RVC OPEN ACCESS REPOSITORY – COPYRIGHT NOTICE

This is the peer-reviewed, manuscript version of the following article:

Gozalo-Marcilla, M., Luna, S. P. L., Crosignani, N., Puoli Filho, J. N. P., Possebon, F. S., Pelligand, L. and Taylor, P. M. 'Sedative and antinociceptive effects of different combinations of detomidine and methadone in standing horses', *Veterinary Anaesthesia and Analgesia.*

© 2017. This manuscript version is made available under the CC-BY-NC-ND 4.0 license <u>http://creativecommons.org/licenses/by-nc-nd/4.0/</u>.

The full details of the published version of the article are as follows:

TITLE: Sedative and antinociceptive effects of different combinations of detomidine and methadone in standing horses

AUTHORS: Gozalo-Marcilla, M., Luna, S. P. L., Crosignani, N., Puoli Filho, J. N. P., Possebon, F. S., Pelligand, L. and Taylor, P. M.

JOURNAL: Veterinary Anaesthesia and Analgesia

PUBLISHER: Elsevier

PUBLICATION DATE: 17 April 2017 (online)

DOI: 10.1016/j.vaa.2017.03.009



1 RESEARCH PAPER

- 2 Running head (author) *M Gozalo-Marcilla et al.*
- 3 Running head (Short title) Detomidine-methadone combinations in horses
- 4 Sedative and antinociceptive effects of different combinations of detomidine and
- 5 methadone in standing horses
- 6 Miguel Gozalo-Marcilla^a, Stelio PL Luna^a, Nadia Crosignani^a, José NP Puoli Filho^b, Fábio S
- 7 Possebon^c, Ludovic Pelligand^d & Polly M Taylor^e
- ^aDepartment of Veterinary Surgery and Anaesthesiology, School of Veterinary Medicine and
- 9 Animal Science, São Paulo State University (UNESP), Botucatu, SP, Brazil
- ¹⁰ ^bDepartment of Animal Production, School of Veterinary Medicine and Animal Science, São
- 11 Paulo State University (UNESP), Botucatu, SP, Brazil
- ¹² ^cDepartment of Veterinary Hygiene and Public Health, School of Veterinary Medicine and
- 13 Animal Science, São Paulo State University (UNESP), Botucatu, SP, Brazil
- ¹⁴ ^dDepartments of Clinical Services and Sciences and Comparative Biomedical Sciences,
- 15 Royal Veterinary College, Hatfield, Herts, UK

^eTaylor Monroe, Ely, Cambs, UK

17

18 Correspondence: Miguel Gozalo-Marcilla, Departamento de Cirurgia e Anestesiologia
19 Veterinária, Faculdade de Medicina Veterinária e Zootecnia, Universidade Estadual Paulista,
20 Rua Prof. Dr. Walter Mauricio Correa s/nº, Botucatu-SP, 18618-681, Brazil. E-mail:
21 miguelgozalomarcilla@gmail.com

22

23

24 Abstract

25 **Objective** To evaluate intravenous (IV) detomidine with methadone in horses to identify a

26 combination which provides sedation and antinociception without adverse effects.

27 Study design Randomized, placebo-controlled, blinded, crossover.

Animals Eight adult healthy horses aged (mean ± standard deviation) 7 ± 2 years and 372 ±
27 kg.

Methods Six treatments were administered IV: saline (SAL); detomidine (5 μ g kg⁻¹; DET); methadone (0.2 mg kg⁻¹; MET) alone or combined with detomidine [2.5 (MLD), 5 (MMD) or 10 (MHD) μ g kg⁻¹]. Thermal, mechanical and electrical nociceptive thresholds (NT) were measured, and sedation, head height above ground (HHAG), cardiopulmonary variables and intestinal motility were evaluated at 5, 15, 30, 45, 60, 75, 90, 120, 180 minutes. Normal data were analyzed by mixed-model ANOVA and non-normal by Kruskal-Wallis (*p* < 0.05).

