



*The reactive species interactome:
evolutionary emergence, biological
significance, and opportunities for redox
metabolomics and personalized medicine*

Article

Accepted Version

Cortese-Krott, M., Koning, A., Kuhnle, G., Nagy, P., Bianco, C., Pasch, A., Wink, D. A., Fukuto, J., Jackson, A. A., van Goor, H., Olson, K. R. and Feelisch, M. (2017) The reactive species interactome: evolutionary emergence, biological significance, and opportunities for redox metabolomics and personalized medicine. *Antioxidants & Redox Signaling*, 27 (10). pp. 684-712. ISSN 1523-0864 doi: <https://doi.org/10.1089/ars.2017.7083> Available at <http://centaur.reading.ac.uk/70039/>

It is advisable to refer to the publisher's version if you intend to cite from the work.

To link to this article DOI: <http://dx.doi.org/10.1089/ars.2017.7083>

Publisher: Mary Ann Leibert Inc

All outputs in CentAUR are protected by Intellectual Property Rights law, including copyright law. Copyright and IPR is retained by the creators or other

copyright holders. Terms and conditions for use of this material are defined in the [End User Agreement](#).

www.reading.ac.uk/centaur

CentAUR

Central Archive at the University of Reading

Reading's research outputs online

Forum Review Article

The Reactive Species Interactome:**Evolutionary Emergence, Biological Significance, and Opportunities for Redox Metabolomics and Personalized Medicine**

Miriam M. Cortese-Krott^{1*}, Anne Koning², Gunter GC Kuhnle³, Peter Nagy⁴, Christopher L. Bianco⁵, Andreas Pasch⁶, David Wink⁷, Jon M. Fukuto⁸, Alan A. Jackson⁹, Harry van Goor², Kenneth R. Olson¹⁰, Martin Feelisch^{9,11*}

¹Cardiovascular Research Laboratory, Department of Cardiology, Pneumology and Angiology, Medical Faculty, Heinrich Heine University of Düsseldorf, Universitätsstrasse 1, 40225 Düsseldorf, Germany. ²Department of Pathology and Medical Biology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands; ³Department of Food & Nutritional Sciences, University of Reading, Reading, RG6 6UR, UK ⁴Molecular Immunology and Toxicology, National Institute Oncology, Ráth György utca 7-9, Budapest, Hungary, 1122. ⁵Department of Chemistry, Johns Hopkins University, Baltimore, MD 21218, United States ⁶Department of Clinical Chemistry and Calciscon AG and University of Bern, Switzerland ⁷Cancer and Inflammation Program, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Frederick, MD, USA ⁸Department of Chemistry, Sonoma State University, Rohnert Park, CA 94928, USA ⁹NIHR Southampton Biomedical Research Center, University of Southampton, Southampton General Hospital, Southampton SO16 6YD, UK ¹⁰Indiana University School of Medicine-South Bend, South Bend, IN, USA ¹¹Clinical & Experimental Sciences, Faculty of Medicine, Southampton General Hospital and Institute for Life Sciences, University of Southampton, Southampton, SO16 6YD, UK.

Running head (max. 50 c): The reactive species interactome

Word count (excluding references and figure legends, max. 8,000 words): 9882

Number of illustrations: grayscale 0, color 6 (on line only)

Reference number (max. 200): 222

***Corresponding authors:**

Prof. Martin Feelisch, Ph.D.

Clinical and Experimental Sciences, Faculty of Medicine, University of Southampton, Southampton General Hospital, South Academic Block, Level F, Mailpoint 810, Tremona Road, Southampton, SO16 6YD, UK

Phone: +44 (0)2381 206891; E-Mail: m.feelisch@soton.ac.uk

Prof. Miriam M. Cortese-Krott, Ph.D.

Cardiovascular Research Laboratory, Department of Cardiology, Pneumology and Angiology, Medical Faculty, Heinrich Heine University of Düsseldorf, Universitätsstrasse 1, 40225 Düsseldorf, Germany

Phone: +49 (0)211 8115115; E-Mail: miriam.cortese@uni-duesseldorf.de

Abstract

Significance. Oxidative stress is thought to account for aberrant redox homeostasis and contribute to aging and disease. However, more often than not administration of antioxidants is ineffective, suggesting our current understanding of the underlying regulatory processes is incomplete. **Recent Advances.** Similar to reactive oxygen and nitrogen species (ROS, RNS), reactive sulfur species (RSS) are now emerging as important signaling molecules, targeting regulatory cysteine redox switches in proteins, affecting gene regulation, ion transport, intermediary metabolism and mitochondrial function. To rationalize the complexity of chemical interactions of reactive species with themselves and their targets and help define their role in systemic metabolic control, we here introduce a novel integrative concept coined the *reactive species interactome* (RSI). The RSI is a primeval multi-level redox-regulatory system whose architecture, together with the physicochemical characteristics of its constituents, allows efficient sensing and rapid adaptation to environmental changes and various other stresses to enhance fitness and resilience at the local and whole-organism level. **Critical Issues.** To better characterise the RSI-related processes that determine fluxes through specific pathways and enable integration, it is necessary to disentangle the chemical biology and activity of reactive species (including precursors and reaction products), their targets, communication systems and effects on cellular, organ and whole-organism bioenergetics using systems-level/network analyses. **Future Directions.** Understanding the mechanisms through which the RSI operates will enable a better appreciation of the possibilities to modulate the entire biological system; moreover, unveiling molecular signatures that characterize specific environmental challenges or other stresses will provide new prevention/intervention opportunities for personalized medicine.

Keywords: Hydrogen sulfide, polysulfides, nitric oxide, systems biology, microbiome, network medicine

1. Introduction

Nothing in biology makes sense except in the light of evolution.
Theodosius Dobzhansky

Life is nothing but an electron looking for a place to rest.
Albert Szent-Györgyi

We are witnessing an unprecedented paradigmatic change in the practice of medicine whereby the concept of intervention is evolving from treating diseases in a one-organ/one-symptom fashion to a systems-based approach that considers a patient's pathophysiological condition including his/her individual genetic blueprint, microbiome, disease history and life-style. Indeed, diseases presenting with similar clinical phenotypes are often heterogeneous conditions of multifactorial origin, involving a multitude of molecular, cellular and organ systems. Their multilevel nature and complexity pose a formidable challenge to identifying the molecular causes; finding the most suitable therapy for each specific case demands a thorough understanding of the fundamental principles of biological regulation and a refined inter-disciplinary systems approach encompassing medicine, pharmacology, biology, chemistry and physics. Recent analyses indicate the existence of disease-specific functional modules that are central hubs in the vast network of human diseases y, offering additional opportunities by embracing mathematical approaches.

A limited number of risk factors (such as poor-quality nutrition or physical inactivity) and chronic conditions (including hypertension, cardiovascular disease, obesity, asthma, diabetes, neurodegenerative diseases and certain forms of cancer) account for the majority of the global burden of disease (114), overall life expectancy and all-cause mortality (203). A common feature of many of these conditions is oxidative stress (176), and some have been re-defined as "redox diseases" (26,206). The term *oxidative stress* was originally described as "an imbalance between oxidants and antioxidants in favor of the oxidants, leading to a disruption of redox signaling and control and/or molecular damage" (91,174); it was initially considered to be triggered by an inflammatory process or mitochondrial dysfunction.

However, the use of selective antioxidants for *redox diseases* has not had the effect anticipated, suggesting that our current understanding of the underlying pathophysiological processes is incomplete (26,176).

Recently, Jones and Sies proposed that besides the *Genetic Code*, allowing reproduction and defining heredity, there exists a *Redox Code*¹ that identifies the regulatory elements and defines the principles through which biological function is enabled and protected (94). Within this concept, the endogenous production of reactive oxygen species (ROS) is a highly regulated enzymatic process, which serves the purpose of *signaling* and can lead to the modification of *cysteine redox switches*. Modification of these *switches* leads to modulation of their functional state, which would result in alterations of protein structure, enzymatic activity or gene transcription. Modifying responses to match a changed environment creates the opportunity for adaptive changes that enhance an organism's fitness for purpose.

However, there is more to this redox network than ROS. Nitric oxide (NO) is a free radical, which is produced endogenously by NO synthases (NOS) and acts as an effector and messenger, regulating a variety of physiological processes. Chemical interactions of NO with ROS form reactive nitrogen species (RNS) constitute the basis for the formation of a multitude of additional oxidative signaling elements (65), including the highly reactive and potentially damaging peroxynitrite (ONOO⁻). Both ROS and RNS may target cysteine thiols leading to oxidative modifications (including formation of sulfinic acid, sulfenic acids and thiyl radicals, and sulfane-sulfur containing molecules, such as persulfides and polysulfides) (131,132,140). By analogy to ROS and RNS, these compounds are identified as reactive sulfur species (RSS) (67). Similar to ROS and RNS, RSS were first considered to be produced only under pathological conditions and not recognized as being involved in

¹ In this context, the word „code“ is used to describe a “set of principles“, rather than a carrier of information like in the genetic code.

signaling functions. More recently, hydrogen sulfide (H₂S) and its sulfane-sulfur derivatives have been shown to participate in fundamental biochemical pathways that control cellular redox homeostasis, signaling, metabolism, and mitochondrial function (145,204), perhaps most intriguingly illustrated by the ‘suspended animation’ observed after H₂S inhalation in small rodents (11). This has led to a renewed interest in sulfide chemistry and biology (37,38,104,126,187), and RSS, together with ROS and RNS, to be considered as important physiological signaling molecules.

The view that placed oxygen at the centre of the redox-regulatory system has been questioned recently by the realization that much of the evolutionary biology of Life evolved in a sulfur-rich atmosphere virtually free of oxygen (135,139). Indeed, it is considered likely that the interaction of RSS with RNS to form S/N-hybrid species participated in forming the building blocks of Life and preceded the advent of aerobic respiration and ROS formation (41). In this model, as the level of atmospheric oxygen rose, enabling the development of larger and more energy-efficient organisms, the ancient mechanisms of sulfur metabolism had to face the new challenge of dealing with rapid oxidation processes superimposed onto those that controlled electron transfer in all life forms until then. This model helps to explain why many regulatory pathways are connected to fundamental sulfur-mediated electron transfer processes. As the levels of complexity increased from unicellular to larger multicellular organisms, the fundamental principles of regulation were conserved.

In this review we provide an integrative biology concept of redox regulation (Fig. 1). We here define the chemical interaction of RSS, RNS and ROS among themselves and with downstream biological targets as the *reactive species interactome* (RSI) (Box 1). We propose that the RSI serves an integrative function to *sense* multiple stressors and adjust bioenergetic/metabolic needs accordingly by activating downstream effector pathways to ensure the organism is able to respond to environmental change and stay fit for purpose (Box 2 and Fig. 1). Within this model, H₂S along with other thiols is considered an important

2. How it all began: Evolution of the reactive species interactome.

Life began nearly 4 billion years ago (bya), and approximately 85% of all ensuing evolution occurred under anoxic or extremely hypoxic conditions. Rather than oxygen, two other gases, H₂S and NO, were present in the early atmosphere and arguably shaped the bulk of the evolution of life on Earth.

Two decades ago, a case was made that NO production by simple life forms may have provided a crucial survival mechanism against ROS at the time of emergence of aerobic life, offering an opportunity for its further utilization as an early signaling molecule (56). Intriguingly, the story of reactive species may have started much earlier. Indeed, more recently H₂S has been implicated in the origin of Life (139). The following section places emphasis on the role of sulfur in the evolution of redox metabolic systems, how at a later stage O₂ replaced some of the roles of sulfur as donor and acceptor of electrons for energy metabolism and signaling, and how reactive species may have contributed to the evolution of Life by enabling environmental sensing, metabolic plasticity, and cell-cell communication. Comparisons with other life origin theories, e.g., the hydrogen hypothesis, the RNA world and panspermia are beyond the scope of this article. For a more comprehensive treatise of how the emerging field of ‘systems chemistry’ has shaped our understanding of the origin of Life and that of metabolism, and the fundamentals of biochemical adaptation the reader is referred elsewhere (20,80,149).

2.1 Prebiotic primordial interactions - Generating the building blocks of Life

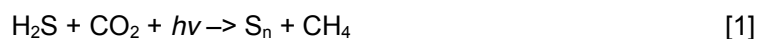
Life requires a set of essential molecular building blocks from which to assemble more complex structures, enzymes to direct these processes, membranes to partition simultaneously occurring events, and energy to overcome inherent entropies (Box 3). The building blocks of Life comprise inorganic or organic precursors of RNA, DNA and proteins

which, it is proposed, were derived from either intense electrical discharge in a “primordial soup” containing basic elements such as carbon, sulfur and nitrogen (125), atmospheric photochemical reactions (158), extraterrestrial sources ranging from collisions with massive objects (22,147) to the fine interstellar dust (which continues to add a large amount of organic compounds to Earth on a daily basis (9)), or through volcanic activity and hydrothermal fissures in the Earth’s crust. However, as only the latter can provide a constant and reliable source of energy, it is most likely that Life emerged here (98,148). The Earth’s earliest atmosphere must have contained large amounts of H₂S (198). Sulfide is an efficient reductant, and its chemical nature was fundamental for driving protometabolic reactions with N₂ and CO₂ to form RNA, amino acids, and lipid precursors (142). Our planet has been likened to a primordial reaction cell (148) where energy in the form of reducing equivalents, namely ferrous iron (Fe²⁺) and sulfide (H₂S, HS⁻, S²⁻), traversed the Earth’s crust through pores (hydrothermal vents) at a steady, and therefore dependable rate. Many of these hydrothermal vents sit on massive sulfide deposits called sulfide lenses (168,169), where the magmatic flow heats the water to over 400° C. The combination of heat, water and high-pressure drives organic synthesis not possible under other conditions, and as the water rises and cools the products become stable.

An argument can be made for the primacy of sulfide in the origin of life: in combination with transition metals, especially iron, copper, zinc, molybdenum and tungsten, allowing 1 and 2-electron transitions, sulfides formed a variety of catalysts that were prototypical enzymes for organic synthesis and created a platform upon which synthesis could occur (33,163,200,201). These metal-sulfide minerals formed primordial “membranes”, allowing compartmentalization of parallel chemical reactions (122). Then, the constant flux of reducing equivalents into the comparatively oxidizing environment of seawater provided a continually renewed and reliable energy source. In addition, some oxidized sulfur coming from the vents gave rise to the formation of defined redox zones.

2.2 Evolution of Life – from sulfur to oxygen

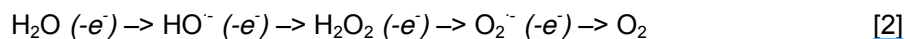
Life is likely to have begun around hydrothermal vents in a ferruginous (anoxic and Fe²⁺ rich) ocean approximately 3.8 bya (153,164) and it was chemolithotrophic, completely dependent upon the Earth for energy (Fig. 2). Within a surprisingly short time, 200-400 million years, photosynthesis appeared, which allowed Life to become independent from Earth's energy. The earliest light gathering antennae were not able to harvest enough light to oxidize water, and the process was anoxygenic. It has been proposed that an intermediate such as H₂O₂ was used as the initial electron donor (154). However, H₂O₂ would have been in short supply, and it is more likely to have been H₂S, H₂S₂ or a related sulfur species (Eq. 1), as seen in modern-day green and purple sulfur bacteria (62).



This reaction is important because H₂S was plentiful and the enzymes that evolved to catalyze this reaction could be readily adapted to oxidize H₂O once sufficient energy could be extracted from the sun.

Oxygenic photosynthesis likely first appeared in cyanobacteria around 3 bya (Fig. 2). This ultimately led to the *great oxidation event* around 2.3 bya when atmospheric O₂ is thought to have risen to 1-2%, which is 5-10% of present atmospheric levels (45,164). However, apart from small oxygen "oases" in the shallows the oceans remained anoxic. Although limited, atmospheric O₂ slowly oxidized exposed elemental sulfur and dissolved H₂S/HS⁻ to sulfate, which was then carried to the oceans, reduced to H₂S by the pervasive Fe²⁺ and, within a hundred million years, vast areas of ocean became euxinic (anoxic and sulfidic). It was in this environment that endosymbiosis, in which a sulfur-reducing Archaea engulfed a sulfide-oxidizing α -protobacterium, produced the mitochondrion around 1.5 bya (111,165). These early eukaryotes would later incorporate cyanobacteria and thus become the ancestors of modern day plants. Combined oxygen production by cyanobacteria and primitive plants eventually oxidized all the oceanic iron and sulfide, and around 600 million

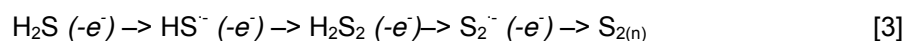
years ago atmospheric O₂ began to increase to present-day levels (Fig. 2). This *oxic* environment is generally thought to have had dire consequences due to formation of hydroxyl radicals (HO[•]), H₂O₂, and superoxide (O₂^{•-}), collectively defined as ROS, (Eq. 2);



According to the *OxTox* hypothesis, organisms either had to develop antioxidant strategies (109), retreat to anoxic niches, or die. But was it really this bad?

