS) prior to seeding aliquots (10 ml, $\approx 1 \times 10^6$ cells) into 75 cm² flasks. Following 24-h incubation, cells were treated (using exposure medium as described above) for a further 24 h with 10⁻⁸ M B[a]P, 0.1 mg/L C₆₀ fullerene or a combination of both; a vehicle control was also included. Thirty min prior to the end of the above incubation period, DCFH-DA was added to each incubation mix; the underlying principle is that upon diffusion across the lipid cell membrane, deacetylation by intracellular esterases generates 2',7'-dichlorodihydrofluorescein (DCFH₂). With intracellular ROS generation, DCFH₂ is oxidized to 2',7'-dichlorofluorescein (DCF), a highly fluorescent end-product. At the end of the incubation period, cells were disaggregated with warm trypsin/EDTA and washed with PBS, upon which intracellular ROS levels were determined using a FACScan flow cytometer (Becton Dickinson) at 488 nm (excitation) and 525 nm (emission). For each treatment, a minimum of 10,000 events were collected and analysed by CellQuest software. Results are presented as the Mean \pm SD of five independent experiments, each performed in duplicate.

Results and discussion

Post-exposure to the test agents studied herein, cells were harvested and interrogated by FTIR spectroscopy. This analysis gave rise to a large spectral dataset containing 12

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classed categories, which were labelled according to the treatments as: Control; C₆₀ 0.1 mg/L; C₆₀ 0.01 mg/L; C₆₀ 0.001 mg/L; B[a]P at 10⁻⁸ M; B[a]P at 10⁻⁷ M; B[a]P at 10⁻⁶ M; Mix 1 (C₆₀ 0.001 mg/L & B[a]P at 10⁻⁶ M); Mix 2 (C₆₀ 0.01 mg/L & B[a]P at 10⁻⁶ M); Mix 3 (C₆₀ 0.1 mg/L & B[a]P at 10⁻⁶ M); Mix 4 (C₆₀ 0.1 mg/L & B[a]P at 10⁻⁷ M); and, Mix 5 (C₆₀ 0.1 mg/L & B[a]P at 10⁻⁸ M). Parallel sets of experiments were conducted in the MCF-7 cell and RTgill-w1 cell lines. Further analyses for *CYP1A1* induction, lipid peroxidation and generation of intracellular ROS were conducted at selected concentrations in MCF-7 cells.

As specific wavenumbers in IR spectra may be used as markers of chemical entities in cells, their intensity absorbance following simple pre-processing (including baseline correction and normalization) may be used to assess treatment-induced alterations in target cells. The intensity absorbance at 1400 cm⁻¹ is used as a protein marker as it corresponds to C=O symmetric stretching of amino acids and 1740 cm⁻¹ is associated with C=O stretching vibrations of lipids. Thus, the protein-to-lipid ratio using the intensity absorbance at these two vibration modes may assess cell proliferation post-exposure³⁸. Similarly, other absorbance ratios are employed including: intensity absorbance ratio of 1650 cm⁻¹ to 1084 cm⁻¹ as ratio of protein/nucleic acid (1650 cm⁻¹ corresponding to Amide I in protein; 1084 cm⁻¹ corresponding to ν_3 PO₂⁻ for DNA/RNA)⁴³, and intensity ratio of (996 cm⁻¹)/(966 cm⁻¹) used as RNA/DNA ratio^{43, 44}.

A tendency for an elevated protein-to-lipid ratio indicates that the test agents used give rise to active cell proliferation⁴⁵. Our findings suggested that most of the treatments in gill cells significantly activated cell proliferation compared with the control, while in MCF-7 cells, only exposure of Mix 4 induced significant proliferation (Figure 1a; see ESI Table S1). In contrast to the protein-to-lipid ratio, the protein-to-nucleic acid ratio exhibited a downregulation following most exposures in gill cells except for the Mix 3 (Figure 1b; see

ESI Table S2). Whereas, the pattern of protein-to-nucleic acid ratio in MCF-7 cells was complicated; only treatment of C₆₀ 0.1 mg/L, Mix 4 or Mix 5 significantly reduced protein-to-nucleic acid levels, while it was significantly elevated by Mix 1 (Figure 1b; see ESI Table S2). Moreover, the ratio of RNA/DNA (Figure 1c; see ESI Table S3) exhibited by IR spectra suggested that in gill cells, treatment with C₆₀ at 0.01 mg/L or 0.1 mg/L could significantly reduce RNA/DNA levels, which potentially indicate an inhibition in gene expression, while B[a]P of 10⁻⁸ M, Mix 1, Mix 4 or Mix 5 showed capability to stimulate gene expression. This supports the observations shown in Table 2 on effects on gene expression; whilst single agent (B[a]P or C₆₀) elevated mRNA transcripts for *P21^{WAF1/CIP1}* or *CYP1A1*, a binary mixture markedly elevated the expression of these gene candidates. The changing trend of RNA/DNA levels in MCF-7 cells following exposure was different from that in gill cells, especially following C₆₀ treatment. It was found that RNA/DNA levels in MCF-7 cells was significantly increased only by treatment with B[a]P at 10⁻⁷ M and 10⁻⁶ M, as well as Mix 4.

As mid-IR spectra from multi-constituent biological samples are rich in biochemical information and complex, using simple intensity absorbance ratios (peak-to-peak ratio) is inadequate for interpretation of biological alterations⁴⁶. Thus, multivariate data-analysis techniques were employed to help with bioinformatics extraction in spectral datasets^{47, 48}. Combinations of different categories with emphasis on different scenarios to explore the post-exposure-effects from single agent or binary treatments were examined. When such spectral datasets are processed by computational analysis, alterations induced by single agents or binary mixtures can be determined. Based on PCA-LDA, dimensional (1-, 2- or 3-D) scores plots are generated for visualisation using the first three LD factors, where most segregation among categories is observed. The first two factors are particularly displayed in 1-D scores plots with their corresponding loadings plots, which identify the wavenumbers responsible for segregation. Additionally, cluster vectors plots are applied to the dataset of binary mixture

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3 exposure, so that the global alterations in cells induced by each treatment could be identified,
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5 compared with the control group.
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10 When the spectral datasets containing whole categories were inputted into PCA-LDA
11 (Dataset Total), 3-D scores plots were generated for visualisation. However, it is difficult to
12 identify segregation in 3-D or 2-D scores plots, with so many categories (see ESI Figure S2).
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14 Thus, displaying the first two LD scores in 1-D plots gives rise to a clearer interpretation of
15 segregation among categories (Figure 2; see ESI Figure S3). For both LD1 and LD2 space,
16 the corresponding loadings plots are displayed with the top seven wavenumbers marked. In
17 LD1 space, the wavenumbers derived from the spectral dataset of gill cells ranks as: 1232 cm^{-1}
18 1 (DNA/RNA; $\nu_{as}\text{PO}_2^-$), 1709 cm^{-1} (lipid), 1664 cm^{-1} (Amide I), 1070 cm^{-1} (DNA/RNA;
19 $\nu_s\text{PO}_2^-$), 985 cm^{-1} (protein phosphorylation), 1417 cm^{-1} (amino acid; $\nu[\text{COO}^-]$), and 1556 cm^{-1}
20 (Amide II). Those derived from MCF-7 cells in LD1 space were similar to gill cells: 1101
21 cm^{-1} (DNA/RNA, $\nu_s\text{PO}_2^-$), 1508 cm^{-1} (Amide II), 1026 cm^{-1} (glycogen), 1566 cm^{-1} (Amide II),
22 983 cm^{-1} (protein phosphorylation), 1406 cm^{-1} (amino acid; $\nu[\text{COO}^-]$), and 1712 cm^{-1} (lipid).
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24 The loadings plots indicate how each variance (*i.e.*, wavenumber) contributes to the
25 discrimination between the categories in the dataset. These wavenumbers correspond to
26 specific chemical entities, which might be used as biomarkers in risk assessment, in which
27 relative importance is identified in loadings plots. However, in LD2 space the loadings plots
28 for each cell line placed emphasis on different biomarkers; most pronounced wavenumbers in
29 loadings plots from gill cells were in the DNA/RNA region ($\sim 1250\text{-}1000 \text{ cm}^{-1}$), while in
30 MCF-7 cells they mostly appeared to be in the lipid / protein (Amide I) region ($\sim 1750\text{-}1500$
31 cm^{-1}). For both cell lines, significant alterations in LD1 and LD2 space were observed
32 between the exposed groups and the control in both LD1 and LD2 space, except following
33 treatment with Mix 3 (see ESI Table S4). For gill cells, its corresponding Dataset Total
34 indicated that in both LD1 and LD2 space, binary treatment with both high-dose of B[a]P and
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C₆₀ is likely to result in highly reduced effects in gill cells, while exposure to a high-dose of the one agent mixed with a low-dose of the other agent could greatly enhance toxicity in gill cells. However, when MCF-7 cells were exposed to the test agents, the response modes in the two LD spaces were presented in different ways. In LD1 space, MCF-7 cells following single-agent exposure exhibit a linear response (high response with high-dose), while in LD2 space low-dose effects were represented with B[a]P treatment. As MCF-7 cells exposed to binary agents, alterations were observed but without obvious enhancement, except that of Mix 1 in LD1. This fits with the effects on lipid peroxidation and intracellular ROS; singly both B[a]P and C₆₀ elevated these markers of oxidative stress, but in combination there is an apparent attenuation of this effect (Table 3). Through *CYP1A1* induction, B[a]P might be expected to increase intracellular ROS generation. C₆₀ is known to generate lipid peroxidation mediated *via* ROS leading to its cytotoxicity⁴⁹, but is also known to be a ROS scavenger through its ability to bind up to six electrons⁴².