Results Nociceptive thresholds in horses administered methadone with the higher doses of 36 detomidine (MMD, MHD) were increased above baseline to a greater degree and for longer 37 duration (MMD: 15-30 minutes, MHD: 30-60 minutes) than in horses administered low dose 38 with methadone or detomidine alone (MLD, DET: 5-15 minutes). No increases in NT were 39 recorded in SAL or MET. Compared with baseline, HHAG was lower for 30 minutes in 40 MMD and DET, and for 45 minutes in MHD. No significant sedation was observed in SAL, 41 MET or MLD. Intestinal motility was reduced for 75 minutes in MHD and for 30 minutes in 42 all other treatments. 43

44 **Conclusions** Methadone (0.2 mg kg⁻¹) potentiated the antinociception produced by 45 detomidine (5 μ g kg⁻¹), with minimal sedative effects.

46 **Clinical relevance** Detomidine (5 μ g kg⁻¹) with methadone (0.2 mg kg⁻¹) produced 47 antinociception without the adverse effects of higher doses of detomidine.

- 49 *Keywords* detomidine, electrical stimulus, equine, mechanical stimulus, methadone, thermal
- 50 stimulus
- 51

52 Introduction

It is common practice to administer drug combinations for chemical restraint of standing horses, avoiding the potential complications and costs of general anaesthesia. The α_2 -agonist detomidine has been extensively used to produce sedation and analgesia. However, adverse effects associated with detomidine administration, especially occurring after intravenous (IV) bolus administration, include ataxia, bradycardia, arrhythmias, increased systemic vascular resistance and reduction in cardiac output, respiratory rate (f_R), arterial oxygen tension and intestinal motility (Yamashita et al. 2000; Daunt & Steffey 2002; Valverde 2010).

Combinations of an α_2 -agonist with an opioid provide synergistic analgesic effects 60 with predictable sedation (Marly et al. 2014; Oliveira et al. 2014; Taylor et al. 2014; Lopes et 61 al. 2016), while potentially reducing adverse effects. Opioids play an important role in 62 analgesia in other species, however, their use in horses is controversial owing to the potential 63 for excitement and intestinal hypomotility, especially when administered IV (Bennett & 64 Steffey 2002; Clutton 2010; Schauvliege 2014). Methadone, a synthetic µ-agonist opioid with 65 similar properties to morphine, also provides analgesia by antagonizing N-methyl-D-aspartate 66 (NMDA) receptors and has delta opioid receptor activity (Pollock et al. 2011). 67

Methadone (0.5 mg kg⁻¹) administered IV alone increased spontaneous locomotor 68 activity and impaired coordination in horses (Combie et al. 1979; Oliveira et al. 2014), and 69 the duration of thermal antinociception was short at 30 minutes (Oliveira et al. 2014). Lower 70 doses (0.2 mg kg⁻¹) administered IV have resulted in less adverse behavioral effect but 71 provided insufficient antinociception (Pippi & Lumb 1979; Oliveira et al. 2014). More recent 72 studies reported that methadone (0.2 mg kg^{-1}) did not produce antinociception when 73 administered alone, but potentiated antinociception induced by detomidine (10 μ g kg⁻¹) 74 (Oliveira et al. 2014; Lopes et al. 2016). No adverse effects were reported after 75 administration of methadone (0.15 mg kg⁻¹) IV, but antinociception was not reported (Linardi 76

et al. 2012). These studies suggest that further evaluation of lower doses of methadone incombination with detomidine in horses may be worthwhile.

Our hypothesis was that the combination of low $(2.5 \ \mu g \ kg^{-1})$ and medium $(5 \ \mu g \ kg^{-1})$ doses of detomidine with methadone $(0.2 \ mg \ kg^{-1})$ would result in sedation and analgesia with less adverse effects when compared with the administration of each drug alone. The aim of the study was to identify a combination resulting in antinociception with minimal adverse effects by evaluating the sedative, antinociceptive and potential side effects of some combinations of detomidine and methadone.