2.3 Antioxidant defense or rather sulfur detoxifying strategies?

Before the atmosphere enriched with O₂, it is quite likely that the early anoxygenic photosynthesis (Eq. 3) initially evolved as stepwise one-electron oxidation of H₂S; (Eq. 3);



Thiyl radicals (HS[•]), disulfane (H₂S₂), and persulfide radicals (S₂^{•-}, “supersulfide”) thus generated can indeed be very reactive and are either potent oxidants or reductants (see section 3), collectively known as RSS.

Like ROS, RSS could have had dire consequences: organisms either had to acquire detoxification capability for coping with RSS or die. These capabilities would have to be different from those found in anaerobic organisms, which could escape the oxygen by retreating to anoxic niches. For organisms carrying out anoxygenic photosynthesis retreating to asulfidic niches was not an option, as these did not exist. Therefore, safely disposing of RSS, or their use for signaling or further metabolism, must have enabled the acquisition of appropriate metabolic pathways long before oxygen became prevalent. This would explain why antioxidant systems like superoxide dismutase (catalyzing reduction of O₂^{•-}), catalase and glutathione peroxidase (catalyzing the reduction of H₂O₂), and the redox systems governed by thioredoxins, peroxiredoxins and glutaredoxins, all appeared with the advent of anoxygenic photosynthesis more than 2 bya before they would be called on to deal with ROS (21,103,124,137,221). Therefore, we suggest that – contrary to common belief (8) – these

systems evolved to detoxify RSS and/or to shuttle reducing equivalents for energy utilization or signaling. With the advent of O₂ it would have been a relatively trivial matter to switch from dealing with RSS to ROS, as their biological chemistries present more similarities than differences. The use of multiple reactive species is in alignment with the need to keep the composition of the internal environment (Claude Bernard's '*milieu interieure*') relatively constant (83).

2.4 RSI for sensing and metabolic plasticity

The ready availability of energy in a usable form is a fundamental requirement for survival and reproductive success. In addition to defense and repair systems suitable to cope with harsh environmental conditions, in a world of finite resources organisms require metabolic flexibility to respond and adapt to changes in environmental conditions (Box 2). The capability of early life forms to adjust their energetic needs and metabolic capability to effectively respond to a variable availability of nutrients/precursors requires an ability to sense and respond to those changes. This would involve the capability to “sniff out” the prevailing conditions in the extracellular environment and adjust metabolic pathways accordingly. The metabolic plasticity required for this responsiveness in living organisms presumes the ability to securely cope with reactive species. Reactive species are formed mainly as enzymatic products from specific organic and inorganic substrates, including amino acids, nitrite, polysulfides, sulfite, sulfate, and O₂ (as discussed in detail in section 4). The RSI captures the interaction at the interface between internal and external milieu that enabled metabolic plasticity of early, unicellular life forms, and persists in regulating the intersection of co-metabolism and pathogenesis in response to bacterial infection today (155).

2.5 From monocellular to multicellular life forms

The development of intercellular communication and the emergence of symbiotic arrangements provided new “collaborative” opportunities to cope with environmental and

infectious threats as well as nutritional shortages. As discussed for NO earlier (Box 3) (56), longer-lived RSI metabolites may later have participated in cell-cell communication and enabled co-metabolic negotiations. As levels of regulatory complexity within those multi-cellular life forms increased, so did the need for communication. Yet, without appropriate protection these symbiotic life forms were still vulnerable to threats and dependent on opportunities provided by their local environment. Gaining independence and resilience against external stressors required formation of cell assemblies allowing robust growth and movement. This may have been a driver for the development of larger multicellular organisms with distributed critical functions and enhanced resilience. With redox processes at the heart of global regulation, all organisms larger than perhaps a few hundred cells would require an internal system to communicate metabolic activity status and perceived threat level throughout the entire system, allowing bioenergetic prioritization to survive and reproduce; in other words an inter-organ communication system (see Section 5 for further details).

3. The reactive species interactome: Sensing and transducing elements

The rich chemistry of the RSI offers a unique opportunity to fine-tune biological reactions, taking advantage of the diverse chemical nature and lifetimes of the intermediary products formed. The interaction of RNS with ROS, exemplified by the formation of ONOO^- , from $\text{O}_2^{\cdot-}$ and NO has been conceptualized in the form of the chemical biology of NO previously (65,190). ROS/RSS interaction leads to production of oxidized sulfur species, some of which can further react with biological targets, including cysteine thiolates, to generate e.g. persulfides. Much less is known about the interaction of RSS with RNS to generate S/N hybrid species. However, HSNO/SNO^- and SSNO^- are prominent examples (37-39) that, along with persulfides/polysulfides (37-39,41), garnered significant interest lately. Persulfides and S/N-hybrid species have a chemical biology with unique characteristics (37). The following section provides a brief overview of known interaction products of biological significance and discusses how their fundamental chemistry dictates their kinetics of formation, action radius and biological reactivity and how they are particularly fit-for-purpose in regulated biological systems (Box 3).

3.1 The chemical biology of the RSI - Interaction of NO, H₂S and O₂ and derived species.

It is becoming increasingly evident that the signaling and physiological functions of NO, H₂S and O₂ should be viewed as components of an integrated whole (219) since they have the potential to interact with each other and effect common biological targets. A comprehensive treatment of all the possible chemical/biochemical interactions between NO, H₂S and O₂ (and derived species) and their potential interactions at common biological targets is an enormous undertaking and beyond the scope of this review; other, more comprehensive treatments are available (5,65). Therefore, the possible interactive nature between NO, H₂S

and O_2 will be discussed in very general terms. However, a more detailed emphasis on sulfide species will be given, because this is an area of significant current activity with understanding of much of the chemical biology of these functional groups coming to light only recently.

Of all the small molecule bio-regulators, the chemical biology of O_2 is clearly the most studied and established. As the ultimate electron acceptor for aerobic life, reduced O_2 species such as superoxide ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2) and hydroxyl radical (HO^{\cdot}) are thought to be generated enzymatically and non-enzymatically, and possess biological relevance. Indeed, all have been proposed to serve as cell signaling agents and/or have pathophysiological consequences. All of these species have been grouped together under the somewhat misleading term ROS even though their reactivities are distinct, highly dependent on the cellular environment and potentially opposing. For the sake of brevity, it is probably best to categorize the different entities according to their predominant chemical attributes in tabular form (**Tab. 2**).

Akin to the term ROS, the equivalent terms RNS and RSS denotes NO-derived and H_2S/RSH -derived species. Undoubtedly, these terms can be equally misleading since the chemical reactivities of RNS and RSS are widely varying and distinct. Regardless, the generation and predominant chemical properties of the RNS and RSS are also listed in **Tab. 2**. It is especially noteworthy that the interaction between ROS, RNS and RSS can lead to products with distinct (and even opposite) chemistry from that of the precursors. For example, the reaction of NO with $O_2^{\cdot-}$ to make peroxynitrite ($ONOO^-$) takes two poor oxidants (NO and $O_2^{\cdot-}$) and generates the potentially potent oxidant, $ONOO^-$. As shown in **Tab. 2**, ROS, RNS and RSS taken together cover a wide array of chemical properties ranging from highly reducing (RSSH, $O_2^{\cdot-}$) to highly oxidizing (HO^{\cdot} , NO_2), from highly electrophilic (RSOH, H_2O_2 , HNO) to highly nucleophilic (RSSH), and from good hydrogen atom donors (HNO, RSSH) to potent hydrogen atom abstractors (HO^{\cdot} , NO_2). This chemical

diversity allows Nature to take advantage of widely varying interactive chemistries provided by a limited number of biochemical precursors (namely O₂, NO, H₂S and derived species). The cellular conditions conducive to formation of these species implies a selective pressure towards systems that enable a high level of control together with the regulation of cellular function with appropriate biochemical transformations. For sensing/signaling purposes the biochemical syntheses of ROS, RNS and RSS must be tightly controlled kinetically, temporally and spatially. For example, NO biosynthesis can occur via three primary pathways involving NOS enzymes that are distinct with regards to their regulation and location (40,58). The generation of NO₂ from NO is kinetically second order in NO and first order in O₂, indicating that significant NO₂ levels (at least made via NO/O₂ chemistry) can only be produced in compartments possessing high levels of both precursors such as lipid membranes (115). Of note, nitrogen oxide-modified lipids (containing nitrated fatty acids) possess potent biological activities (156). Moreover, significant generation of ONOO⁻ requires that both NO and O₂⁻ be made at the same place, rate and time (96). This requirement makes ONOO⁻ generation rather difficult, possibly protecting cells from inadvertent formation and narrowing its action radius.

S-Nitrosothiol formation can occur in several ways (37,65). One possibility is the reaction of a free thiol with a nitrosating species, i.e. an entity that donates the equivalent of “NO⁺” such as N₂O₃. Another possibility is the reaction of a thiyl radical with NO. Importantly, N₂O₃ generation is kinetically restricted (214) (for similar reasons as NO₂ generation); as a one-electron oxidant thiyl radical is very reactive, and its formation can only take place under very specific conditions, such as at the active site of enzymes like ribonucleotide reductase (184). These strict chemical requirements offer a selective advantage by limiting the generation of unwanted reactive and/or deleterious species, which ensures that they are only formed under specific conditions for a particular purpose. By contrast, inadvertent or aberrant generation of any ROS, RNS or RSS carries the risk of pathophysiological consequences.

Finally, RSS receiving considerable recent attention are hydropersulfides (RSSH). Generation of hydropersulfides from the corresponding thiol represents an oxidation (an RSSH species is at the same oxidation state as a disulfide, RSSR) and can be mediated by several of the oxidants listed in **Table 2** (e.g. H₂O₂ in the presence of H₂S). Interestingly, a hydropersulfide is a superior reductant compared to the corresponding thiol. Thus, an extremely potent reductant (RSSH) is made primarily under oxidizing conditions, a fact that seems to have been taken advantage of in Nature as it has been proposed that RSSH formation can be protective against oxidative stress (140).

Since RSS are relatively new players in the RSI some more space is allotted here to the discussion of these species.

3.2 The chemistry of persulfides/polysulfides

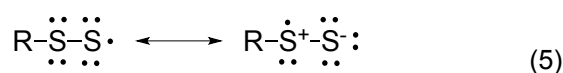
One of the most well established reactions of H₂S in biochemistry is that with disulfides (10,60,140). Reaction of H₂S with RSSR yields an equilibrated system involving the corresponding persulfide (RSSH) and thiol (RSH) species (Eq. 4).



RSSH species display a unique chemistry that differs from that of RSH and H₂S, conferring potential advantages in biology.

In comparison to RSH, RSSH is both more nucleophilic and reducing. The greater nucleophilicity of RSSH can be explained by 1) the α -effect, in which the electrons of the internal sulfur atom repel those of the external sulfur atom, thus enhancing nucleophilic reactivity – a characteristic lacking in RSH and, 2) the pK_a of RSSH typically being 1-2 units lower than analogous RSH species, making the anionic RSS⁻ present in greater concentrations than RS⁻ at physiological pH.

RSSH is also a more potent one- and two-electron reductant than RSH. The greater two-electron reducing ability of RSSH can be explained by its greater nucleophilic character. The fact that RSSH is a better one-electron reductant than RSH is explained by the stability of the corresponding perthiyl radical (RSS \cdot) over that of the thiyl radical (RS \cdot). Formation of RSS \cdot leads to a resonance-stabilized unpaired electron (shown below) that does not exist for RS \cdot (Eq. 5)



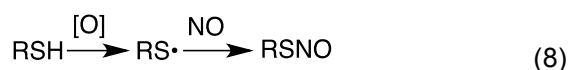
Lastly and perhaps most intriguingly, RSSH are also electrophilic species whereas RSH are not. Consideration of the oxidation state of the sulfur atoms in RSSH reveals that each are both in a -1 oxidation state. By comparison, the oxidation state of the sulfur atom of RSH is -2 and therefore, RSSH is oxidized with respect to RSH. In this light, RSSH are similar to RSSR (in which both sulfur atoms are also in the -1 oxidation state) and thus, are also able to act as an electrophile. Indeed, the electrophilic ability of RSSH is expected to be a function of pH, as the deprotonated RSS $^-$ species is considered to be less electrophilic (and more nucleophilic) than the protonated RSSH. Electrophilic reactivity of RSSH can yield H $_2$ S (via nucleophilic attack on the internal sulfur), or result in a transsulfuration process (via nucleophilic attack at the terminal sulfane-sulfur), yielding another RSSH species (Eq. 6) or inorganic polysulfides (i.e. HSSH; Eq. 7).



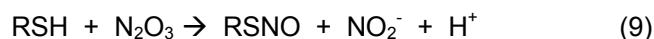
3.3 S/N hybrid species

Although several groups have investigated the interaction of RSH and nitrogen oxides, specifically NO, mechanisms for the formation of resulting species in biological systems are still controversial. Therefore, the possible reactions of RSH and other related

species with NO will be discussed here from a chemical standpoint and implications for biological relevance will be given based on this. As alluded to above, *S*-nitrosothiols have been reported to have important biological function and serve as biological signaling molecules (17,59,179). However, no direct reaction between RSH and NO should be expected to produce such RSNO species because NO has an unpaired electron that occupies an NO antibonding orbital, preventing nucleophilic attack by RSH. However, oxidation of RSH to the corresponding RS[•] allows for reaction with NO, yielding RSNO (Eq. 8).



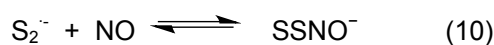
Other pathways leading to RSNO formation include RSH reaction with products from the reaction of O₂ and NO (i.e. N₂O₃, Reaction 9), or by reaction with metal-nitrosyl complexes in which the NO ligand acts as a nitrosonium ion (i.e. Fe²⁺-NO⁺, Eq. 10). For example, coordination of NO to a ferric iron species (Fe³⁺) can generate a ferric nitrosyl complex (Fe³⁺-NO; also described as {Fe(NO)}⁶ using the Enemark-Feltham notation for metal nitrosyls). This species can be viewed as having significant ferrous nitrosonium character (Fe²⁺-NO⁺) and thus can serve as a source of “NO⁺” when reacting with appropriate nucleophiles.



Likewise, similar reactivity is predicted for HS⁻ (in comparison to RSH), theoretically leading to formation of HSNO. For the same reasons as outlined for RSH above, no direct reaction between HS⁻ and NO should occur to any significant extent.

Like RSH, RSSH is not expected to react directly with NO. Although one-electron oxidation of RSSH to RSS[•] might be expected to yield the corresponding alkyl-S-nitrosopersulfide (RSSNO) via reaction with NO, recent studies indicate this either does not occur to any great extent (10) or the product has a short lifetime (2). For this reason, RSSNO may not be expected to serve as a biological signaling molecule or NO transporter. Curiously (and unlike thiyl radicals), RSS[•] is rather stable even in the presence of O₂ (10), offering potential opportunities for electron transfer reactions under aerobic conditions (see Section 5.3).

Contrary to the presumed instability/reversibility of RSSNO, SSNO⁻² (37) appears to be relatively stable (a result of the resonance-stabilized anion (10,120)), existing for extended periods of time even in the presence of other RSH species (39,41). Although SSNO⁻ has been observed to form under various conditions including reaction of NO with H₂S and polysulfides (HSS_nH, n≥2) (41,120), the exact mechanism for SSNO⁻ formation is unknown. However, it is reasonable to consider that SSNO⁻ is made via reaction of NO with trace polysulfide contaminants present in H₂S sources. For example, presence of trace S₂⁻ (a possible result of one-electron oxidation of S₂²⁻ or homolytic cleavage of S₄²⁻), which is a species well recognized by sulfur chemists to exist in salt melts and heated non-aqueous solutions of sulfur (181), could be expected to react directly with NO, yielding SSNO⁻ (37), (Eq. 10).



It should be noted however that to date, SSNO⁻ has yet to be observed in a biological system, leaving its relevance and biological formation still uncertain. Nevertheless, pharmacological SSNO⁻ has been shown to release NO, dilate blood vessels and activate the prototypical Nrf2 stress-response pathway (39,41,42).

² We here use the most traditional notation of perthionitrite (SSNO⁻), as the molecule was originally named by Seel *et al* (see reference 37 for a comprehensive review); however, the notation as nitrosopersulfide (ONSS⁻) is also correct.

3.4 Cysteine-based redox switches and redox relays

Free sulfhydryl (-SH) groups in low-molecular-weight thiols such as cysteine, peptides (like GSH), and proteins (e.g. albumin) are predominant targets of RSI signal transduction; others include methionine, tryptophan, tyrosine, and histidine moieties, but their functional significance is not fully understood.

Cysteines may serve structural, catalytic and regulatory functions in proteins and are considered *redox switches* as they are targeted for oxidation, nitrosation, thiolation and sulfidation (also termed “sulfhydration”). Therefore, rather than “on/off” switches, protein cysteines may act as multistage cysteine relays (Fig. 3), allowing cells to dynamically adjust protein structure and enzymatic function according to the local redox state (100,216). In addition to protein thiols, low-molecular-weight thiols including cysteine and glutathione are important contributors to intracellular and, via mixed disulfide formation, possibly also extracellular redox status (Fig. 5). In order to function as regulatory elements, those thiol-based post-translational modifications must be also under kinetic control. This is achieved by coupling cysteine-based modifications to a battery of target-specific reductases, denitrosylases, and desulfurases which together are able to maintain steady-state concentrations of thiol modifications low; these include thioredoxin/thioredoxin reductase, glutaredoxin, peroxiredoxins and other enzymes (68). Both thiol modifications and their regeneration are dynamically linked to global redox and nutritional status (see Section 4, and Fig. 1).