The dataset of the control category compared to those from single treatment with either C₆₀ or B[a]P was explored to examine for single-agent effects (Dataset C₆₀ or Dataset B[a]P). When cells were exposed to C₆₀, both gill cells and MCF-7 cells were likely to show a linear dose-response in the LD1 space (Figure 3). Gill cells were significantly affected by C₆₀ at each dose in both LD spaces, while with MCF-7 cells only treatment of 0.1 mg/L in LD1 space and treatment of 0.01 mg/L in LD2 space appeared to be significant (see ESI Table S4). In the LD1 loadings plot derived from gill cells, the most pronounced wavenumbers were related to Amide I, glycogen, DNA/RNA, and lipid regions (Figure 3a, ESI Table S6). This is similar for MCF-7 cells in that segregation in LD1, mostly resulted from alterations in Amide I, lipid, Amide II and DNA/RNA (Figure 3c, see ESI Table S8). These spectral profiles indicate that C₆₀ is capable of not only inducing alterations in outer cellular structures (lipids and proteins), but also in internal components of DNA/RNA,

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namely genotoxicity^{50, 51}. It is suggested that the genotoxicity of C₆₀ is possibly caused by a ROS pathway⁵². Further work will be needed to delineate the time course of ROS generation and consequent cytotoxicity; it would be important to guard against missing transient effects.

However, B[a]P treatment was likely to result in a non-linear dose-response in both cell lines, and in both LD spaces significant segregation was observed in the treated categories compared with the control category, except that of gill cells exposed to B[a]P at 10⁻⁸ M (Figure 3b & 3d, ESI Table S4). B[a]P specifically induces alterations in DNA/RNA, as loadings plots in both LD1 and LD2 show obviously pronounced peaks in corresponding wavenumbers (Figure 3, see ESI Figure S4 and Table S6). This indicates that B[a]P is a genotoxic compound inducing DNA damage^{25, 53}, which is consistent with previous studies from our group³². Moreover, a low-dose effect is observed in cells following B[a]P exposure.

To gain insights into the mechanism underlying the action of binary exposure in cells, specific categories were combined as an associated dataset (Dataset Mix). In these datasets, cluster vectors plots were employed to indicate the most pronounced wavenumbers corresponding to alterations in each treatment category compared to control. When spectral data from 0.1 mg/L C₆₀, 10⁻⁶ M B[a]P and their mixture were processed by PCA-LDA, 2-D and 3-D scores plots were derived for visualisation (Figure 4). For gill cells, both 2-D scores and cluster vectors plots show that treatment with 10⁻⁶ M B[a]P led to the most pronounced alterations, mostly associated with the DNA/RNA region, while C₆₀ exerted a lower level of alteration (Figure 4a). However, the 1-D scores plots (see ESI) in the first two LD spaces indicate that co-exposure with these two treatments dramatically reduces their effects in gill cells, but it is still suggestive by the cluster vectors plot that slight genotoxicity was induced (see ESI Table S7). A similar situation occurred in MCF-7 cells (Figure 4b). Cluster vectors plots show that all treatments caused marked alterations in cells including lipids, protein and

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3 DNA/RNA; it was found that both 0.1 mg/L C₆₀ and 10⁻⁶ M B[a]P induced marked
4 alterations in cells, while treatment with Mix 3 appears relatively limited in its effect. View Article Online
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8 Additionally, the cluster vectors peak shows C₆₀ induced higher alterations in protein rather
9 than in DNA/RNA, while B[a]P mostly altered the DNA/RNA region (see ESI Table S8).
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11 However, the cluster vectors peak plot suggests that the action of co-exposure is likely to be
12 limited to the lipid and protein region of MCF-7 cells.
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18 Additional analyses were performed to explore the differences in alterations in cells
19 when the binary treatment varied (Dataset C₆₀ mix or Dataset B[a]P mix). When gill cells
20 were treated with both C₆₀ and B[a]P, the alterations were observed to be elevated with
21 decreasing C₆₀ exposure (B[a]P at 10⁻⁶ M) (Dataset C₆₀ mix, Figure 5a). As shown in the
22 cluster vectors plot, the Mix 1 (C₆₀ 0.001 mg/L and B[a]P 10⁻⁶ M) caused the most distinct
23 alterations in gill cells, and the effects seem to be a combination of both C₆₀ and B[a]P,
24 giving enhanced alterations. However, when gill cells were exposed to C₆₀ at 0.1 mg/L with
25 B[a]P at different level, the limited alterations induced by binary exposure were increased
26 with B[a]P decreasing (Figure 6a). Additionally, the action of co-exposure in MCF-7 cells
27 appeared to be similar with that in gill cells (Figure 5b and 6b). However, higher alterations
28 in the DNA/RNA region were observed in MCF-7 cells than in gill cells, as MCF-7 cells are
29 mammalian and more sensitive to genotoxicity of B[a]P. Generally, these datasets indicate
30 that high-dose C₆₀ may limit the toxicity of B[a]P. Table 3 suggests that this might be
31 through attenuation of oxidative damage, although C₆₀ may deliver more B[a]P to the cell
32 resulting in elevated *CYP1A1* induction (Table 2).
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55 In general, biological effects resulting from binary exposure are difficult to predict⁵⁴.
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57 Particularly, when NPs encounter environmental chemical compounds, this issue become
58 more intractable⁵⁵, as more factors come into account⁵⁶. Binary effects of C₆₀ co-exposure
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with other chemical compounds are controversial. It is reported that association of Hg²⁺ with C₆₀ could increase the bioavailability of Hg²⁺ in zebrafish⁵⁷. Similar investigations also determined that co-exposure with C₆₀ may enhance the effects of organic industrial chemicals⁵⁸. Another study in ZF-L cells that also focused on co-exposure of C₆₀ and B[a]P (using one high-dose of C₆₀ at 1.0 mg/L in co-exposure) suggests that C₆₀ may enhance toxicity by increasing B[a]P intake⁵⁹. Other studies draw a different conclusion. It was observed that association between C₆₀ and EE₂ reduced EE₂ bioavailability in zebrafish^{60, 61}. Additionally, a reduced histological damage induced by fluoroanthene occurred when co-exposed with C₆₀ under UV radiation⁶². Using *in vivo* models, susceptible target organ based on molecular characteristics such as lipid composition, will need to be identified.^{63, 64} Within *in vitro* models, other underlying mechanisms such as epigenetic alterations can be further investigated⁶⁵. Our results suggest that in some exposure scenarios with a particular endpoint a synergistic response is observed (Table 2), whereas in other cases the response might be additive or individual agents in a binary mixture cancel out each other's effects (Table 3). Dis-entangling such complex responses will likely require a systems biology-based approach using "omics" tools⁶³. Herein, the spectral data indicate that low-dose C₆₀ may elevate B[a]P toxicity, while high concentration of C₆₀ limit effects. Biospectroscopy also interprets the toxic action mode of such test agents even at low levels, both single and binary treatments. However, the mechanisms underlying the different actions from co-exposure with diverse combinations still requires further investigation.