85

86 Materials and methods

The study was designed as a prospective, randomized, placebo-controlled, observer blinded
crossover, with a washout period of ≥1 week between treatments. It was approved by the
Institutional Ethical Committee for the Use of Animals in Research of the Faculty of
Veterinary Medicine and Animal Science of the State of São Paulo University (UNESP),
Botucatu (EC no. 08/2015).

92

93 Animals

Eight healthy adult cross-bred horses (four castrated males, four females), aged [mean \pm standard deviation (SD) (range)] 7 \pm 2 years (3-9 years), weighing 372 \pm 27 kg (325-420 kg) were studied. The animals were kept at grass and brought to covered pens with outdoor access at least 24 hours before each experiment. Commercial horse feed, hay and *ad libitum* water were provided. The horses were considered healthy based upon physical examination and laboratory investigations performed 1-2 weeks before the study started. No sedatives or analgesics were administered for at least 1 month before the beginning of the study.

101 The sample size was estimated using OpenEpi Software (<u>www.openepi.com</u>) based on 102 the results of a pilot study with three horses and from previous data (Oliveira et al. 2014; 103 Lopes et al. 2016). The expected mean differences between the antinociceptive variables in 104 the different treatments were assessed. A test power of 80% and a significance level of 5% 105 were employed.

107 Study design

The horses were weighed on the day of each experiment and fly repellent was applied to the 108 skin. The hair over both jugular veins was clipped for catheter placement. After skin 109 disinfection, 1 mL of 2% lidocaine (Xylestesin 2%; Cristália Produtos Químicos 110 Farmacêuticos Ltda, SP, Brazil) was injected subcutaneously and two 14 gauge catheters 111 were placed at each site, in the direction of the blood flow. The one in the right jugular vein 112 was used for drug administration (14 gauge, 48 mm BD Angiocath; Becton Dickinson Ind., 113 MG, Brazil) and in the left (14 gauge, 70 mm, Delta Med Srl, Italy) for blood sampling for 114 pharmacokinetic analysis (results reported elsewhere). Hair was clipped over the dorsal 115 aspects of both metacarpi for the thermal and mechanical devices, and immediately proximal 116 to the coronary band of the left thoracic limb for the electrical electrodes. To obtain a good 117 contact for the electrodes and to reduce resistance, the clipped coronary skin was washed 118 with water and degreased with chlorhexidine containing surfactants (RIOHEX 2%; Industria 119 Farmaceutica Rioquimica Ltda, SP, Brazil). This process was repeated twice. 120

After that, the horse was brought to the experimental room (6 m², without windows), previously sprayed with fly repellent, and allowed 30 minutes for familiarization. A blood pressure cuff [DURA-CUF CRITIKON (12–19 and 17–25 cm); GE Healthcare, Finland] was placed at the base of the tail, to measure heart rate (HR) and systolic arterial blood pressure (SAP) with a noninvasive Doppler (Model 812; Parks Medical Electronics, Inc., OR, USA).

¹⁰⁶

126 The prepared area at the left coronary band was twice cleaned with alcohol (Álcool 96; Farmácia Santa Cruz, SP, Brazil) and dried before attaching two adhesive electrodes 127 (2223BRQ; 3M do Brasil Ltda, SP, Brazil) 8 cm apart, secured with four adhesive wrap strips 128 around the hoof. The thermal (WTT2; Topcat Metrology Ltd, UK) and mechanical (WMT1; 129 Topcat Metrology Ltd) control units (Topcat Metrology Ltd) were attached with Velcro onto 130 a commercial blanket (Topcat Metrology Ltd) on the horse's back. The units were remotely 131 controlled by infrared signals. Temperature and force were recorded in degrees Celsius (°C) 132 and newtons (N), respectively. Finally, the thermal sensor on the right clipped metacarpus 133 and mechanical pin on the left were placed and connected to the control units. 134