3.5 Biological targets of the RSI

The net biological effects of the reactive species are determined by the nature, level of expression and function of the biological targets carrying functional cysteine redox switches (Box 1 and Fig. 1). Examples include protein kinases and phosphatases, ion channels, transporters, and enzymes (e.g. those involved in intermediary metabolism), allowing rapid short-term adjustments (Fig. 1). In addition, longer-term regulation is achieved

by interaction with redox-sensitive transcription factors, e.g. Nrf2/Keap1, NFkb, and HIF (54). Even longer-persisting effects are achieved by redox regulation of gene expression under epigenetic control, making redox effects transmissible to the progeny. This notion is consistent with the *developmental origin of health and disease* (DOHaD) paradigm, which provides a mechanistic explanation for the pathophysiological basis of how environmental influences experienced during early embryonic development may influence the risk of non-communicable diseases later in life and across generations (71,77,217)(see also Section 6).

3.6 Functional significance of the RSI

A corollary of the RSI concept is that reactive species can no longer be regarded as mere stressors (Box 1). Rather, they should be considered controlled reaction products, which serve to sense and transduce information about any changes of internal and/or external conditions; as such they may be considered as elements of a regulatory system (the RSI) that enable an integrated response to various forms of *stress*, e.g. changes in metabolic, nutritional and redox status, and environmental conditions (Box 4). See also Boxes 1, 2, and 5.

4. RSI precursors in the context of intermediary metabolism and nutrition

The RSI is driven by specific substrates for enzymatic production of individual ROS, RNS and RSS. Local production of reactive species depends on the availability of O₂, certain amino acids and cofactors as well as the activity of specific enzymes, which together are embedded within an intricate system of intermediary metabolism that determines the pattern and rate of fluxes according to synthetic and energetic needs (Fig. 4). Given the fundamental role redox regulation plays in cellular defense, repair and survival, the balance of metabolic fluxes must be prioritized to first support an adequate redox status before fulfilling local metabolic needs. The following section provides a short overview of the precursors needed for ROS, RNS and RSS synthesis in the context of dietary intake and human nutrition (Fig. 4).

4.1 Precursors of ROS, RNS and RSS – The oxygen-arginine-methionine metabolome

Unsurprisingly, in aerobic organisms oxygen sensing is intimately linked to intermediary metabolism (1). ROS production involves a variety of different enzymes and organelles utilizing oxygen, but the relationship is not straightforward; counter-intuitively, more mitochondrial O₂^{•-} is produced in hypoxia than under normoxia (207). O₂ is also the substrate of various NADPH oxidases producing either O₂^{•-} or H₂O₂ (16,199). Other sources include xanthine oxidoreductase, 5-lipoxygenase and cytochrome P₄₅₀. Non-enzymatic ROS production may also be generated in an unregulated fashion through the metal-driven Haber-Weiss reaction leading to the formation of OH[•]. Enzymatic generation of NO requires both a source of N and O, specifically in the form of arginine (and possibly homoarginine) and O₂ (72,127,167), whereas NOS-independent reactions leading to NO formation include nitrite/nitrate reduction (43,90,116,172,192). NOS activity is also dependent on tetrahydrobiopterin and reducing equivalents in the form of NADPH (72,185). RSS

production relies on the availability of methionine, homocysteine and cysteine serving as substrates in the methionine recycling and transsulfuration pathway. The enzymes of the transsulfuration pathway are responsible for the formation of cysteine from the essential amino acid methionine and serine. Cysteine is crucial for defining protein structure (disulfide bonds), function (e.g. enzymatic activity) and redox signaling (e.g. by acting as *redox switches*, see Section 3), and as a building block for glutathione (GSH) production. The tripeptide GSH (Glu-Cys-Gly) is less toxic for cells than cysteine itself, is present in millimolar concentrations intracellularly and buffers the cellular antioxidant network, together with ascorbate (vitamin C), ubiquinol (coenzyme Q) and α -tocopherol (vitamin E) (175). The two other amino acids critical to GSH synthesis are glycine (itself in part derived from serine) and glutamine (formed from glutamate and interaction with proline) (see Suppl. Fig.1).

Cofactors like folate, choline, vitamin B₆ (pyridoxal phosphate) and B12 (cobalamin) are critically important for adequate methionine recycling. Interestingly, several metabolic aberrations in either the tetrahydrofolate cycle, the methionine recycling capacity, or flux through the transsulfuration pathway are associated with elevated homocysteine concentrations in blood. The latter marks metabolic imbalance and/or inadequate nutrient availability, and hence is a marker of risk for cardiovascular disease (95,138). The transsulfuration pathway enzymes cysteine- β -synthase (CBS), cysteine- γ -lyase (CSE) and 3-mercaptosulfotransferase (MST) are also responsible for the endogenous production of H₂S (144) as well as organic persulfides (CBS) (86) and polysulfides (3-MST)(101,102). CBS is functionally regulated by NO, its expression enhanced by oxidative stress and gene transcription hormonally regulated in response to fuel supply (182). These pathways therefore can be considered to be a central hub for intermediary metabolism and a point of intersection for the production of proteins (as building block for tRNA and ribosomal protein synthesis), lipids (via S-adenosylmethionine and choline), methylation reactions (via S-adenosylmethionine) as well as GSH production and H₂S/persulfide signaling. This is in accordance with the recent discovery that the nearly 4 billion year old metabolism of the last

universal common ancestor (LUCA), the forerunner of all contemporary life forms on Earth, already relied on S-adenosylmethionine-dependent 1-carbon metabolism to make a living by harnessing energy from its primordial geological environment (208).

The interaction of H₂S, NO and O₂ is tightly linked to bioenergetics through their convergence in the regulation of mitochondrial function. In cultured cells, hypoxic stress induces CSE translocation from the cytosol to mitochondria to sustain ATP production, presumably via fine-tuning of the electron transport chain and use of sulfide as a mitochondrial substrate (63,78,188). Marked changes in metabolic needs and/or mitochondrial function are likely to affect precursor/cofactor availabilities and therefore RSI-mediated sensing and adaptation processes (Box 5).

4.2 How nutrition affects precursor availability

4.2.1 L-Arginine uptake and metabolism in the human body

In addition to its role in protein biosynthesis, arginine is a precursor for creatine and NO production. There is the need for endogenous formation of arginine, and for young, growing mammals it has to be provided in the diet (i.e. it is an essential amino acid), but less so in adulthood, where it can be considered to be conditionally essential. (189). Inadequate availability of arginine has been associated with T-cell and endothelial dysfunction (129); these effects are not usually observed in healthy adults (24) as endogenous synthesis is sufficient to meet usual demands, except in situations of catabolic stress (e.g. inflammation or infection) (128). The net rate of endogenous *de novo* arginine synthesis is modulated in relation to provision from the diet and the breakdown of proteins (28).

There is evidence for *de novo* synthesis of arginine in enterocytes up to the age of 3-5 years (202,212). Beyond this, a more complex inter-organ amino acid cooperativity is required, which involves enterocytes and the renal cortex (14,48,213) (known as the *intestinal-renal axis*). In enterocytes, endogenous and dietary glutamine is converted into

citrulline via glutamate and ornithine (218). Circulating citrulline is then taken up by cells in the renal cortex and converted into arginine (14,48,213). The conversion of arginosuccinate to arginine, the final step in arginine *de novo* synthesis, requires arginosuccinate lyase (ASL), which is almost exclusively found in the renal cortex. Hepatic arginine synthesis is embedded in the metabolic pathway of the urea cycle and therefore results in high flux but low net production (218).

Approximately 60% of net arginine synthesis occurs in the kidney. However, renal insufficiency does not result in decreased plasma arginine concentration, but in increased citrulline levels (112,218). The mechanisms underlying the maintenance of plasma arginine concentration are poorly understood, but may involve a compensatory decrease in arginine utilization (28).

Only a small proportion (~1%) of the overall arginine turnover but a considerable amount (54%) of circulating arginine is used for NO production (27). In healthy human adults, the production of NO from L-arginine corresponds to ~1 mmole/day (173). Citrulline, one of the products of NOS, can be recycled by transamination to arginine via the so-called “citrulline/NO cycle” or “arginine/citrulline cycle” (76,218), although *in vitro* this cycle is much less efficient than the hepatic urea cycle (218). Of note, the guanidino nitrogen group used to form NO is mostly not derived from dietary arginine, but from carbamoylphosphate and aspartate (see Suppl. Fig.1).

Arginine moieties in proteins can be methylated to form mono- and dimethylated derivatives, which are released into the circulation upon proteolysis. Circulating concentrations of two of these methylated arginine derivatives (L-N^G-methylarginine and asymmetric, but not symmetric dimethylarginine) are effective inhibitors of cellular arginine uptake and NOS activity (113). While symmetric dimethylarginine does not act as direct NOS inhibitor it can reduce NO production by competing with arginine transport (13)..

4.2.3. Methionine recycling, transsulfuration and one-carbon metabolism

Methionine and cysteine are the two sulfur-containing amino acids (SAA) incorporated into proteins. Methionine is one of the most hydrophobic amino acids. It has important physiological roles including the initiation of translation via initiation tRNA (met-tRNA_i^{met}) and methylation pathways via *S*-adenosylmethionine (18,182), which are important for the formation of co-factors such as biotin and lipoic acid. Despite its importance in physiology and being the 7th most abundant element in higher vertebrates, the extent to which the dietary provision of sulfur-related components adequately supports the needs of sulfur metabolism has received inadequate attention (87,134).

The main sources of sulfur in the diet are inorganic sulfate (SO₄²⁻) and SAAs. Methionine can be converted into cysteine, and with a sufficient supply of the former, adequate amounts of the latter can be formed endogenously from serine. However, as this reaction is irreversible, methionine has to be provided preformed in the diet regardless of cysteine status (152). Dietary methionine is absorbed rapidly and almost completely and only small amounts are excreted directly following bolus administration. It is eliminated from plasma with a half-life of approximately 150 minutes, and a 3-fold increase in urinary SO₄²⁻, another important product of transsulfuration (85).

Healthy adults are in sulfur balance with equilibrium between intake, transsulfuration and excretion. The conversion of methionine to cysteine via homocysteine, is the only catabolic pathway of methionine. Sulfur is excreted via the kidney mainly as free sulfate (SO₄²⁻, 77% – 92%), esterified sulfate (7% – 9%), taurine (2% – 6%), cyst(e)ine (0.6 – 0.7% and minor amounts of methionine, homocysteine, cystathionine, N-acetylcysteine, mercaptolactate, mercaptoacetate, thiosulphate and thiocyanate (182). The net changes represented by external balance do not adequately capture the considerable internal flux associated with the turnover of methionine and cysteine into and from protein and peptide pools, estimated as 32 mmol and 38 mmol per day, respectively (19). Although the daily production of H₂S has not been species quantified it is likely considerably higher than that of NO.

Homocysteine represents a determinant branch point for methionine flow either to cysteine through transsulfuration via pyridoxal-phosphate dependent CBS and CSE to cystathionine and cysteine, or re-methylation via betaine, folate or vitamin B₁₂ (cobalamin) dependent pathways (89,177) (Fig. 4 and Fig. S1). Under physiological conditions, there is a similar flow through each pathway (183), but changes in methionine supply or the relative availability of donors for methylation modify the flow through these pathways. Low methionine availability results in high transsulfuration rates (presumably secondary to systemically increased ROS production), whereas a replacement of methionine by cysteine results in increased re-methylation (49). Importantly, modulation of dietary SAA intake can affect plasma redox status (93).

Methionine homeostasis is achieved by modulation of the balance of protein turnover and the relative rates of transsulfuration and re-methylation (64,151,152,183). The reductive adaptations developed during malnutrition limit the capacity for handling large doses of methionine leading to a high plasma concentration of methionine (178) and increased concentration of homocysteine (88). Similar to poor vitamin B6 status and limited serine availability perturbations in amino acid status can result in homocysteine accumulation, increased re-methylation to methionine and a concomitant reduction in flux through the transsulfuration pathway. Overall, methionine maintains a very stable plasma concentration at the expense of endogenous sulfate production (87).

4.2.4 The sulfur / nitrogen relationship

In adults consuming a normal diet there is a strong correlation between urinary sulfate and dietary sulfur-containing amino acid intake, and urinary S:N ratio reflects dietary S:N ratio (157,171). This relationship is however modulated by unusual dietary patterns, for example during starvation or on low-protein diets when post-prandial urinary sulfate excretion is reduced to a greater extent than urea excretion (74,97,118,119). This suggests that protein restriction results in preservation of SAA and replenishment of the non-protein

SAA pool, but also complex and tight interconnections between the metabolic pathways of nitrogen and sulfur metabolism to maintain constant plasma concentrations of methionine. Thus, alterations in the S:N ratio of urinary metabolites may hold promise as indicators of a stressed system with unusual metabolic demands.

4.3 Impact of microbial-host co-metabolism on components of the RSI

Besides the above mammalian pathways, H₂S may also be generated from isothiocyanates, which are particularly prevalent in Brassica vegetables such as broccoli (23,32), polysulfides contained in garlic (7), and via gut microbial reduction of dietary sulfate (SO₄²⁻) (23), cysteine and protein (23,119). Microbial H₂S generation may contribute to the total body pool of sulfide (170) and may even have blood pressure lowering effects (191). Likewise, dietary nitrite and nitrate can be reduced by the oral and gut microbial flora to NO, contributing to circulating nitrite levels and mildly lower blood pressure (25,117); together with the enterosalivary recirculation pathway this has become known as the mammalian N-oxide cycle (25,116). It is conceivable that a similar sulfur cycle exists in mammalian organisms and those pools of H₂S and NO may give rise to reactive species, including ONOO⁻ and possibly SSNO⁻ (37,39,41). Fluctuations in intestinal oxygen gradients may further shape the redox relationships between the gut microbiome and the host metabolome (53).

5. Inter-organ redox communication systems

One of the primary functions of the circulatory system in mammals is to efficiently transport oxygen, nutrients and waste products around the system. With one exception (136) little consideration has been given to its further potential role in acting to communicate and maintain whole-body redox status in relation to external environmental conditions and internal metabolic needs .

Individual cells within a given tissue/organ need to sense their microenvironment in relation to that of the entire organism in order to achieve a metabolic status that adequately enables the needs of their preferred activity (Box 4). Similarly, the cells need to relate their individual redox status within a composite redox state that matches with other cells and organs. This necessitates reciprocal sensing of intra- and extracellular redox poise. This has to embrace cell membrane behavior, and an inter-organ communication system that connects the various contributing elements and provides a read-out of the global redox poise. Extracellular fluids such as interstitial fluid, lymph and blood are especially well suited as the connecting medium (36). We propose that the entirety of protein thiols serves as an important redox buffer and that extracellular thiol status helps to mark the global redox state in health and disease.

5.1 Quantifying “oxidative stress” : early attempts

In vitro, oxidative stress is typically assessed by determining the concentration of reduced/oxidized glutathione, antioxidant enzyme (SOD, catalase, glutathione peroxidase) activity and/or the levels of (anti)oxidants and the potential for free radical scavenging using fluorescent molecules with all their limitations (205), including their lack of specificity for ROS vs. RSS (47). More often than not, little distinction is made between assessing capacity to cope with an oxidant burden, determining the magnitude of exposure to an oxidant, and the balance between exposure and antioxidant capacity. Earlier attempts at quantifying oxidative

stress in vivo by measuring levels of select products of lipid, protein or DNA oxidation (e.g., 8-isoprostanes, malondialdehyde, protein carbonyls, 8-hydroxyguanosine) often met with disappointment in that all these biomarkers showed distinct profiles scaling with oxidative stress burden but considerable temporal heterogeneity. The reason for this divergence is not immediately apparent, and is likely due to the fact that some “markers” are only bystanders of oxidative damage, while others are actual regulatory nodes of the antioxidant network. Insights into the system architecture of the redox network and its modus operandi are required to interpret this information. Valuable insight might be derived from information about metabolic fluxes (the direction of travel within metabolic pathways), which can be achieved by either applying stable isotope labeling methodology or monitoring natural isotopic fractionation using high-sensitivity, high-resolution mass spectrometry (“fluxomics”).

5.2 Assessment of low molecular weight and protein thiols in plasma/serum

Based upon the assumption that thiols play a determinant role in redox regulation, measuring the ratio of reduced over oxidized forms of small aminothiols such as cysteine or glutathione is potentially more powerful than measuring levels of individual oxidation products inasmuch as it informs us about the status of a dynamic system that serves to shuttle nutrients between cells/organs and electrons between the intracellular and extracellular milieu. Often misunderstood, it is not the electrochemical redox potentials (of e.g. GSH/GSSG) that drive the biology; those concentration ratios are simply the outcome of fast enzymatic processes related to thiol transport, degradation and regeneration (57). The ratios of reduced/oxidized thiols show diurnal variation (12) and decrease with age, but the redox couples of cysteine and glutathione, for example, are not in equilibrium (92). This indicates that ratios are maintained at defined levels, which might be presumed to confer benefits that are as yet unclear. However, even those measurements reflect only a small part of the overall thiol redox network, as it fails to capture the large protein-bound thiol pool (196) and kinetically controlled exchange reactions with free thiols in the intra- and extracellular compartment. While the overall complexity of this system has been appreciated

already some time ago (196), little is known about central regulatory nodes governing these equilibria.