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Table 1. Primers used for quantitative real-time RT-PCR analysesView Article Online
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Assay	Name	Sequence (5' to >3')
<i>P21^{WAF1/CIP1}</i>	<i>P21^{WAF1/CIP1}-F</i>	GAC CAG CAT GAC AGA TTT CTA CCA
	<i>P21^{WAF1/CIP1}-R</i>	TTC CTG TGG GCG GAT TAG G
<i>CYP1A1</i>	<i>CYP1A1-F</i>	ACT TCA TCC CTA TTC TTC GCT ACC T
	<i>CYP1A1-R</i>	CGG ATG TGG CCC TTC TCA
<i>β-Actin</i>	<i>β-Actin-F</i>	CCT GGC ACC CAG CAC AAT
	<i>β-Actin-R</i>	GCC GAT CCA CAC GGA GTA CT

F, forward primer; R, reverse primer.

Table 2. mRNA transcript levels in MCF-7 cells treated with or without B[a]P in the presence or absence of C₆₀ fullerene

Treatment	Relative expression levels	
	<i>P21^{WAF1/CIP1}</i>	<i>CYP1A1</i>
Vehicle control	1 ©	1 ©
0.01 M B[a]P	1.3 ± 0.3	1.9 ± 0.2
C ₆₀ fullerene	1.9 ± 0.3	1.5 ± 0.3
0.01 M B[a]P + 0.1 mg/L C ₆₀ fullerene	7.4 ± 1.9	21.1 ± 5.8

©, calibrator, which for the purposes of these experiments were vehicle controls. MCF-7 cells in aliquots of complete medium (5 ml, $\approx 1 \times 10^5$ cells) were seeded into 60-mm Petri dishes. Following reverse transcription of total RNA, quantitative real-time RT-PCR was carried out. Results are the means \pm SD of three separate experiments. Each experimental medium contained a final level of 0.1% (vol/vol) 1% BSA solution and DMSO.

Table 3. Oxidative damage in MCF-7 cells treated with or without B[a]P in the presence or absence of C₆₀ fullerene

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Treatment	Lipid peroxidation levels	
	TBARS	DCF fluorescence
	(nmol/mg protein)	(arbitrary units)
Vehicle control	99.1 ± 46.0	203.6 ± 33.5
0.01 M B[a]P	143.9 ± 48.1	276.0 ± 30.3
0.1 mg/L C ₆₀ fullerene	200.6 ± 90.0	306.1 ± 52.0
0.01 M B[a]P + 0.1 mg/L C ₆₀ fullerene	106.14 ± 15.7	200.6 ± 35.7

Routinely cultured MCF-7 cells were disaggregated and re-suspended in complete medium (DMEM, 10% FCS) prior to seeding aliquots (10 ml, $\approx 1 \times 10^6$ cells) into 75 cm² flasks. Following 24-h incubation, cells were treated for a further 24 h 10⁻⁸ M B[a]P, 0.1 mg/L C₆₀ fullerene or a combination of both; a vehicle control was also included. Levels of lipid peroxidation were determined using the TBARS assay. Relative fluorescence intensity was quantified by flow cytometry, using DCFH-DA as a probe. Results are the means ± SD of five separate experiments, each performed in duplicate. Each experimental medium contained a final level of 0.1% (vol/vol) 1% BSA solution and DMSO

References

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DOI: 10.1039/C7EN00164A

1. A. Kahru and H.-C. Dubourguier, From ecotoxicology to nanoecotoxicology, *Toxicology*, 2010, **269** (2–3), 105-119.
2. K. Savolainen, H. Alenius, H. Norppa, T. Tuomi and G. Kasper, Risk assessment of engineered nanomaterials and nanotechnologies--a review, *Toxicology*, 2010, **269** (2-3), 92-104.
3. N. A. Monteiro-Riviere, A. O. Inman and L. W. Zhang, Limitations and relative utility of screening assays to assess engineered nanoparticle toxicity in a human cell line, *Toxicol. Appl. Pharmacol.*, 2009, **234** (2), 222-235.
4. M. Crane, R. Handy, J. Garrod and R Owen, Ecotoxicity test methods and environmental hazard assessment for engineered nanoparticles, *Ecotoxicology*, 2008, **17** (5), 421-437.
5. M. Hassellöv, J. Readman, J. Ranville and K. Tiede, Nanoparticle analysis and characterization methodologies in environmental risk assessment of engineered nanoparticles, *Ecotoxicology*, 2008, **17** (5), 344-361.
6. K. Aschberger, C. Micheletti, B. Sokull-Klüttgen and F. M. Christensen, Analysis of currently available data for characterising the risk of engineered nanomaterials to the environment and human health - Lessons learned from four case studies, *Environ. Int.*, 2011, **37** (6), 1143-1156.
7. G. A. Hughes, Nanostructure-mediated drug delivery, *Nanomed. Nanotechnol. Biol. Med.*, 2005, **1** (1), 22-30.
8. R. Singh and J. W. Lillard Jr, Nanoparticle-based targeted drug delivery, *Exp. Mol. Pathol.*, 2009, **86** (3), 215-223.
9. A. Montellano, T. Da Ros, A. Bianco and M. Prato, Fullerene C₆₀ as a multifunctional system for drug and gene delivery, *Nanoscale*, 2011, **3** (10), 4035-4041.
10. T. Y. Zakharian, A. Seryshev, B. Sitharaman, B. E. Gilbert, V. Knight and L. J. Wilson, A Fullerene-Paclitaxel Chemotherapeutic: Synthesis, Characterization, and Study of Biological Activity in Tissue Culture, *J. Am. Chem. Soc.*, 2005, **127** (36), 12508-12509.

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11. K. Aschberger, H. J. Johnston, V. Stone, R. J. Aitken, C. L. Tran, S. M. Hankin, S. A. K. Peters and F. M. Christensen, Review of fullerene toxicity and exposure - Appraisal of a human health risk assessment, based on open literature, *Regul. Toxicol. Pharmacol.*, 2010, **58** (3), 455-473.
12. P. Spohn, C. Hirsch, F. Hasler, A. Bruinink, H. F. Krug and P. Wick, C₆₀ fullerene: A powerful antioxidant or a damaging agent? The importance of an in-depth material characterization prior to toxicity assays, *Environ. Pollut.*, 2009, **157** (4), 1134-1139.
13. X. R. Xia, N. A. Monteiro-Riviere and J. E. Riviere, Intrinsic biological property of colloidal fullerene nanoparticles (nC₆₀): Lack of lethality after high dose exposure to human epidermal and bacterial cells, *Toxicol. Lett.*, 2010, **197** (2), 128-134.
14. P. Christian, F. Von der Kammer, M. Baalousha and T Hofmann, Nanoparticles: structure, properties, preparation and behaviour in environmental media, *Ecotoxicology*, 2008, **17** (5), 326-343.
15. L. K. Limbach, P. Wick, P. Manser, R. N. Grass, A. Bruinink and W. J. Stark, Exposure of Engineered Nanoparticles to Human Lung Epithelial Cells: Influence of Chemical Composition and Catalytic Activity on Oxidative Stress, *Environ. Sci. Technol.*, 2007, **41** (11), 4158-4163.
16. W. Fan, M. Cui, H. Liu, C. Wang, Z. Shi, C. Tan and X. Yang, Nano-TiO₂ enhances the toxicity of copper in natural water to *Daphnia magna*, *Environ. Pollut.*, 2011, **159** (3), 729-734.
17. B. Pan and B. Xing, Adsorption Mechanisms of Organic Chemicals on Carbon Nanotubes, *Environ. Sci. Technol.*, 2008, **42** (24), 9005-9013.
18. B. Pan, D. Lin, H. Mashayekhi and B. Xing, Adsorption and Hysteresis of Bisphenol A and 17 α -Ethinyl Estradiol on Carbon Nanomaterials, *Environ. Sci. Technol.*, 2008, **42** (15), 5480-5485.
19. X. Li, B. Gámiz, Y. Wang, J. J. Pignatello and B. Xing, Competitive Sorption Used To Probe Strong Hydrogen Bonding Sites for Weak Organic Acids on Carbon Nanotubes, *Environ. Sci. Technol.*, 2015, **49** (3), 1409-1417.