A recent report suggests that cysteine and glutathione redox status are associated with mortality from coronary artery disease (143). Another highly significant association was found between total free thiol status in serum and clinical outcome in unrelated clinical conditions (61,105). Given the overall complexity of the extracellular antioxidant network and its link to intracellular redox status the latter was utterly unexpected. A simplistic view of total free thiol levels in a given compartment could be interpreted as a direct reflection of the balance between oxidants and antioxidant capacity (or overall redox poise). Indeed, a decrease of reduced thiols and/or an increase of oxidized thiols has been found in patients with blood disorders, cancer, cardiovascular disease, diabetes, inflammatory disease, kidney disease, metabolic disease, neurological disease, skin disease and thyroid disease (3,108,110,123,133,180,220). Thiol oxidation has also been associated with risk factors, including aging, smoking, obesity and alcohol abuse (69). Both within cells (75) and in blood (195), proteins constitute by far the largest pool of redox-active thiols. Approx. 60% of the total thiol groups in serum/plasma are accounted for by the single free cysteine (Cys³⁴) of albumin (195). Thus, when instead of the ratio of free and oxidized thiols only free thiols are measured, adjustment for total serum protein can be seen as an indirect way of accounting for total thiol content.

In renal transplant recipients, protein-adjusted serum free thiols were found to predict graft failure and patient survival (61,105). In a small cohort of stable chronic heart failure patients there was a positive association between protein-adjusted serum free thiols and a favorable disease outcome (61,105). Interestingly, a study evaluating the concentration of serum protein thiols in a wide range of species concluded that free thiol levels are positively associated with life span, suggesting that control of redox status has retained its importance from evolution to modern-day (patho)physiology (146).

Studies relating serum free thiol levels to other components of the redox network are lacking. In this context, disentangling the relationship between overall thiol redox status (free reduced and oxidized as well as protein-bound thiols), which is likely affected by cellular uptake and reduction processes, and production/metabolism of both NO and H₂S/sulfide would seem to be important. We here propose that protein thiols in the extracellular fluid play a fundamental role in communication between different body compartments, acting as sentinels of distant danger, transporters of specific substrates and as dynamic entities that reflect a readout of global thiol redox status. Rather than relying on the integrity of a single protein to fulfill this important function, there is greater security with greater buffering capability and in Nature the entire protein thiol pool may play a role for this purpose. Nevertheless, albumin is likely to play a more important role quantitatively, simply based on its abundance in the extracellular compartment (195) and the extent to which it transports small aminothiols. While mixed disulfide formation can occur non-enzymatically via attack of reactive protein thiolates (e.g. Cys³⁴ of albumin) at the disulfide bond of oxidized thiols, the reverse process that would regenerate the free thiol is very slow. This implies that a significant portion of these reactions must be controlled through the activity of specific thiol oxidoreductases such as glutaredoxin, protein disulfide isomerase, and thioredoxin/thioredoxin reductase (70). Besides mixed disulfide formation (S-thiolation), other sulfhydryl modifications including S-nitrosylation, S-sulfuration (“sulfhydration”), S-oxidation, and S-acylation (e.g. S-palmitoylation) may also reversibly decrease free thiol availability, but information on the practical relevance of these processes is limited. This may be particularly relevant for cysteinylolation and persulfidation of protein thiols as micromolar concentrations of per/polysulfides were detected in biological tissues (86) and therefore have the potential to affect the measurement of total free thiols.

5.3 Intracellular redox regulation, bioenergetics and intermediary metabolism

The same fundamental regulatory principles that operate in the extracellular compartment likely apply to intracellular redox state (Fig. 5). Much emphasis has been devoted to the process of S-cysteinylation and S-glutathionylation (46,66), although other posttranslational modifications may be of similar importance. Cross-talk between cell compartments of widely different redox status (nucleus, endoplasmic reticulum, peroxisomes, mitochondria) is of significance in this context (30,51,161), since the same reactive species that modulate mitochondrial activity and dynamics (130,211) may also link overall redox poise to intracellular signaling, metabolic control and bioenergetic status (188). Mitochondrial dysfunction has recently been demonstrated to remodel one-carbon metabolism (4), and the latter is fundamental for mammalian health and disease (50).

These regulatory principles would provide the opportunity to prioritize options for metabolic adjustments in individual cells in relation to the overall redox status of the organism, which might be achieved as a consequence of exchange of redox information across cell membranes (Fig. 5). Although the cystine/glutamate antiporter has been implicated (34) the elements connecting intra- and extracellular space to exchange this information are currently unknown. An effective communication across cell membranes requires moieties that reliably operate under widely different redox conditions, i.e. under oxidative as well as reductive stress. There are limited choices chemically that fulfill the need of electron exchange under those extremes, but one interesting possibility may involve the persulfide/perthiyl radical couple (10).

5.4 Redox state and cell survival

Redox status is also linked to cell survival (193,222). Effective repair of damage is cardinal to survival and resilience. DNA repair mechanisms have been studied most extensively, not least because the survival of organisms depends on faithful transmission of genetic information from one cell to the next (and across generations), from the level of DNA replication over chromosomal distribution to the repair of damage incurred. This involves

surveillance systems for structural monitoring and orchestration of the sophisticated repair processes during normal functioning (31). Spontaneous DNA lesions are common events (82), and the response to DNA damage is principally orchestrated through activation of *sensors*, *transducers* and *effectors*. This allows to resolve problems induced by physical or chemical stresses while limiting unnecessary maintenance. Several DNA repair enzymes including poly ADP-ribose polymerase are under redox control (3), which may provide a mechanistic explanation for the observed association between plasma free thiols and active disease. Similar processes are at work under conditions of strong emotional stresses, which are known to be associated with oxidative stress, poor immune function and health, lower telomerase activity and telomere shortening (52). These observations are akin to what happens during normal ageing, just at a higher pace. Thus, life stresses of any sort, perceived or real, seem to lead to accelerated (premature) ageing. Since parallel monitoring of multiple stresses is energetically costly, integrated stress sensing will be a preferred option; bacteria realize this through distributed sensing of metabolic fluxes (106). A sizeable portion of an organism's energy is spent processing sensory information (162), which enables stress tolerance: as the ability of cells to generate adequate levels of energy declines, housekeeping and acute repair processes are compromised and physiological function starts failing.

6. Perspective: How redox biology and insights into the regulation of the RSI may transform personalized medicine.

The regulatory concepts outlined above, as viewed from the perspective of systems biology, may provide useful guidance in any search for prevention and treatment approaches in the emerging area of *redox medicine*. Based upon the notion that 1.) a set of

factors plays a dominant role in sensing environmental changes (O₂, amino acid substrates around the arginine-methionine metabolome and its cofactors), 2.) another set of factors acts as transducing elements (cysteine-based redox relays and the dynamic interplay of free reduced and oxidized, and protein-bound thiols), and that 3.) there are stable end products of the RSI (S,N,O-based stable metabolites), it is possible to create a conceptual framework, which places the complexities of these interactions into context and allows to interrogate the quality and quantity of the balance of forces amongst the different components. The specificity of this approach will be guided not by pre-defined levels of normality of particular read-outs (as commonly practiced in current medicine), but rather by understanding what determines the metabolic fluxes through these pathways (Fig. 6). Due to the interacting nature of the pattern and direction of flow through many pathways within a complex system, the nature and outcome of these interactions will differ depending upon context and circumstance from individual to individual. This underlies the basis of personalized medicine and indicates the diagnostic basis needed to develop rational therapeutic interventions that adequately meet the individual situation (Box 6). In the following, we will present our understanding of redox regulation in the context of health and disease, suggest methods to interrogate the activity status of its components and how to utilize these insights for the benefit of a personalized medicine approach.

6.1 What defines health?

Contemporary medicine still uses the same organ-based classification system physicians developed centuries ago to define disease symptoms. We have become better and better at treating diseases, but not necessarily at identifying the root cause of illness or just “feeling sick” (166). More often than not ‘disease’ itself is equated with dysfunction. The converse is our limited ability to define and measure what constitutes health. In medical diagnostic terms it often means staying within perceived limits of the distribution of variations in simple physiological readouts (such as blood pressure, heart rate, peripheral oxygen saturation, white blood count etc.) and the absence of overt signs of organ/tissue damage. Systems

biology has taught us much about the emergence of unpredicted states arising from the complex interaction of multiple, non-linear interconnected systems, and these lessons are now being incorporated into what constitutes *systems medicine*. To this end, good health is not merely a state defined by the absence of overt symptoms of disease, but includes overall mental and physical wellbeing and resilience to stress. Biomarkers of good health are in short supply and limited to a few markers of nutritional adequacy (e.g. amino acid, protein and micronutrient status) and perceived levels of normality of markers associated with inflammation, oxidative stress cell aging (telomere length) and DNA methylation. However, the latter two tend to differ markedly between individuals and ethnicities and provide limited information about the origin of the problem or the reserve capacity of the regulatory systems affected. From a systems perspective, as long as an organism is alive, one can consider its sub-systems to be functioning; however, perturbations may force them to deviate from their normal equilibria and setpoints. If severe enough, these perturbations may become obvious and observable as disease phenotype (e.g. elevated blood pressure, hyperglycemia), which may be interpreted as the price paid by the organism to stay fully operational. To this end, accelerated aging (and aging *per se*) may be viewed as a process that leads to a compromised capacity for repair and/or adaptive change, eventually culminating in system failure and death (217)

6.2 Personalized medicine

Personalized medicine aims at providing individually tailored prevention and treatment strategies for patients. One important driver for this development is the realization that generic treatment approaches are not optimally addressing patients' needs. With an ageing population and ever-rising healthcare costs the idea is to provide tailored solutions matched to specific deviations from the desired equilibrium with the aim to improve population health. In the words of the Horizon2020 Advisory Group personalized medicine is "a medical model using characterization of individuals' phenotypes and genotypes (e.g. molecular profiling, medical imaging, lifestyle data) for tailoring the right therapeutic strategy

for the right person at the right time, and/or to determine the predisposition to disease and/or to deliver timely and targeted prevention" (see <https://ec.europa.eu/research/health/index.cfm?pg=policy&policyname=personalised>). The fascination with this concept transcends medicine as it goes well beyond the treatment with medical products to include a better understanding of how particular biological mechanisms and interactions of our body with environmental, nutritional and infectious stresses (known as the 'exposome' (210)) affect health and disease along the life course. How can these high expectations be fulfilled?

6.3 Targeted redox metabolomics for understanding disease processes

Great methodological strides have been made in recent years to capture the entirety of genetic and transcriptomic events in an affordable manner. Beyond the use of multiple 'omics' techniques for deep phenotyping, however, a sound understanding of the key drivers and mechanisms underlying disease processes and resilience is required. Importantly, the mere integration of genomics, transcriptomics and proteomics does not provide any information about enzyme function/activity and metabolic pathways. *Metabolomics*, i.e. the analysis of stable products of metabolic pathways, may offer a deeper level of information about the state of an organ or the entire organism, and may therefore be a key enabling technique for personal/precision medicine (6). Much of the recent success of metabolomic approaches is linked to the rapid advancement in mass spectrometry-based techniques over the last two decades, but many technical obstacles are still to overcome to extend the level of coverage typical of genomic approaches to metabolic events. Measuring the entire metabolome of every patient may remain beyond reach for some time, but a targeted approach covering key aspects appears realistic even today.

Despite the advancements in untargeted metabolomics to identify and analyze complex patterns of metabolites, some of the low-abundance, polar or volatile metabolites still pose analytical challenges; in particular, many of the highly reactive species of fleeting

existence, as for example H₂S and its per- and polysulfides belonging to the RSI, are currently only reliably quantifiable after chemical trapping, providing more stable derivatives (86,99). A disadvantage of such approaches is that they tend to perturb natural equilibria, not only complicating the interpretation of changes recorded but also introducing bias as their disappearance can elicit adaptive reactions not normally seen under physiological conditions.

6.4 What to measure where and how?

To adequately capture the substrates utilized by the RSI for sensing and transduction processes, we propose to concentrate on those parts of intermediary metabolism that are central to the regulation of redox status and production of essential building blocks of life such as DNA/RNA, vitamins, amino acids/proteins and fatty acids/lipids. Emphasis should be placed on integrative biomarkers faithfully reporting on multiple mechanisms operating in synchrony. Circulating homocysteine concentrations are a pertinent example as they represent a combined read-out of methionine and vitamin B_{6/12} availability, one-carbon metabolism (methionine recycling and tetrahydrofolate pathways), and flux through the transsulfuration pathway, an ancient pathway established almost 4 bya (208); there may well be many more among the markers we routinely assess today. Thus, besides capturing as many species as possible around these pathways to monitor bodily sulfur/SAA handling and H₂S production the arginine metabolome will be another sensible target to capture as it provides insight into nitrogen handling and NO-related processes. To describe the dynamics of the thiol regulatory system, we suggest quantifying the entirety of free and protein-bound thiols; differences in concentration of stable end-products of the RSI in different compartments may serve as readouts of the sensing and adaptation system in operation. For sulfur hydrosulfide, per- and polysulfides, thiosulfate and sulfate would seem to be useful readouts, complemented by ammonium, urea, nitrite, nitrate, nitroso and nitrosyl species for the nitrogen products. Relevant, clinically accessible compartments to analyze include blood (plasma and cellular components), saliva, sweat, urine and exhaled breath.

To gain insight into the regulation of the entire system whole-body responses to various stressors ought to be explored first in healthy individuals subjected to e.g. exercise vs. rest, hypoxia vs. normoxia, heat vs. cold, fasting vs. controlled feeding, and exposure to frequently prescribed or over-the-counter drugs such as anti-inflammatories. This will form the basis for subsequent interrogations in diseased individuals and patients on specific drug regimens and allow disentangling the underlying redox network architecture using specific patient cohorts. Once we understand how the system is structured, organized and able to sense in a way that enables control and regulation we will be in a position to intervene causally to achieve defined objectives.

7 Summary and outlook: Disentangling the RSI using redox metabolomics to enable precision medicine

The interactions between individual constituents of the RSI and key biological targets of the antioxidant network are complex. Hoping to disentangle the *in vivo* relationships for the benefit of defining treatment opportunities for *redox medicine* will require quantifying stable decomposition products and downstream metabolites of RSS, RNS and ROS interaction, along with their substrates, in multiple biofluids, in health and disease. Since we are not dealing with 'on/off reactions', it would be naive to expect that measuring a few biomarkers in health and comparing them to levels one observes in overt disease would suffice. The perils of oversimplification are nowhere more apparent than in the interpretation of circulating homocysteine concentrations (197). To capture the dynamics of regulation, we ought to measure the same set of readouts in healthy individuals of different sex/age under varying levels of physiological perturbation, (sedentary/exercise, normoxia/hypoxia, starved/fed, etc.) and compare their steady-state levels under these conditions to concentrations in sex/age-matched patients suffering from specific diseases. By carefully matching the physiologic/metabolic phenotypes to the profile of biochemical markers we should be able to learn enough to decode the underlying network architecture. In turn, this will enable us to identify new avenues for targeted therapeutic modulation of redox signaling. Besides guiding pharmacological treatments, those insights may also aid risk-stratifying patients before undergoing major surgery, improve recovery from critical illness or major trauma and inform nutritional priorities for healthy ageing.

Acknowledgements

We are indebted to many of our colleagues for stimulating discussions over the years; special thanks go to Takaaki Akaike, Giuseppe Cirino, Michael P Frenneaux, Michael P Grocott, Dean Jones, Malte Kelm, Hideo Kimura, John Martin, Salvador Moncada, Jerome Santolini, Helmut Sies, Mervyn Singer, David Whitlock and Steven Wootton. CLB acknowledges support by the US National Science Foundation (CHE-1566065), MCK financial support from the German Research Council (DFG CO 1305/2-1, SFB1116, IRTG1902) and the Forschungskommission of the Universitätsklinikum Düsseldorf, and PN from the Hungarian National Science Foundation (OTKA; grant No.: K 109843). HvG was supported by a grant from the Dutch Kidney Foundation (IP13-114). MF is supported by the UK Medical Research Council (G1001536) and the National Institute for Health Research through the NIHR Southampton Biomedical Research Centre. Artwork for the figures was adapted from Serviers Medical Art (<http://www.servier.com/Powerpoint-image-bank>).

Conflicts of interest

MF is a member of the Scientific Advisory Board of AOBiome LLC, a company commercializing the use of ammonia oxidizing bacteria for the treatment of inflammatory skin diseases and hypertension. None of the other authors has any conflict of interests to declare.

List of Abbreviations

arginosuccinate lyase (ASL); billion years ago (bya); cysteine- β -synthase (CBS), cysteine- γ -lyase (CSE); 3-mercaptosulfotransferase (MST); glutathione (GSH); last universal common ancestor (LUCA); nitric oxide (NO); nitric oxide synthases (NOS); reactive nitrogen species (RNS); reactive oxygen species (ROS); reactive species interactome (RSI); reactive sulfur species (RSS); sulfur to nitrogen ratio (S:N ratio); sulfur-containing amino acids (SAA)

References

1. Aragonés J, Fraisl P, Baes M, Carmeliet P. Oxygen sensors at the crossroad of metabolism. *Cell Metab.* 9: 11-22, 2009.
2. Bailey TS, Henthorn HA, Pluth MD. The intersection of NO and H₂S: persulfides generate NO from nitrite through polysulfide formation. *Inorg. Chem.* 55: 12618-12625, 2016.
3. Banne AF, Amiri A, Pero RW. Reduced level of serum thiols in patients with a diagnosis of active disease. *J. Anti Aging Med.* 6: 327-34, 2003.
4. Bao XR, Ong S-E, Goldberger O, Peng J, Sharma R, Thompson DA, Vafai SB, Cox AG, Marutani E, Ichinose F. Mitochondrial dysfunction remodels one-carbon metabolism in human cells. *Elife* 5: e10575, 2016.
5. Basudhar D, Ridnour LA, Cheng R, Kesarwala AH, Heinecke J, Wink DA. Biological signaling by small inorganic molecules. *Coord. Chem. Rev.* 306: 708-723, 2016.
6. Beger RD, Dunn W, Schmidt MA, Gross SS, Kirwan JA, Cascante M, Brennan L, Wishart DS, Oresic M, Hankemeier T. Metabolomics enables precision medicine: "A White Paper, Community Perspective". *Metabolomics* 12: 149, 2016.
7. Benavides GA, Squadrito GL, Mills RW, Patel HD, Isbell TS, Patel RP, Darley-Usmar VM, Doeller JE, Kraus DW. Hydrogen sulfide mediates the vasoactivity of garlic. *Proc. Natl. Acad. Sci. U. S. A.* 104: 17977-17982, 2007.
8. Benzie IF. Evolution of antioxidant defence mechanisms. *Eur. J. Nutr.* 39: 53-61, 2000.
9. Bernstein M. Prebiotic materials from on and off the early Earth. *Philos. Trans. R. Soc. Lond B Biol. Sci* 361: 1689-1700, 2006.
10. Bianco CL, Chavez TA, Sosa V, Saund SS, Nguyen QN, Tantillo DJ, Ichimura AS, Toscano JP, Fukuto JM. The chemical biology of the persulfide (RSSH)/perthiyl (RSS.) redox couple and possible role in biological redox signaling. *Free Radic. Biol. Med.* 101: 20-31, 2016.
11. Blackstone E, Morrison M, Roth MB. H₂S induces a suspended animation-like state in mice. *Science* 308: 518, 2005.
12. Blanco RA, Ziegler TR, Carlson BA, Cheng PY, Park Y, Cotsonis GA, Accardi CJ, Jones DP. Diurnal variation in glutathione and cysteine redox states in human plasma. *Am. J. Clin. Nutr.* 86: 1016-23, 2007.
13. Bode-Böger SM, Scalera F, Kielstein JT, Martens-Lobenhoffer J, Breithardt G, Fobker M, Reinecke H. Symmetrical dimethylarginine: a new combined parameter for renal function and extent of coronary artery disease. *J. Am. Soc. Nephrol.* 17: 1128-1134, 2006.
14. Borsook H, Dubnoff JW. The conversion of citrulline to arginine in the kidney. *J. Biol. Chem.* 141: 717-738, 1941.
15. Brandes N, Schmitt S, Jakob U. Thiol-based redox switches in eukaryotic proteins. *Antioxid Redox Signal* 11: 997-1014, 2009.
16. Brandes RP, Weissmann N, Schroder K. Nox family NADPH oxidases: Molecular mechanisms of activation. *Free Radic. Biol. Med.* 76: 208-26, 2014.
17. Broniowska KA, Hogg N. The chemical biology of S-nitrosothiols. *Antioxid Redox Signal* 17: 969-980, 2012.

18. Brosnan JT, Brosnan ME. The sulfur-containing amino acids: an overview. *J. Nutr.* 136: 1636S-1640S, 2006.
19. Bunker VW, Lawson MS, Stansfield MF, Clayton BE. Nitrogen balance studies in apparently healthy elderly people and those who are housebound. *Br. J. Nutr.* 57: 211-21, 1987.
20. Caetano-Anolles G, Yafremava LS, Gee H, Caetano-Anolles D, Kim HS, Mittenthal JE. The origin and evolution of modern metabolism. *Int. J. Biochem. Cell Biol.* 41: 285-97, 2009.
21. Cannio R, Fiorentino G, Morana A, Rossi M, Bartolucci S. Oxygen: friend or foe? Archaeal superoxide dismutases in the protection of intra- and extracellular oxidative stress. *Front. Biosci.* 5: D768-D779, 2000.
22. Capaccioni F, Coradini A, Filacchione G, Erard S, Arnold G, Drossart P, De Sanctis MC, Bockelee-Morvan D, Capria MT, Tosi F, Leyrat C, Schmitt B, Quirico E, Cerroni P, Mennella V, Raponi A, Ciarniello M, McCord T, Moroz L, Palomba E, Ammannito E, Barucci MA, Bellucci G, Benkhoff J, Bibring JP, Blanco A, Blecka M, Carlson R, Carsenty U, Colangeli L, Combes M, Combi M, Crovisier J, Encrenaz T, Federico C, Fink U, Fonti S, Ip WH, Irwin P, Jaumann R, Kuehrt E, Langevin Y, Magni G, Mottola S, Orofino V, Palumbo P, Piccioni G, Schade U, Taylor F, Tiphene D, Tozzi GP, Beck P, Biver N, Bonal L, Combe JP, Despan D, Flamini E, Fornasier S, Frigeri A, Grassi D, Gudipati M, Longobardo A, Markus K, Merlin F, Orosei R, Rinaldi G, Stephan K, Cartacci M, Cicchetti A, Giuppi S, Hello Y, Henry F, Jacquinod S, Noschese R, Peter G, Politi R, Reess JM, Semery A. Cometary science. The organic-rich surface of comet 67P/Churyumov-Gerasimenko as seen by VIRTIS/Rosetta. *Science* 347: aaa0628, 2015.
23. Carbonero F, Benefiel AC, Alizadeh-Ghamsari AH, Gaskins HR. Microbial pathways in colonic sulfur metabolism and links with health and disease. *Front. Physiol.* 3: 448, 2012.
24. Carey GP, Kime Z, Rogers QR, Morris JG, Hargrove D, Buffington CA, Brusilow SW. An arginine-deficient diet in humans does not evoke hyperammonemia or orotic aciduria. *J. Nutr.* 117: 1734-1739, 1987.
25. Carlstrom M, Liu M, Yang T, Zollbrecht C, Huang LY, Peleli M, Borniquel S, Kishikawa H, Hezel M, Persson AEG, Weitzberg E, Lundberg JO. Cross-talk between nitrate-nitrite-NO and NO synthase pathways in control of vascular NO homeostasis. *Antioxid Redox Signal* 23: 295-306, 2015.
26. Casas AI, Dao VT-V, Daiber A, Maghzal GJ, Di Lisa F, Kaludercic N, Leach S, Cuadrado A, Jaquet V, Seredenina T. Reactive oxygen-related diseases: therapeutic targets and emerging clinical indications. *Antioxid Redox Signal* 23: 1171-1185, 2015.
27. Castillo L, Beaumier L, Ajami AM, Young VR. Whole body nitric oxide synthesis in healthy men determined from [15N] arginine-to-[15N]citrulline labeling. *Proc. Natl. Acad. Sci. U. S. A.* 93: 11460-5, 1996.
28. Castillo L, Chapman TE, Sanchez M, Yu YM, Burke JF, Ajami AM, Vogt J, Young VR. Plasma arginine and citrulline kinetics in adults given adequate and arginine-free diets. *Proc. Natl. Acad. Sci. U. S. A.* 90: 7749-53, 1993.
29. Chacko BK, Kramer PA, Ravi S, Benavides GA, Mitchell T, Dranka BP, Ferrick D, Singal AK, Ballinger SW, Bailey SM, Hardy RW, Zhang J, Zhi D, Darley-Usmar VM. The Bioenergetic Health Index: a new concept in mitochondrial translational research. *Clin. Sci. (Lond.)* 127: 367-73, 2014.

47. DeLeon ER, Gao Y, Huang E, Arif M, Arora N, Divietro A, Patel S, Olson KR. A case of mistaken identity: are reactive oxygen species actually reactive sulfide species? *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 310: R549-60, 2016.
48. Dhanakoti SN, Brosnan JT, Herzberg GR, Brosnan ME. Renal arginine synthesis: studies in vitro and in vivo. *Am. J. Physiol.* 259: E437-42, 1990.
49. Di Buono M, Wykes LJ, Cole DEC, Ball RO, Pencharz PB. Regulation of sulfur amino acid metabolism in men in response to changes in sulfur amino acid intakes. *J. Nutr.* 133: 733-739, 2003.
50. Ducker GS, Rabinowitz JD. One-carbon metabolism in health and disease. *Cell Metab.*, 2016.
51. Elsasser TH, Kahl S, Capuco AV, Schmidt W. Effects of stress on endocrine and metabolic processes and redirection: cross talk between subcellular compartments. *Domest. Anim. Endocrinol.* 43: 132-45, 2012.
52. Epel ES, Blackburn EH, Lin J, Dhabhar FS, Adler NE, Morrow JD, Cawthon RM. Accelerated telomere shortening in response to life stress. *Proc. Natl. Acad. Sci. U. S. A.* 101: 17312-17315, 2004.
53. Espey MG. Role of oxygen gradients in shaping redox relationships between the human intestine and its microbiota. *Free Radic. Biol. Med.* 55: 130-40, 2013.
54. Espinosa-Diez C, Miguel V, Mennerich D, Kietzmann T, Sanchez-Perez P, Cadenas S, Lamas S. Antioxidant responses and cellular adjustments to oxidative stress. *Redox Biol* 6: 183-97, 2015.
55. Feelisch M, Fernandez BO, Bryan NS, Garcia-Saura MF, Bauer S, Whitlock DR, Ford PC, Janero DR, Rodriguez J, Ashrafiyan H. Tissue processing of nitrite in hypoxia an intricate interplay of nitric oxide-generating and-scavenging systems. *J. Biol. Chem.* 283: 33927-33934, 2008.
56. Feelisch M, Martin JF. The early role of nitric oxide in evolution. *Trends Ecol. Evol.* 10: 496-9, 1995.
57. Flohe L. The fairytale of the GSSG/GSH redox potential. *Biochim. Biophys. Acta* 1830: 3139-42, 2013.
58. Förstermann U, Sessa WC. Nitric oxide synthases: regulation and function. *Eur. Heart J.* 33: 829-837, 2012.
59. Foster MW, McMahon TJ, Stamler JS. S-nitrosylation in health and disease. *Trends Mol. Med.* 9: 160-8, 2003.
60. Francoleon NE, Carrington SJ, Fukuto JM. The reaction of H(2)S with oxidized thiols: generation of persulfides and implications to H(2)S biology. *Arch. Biochem. Biophys.* 516: 146-53, 2011.
61. Frenay AS, de Borst MH, Bachtler M, Tschopp N, Keyzer CA, van den Berg E, Bakker SJ, Feelisch M, Pasch A, van Goor H. Serum free sulfhydryl status is associated with patient and graft survival in renal transplant recipients. *Free Radic. Biol. Med.* 99: 345-351, 2016.
62. Frigaard NU, Dahl C. Sulfur metabolism in phototrophic sulfur bacteria. *Adv. Microb. Physiol* 54: 103-200, 2009.
63. Fu M, Zhang W, Wu L, Yang G, Li H, Wang R. Hydrogen sulfide (H₂S) metabolism in mitochondria and its regulatory role in energy production. *Proc. Natl. Acad. Sci. U. S. A.* 109: 2943-8, 2012.

82. Hoeijmakers JHJ. DNA damage, aging, and cancer. *N. Engl. J. Med.* 361: 1475-1485, 2009.
83. Holmes FL. Claude Bernard, the "Milieu Intérieur", and regulatory physiology. *History and philosophy of the life sciences*: 3-25, 1986.
84. Hood L, Friend SH. Predictive, personalized, preventive, participatory (P4) cancer medicine. *Nat. Rev. Clin. Oncol.* 8: 184-187, 2011.
85. Horowitz JH, Rypins EB, Henderson JM, Heymsfield SB, Moffitt SD, Bain RP, Chawla RK, Bleier JC, Rudman D. Evidence for impairment of transsulfuration pathway in cirrhosis. *Gastroenterology* 81: 668-75, 1981.
86. Ida T, Sawa T, Ihara H, Tsuchiya Y, Watanabe Y, Kumagai Y, Suematsu M, Motohashi H, Fujii S, Matsunaga T, Yamamoto M, Ono K, Devarie-Baez NO, Xian M, Fukuto JM, Akaike T. Reactive cysteine persulfides and S-polythiolation regulate oxidative stress and redox signaling. *Proc. Natl. Acad. Sci. U. S. A.* 111: 7606-11, 2014.
87. Ingenbleek Y. The nutritional relationship linking sulfur to nitrogen in living organisms. *J. Nutr.* 136: 1641S-1651S, 2006.
88. Ingenbleek Y, Hardillier E, Jung L. Subclinical protein malnutrition is a determinant of hyperhomocysteinemia. *Nutrition* 18: 40-6, 2002.
89. Ingenbleek Y, Young VR. The essentiality of sulfur is closely related to nitrogen metabolism: a clue to hyperhomocysteinemia. *Nutr. Res. Rev.* 17: 135-151, 2004.
90. Jansson EA, Huang L, Malkey R, Govoni M, Nihlen C, Olsson A, Stensdotter M, Petersson J, Holm L, Weitzberg E, Lundberg JO. A mammalian functional nitrate reductase that regulates nitrite and nitric oxide homeostasis. *Nat. Chem. Biol.* 4: 411-417, 2008.
91. Jones DP. Redefining oxidative stress. *Antioxid Redox Signal* 8: 1865-79, 2006.
92. Jones DP, Mody VC, Jr., Carlson JL, Lynn MJ, Sternberg P, Jr. Redox analysis of human plasma allows separation of pro-oxidant events of aging from decline in antioxidant defenses. *Free Radic. Biol. Med.* 33: 1290-300, 2002.
93. Jones DP, Park Y, Gletsu-Miller N, Liang Y, Yu T, Accardi CJ, Ziegler TR. Dietary sulfur amino acid effects on fasting plasma cysteine/cystine redox potential in humans. *Nutrition* 27: 199-205, 2011.
94. Jones DP, Sies H. The redox code. *Antiox Redox Signaling* 23: 734-746, 2015.
95. Joseph J, Handy DE, Loscalzo J. Quo vadis: whither homocysteine research? *Cardiovasc. Toxicol.* 9: 53-63, 2009.
96. Jourdain D, Jourdain FL, Kutchukian PS, Musah RA, Wink DA, Grisham MB. Reaction of Superoxide and Nitric Oxide with Peroxynitrite implications for peroxynitrite-mediated oxidation reactions in vivo *J. Biol. Chem.* 276: 28799-28805, 2001.
97. Jourdan M, Glock C, Margen S, Bradfield RB. Sulfate, acid-base, and mineral balances of obese women during weight-loss. *Am. J. Clin. Nutr.* 33: 236-243, 1980.
98. Kalapos MP. A theoretical approach to the link between oxidoreductions and pyrite formation in the early stage of evolution. *Biochim. Biophys. Acta* 1553: 218-222, 2002.
99. Kamysny A, Jr., Gun J, Rizkov D, Voitsekovski T, Lev O. Equilibrium distribution of polysulfide ions in aqueous solutions at different temperatures by rapid single phase derivatization. *Environ. Sci. Technol.* 41: 2395-2400, 2007.