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20. Y. Su, X. Yan, Y. Pu, F. Xiao, D. Wang and M. Yang, Risks of Single-Walled Carbon Nanotubes Acting as Contaminants-Carriers: Potential Release of Phenanthrene in Japanese Medaka (*Oryzias latipes*), *Environ. Sci. Technol.*, 2013, **47** (9), 4704-4710. View Article Online
DOI: 10.1039/C3EN00164A
21. M. Song, F. Wang, L. Zeng, J. Yin, H. Wang and G. Jiang, Co-exposure of Carboxyl-Functionalized Single-Walled Carbon Nanotubes and 17 α -Ethinylestradiol in Cultured Cells: Effects on Bioactivity and Cytotoxicity, *Environ. Sci. Technol.*, 2014, **48** (23), 13978-13984.
22. K. Srogi, Monitoring of environmental exposure to polycyclic aromatic hydrocarbons: a review, *Environ. Chem. Lett.*, 2007, **5** (4), 169-195.
23. K. W. Bock, Aryl hydrocarbon or dioxin receptor: Biologic and toxic responses. In *Reviews of Physiology, Biochemistry and Pharmacology*, Springer Berlin Heidelberg: 1994, **Vol. 125**, pp 1-42.
24. W. M. Baird, L. A. Hooven and B. Mahadevan, Carcinogenic polycyclic aromatic hydrocarbon-DNA adducts and mechanism of action, *Environ. Mol. Mutagen.*, 2005, **45** (2-3), 106-114.
25. W. Xue and D. Warshawsky, Metabolic activation of polycyclic and heterocyclic aromatic hydrocarbons and DNA damage: A review, *Toxicol. Appl. Pharmacol.*, 2005, **206** (1), 73-93.
26. D. W. Nebert and T. P. Dalton, The role of cytochrome P450 enzymes in endogenous signalling pathways and environmental carcinogenesis, *Nat. Rev. Cancer*, 2006, **6** (12), 947-960.
27. M. J. Baker, J. Trevisan, P. Bassan, R. Bhargava, H. J. Butler, K. M. Dorling, P. R. Fielden, S. W. Fogarty, N. J. Fullwood, K. A. Heys, C Hughes, P. Lasch, P. L. Martin-Hirsch, B. Obinaju, G. D. Sockalingum, J. Sulé-Suso, R. J. Strong, M. J. Walsh, B. R. Wood, P. Gardner, and F. L. Martin, Using Fourier transform IR spectroscopy to analyze biological materials, *Nat. Protoc.*, 2014, **9** (8), 1771-1791.
28. D. I. Ellis and R. Goodacre, Metabolic fingerprinting in disease diagnosis: biomedical applications of infrared and Raman spectroscopy, *Analyst*, 2006, **131** (8), 875-885.
29. M. J. Walsh, T. G. Fellous, A. Hammiche, W.-R. Lin, N. J. Fullwood, O. Grude, F. Bahrami, J. M. Nicholson, M. Cotte, J. Susini, H. M. Pollock, M. Brittan, P. L. Martin-Hirsch,

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- M. R. Alison and F. L. Martin, Fourier Transform Infrared Microspectroscopy Identifies Symmetric PO₂⁻ Modifications as a Marker of the Putative Stem Cell Region of Human Intestinal Crypts, *Stem Cells*, 2008, **26** (1), 108-118. View Article Online
DOI: 10.1039/C7EN00164A
30. L. Corte, P. Rellini, L. Roscini, F. Fatichenti and G. Cardinali, Development of a novel, FTIR (Fourier transform infrared spectroscopy) based, yeast bioassay for toxicity testing and stress response study, *Anal. Chim. Acta*, 2010, **659** (1-2), 258-265.
31. J. G. Kelly, J. Trevisan, A. D. Scott, P. L. Carmichael, H. M. Pollock, P. L. Martin-Hirsch and F. L. Martin, Biospectroscopy to metabolically profile biomolecular structure: a multistage approach linking computational analysis with biomarkers, *J. Proteome Res.*, 2011, **10** (4), 1437-1448.
32. W. Pang, J. Li, A. A. Ahmadzai, L. D. Heppenstall, V. Llabjani, J. Trevisan, X. Qiu and F. L. Martin, Identification of benzo[*a*]pyrene-induced cell cycle-associated alterations in MCF-7 cells using infrared spectroscopy with computational analysis, *Toxicology*, 2012, **298** (1-3), 24-29.
33. J. Li, R. Strong, J. Trevisan, S. W. Fogarty, N. J. Fullwood, K. C. Jones and F. L. Martin, Dose-Related Alterations of Carbon Nanoparticles in Mammalian Cells Detected Using Biospectroscopy: Potential for Real-World Effects, *Environ. Sci. Technol.*, 2013, **47** (17), 10005-10011.
34. P. A. Walker, N. R. Bury and C. Hogstrand, Influence of Culture Conditions on Metal-Induced Responses in a Cultured Rainbow Trout Gill Epithelium, *Environ. Sci. Technol.*, 2007, **41** (18), 6505-6513.
35. K. Schirmer, A. G. J. Chan, B. M. Greenberg, D. G. Dixon and N. C. Bols, Methodology for demonstrating and measuring the photocytotoxicity of fluoranthene to fish cells in culture, *Toxicol. In Vitro*, 1997, **11** (1-2), 107-119.
36. K. Tanneberger, A. Rico-Rico, N. I. Kramer, F. J. M. Busser, J. L. M. Hermens and K. Schirmer, Effects of Solvents and Dosing Procedure on Chemical Toxicity in Cell-Based in Vitro Assays, *Environ. Sci. Technol.*, 2010, **44** (12), 4775-4781.
37. R. Gautam, S. Vanga, F. Ariese and S. Umopathy, Review of multidimensional data processing approaches for Raman and infrared spectroscopy, *EPJ Techn. Instrum.*, 2015, **2** (1), 1-38.