100. [Kemp M, Go YM, Jones DP. Nonequilibrium thermodynamics of thiol/disulfide redox systems: a perspective on redox systems biology. *Free Radic. Biol. Med.* 44: 921-37, 2008.](#)
101. [Kimura H. Signaling of hydrogen sulfide and polysulfides. *Antioxidants and Redox Signaling* 22: 347-349, 2014.](#)
102. [Kimura Y, Toyofuku Y, Koike S, Shibuya N, Nagahara N, Lefer D, Ogasawara Y, Kimura H. Identification of H₂S₃ and H₂S produced by 3-mercaptopyruvate sulfurtransferase in the brain. *Sci. Rep.* 5: 14774, 2015.](#)
103. [Knoops B, Loumaye E, Van DE, V. Evolution of the peroxiredoxins. *Subcell. Biochem* 44: 27-40, 2007.](#)
104. [Kolluru GK, Shen X, Bir SC, Kevil CG. Hydrogen sulfide chemical biology: Pathophysiological roles and detection. *Nitric Oxide* 35: 5-20, 2013.](#)
105. [Koning AM, Meijers WC, Pasch A, Leuvenink HG, Frenay AR, Dekker MM, Feelisch M, de Boer RA, van Goor H. Serum free thiols in chronic heart failure. *Pharmacol. Res.* 111: 452-8, 2016.](#)
106. [Kotte O, Zaugg JB, Heinemann M. Bacterial adaptation through distributed sensing of metabolic fluxes. *Mol. Syst. Biol.* 6: 355, 2010.](#)
107. [Kultz D. Molecular and evolutionary basis of the cellular stress response. *Annu. Rev. Physiol.* 67: 225-57, 2005.](#)
108. [Kundi H, Ates I, Kiziltunc E, Cetin M, Cicekcioglu H, Neselioglu S, Erel O, Ornek E. A novel oxidative stress marker in acute myocardial infarction; thiol/disulphide homeostasis. *Am. J. Emerg. Med.* 33: 1567-1571, 2015.](#)
109. [Kurland CG, Andersson SG. Origin and evolution of the mitochondrial proteome. *Microbiol. Mol. Biol. Rev* 64: 786-820, 2000.](#)
110. [Lalwani P, de Souza GKBB, de Lima DSN, Passos LFS, Boechat AL, Lima ES. Serum thiols as a biomarker of disease activity in lupus nephritis. *PLoS One* 10: e0119947, 2015.](#)
111. [Lane N. *Power, sex, suicide: mitochondria and the meaning of life*: Oxford University Press; 2006.](#)
112. [Lau T, Owen W, Yu YM, Noviski N, Lyons J, Zurakowski D, Tsay R, Ajami A, Young VR, Castillo L. Arginine, citrulline, and nitric oxide metabolism in end-stage renal disease patients. *J. Clin. Invest.* 105: 1217-25, 2000.](#)
113. [Leiper J, Nandi M. The therapeutic potential of targeting endogenous inhibitors of nitric oxide synthesis. *Nat. Rev. Drug Discov.* 10: 277-291, 2011.](#)
114. [Lim SS, Vos T, Flaxman AD, Danaei G, Shibuya K, Adair-Rohani H, Amann M, Anderson HR, Andrews KG, Aryee M, Atkinson C, Bacchus LJ, Bahalim AN, Balakrishnan K, Balmes J, Barker-Collo S, Baxter A, Bell ML, Blore JD, Blyth F, Bonner C, Borges G, Bourne R, Boussinesq M, Brauer M, Brooks P, Bruce NG, Brunekreef B, Bryan-Hancock C, Bucello C, Buchbinder R, Bull F, Burnett RT, Byers TE, Calabria B, Carapetis J, Carnahan E, Chafe Z, Charlson F, Chen H, Chen JS, Cheng AT, Child JC, Cohen A, Colson KE, Cowie BC, Darby S, Darling S, Davis A, Degenhardt L, Dentener F, Des Jarlais DC, Devries K, Dherani M, Ding EL, Dorsey ER, Driscoll T, Edmond K, Ali SE, Engell RE, Erwin PJ, Fahimi S, Falder G, Farzadfar F, Ferrari A, Finucane MM, Flaxman S, Fowkes FG, Freedman G, Freeman MK, Gakidou E, Ghosh S, Giovannucci E, Gmel G, Graham K, Grainger R, Grant B, Gunnell D, Gutierrez HR, Hall W, Hoek HW, Hogan A, Hosgood HD, 3rd, Hoy D, Hu H, Hubbell BJ, Hutchings SJ, Ibeanusi SE, Jacklyn GL, Jasrasaria R, Jonas JB, Kan H, Kanis JA, Kassebaum N, Kawakami N, Khang YH, Khatibzadeh S,](#)

- Khoo JP, Kok C, Laden F, Laloo R, Lan Q, Lathlean T, Leasher JL, Leigh J, Li Y, Lin JK, Lipshultz SE, London S, Lozano R, Lu Y, Mak J, Malekzadeh R, Mallinger L, Marcenos W, March L, Marks R, Martin R, McGale P, McGrath J, Mehta S, Mensah GA, Merriman TR, Micha R, Michaud C, Mishra V, Mohd Hanafiah K, Mokdad AA, Morawska L, Mozaffarian D, Murphy T, Naghavi M, Neal B, Nelson PK, Nolla JM, Norman R, Olives C, Omer SB, Orchard J, Osborne R, Ostro B, Page A, Pandey KD, Parry CD, Passmore E, Patra J, Pearce N, Pelizzari PM, Petzold M, Phillips MR, Pope D, Pope CA, 3rd, Powles J, Rao M, Razavi H, Rehfuss EA, Rehm JT, Ritz B, Rivara FP, Roberts T, Robinson C, Rodriguez-Portales JA, Romieu I, Room R, Rosenfeld LC, Roy A, Rushton L, Salomon JA, Sampson U, Sanchez-Riera L, Sanman E, Sapkota A, Seedat S, Shi P, Shield K, Shivakoti R, Singh GM, Sleet DA, Smith E, Smith KR, Stapelberg NJ, Steenland K, Stockl H, Stovner LJ, Straif K, Straney L, Thurston GD, Tran JH, Van Dingenen R, van Donkelaar A, Veerman JL, Vijayakumar L, Weintraub R, Weissman MM, White RA, Whiteford H, Wiersma ST, Wilkinson JD, Williams HC, Williams W, Wilson N, Woolf AD, Yip P, Zielinski JM, Lopez AD, Murray CJ, Ezzati M, AlMazroa MA, Memish ZA. A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 380: 2224-60, 2012.
115. [Liu X, Miller MJ, Joshi MS, Thomas DD, Lancaster JR. Accelerated reaction of nitric oxide with O₂ within the hydrophobic interior of biological membranes. *Proc. Nat. Acad. Sci. USA* 95: 2175-2179, 1998.](#)
 116. [Lundberg JO, Gladwin MT, Ahluwalia A, Benjamin N, Bryan NS, Butler A, Cabrales P, Fago A, Feelisch M, Ford PC, Freeman BA, Frenneaux M, Friedman J, Kelm M, Kevil CG, Kim-Shapiro DB, Kozlov AV, Lancaster JR, Jr., Lefer DJ, McColl K, McCurry K, Patel RP, Petersson J, Rassaf T, Reutov VP, Richter-Addo GB, Schechter A, Shiva S, Tsuchiya K, van Faassen EE, Webb AJ, Zuckerbraun BS, Zweier JL, Weitzberg E. Nitrate and nitrite in biology, nutrition and therapeutics. *Nat. Chem. Biol.* 5: 865-869, 2009.](#)
 117. [Lundberg JO, Gladwin MT, Weitzberg E. Strategies to increase nitric oxide signalling in cardiovascular disease. *Nat. Rev. Drug Discov.* 14: 623-41, 2015.](#)
 118. [Magee EA, Curno R, Edmond LM, Cummings JH. Contribution of dietary protein and inorganic sulfur to urinary sulfate: toward a biomarker of inorganic sulfur intake. *Am. J. Clin. Nutr.* 80: 137-142, 2004.](#)
 119. [Magee EA, Richardson CJ, Hughes R, Cummings JH. Contribution of dietary protein to sulfide production in the large intestine: an in vitro and a controlled feeding study in humans. *Am. J. Clin. Nutr.* 72: 1488-94, 2000.](#)
 120. [Marcolongo JP, Morzan UN, Zeida A, Scherlis DA, Olabe JA. Nitrosodisulfide \[S₂NO\]-\(perthionitrite\) is a true intermediate during the "cross-talk" of nitrosyl and sulfide. *PCCP* 18: 30047-30052, 2016.](#)
 121. [Mattson MP. Hormesis and disease resistance: activation of cellular stress response pathways. *Hum. Exp. Toxicol.* 27: 155-62, 2008.](#)
 122. [McGlynn SE, Kanik I, Russell MJ. Peptide and RNA contributions to iron-sulphur chemical gardens as life's first inorganic compartments, catalysts, capacitors and condensers. *Philos. Trans. A Math. Phys. Eng Sci* 370: 3007-3022, 2012.](#)
 123. [Milevoj Kopčinović L, Domijan A-M, Posavac K, Čepelak I, Žanić Grubišić T, Rumora L. Systemic redox imbalance in stable chronic obstructive pulmonary disease. *Biomarkers*: 1-7, 2016.](#)
 124. [Miller AF. Superoxide dismutases: ancient enzymes and new insights. *FEBS Lett.* 586: 585-595, 2012.](#)

- biomarker of oxidative stress is associated with risk of death in patients with coronary artery disease. *Circulation* 133: 361-9, 2016.
144. Paul BD, Snyder SH. Modes of physiologic H₂S signaling in the brain and peripheral tissues. *Antioxid Redox Signal*, 2014.
 145. Paul BD, Snyder SH. H₂S: a novel gasotransmitter that signals by sulfhydration. *Trends Biochem. Sci* 40: 687-700, 2015.
 146. Pero RW, Hoppe C, Sheng Y. Serum thiols as a surrogate estimate of DNA repair correlates to mammalian life span. *J. Anti Aging Med.* 3: 241-249, 2000.
 147. Pizzarello S. Molecular asymmetry in prebiotic chemistry: an account from meteorites. *Life (Basel)* 6, 2016.
 148. Pratt AJ. Prebiological evolution and the metabolic origins of life. *Artif. Life* 17: 203-217, 2011.
 149. Pross A. What is Life?: How chemistry becomes biology: Oxford University Press; 2016.
 150. Quirós PM, Mottis A, Auwerx J. Mitonuclear communication in homeostasis and stress. *Nat. Reviews Mol. Cell Biol.* 17: 213-226, 2016.
 151. Raguso CA, Ajami AM, Gleason R, Young VR. Effect of cystine intake on methionine kinetics and oxidation determined with oral tracers of methionine and cysteine in healthy adults. *Am. J. Clin. Nutr.* 66: 283-292, 1997.
 152. Raguso CA, Regan MM, Young VR. Cysteine kinetics and oxidation at different intakes of methionine and cystine in young adults. *Am. J. Clin. Nutr.* 71: 491-499, 2000.
 153. Raiswell R, Canfield DE. The iron biogeochemical cycle past and present. *Geochem Perspectives* 1: 1-220, 2012.
 154. Raymond J, Blankenship RE. The origin of the oxygen-evolving complex. *Coord. Chem. Rev* 252: 377-383, 2008.
 155. Richardson AR, Somerville GA, Sonenshein AL. Regulating the intersection of metabolism and pathogenesis in gram-positive bacteria. *Microbiol Spectr* 3, 2015.
 156. Rudolph V, Freeman BA. Cardiovascular consequences when nitric oxide and lipid signaling converge. *Circ. Res.* 105: 511-522, 2009.
 157. Sabry ZI, Shadarevian SB, Cowan JW, Campbell JA. Relationship of dietary intake of sulphur amino-acids to urinary excretion of inorganic sulphate in man. *Nature* 206: 931-3, 1965.
 158. Sagan C, Khare BN. Long-wavelength ultraviolet photoproduction of amino acids on the primitive Earth. *Science* 173: 417-420, 1971.
 159. Saks VA. *Molecular system bioenergetics: energy for life*: John Wiley & Sons; 2007.
 160. Salminen A, Kaarniranta K. AMP-activated protein kinase (AMPK) controls the aging process via an integrated signaling network. *Ageing Res Rev* 11: 230-41, 2012.
 161. Sandalio LM, Romero-Puertas MC. Peroxisomes sense and respond to environmental cues by regulating ROS and RNS signalling networks. *Ann. Bot.* 116: 475-485, 2015.
 162. Sartori P, Granger L, Lee CF, Horowitz JM. Thermodynamic costs of information processing in sensory adaptation. *PLoS Comput. Biol.* 10: e1003974, 2014.

182. [Stipanuk MH. Sulfur amino acid metabolism: pathways for production and removal of homocysteine and cysteine. *Annu. Rev. Nutr.* 24: 539-77, 2004.](#)
183. [Storch KJ, Wagner DA, Burke JF, Young VR. \[1-13C; methyl-2H3\]methionine kinetics in humans: methionine conservation and cystine sparing. *Am. J. Physiol.* 258: E790-8, 1990.](#)
184. [Stubbe J, van der Donk WA. Protein radicals in enzyme catalysis. *Chem. Rev.* 98: 705-762, 1998.](#)
185. [Stuehr DJ. Enzymes of the L-arginine to nitric oxide pathway. *J. Nutr.* 134: 2748S-2751S- discussion 2765S-2767S, 2004.](#)
186. [Suhm T, Ott M. Mitochondrial translation and cellular stress response. *Cell Tissue Res.*, 2016.](#)
187. [Szabo C. Hydrogen sulphide and its therapeutic potential. *Nat. Rev. Drug Discov.* 6: 917-935, 2007.](#)
188. [Szabo C, Ransy C, Módis K, Andriamihaja M, Murghes B, Coletta C, Olah G, Yanagi K, Bouillaud F. Regulation of mitochondrial bioenergetic function by hydrogen sulfide. Part I. Biochemical and physiological mechanisms. *Br. J. Pharmacol.* 171: 2099-2122, 2014.](#)
189. [Tapiero H, Mathe G, Couvreur P, Tew KD. Dossier: Free amino acids in human health and pathologies - I. Arginine. *Biomed. Pharmacother.* 56: 439-445, 2002.](#)
190. [Thomas DD, Ridnour LA, Isenberg JS, Flores-Santana W, Switzer CH, Donzelli S, Hussain P, Vecoli C, Paolocci N, Ambs S, Colton CA, Harris CC, Roberts DD, Wink DA. The chemical biology of nitric oxide: implications in cellular signaling. *Free Radic. Biol. Med.* 45: 18-31, 2008.](#)
191. [Tomasova L, Dobrowolski L, Jurkowska H, Wrobel M, Huc T, Ondrias K, Ostaszewski R, Ufnal M. Intracolonic hydrogen sulfide lowers blood pressure in rats. *Nitric Oxide* 60: 50-58, 2016.](#)
192. [Totzeck M, Hendgen-Cotta UB, Luedike P, Berenbrink M, Klare JP, Steinhoff HJ, Semmler D, Shiva S, Williams D, Kipar A, Gladwin MT, Schrader J, Kelm M, Cossins AR, Rassaf T. Nitrite regulates hypoxic vasodilation via myoglobin-dependent nitric oxide generation. *Circulation* 126: 325-334, 2012.](#)
193. [Trachootham D, Lu W, Ogasawara MA, Nilsa RD, Huang P. Redox regulation of cell survival. *Antioxid Redox Signal* 10: 1343-74, 2008.](#)
194. [Trusheim MR, Burgess B, Hu SX, Long T, Averbuch SD, Flynn AA, Lieftucht A, Mazumder A, Milloy J, Shaw PM, Swank D, Wang J, Berndt ER, Goodsaid F, Palmer MC. Quantifying factors for the success of stratified medicine. *Nat. Rev. Drug Discov.* 10: 817-833, 2011.](#)
195. [Turell L, Radi R, Alvarez B. The thiol pool in human plasma: The central contribution of albumin to redox processes. *Free Radical Biol. Med.* 65: 244-253, 2013.](#)
196. [Ueland PM. Homocysteine species as components of plasma redox thiol status. *Clin. Chem.* 41: 340-342, 1995.](#)
197. [Ueland PM, Loscalzo J. Homocysteine and cardiovascular risk: the perils of reductionism in a complex system. *Clin. Chem.* 58: 1623-5, 2012.](#)
198. [Urey HC. On the early chemical history of the Earth and the origin of Life. *Proc. Natl. Acad. Sci. U. S. A.* 38: 351-63, 1952.](#)

217. Wootton S, Jackson A. Influence of under-nutrition in early life on growth, body composition and metabolic competence. In: *Long-term consequences of early environment; growth, development and the lifespan developmental perspective (Society for the Study of Human Biology Symposium Series; 37)*. 1996. pp. 109-123.
218. Wu G, Morris SM, Jr. Arginine metabolism: nitric oxide and beyond. *Biochem. J* 336 (Pt 1): 1-17, 1998.
219. Yamasaki H, Cohen MF. Biological consilience of hydrogen sulfide and nitric oxide in plants: Gases of primordial earth linking plant, microbial and animal physiologies. *Nitric Oxide* 55-56: 91-100, 2016.
220. Yazici C, Köse K, Utaş S, Tanrikulu E, Taşlıdere N. A novel approach in psoriasis: first usage of known protein oxidation markers to prove oxidative stress. *Arch. Dermatol. Res.* 308: 207-212, 2016.
221. Zamocky M, Gasselhuber B, Furtmuller PG, Obinger C. Molecular evolution of hydrogen peroxide degrading enzymes. *Arch. Biochem. Biophys* 525: 131-144, 2012.
222. Zhang Y, Du Y, Le W, Wang K, Kieffer N, Zhang J. Redox control of the survival of healthy and diseased cells. *Antioxid Redox Signal* 15: 2867-908, 2011.

Table captions

Tab. 1 **Definition of terms**. Scientific terms used in the text and marked in *italics*.