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38. V. Llabjani, J. Trevisan, K. C. Jones, R. F. Shore and F. L. Martin, Derivation by Infrared Spectroscopy with Multivariate Analysis of Bimodal Contaminant-Induced Dose-Response Effects in MCF-7 Cells, *Environ. Sci. Technol.*, 2011, **45** (14), 6129-6135.
39. V. Llabjani, J. D. Crosse, A. A. Ahmadzai, I. I. Patel, W. Pang, J. Trevisan, K. C. Jones, R. F. Shore and F. L. Martin, Differential Effects in Mammalian Cells Induced by Chemical Mixtures in Environmental Biota As Profiled Using Infrared Spectroscopy, *Environ. Sci. Technol.*, 2011, **45** (24), 10706-10712.
40. H. Jiao, S. L. Allinson, M. J. Walsh, R. Hewitt, K. J. Cole, D. H. Phillips and F. L. Martin, Growth kinetics in MCF-7 cells modulate benzo[*a*]pyrene-induced *CYP1A1* up-regulation, *Mutagenesis*, 2007, **22** (2), 111-116.
41. P. Nusuetrong, T. Pengsuparp, D. Meksuriyen, M. Tanitsu, H. Kikuchi, M. Mizugaki, K. Shimazu, Y. Oshima, N. Nakahata and M. Yoshida, Satratoxin H generates reactive oxygen species and lipid peroxides in PC12 cells, *Biol. Pharm. Bull.*, 2008, **31** (6), 1115-1120.
42. Y. I. Prylutsky, I. V. Vereshchaka, A. V. Maznychenko, N. V. Bulgakova, O. O. Gonchar, O. A. Kyzyma, U. Ritter, P. Scharff, T. Tomiak, D. M. Nozdrenko, I. V. Mishchenko and A. I. Kostyukov, C₆₀ fullerene as promising therapeutic agent for correcting and preventing skeletal muscle fatigue, *J. Nanobiotechnol.*, 2017, **15**, 8.
43. P. G. Andrus, Cancer monitoring by FTIR spectroscopy, *Technol. Cancer Res. Treat.*, 2006, **5**, (2), 157-167.
44. R. K. Sahu, S. Argov, A. Salman, M. Huleihel, N. Grossman, Z. Hammody, J. Kapelushnik and S. Mordechai, Characteristic Absorbance of Nucleic Acids in the Mid-IR Region as Possible Common Biomarkers for Diagnosis of Malignancy, *Technol. Cancer Res. Treat.*, 2004, **3** (6), 629-638.
45. J. R. Mourant, Y. R. Yamada, S. Carpenter, L. R. Dominique and J. P. Freyer, FTIR Spectroscopy Demonstrates Biochemical Differences in Mammalian Cell Cultures at Different Growth Stages, *Biophys. J.*, 2003, **85** (3), 1938-1947.
46. L. Wang and B. Mizaikoff, Application of multivariate data-analysis techniques to biomedical diagnostics based on mid-infrared spectroscopy, *Anal. Bioanal. Chem.*, 2008, **391** (5), 1641-1654.

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53
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56
57
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60
47. J. Trevisan, P. P. Angelov, I. I. Patel, G. M. Najand, K. T. Cheung, V. Llabjani, H. M. Pollock, S. W. Bruce, K. Pant, P. L. Carmichael, A. D. Scott and F. L. Martin, Syrian hamster embryo (SHE) assay (pH 6.7) coupled with infrared spectroscopy and chemometrics towards toxicological assessment, *Analyst*, 2010, **135** (12), 3266-3272.
48. J. Trevisan, P. P. Angelov, P. L. Carmichael, A. D. Scott and F.L. Martin, Extracting biological information with computational analysis of Fourier-transform infrared (FTIR) biospectroscopy datasets: current practices to future perspectives, *Analyst*, 2012, **137** (14), 3202-3215.
49. C. M. Sayes, A. M. Gobin, K. D. Ausman, J. Mendez, J. L. West and V. L. Colvin, Nano-C₆₀ cytotoxicity is due to lipid peroxidation, *Biomaterials*, **26**, 7587-7595.
50. A. Dhawan, J. S. Taurozzi, A. K. Pandey, W. Shan, S. M. Miller, S. Hashsham and V. V. Tarabara, Stable Colloidal Dispersions of C60 Fullerenes in Water: Evidence for Genotoxicity. *Environ. Sci. Technol.*, 2006, **40** (23), 7394-7401.
51. J. K. Folkmann, L. Risom, N. R. Jacobsen, H. Wallin, S. Loft and P. Møller, Oxidatively damaged DNA in rats exposed by oral gavage to C60 fullerenes and single-walled carbon nanotubes, *Environ. Health Perspect.*, 2009, **117** (5), 703-708.
52. F. Wang, C. Jin, H. Liang, Y. Tang, H. Zhang and Y. Yang, Effects of fullerene C₆₀ nanoparticles on A549 cells, *Environ. Toxicol. Pharmacol.*, 2014, **37** (2), 656-661.
53. A. Castaño and C. Becerril, In vitro assessment of DNA damage after short- and long-term exposure to benzo(a)pyrene using RAPD and the RTG-2 fish cell line, *Mutat. Res.*, 2004, **552** (1-2), 141-151.
54. V. Llabjani, J. Trevisan, K. C. Jones, R. F. Shore and F. L. Martin, Binary Mixture Effects by PBDE Congeners (47, 153, 183, or 209) and PCB Congeners (126 or 153) in MCF-7 Cells: Biochemical Alterations Assessed by IR Spectroscopy and Multivariate Analysis, *Environ. Sci. Technol.*, 2010, **44** (10), 3992-3998.
55. L. Canesi, C. Ciacci and T. Balbi, Interactive effects of nanoparticles with other contaminants in aquatic organisms: Friend or foe? *Mar. Environ. Res.*, 2015, **111**, 128-134.

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57
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59
60
56. A. Baun, S. N. Sørensen, R. F. Rasmussen, N. B. Hartmann and C. B. Koch, Toxicity and bioaccumulation of xenobiotic organic compounds in the presence of aqueous suspensions of aggregates of nano-C₆₀, *Aquat. Toxicol.*, 2008, **86** (3), 379-387. View Article Online
DOI: 10.1039/C7EN00164A
57. T. B. Henry, S. J. Wileman, H. Boran and P. Sutton, Association of Hg²⁺ with Aqueous (C₆₀)_n Aggregates Facilitates Increased Bioavailability of Hg²⁺ in Zebrafish (*Danio rerio*), *Environ. Sci. Technol.*, 2013, **47** (17), 9997-10004.
58. M. Lehto, T. Karilainen, T. Róg, O. Cramariuc, E. Vanhala, J. Tornaesus, H. Taberman, J. Jänis, H. Alenius, I. Vattulainen and O. Laine, Co-Exposure with Fullerene May Strengthen Health Effects of Organic Industrial Chemicals, *PLoS ONE*, 2014, **9** (12), e114490.
59. J. L. R. Ferreira, M. N. Lonné, T. A. França, N. R. Maximilla, T. H. Lugokenski, P. G. Costa, G. Fillmann, F. A. Antunes Soares, F. R. de la Torre and J. M. Monserrat, J. M., Co-exposure of the organic nanomaterial fullerene C₆₀ with benzo[a]pyrene in *Danio rerio* (zebrafish) hepatocytes: Evidence of toxicological interactions, *Aquat. Toxicol.*, 2014, **147**, 76-83.
60. J.-W. Park, T. B. Henry, F.-M. Menn, R. N. Compton and G. Sayler, No bioavailability of 17 α -ethinylestradiol when associated with nC₆₀ aggregates during dietary exposure in adult male zebrafish (*Danio rerio*), *Chemosphere*, 2010, **81** (10), 1227-1232.
61. J.-W. Park, T. B. Henry, S. Ard, F.-M. Menn, R. N. Compton and G. S. Sayler, The association between nC₆₀ and 17 α -ethinylestradiol (EE2) decreases EE2 bioavailability in zebrafish and alters nanoaggregate characteristics, *Nanotoxicology*, 2011, **5** (3), 406-416.
62. X. Y. Yang, R. E. Edelman and J. T. Oris, Suspended C₆₀ nanoparticles protect against short-term UV and fluoranthene photo-induced toxicity, but cause long-term cellular damage in *Daphnia magna*, *Aquat. Toxicol.*, 2010, **100** (2), 202-210.
63. E. Gorrochategui, J. Li, N. J. Fullwood, G. G. Ying, M. Tian, L. Cui, H. Shen, S. Lacorte, R. Tauler and F. L. Martin, Diet-sourced carbon-based nanoparticles induce lipid alterations in tissues of zebrafish (*Danio rerio*) with genomic hypermethylation changes in brain, *Mutagenesis*, 2017, **32** (1), 91-103.