Tab. 2 **Chemical Attributes of ROS, RNS and RSS** (interactions with metals and metalloproteins not included in this overview)

Figure legends

Antioxidants & Redox Signaling
 The Reactive Species Interactome
 Evolutionary Emergence, Biological Significance, and Opportunities for Redox Metabolomics and Personalized Medicine
 (doi: 10.1089/ars.2017.7083)

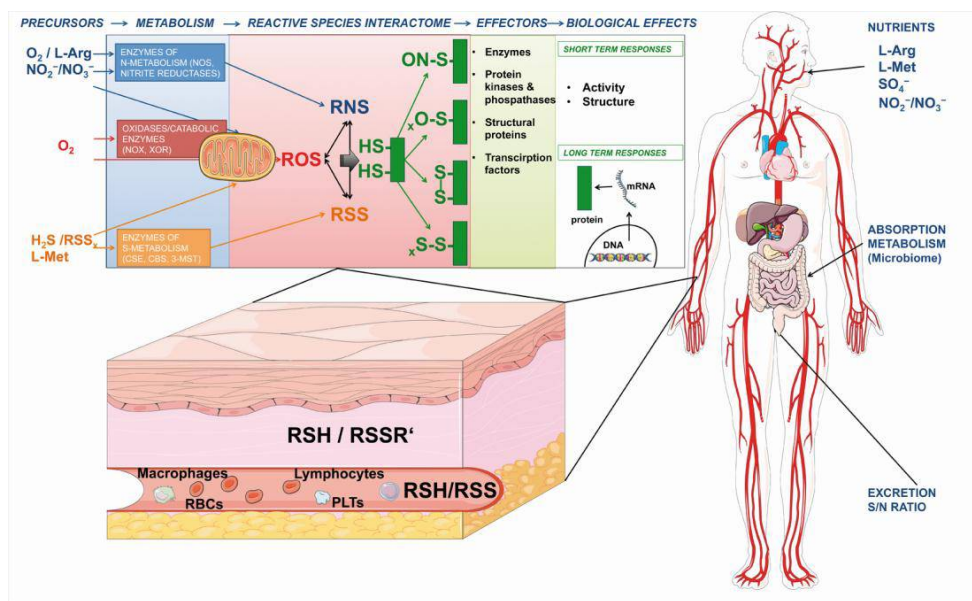


Fig. 1 – Intracellular, extracellular and inter-organ/systemic role of the RSI. Precursors of the RSI are organic and inorganic substrates and cofactors including amino acids (e.g., arginine, methionine), vitamins (B6, B12, C), xanthine as well as oxygen, nitrite, polysulfides, thiosulfate and sulfate, which are transformed by mitochondrial or cytoplasmic enzymes into reactive oxygen species (ROS), reactive nitrogen species (RNS) and reactive sulfur species (RSS). The chemical interactions among ROS, RNS and RSS lead to formation of a number of products with different reactivities, stabilities, half-lives, and therefore different lifetimes defined by their physicochemical properties, covering a wide range of maximal travel distances. A common target of the RSI are cysteine thiols in proteins, acting as redox switches, able to fine-tune activity of signaling molecules and leading to short-term responses (e.g., protein kinases and phosphatases inducing changes in signaling and glucose metabolism) or long-term adaptation (by modifying redox switches responsible for gene expression regulation, like the HIF, NFkB, and Keap-1/Nrf2 pathways). The RSI serves also as a local and systemic heterocellular communication system mediated by actions of longer-lasting products of the RSI (e.g. nitrite, polysulfides) and circulating thiols. The nutritional and physiological status of the organism affects the RSI by reciprocally regulating precursor availability, metabolism, signaling and mitochondrial function. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars)

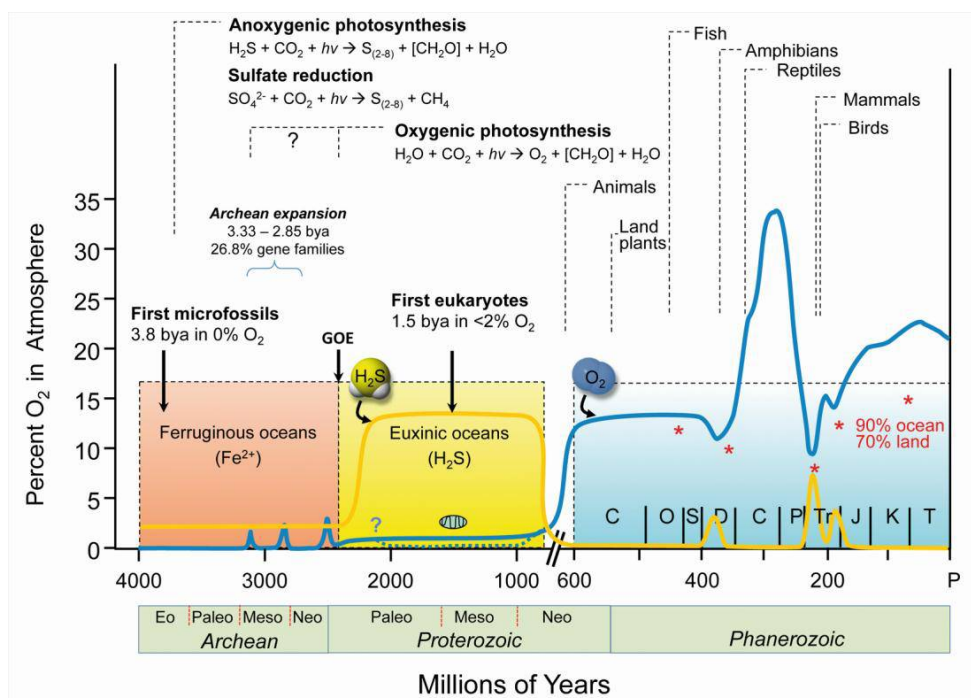


Fig. 2 – Evolution of sulfur and oxygen metabolism. The lines indicate fluctuations in concentration of atmospheric oxygen (blue) and oceanic sulfide (orange) over evolutionary times. Atmospheric O_2 was essentially absent from the environment at the onset of life ca. 3.8 billion years ago (bya). After the great oxidation event (GOE) the concentration of O_2 in the atmosphere increased, which was accompanied by a substantial increase in H_2S . The first eukaryotes appeared in oceans and developed in anoxic and sulfidic (euxinic) conditions for hundreds of millions of years using sulfur as their energy source, producing RSS. During this time, defense mechanisms against RSS evolved improving cell survival and minimizing the need for repair of damaged cell constituents. Appearance of oxygenic cyanobacteria and plants lead to increases in O_2 levels and oxidation of H_2S and Fe^{2+} approximately 0.6 bya. Those changes were accompanied by a significant decrease in dissolved H_2S and a repurposing of enzymatic systems that originally evolved to protect organisms against RSS to serve additional antioxidative protective functions. Mass extinctions (*, percentage of marine and land life) were often associated with a fall in ambient O_2 and increases in H_2S , perhaps providing a biological filter for descendants that retained some degree of tolerance to hypoxia and sulfide. Modified with permission from Olson & Straub (139). (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars)

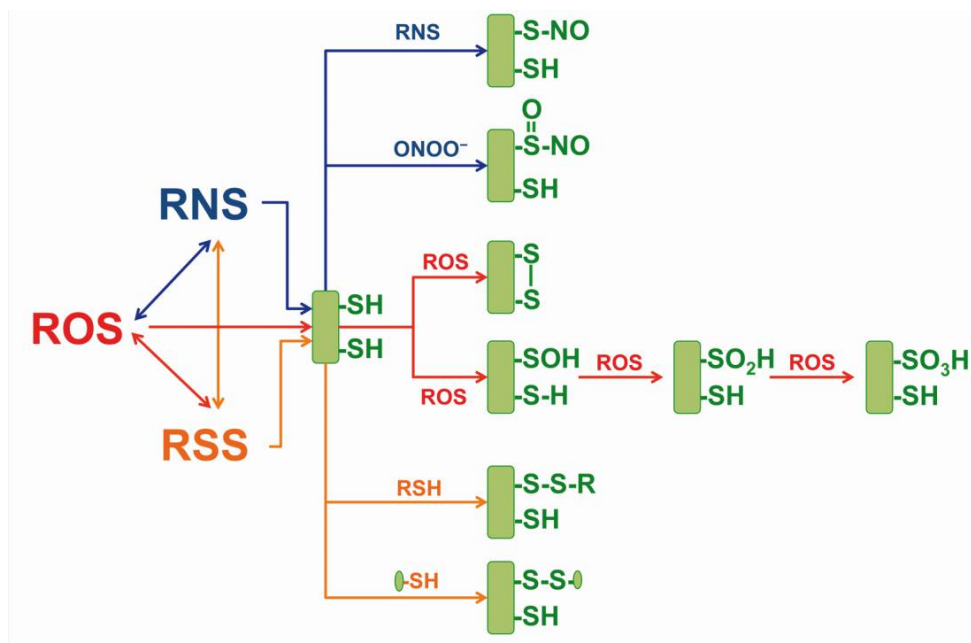


Fig. 3 – Cysteine modifications induced by the interaction with reactive oxygen, nitrogen and sulfur species. The reactive species interactome consists of the interaction of reactive species (ROS, RNS, RSS) with one another and with cysteine thiols as redox switches (reactions with other functional groups omitted here for the sake of simplicity). The outcome of these interactions depends on the chemical characteristics of the species inducing the modification (e.g. $O_2^{\square-}$, H_2O_2 , NO, $ONOO^-$) and their fluxes, the environmental conditions (e.g. pO_2 , pH) as well as on the reactivity and localization of the targeted cysteines. The lines indicate the outcome of protein cysteine modifications induced by RNS (blue), ROS (red) or RSS (orange). (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars)

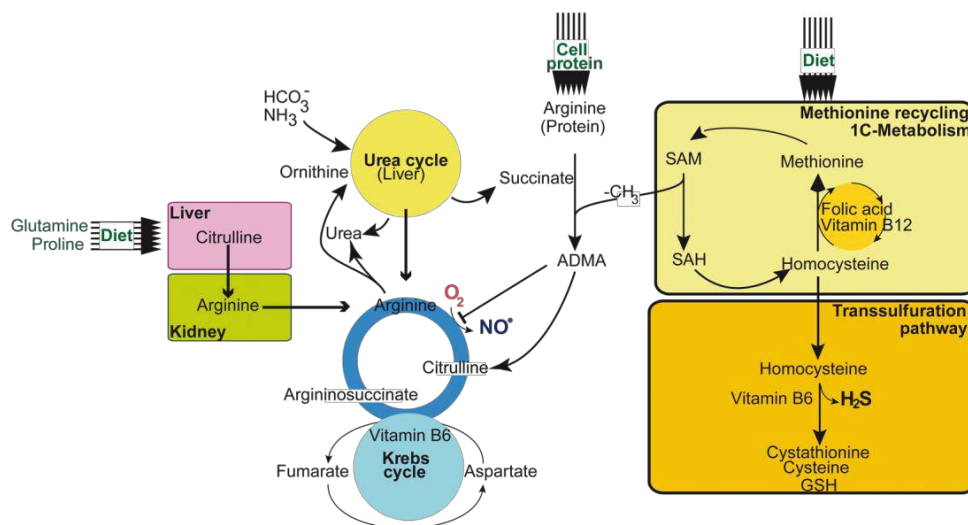


Fig. 4 – Metabolic pathways fuelling the RSI. In mammals, L-arginine is formed from citrulline, derived either from dietary glutamine or proline via ornithine and carbamoylphosphate in the mitochondria, or from bicarbonate (HCO_3^-) and ammonia (NH_3) via the hepatic urea cycle. Citrulline is then transported via the blood to the kidney where it is converted into arginine. Arginine used for protein formation and (O_2 -dependent) NO synthesis can be recycled via the arginine/citrulline cycle. NO synthase activity is inhibited by different methylated arginine residues released by proteolysis (e.g. ADMA, asymmetric dimethylarginine). A key interaction between nitrogen and sulfur metabolism is the methylation of arginine using S-adenosyl-methionine (SAM)-dependent methyltransferases. SAM is a cofactor produced from methionine and used for the methylation of a large number of biomolecules; in the methionine recycling pathway, the removal of one methyl group ($-\text{CH}_3$), resulting in the formation of homocysteine. Depending on the availability of methionine, homocysteine is either recycled to methionine with the help of vitamin B12 and folic acid, or is degraded to cystathionine and cysteine. While the latter also serves as precursor of cellular glutathione production both compounds can generate H_2S in the transsulfuration pathway. Not shown here is the formation of ROS via NADPH oxidases, the mitochondrial respiratory chain and other sources. See Supplementary Fig. 1 for more details. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars)

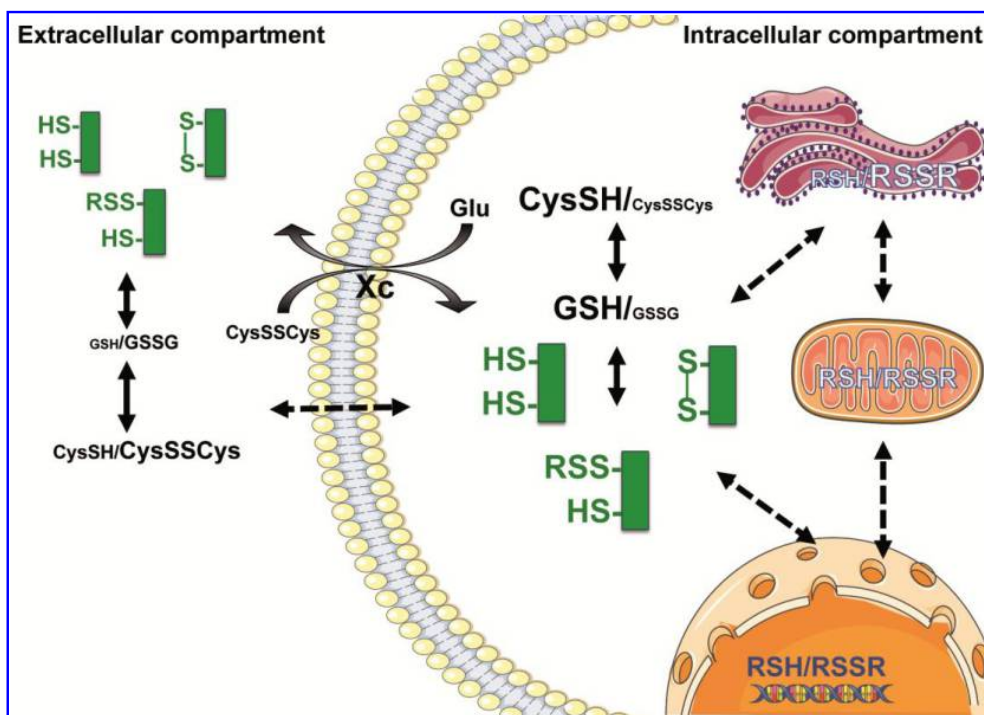


Fig. 5 – Thiol transport between cell organelles and exchange between the intra- and extracellular compartment. In human plasma, amino thiols such as cysteine, homocysteine and glutathione exist in free (reduced and oxidized) and protein-bound form, but little is known about the dynamics of their regulation and relationship with each other. As documented for cysteine and glutathione, amino thiols are transported across cell membranes and exchanged between cell organelles, with specific transporters such as the cysteine (CysSSCys)/glutamate (Glu) antiporter (Xc), which plays an important role in the regulation of cell surface redox. Continuous arrows indicate known relationships, interrupted arrows represent unknown relationships. In both the intracellular and extracellular compartment protein thiols represent the main pool of sulfhydryl (-SH) groups. (Note that different font sizes in the figure denote relative concentrations and that mixed disulfides of low-molecular-weight thiols and post-translational thiol modifications are omitted here for the sake of simplicity.) The cytosol is considerably more reduced compared to the extracellular space or the endoplasmic reticulum (where proper protein folding requires more oxidizing conditions). The redox couples cysteine/cystine, GSH/GSSG and protein bound thiols are not in equilibrium with each other, which suggests the involvement of specific enzyme systems that determine the steady-state levels of these species. Maintaining dysequilibria requires energy, and energy tends to be allocated according to criteria that confer robustness of organisms along the evolutionary selection process. First steps into the direction of decoding what determines ATP utilization hierarchies at a systems level are being taken, but the mechanisms of regulation of systemic thiol/disulfide status remain largely obscure. Considering the inverse association of free thiols with risk of death, a further assessment of these relationships and their significance for the cellular stress responses, DNA repair processes and other hard clinical endpoints seems to be justified. However, no number of observational studies will ever be able to establish causality; this will require prospective and interventional studies. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars)