1
2
3
4 64. J. Li, G. G. Ying, K. C. Jones and F. L. Martin, Real-world carbon nanoparticle View Article Online
5 exposures induce brain and gonadal alterations in zebrafish (*Danio rerio*) as determined by DOI: 10.1039/C7EN00164A
6 biospectroscopy techniques, *Analyst*, **140** (8), 2687-2695.
7
8

9
10 65. J. Li, M. Tian, L. Cui, J. Dwyer, N. J. Fullwood, H. Shen and F. L. Martin, Low-dose
11 carbon-based nanoparticle-induced effects in A549 lung cells determined by biospectroscopy
12 are associated with increases in genomic methylation. *Sci. Rep.*, 2016, **6**, 20207.
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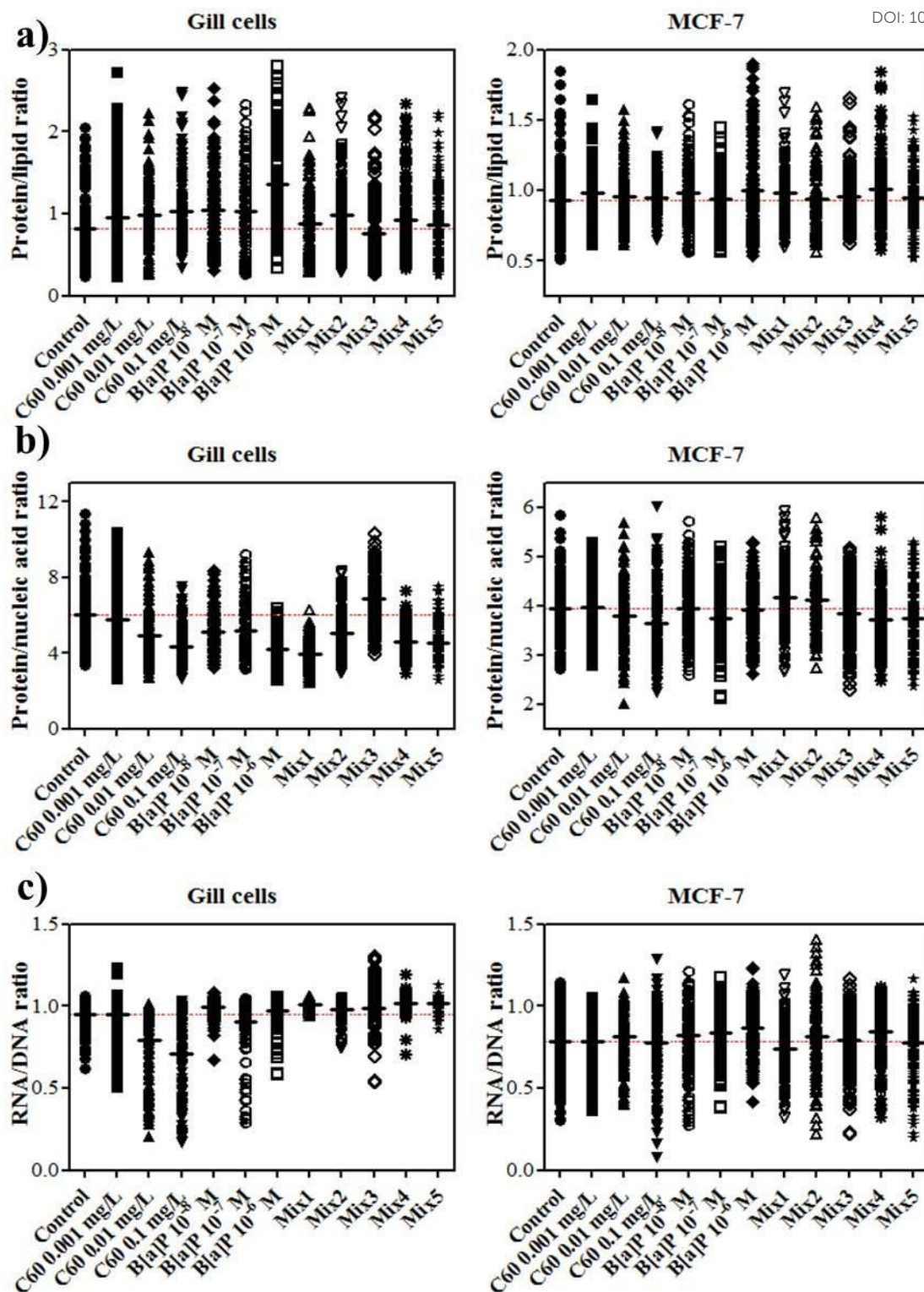


Figure 1 Intensity absorbance ratio derived from IR spectra of the cells: *a*) Ratio of protein-to-lipid; *b*) Ratio of protein-to-nucleic acid; and, *c*) Ratio of RNA-to-DNA.

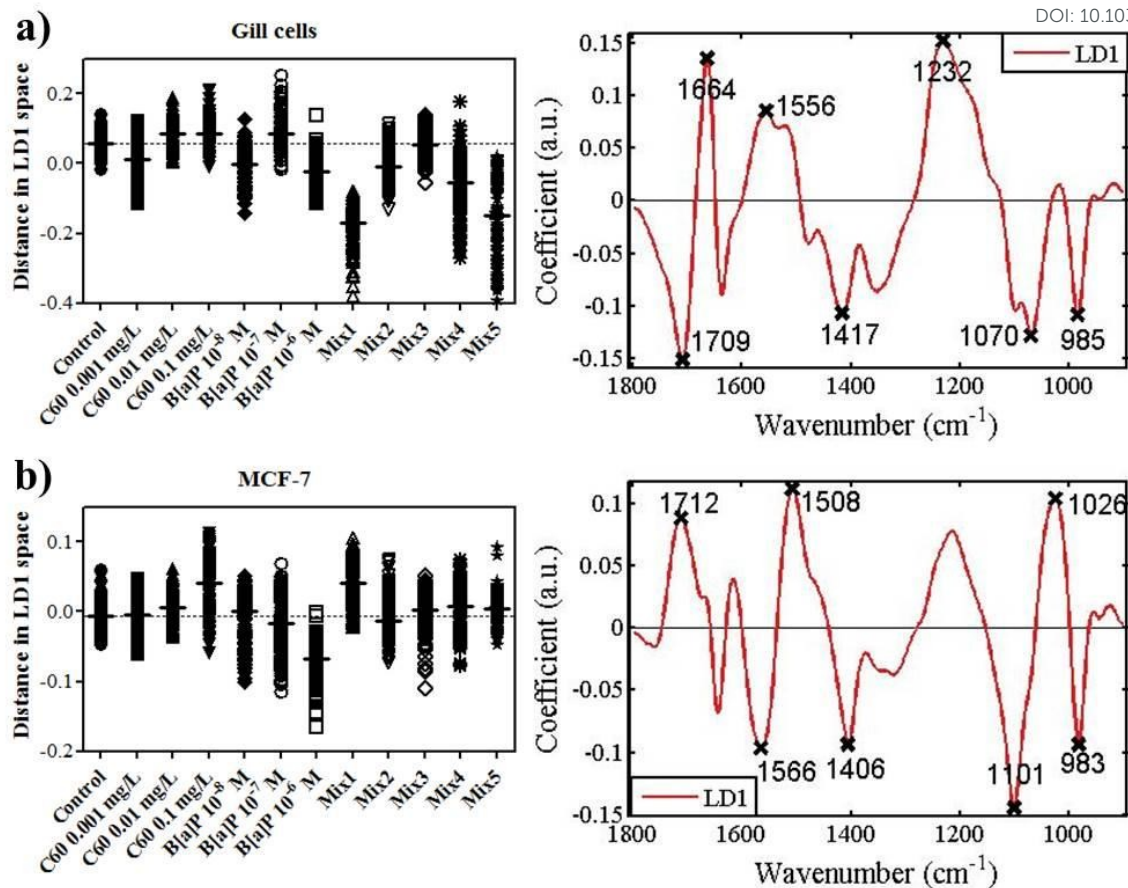


Figure 2 One-D scores plot in LD1 derived from PCA-LDA of spectral dataset (Dataset Total), with corresponding loadings plot: *a*) Gill cells; and, *b*) MCF-7 cells.