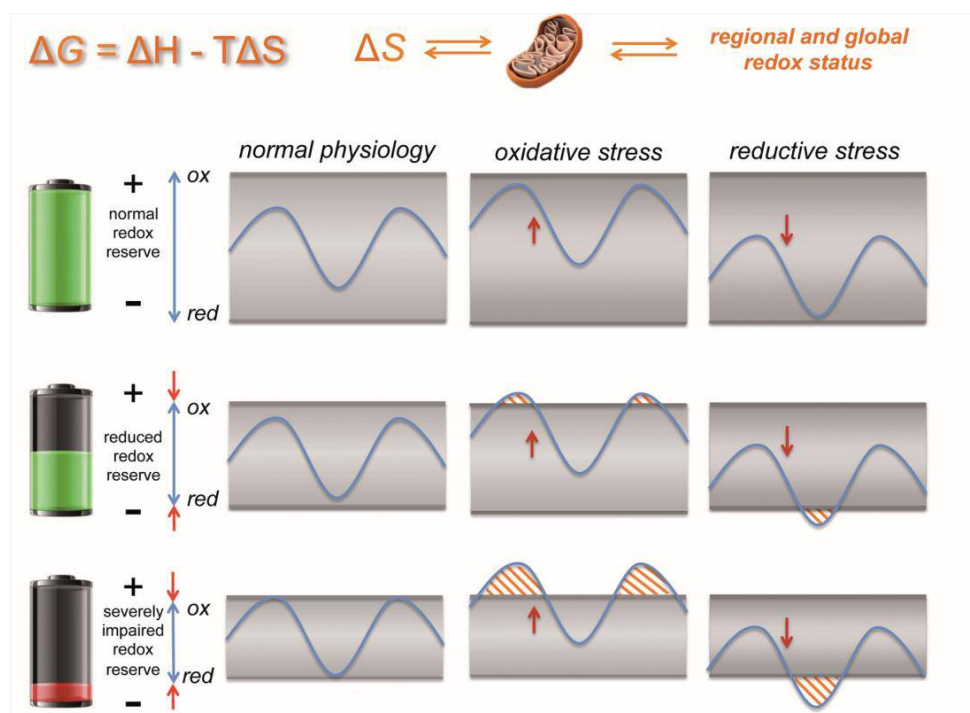


Fig. 6 – Simplified scheme visualizing the consequences of a reduced redox buffering capacity (“redox reserve”) due to bioenergetic limitations. Under normal physiological conditions, extracellular redox poise is subject to diurnal variations, fluctuating between more reducing (-) and more oxidizing (+) conditions (12). The capacity of a cell/organism to deal with changes in reductive and oxidative load is intimately linked to mitochondrial function. Enhanced mitochondrial activity is associated with higher oxidative stress, which affects the redox status of the local tissue microenvironment, while the activity of the mitochondrial respiratory chain itself and mitochondrial intermediary metabolism are modulated by the local and global redox status (upper cartoon). An adequate bioenergetic reserve capacity allows redox stresses (oxidative stress induced by e.g., strenuous physical activity or reductive stress due to chronic overfeeding) into either direction to be comfortably accommodated without incurring damage to cellular constituents (upper panels); physiological redox fluctuations experienced in daily life situations are well within the normal buffering capacity. Mitochondrial dysfunction leads to impaired cellular bioenergetics, resulting in a narrowing of a cell’s or an organism’s ability to buffer redox stresses, inflicting damage and/or compromising adaptive capacities (middle panels), which can result in severe impairment of redox regulatory events upon further bioenergetic challenge (lower panels). Under these conditions, significant damage may be inflicted (shaded areas), demanding the allocation of additional energy to cellular repair processes. According to these relationships, chronic stress triggers a vicious cycle that can lead to a condition associated with severely limited redox reserve capacity, compromising cellular surveillance and repair mechanisms and inviting cascading network failures. This notion is consistent with the links of AMP-activated protein kinase and aberrant mitochondrial gene expression with cellular stress, aging/degenerative processes and immune processes (186), molecular systems energetics (159,160) in general, and the emerging concept of *bioenergetic health*, where redox biology controls the interface between bioenergetics, autophagy and circadian control of metabolism (209)(29). See also Box 6. (To see this illustration in color the reader is referred to the web version of this article at www.liebertonline.com/ars)

Tab. 1		
Term	Definition	References
Stress and Adaptation		
Stress / adaptation to stress	the term stress, as used in the biomedical literature, was coined by Hans Selye in the 1930s, who defined it as "the non-specific response of the body to any demand for change".	(166)
Cellular stress response	a set of cellular responses (including the down-regulation of protein synthesis and up-regulation of transcription factors involved in defense and repair mechanisms) common to all eukaryotes aimed at coping with various forms of stress; also known as "integrated stress response"	(141,150)
Hormesis	beneficial effects of low levels of stress ("what does not kill us makes us stronger" TM)	(121)
Redox Chemistry and Biology		
Oxidative stress	an imbalance between oxidants and antioxidants in favour of the oxidants, leading to a disruption of redox signaling and control and/or molecular damage	(176)
Reactive species	compounds with high chemical reactivity produced enzymatically or non-enzymatically; the interaction of reactive species may produce other reactive species	(73)
Reactive oxygen species (ROS)	chemically reactive compounds derived from oxygen	
Reactive nitrogen species (RNS)	chemically reactive compounds originating from the reaction of nitric oxide with oxygen or oxygen-derived compounds	
Reactive sulfur species (RSS)	according to Jacobs et al produced by "toxification" of thiols by reactive oxygen and nitrogen species, including disulfide-S-oxides, sulfenic acids and thiyl radicals and predicted to modulate the redox status of biological thiols and disulfides; more recently, RSS were discussed as putative regulatory entities of biological significance	(67,126)
Reactive species interactome (RSI)	chemical interaction of RSS, RNS and ROS (possibly including other short-lived species to be discovered in the future) among themselves and with downstream biological targets. The RSI is characterized by a) high variability/adaptability; b) rapid responsiveness; c) flexibility, which is required for fine-tuning of biological functions and communication at multiple levels; and d) high redundancy (explaining why antioxidants do not work)	This review
Redox regulation	a term used to define the control of redox signalling	(176)
Redox code	a set of principles that defines the positioning of the nicotinamide adenine dinucleotide (NAD, NADP), thiol/disulfide and other redox systems as well as the thiol redox proteome in space and time in biological systems	(94)
Personalized and Redox medicine		
Personalized medicine	health care that is tailored on individual condition, needs and life-style (in contrast to one-size-fits-all approach)	
Precision medicine	health care that is tailored on the basis of an individual's genes, lifestyle and environment	(81)
Stratified medicine	a therapy that is targeted to a specific patient population on the basis of a clinical characteristic such as a biomarker that predicts treatment response	(194)
Redox medicine / redox diseases	diseases with involvement of aberrant redox regulation/processes ranging from oxidative to reductive stress	(26,206)
P4 medicine	predictive, personalized, preventive and participatory (P4) medicine. P4 medicine is fueled by systems approaches to disease, emerging technologies and analytical tools	(84)
Developmental origins of health and disease	a paradigm affirming that environmental influences experienced during early embryonic development may influence the risk of non-communicable diseases later in life and across generations	(77)

Box 2 – The RSI in sensing, signaling and adaptation to stress – Principles of regulation in the context of origins of life, evolution and adaptation

Organisms observed today represent a snapshot of *now* – i.e. a cross-sectional view encompassing historical experience of preferred life forms that survived past evolutionary stresses, are compatible with the prevailing environment and fit for purpose. Assuming the overarching biological purpose is reproduction, this requires faithful replication of complex structures and molecular forms, which need to be dynamic yet sufficiently stable to assure structural and functional integrity of the system as a whole.

Biological flexibility in response provides resilience to all sorts of stressors in a constantly changing environment and can be identified at all levels of organization, which can be rationalized in terms of mathematics (networks), (bio)physical, (bio)chemical, physiological principles, as well as individual behavior and function, group behavior, and social realities on a global scale.

Influenced by Claude Bernard’s concept of the “*milieu interieure*” and Walter Cannon’s notion of *homeostasis*, the term *stress* was coined by Hans Selye in the middle of the last century; using experimental animal models, Selye also observed that persistent stress could lead to the development of various diseases (166). Mechanisms enabling to cope with stress are crucial checkpoints for resilience. Adaptation to (perceived or real) environmental, nutritional, life-style related or mental stresses serves the purpose to improve the fitness of a biological organism to deal with those stresses in the future. The concept of *hormesis* describes the ability of small

Table 2

Species	Occurrence/Formation	Chemistry
Dioxygen, O ₂	aerobic life	Has unpaired electrons - reacts readily with other radicals, poor 1e ⁻ oxidant but otherwise easily reduced by 2,3 and 4e ⁻ .

Superoxide, O_2^-	$1e^-$ reduction of O_2	Has one unpaired electron. Good reductant. Under acidic conditions can be an oxidant. Reacts with other radicals.
Hydrogen peroxide, H_2O_2	$1e^-$ reduction of O_2^-	Two-electron oxidant. Electrophilic. Not a radical species. Can modify RSH (below).
Hydroxyl radical, $HO\cdot$	$1e^-$ reduction of H_2O_2	Potent $1e^-$ oxidant. A short-lived radical species capable of abstracting an e^- or hydrogen atom from most biological molecules.
Nitric oxide, NO	enzymatic, NO_2^- reduction	Has unpaired e^- . Poor oxidant. Reacts with other radicals (O_2 and O_2^- and other organic radicals ($R\cdot$)). Can quench radical chemistry.
Nitrogen dioxide, NO_2	oxidation of NO	Good $1e^-$ oxidant. Radical species. Reacts with other radicals (can make $R-NO_2$ when reacted with $R\cdot$).
Dinitrogen trioxide, N_2O_3	oxidation of NO by O_2	Not a radical. Electrophilic and can nitrosate nucleophiles (add equivalent of " NO^+ "). Synthesis only relevant at high concentrations of NO.
Peroxynitrite, $ONOO^-$	reaction of NO and O_2^-	Not a radical but can generate both $HO\cdot$ and NO_2 . Rearranges to give nitrate (NO_3^-). Can oxidize by $2e^-$ (via peroxide-like chemistry).
Nitroxyl, HNO	S-nitrosothiol reduction	Reacts readily with thiols. A good hydrogen atom donor. Can act as an anti-oxidant by quenching radical reactions.
Nitrite, NO_2^-	oxidation of NO, dietary	Unreactive at neutral pH. Not a radical. Nitrosating agent under acidic conditions. Can be reduced to NO (under acidic conditions).
Hydrogen sulfide, H_2S	geochemical, enzymatic	Good metal ligand. Not a radical. Can react with other biological electrophilic sulfur species (e.g. RSSR, RSOH).
Thiol, RSH	endogenous (e.g. cysteine)	Good metal ligand. Not a radical. Can be oxidized to give other biologically relevant sulfur species.
Thiyl radical, $RS\cdot$	$1e^-$ oxidation of RSH	Radical species. Good $1e^-$ oxidant. Will react with other radical species such as NO.
Disulfide, RSSR	oxidation of RSH	Not a radical. Electrophilic. Can be reduced back to RSH under biological conditions.
S-Nitrosothiols, RSNO	Nitrosation of RSH	Not a radical. Can be reduced to RSH and HNO. Can transfer " NO^+ " to another thiol (transnitrosation).

Box 1 – The reactive species interactome (RSI)

The *reactive species interactome* (RSI) is a redox system consisting of chemical interactions of RSS, RNS and ROS among themselves and with downstream biological targets.

The RSI is characterized by a) robustness and flexibility; b) adaptability; c) rapid responsiveness; d) ability to sense the environment; and e) the ability to transduce signals that are required for fine-tuning of biological functions and communication at multiple levels.

The richness of chemical products of the RSI affords the unique redundancy and flexibility of the system. The products of the RSI are continually generated by enzymatic reactions and are as varied as the chemistries of the reactive species themselves. Chemical interactions of the RSI include one- and two-electron oxidations, nitrosation, nitration and sulfuration/polysulfidation reactions. Each of the species of this interactome has a distinct reactivity and lifetime that is

Box 3 – The RSI in sensing, signaling and adaptation to stress – Principles of regulation in the context of origins of life, evolution and adaptation

Organisms observed today represent a snapshot of *now* – i.e. a cross-sectional view encompassing historical experience of preferred life forms that survived past evolutionary stresses, are compatible with the prevailing environment and fit for purpose. Assuming the overarching biological purpose is reproduction, this requires faithful replication of complex structures and molecular forms, which need to be dynamic yet sufficiently stable to assure structural and functional integrity of the system as a whole.

Biological flexibility in response provides resilience to all sorts of stressors in a constantly changing environment and can be identified at all levels of organization, which can be rationalized in terms of mathematics (networks), (bio)physical, (bio)chemical, physiological principles, as well as individual behavior and function, group behavior, and social realities on a global scale.

Influenced by Claude Bernard's concept of the "*milieu interieure*" and Walter Cannon's notion of *homeostasis*, the term *stress* was coined by Hans Selye in the middle of the last century; using experimental animal models, Selye also observed that persistent stress could lead to the development of various diseases (166). Mechanisms enabling to cope with stress are crucial checkpoints for resilience. Adaptation to (perceived or real) environmental, nutritional, life-style related or mental stresses serves the purpose to improve the fitness of a biological organism to deal with those stresses in the future. The concept of *hormesis* describes the ability of small

Box 4 – Chemical biology and functional significance of the RSI.

ROS were initially viewed as mere by-products of redox reactions, especially mitochondrial respiration and certain pathological conditions, leading to oxidative damage of biological targets (protein, lipids, DNA). Further oxidative reactions were believed to be mediated by RNS, mainly produced by the oxidation of NO. Similarly, cysteine oxidative modification and formation of RSS were first considered only as a consequence of pathological conditions (67). Today, reactive species are considered part of a complex redox signaling network that interacts with protein thiol targets, which act as *redox switches* to control protein structure and function in dependence of local and global redox and environmental/nutritional status. The analysis of the chemical biology of H₂S and related sulfane sulfur species, and their interaction with NO and ROS indicate that the RSI is a tightly intertwined redox network that enables rapid sensing and adaptation of the internal cellular milieu to a changing environment. As indicated in Box 1 it is

Box 5 – RSI precursors and cofactors

The precursors and cofactors required to support the functioning of the RSI belong to the oxygen-arginine-methionine metabolome and originate from the same pathways that provide the basic building blocks for proteins, lipids, methyl groups, DNA/RNA synthesis and are thus important for cell proliferation and repair; this suggests competition between anabolic events and redox signaling. The RSI also regulates the expression and activity of enzymes belonging to intermediary metabolism and stress response, highlighting the interactions between catabolism, bioenergetics and redox status. The reciprocal nature of these relationships indicates that RSI

Box 6 – Chemical biology of the reactive species interactome - The long and winding road from evolution to personalised medicine

The challenge of developing a dynamic map of redox interactions is related to placing the fundamental chemistry within a given physiological process or disease mechanism into spatial and temporal context. Although the RSI concept begins disentangling the fundamental interplay between the different reactive species and their biological targets, before their use in *personalized medicine* specific mechanism-based biomarker panels need to be developed. To this end, the evolutionary origin of redox regulation based fundamentally on the handling of various sulfur species may provide a useful pointer since “Nature does not waste a good chemical reaction”.

A case in point is nitrogen fixation and nitrate/nitrite reduction to make NO. Bacteria can produce NO either via ammonia oxidation (e.g. Nitrosomonas) or denitrification (e.g. Pseudomonas), giving rise to NO emanations from soil; plants use NO to stimulate cyclic GMP. Nitrite reductase is a P₄₅₀-like enzyme that evolved into a NO synthase (NOS) using another source that connects important proliferative and immune responses to arginine metabolism. This evolution helps today's mammalian organisms regulate host defense mechanisms via modulation of mitochondrial respiration and intermediary metabolism (affecting bioenergetics and immunometabolism).

The ultimate thermodynamic equation that connects evolution with precision medicine is that describing Gibbs free energy: $\Delta G = \Delta H - T\Delta S$ where the fundamental reactions of small molecules depend mostly on ΔH or the energy of covalent bond making. However, in the biological context ΔS means there are ever-increasing differentiated states/niches defined by the process of evolution. Precision medicine is by definition where ΔH meets ΔS . To get there involves a series of approximations: 1st the ΔH of the fundamental chemical reactions (section 3); 2nd the interaction of the reactive species metabolome with macromolecules; 3rd cellular compartmentalization; 4th tissue interactions; 5th systemic physiological and inflammatory; 6th is the genomic and epigenetic interaction with these processes; and 7th the interaction between macroorganisms and microbiome, virome and exposome (210).

Thermodynamics can tell us into which direction redox reactions may proceed, but it cannot tell us anything about their speed or indeed whether those reactions can ever be fast enough to be of biological significance. Overcoming energetic barriers by using substrate-specific enzymatic catalysis is an important first step, but just the beginning. Redox tunes systems through reaction rates (affecting concentrations), timing, and location - these factors provide precise targets that in Nature are exploited to improve biological fitness. The challenge is to unveil how this chemical redox biology operates in health and disease using multi-biomarker analysis (via “omics”) and imaging tools to determine location and timing. Such redox mapping will hold the key to novel therapeutics.

Supplementary figure legend

Supplementary Fig. 1: Metabolic pathways fuelling the RSI. In adult mammals, L-arginine is formed either via the entero-renal axis from dietary glutamine or proline, or via the hepatic urea cycle. Enterocyte mitochondria form citrulline from proline and glutamine via ornithine and carbamoylphosphate, and export it to the kidney where it is converted into arginine. The hepatic urea cycle also uses ornithine and carbamoylphosphate as precursor for arginine synthesis. As in enterocytes, the formation of carbamoylphosphate, from ammonia and bicarbonate, and citrulline synthesis takes place in the mitochondria. Arginine used for NO synthesis can then be recycled via the arginine/citrulline cycle. NO synthase activity is inhibited by different methylated arginine species (ADMA, NMA), which themselves are precursors for arginine biosynthesis via citrulline. A key interaction between nitrogen and sulfur metabolism is the methylation of arginine using SAM-dependent methyl transferases. SAM is formed from methionine and used for the methylation of a large number of different molecules, resulting in the formation of homocysteine (Hcy). Depending on the availability of methionine, homocysteine is either recycled to methionine via vitamin B12 and folate-dependent methionine synthase or betaine-homocysteine S-methyltransferase (BHMT), or is degraded to cystathionine, cysteine and H₂S via the transsulfuration (TS) pathway.