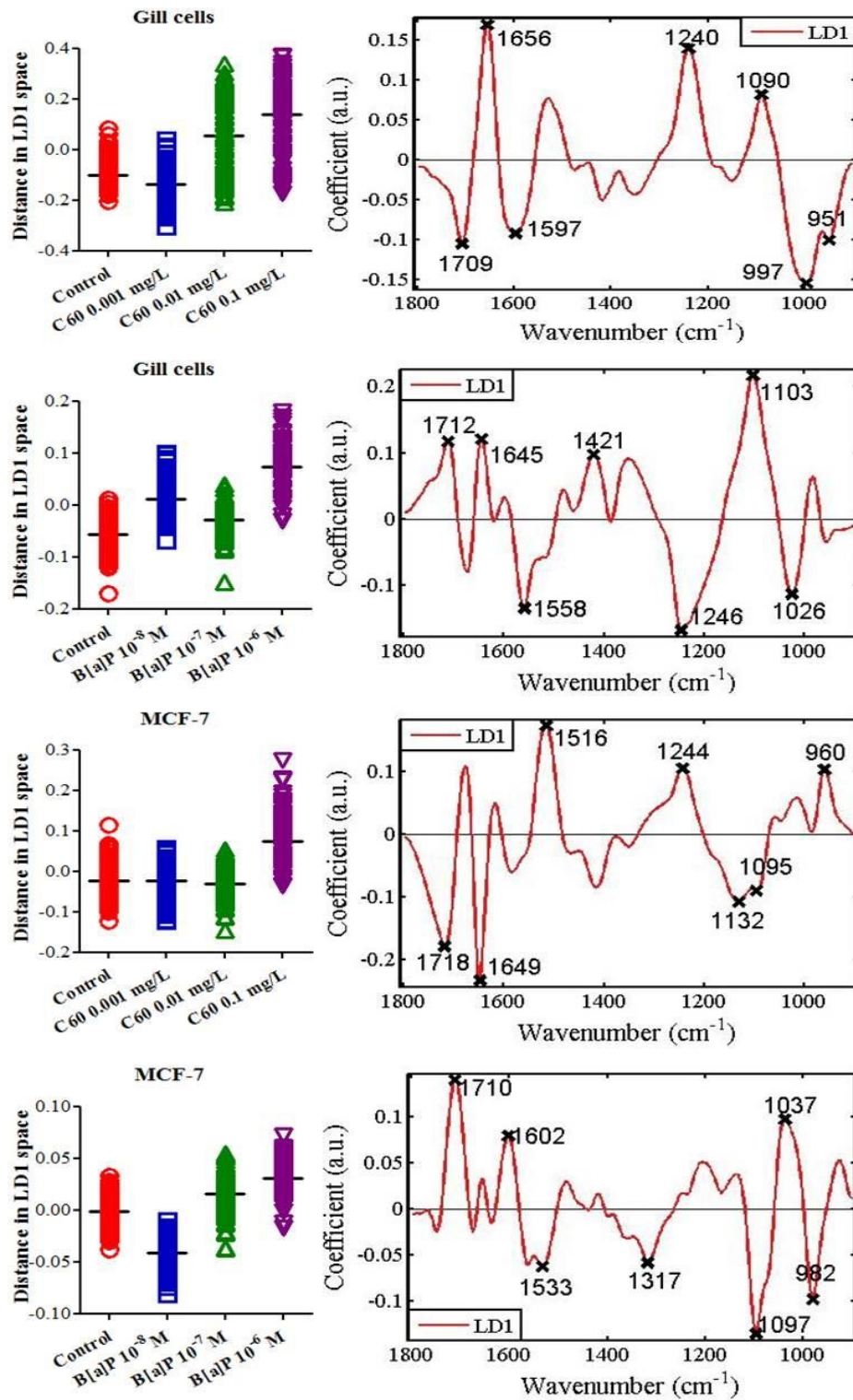


Figure 3 One-D scores plot in LD1 derived from PCA-LDA of spectral dataset for single treatment, with corresponding loadings plot.

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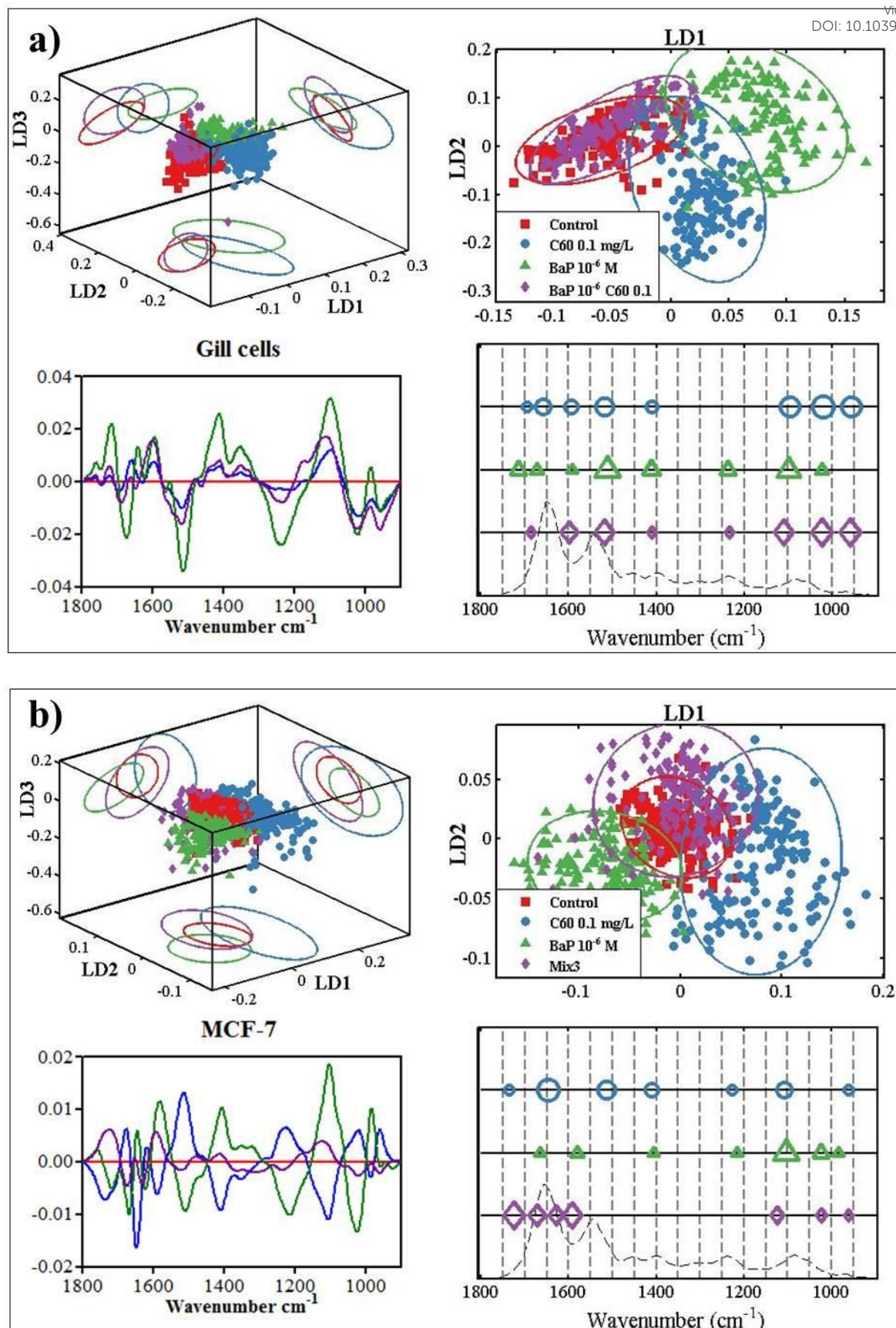


Figure 4 Scores plot and cluster vector derived from PCA-LDA of spectral dataset (Dataset mix). *a)* Gill cells; and, *b)* MCF-7 cells.

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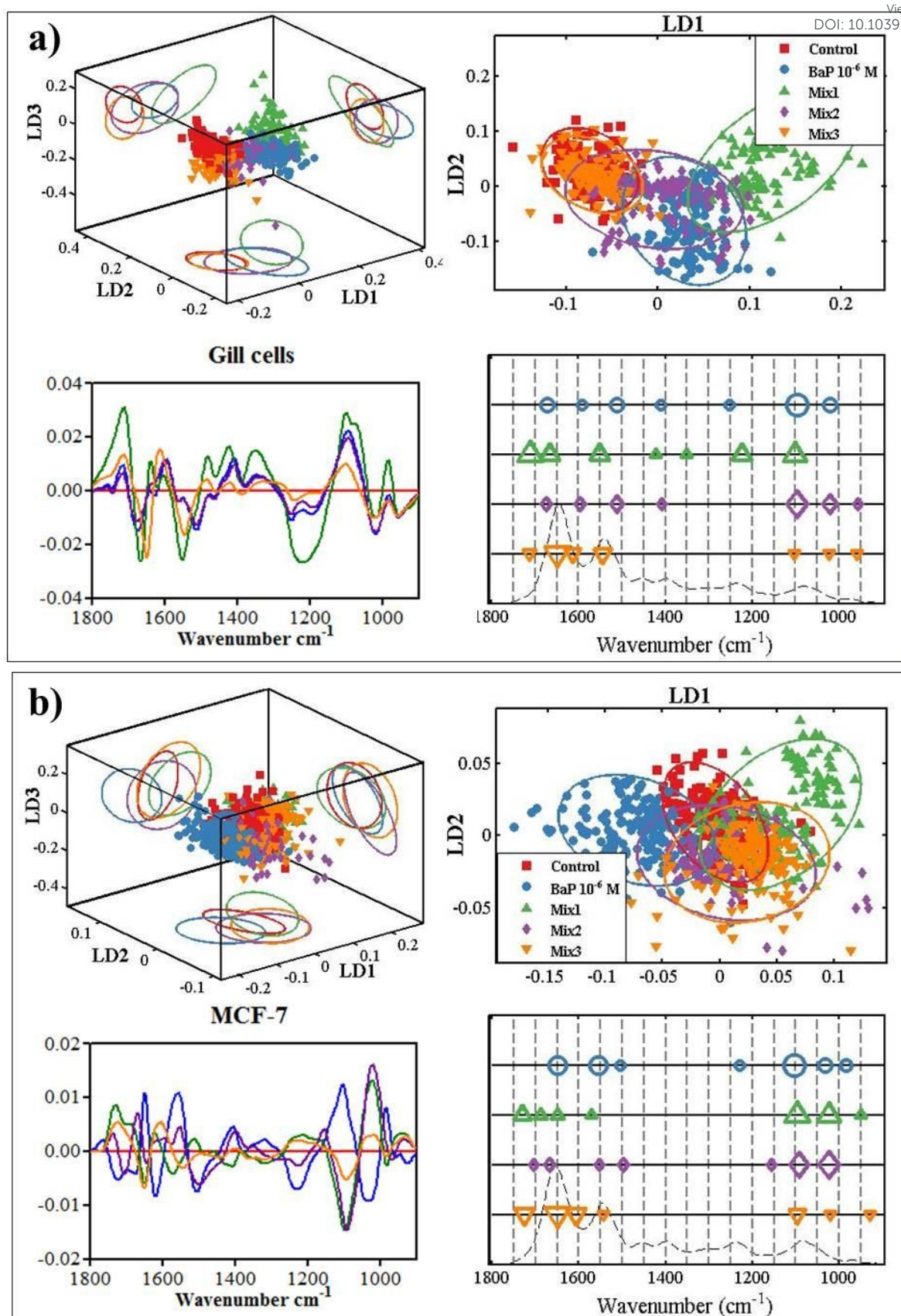


Figure 5 Scores plot and cluster vectors derived from PCA-LDA of spectral dataset (Dataset B[a]P mix). *a*) Gill cells; and, *b*) MCF-7 cells.

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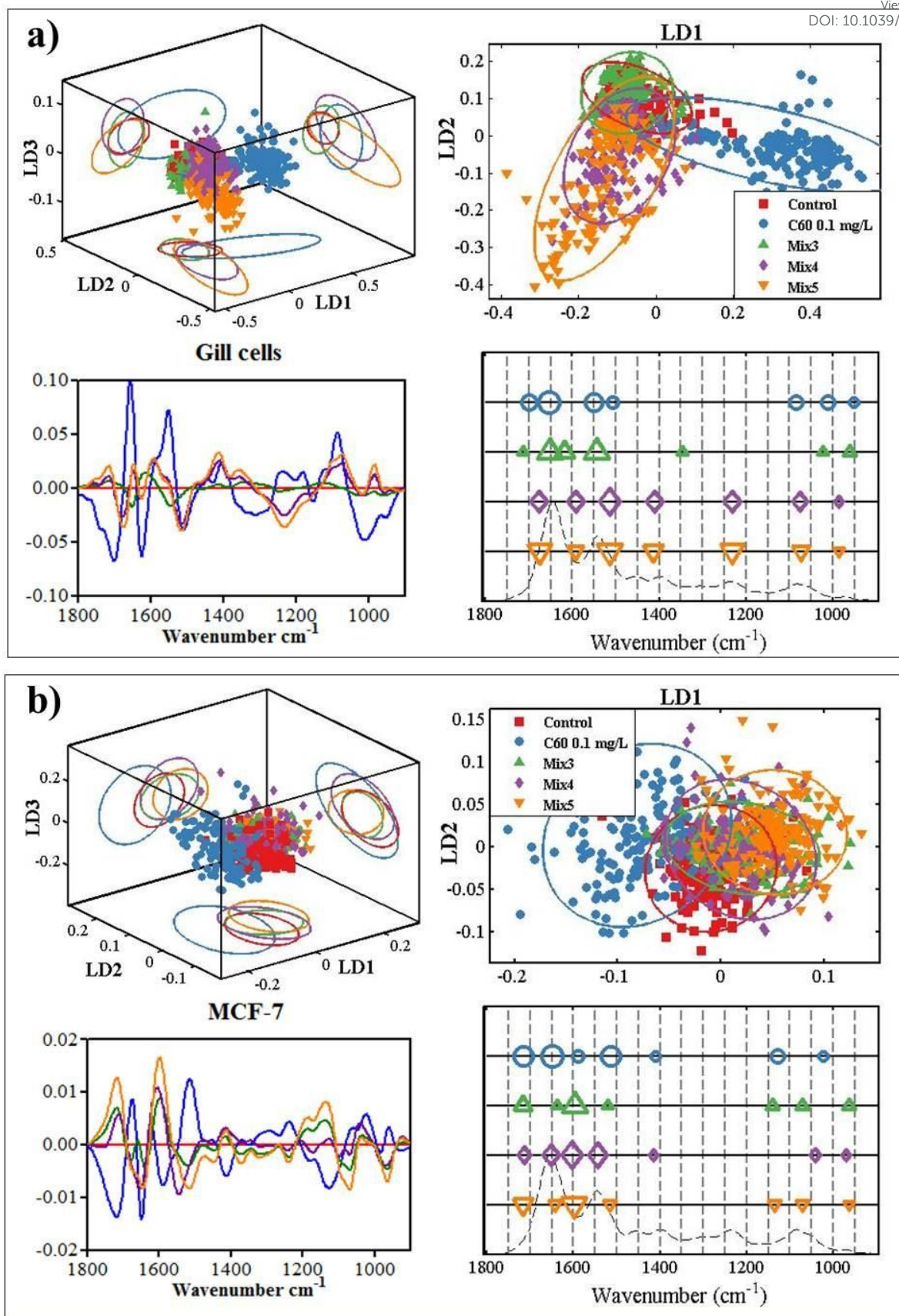


Figure 6 Scores plot and cluster vectors derived from PCA-LDA of spectral dataset (Dataset C₆₀ mix). *a*) Gill cells; and, *b*) MCF-7 cells.