Six treatments were randomly assigned using Microsoft Excel: saline (Cloreto de 135 sódio 0.9%; Fresenius Kabi, SP, Brazil; treatment SAL), detomidine (5 µg kg⁻¹; Eqdomin, 10 136 mg mL⁻¹; Ourofino Saúde Animal, SP, Brazil; treatment DET), methadone (0.2 mg kg⁻¹; 137 Mytedom 10 mg mL⁻¹, Cristália Produtos Químicos Farmacêuticos Ltda, SP, Brazil; 138 treatment MET), and methadone at the same dose combined with detomidine (2.5 μ g kg⁻¹; 139 treatment MLD), detomidine (5 μ g kg⁻¹; treatment MMD) and detomidine (10 μ g kg⁻¹; 140 treatment MHD). The final volume was adjusted to 10 mL by adding saline and administered 141 IV by hand at T0 over 10 seconds. 142

143 Sedation and responses to antinociceptive stimuli were evaluated in triplicate at 144 baseline (T0) before and at 5, 15, 30, 45, 60, 75, 90, 120 and 180 minutes after drug 145 administration. Intestinal motility was scored at the same time points starting at 15 minutes.

146

147 Degree and quality of sedation

Sedation was evaluated using the height of the head above the ground (HHAG) by measuring
the level of the lower lip against a scale on the wall (Ringer et al. 2012; Marly et al. 2014).
Quality of sedation was scored first by evaluating the degree of ataxia and second by

151 responses to tactile and audiovisual stimulation, always in the same order. The scoring system was a numerical rating scale (NRS) ranging from 0 to 3 for ataxia (0 no ataxia, 3 152 maximal ataxia) and responses to audiovisual stimuli (0 no response, 3 maximal response) 153 (Appendix A). A subjective visual analogue scale (VAS) was assigned consisting of a 10 cm 154 line where 0 cm represented no sedation or ataxia and 10 cm represented maximal sedation 155 and/or maximal ataxia. All the sedation variables were always evaluated by the main 156 investigator (MGM) unaware of the treatment. 157

158

159 Cardiopulmonary variables

SAP was corrected according to the height difference between the cuff and shoulder joint, the 160 latter location representing the level of the right atrium of the heart. The f_R was measured by 161 observation of chest movements over 30 seconds.

163

162

Nociceptive threshold testing 164

Nociceptive stimuli were applied at each time point immediately after sedation was scored 165 and cardiopulmonary variables had been recorded. Stimuli were always applied in the same 166 order; first electrical, followed by thermal and mechanical stimulation. Aversive responses 167 were recorded when the horse lifted its foot, pawed the ground, stamped, flexed the limb or 168 walked to avoid the stimulus (Luna et al. 2015). The stimulus was immediately withdrawn at 169 this end point and the value in volts (V), °C and N at threshold recorded. If cut-out was 170 reached, this value was recorded as the threshold. 171

172

Electrical threshold testing 173

The resistance between electrodes was measured using a digital multimeter (ET 1100; 174 Minipa do Brasil Ltda, SP, Brazil) to confirm it was below 3 k Ω . The electrodes were 175

connected to an electrical stimulator (Grass S-48; Astro-Med, Inc., RI, USA) adjusted to
deliver pulsatile current square waves of 10 ms at 10 Hz. The voltage started at 1 V, then was
increased by 1 V every 5 seconds. Stimulation was stopped immediately when an avoidance
response was observed or the voltage reached 20 V.

180

181 *Thermal threshold testing*

A thermal probe with a heating element encased in an 8 mm long brass tube (internal 182 diameter 2.4 mm and external diameter 3.2 mm) (probe 3; Dixon et al. 2016) was placed on 183 the clipped area of the right metacarpus and attached with an elasticated band secured by 184 Velcro. The probe includes a heater and a temperature sensor and is connected to the thermal 185 control unit. At least 5 minutes were allowed for equilibration with body temperature. After 186 recording skin temperature, the ramped stimulus was applied, heating at 0.8 °C second⁻¹ until 187 a positive response was observed or the cut-out at 60 °C was reached (Luna et al. 2015; 188 Lopes et al. 2016). After each stimulus, the probe was moved 1-2 cm proximally on the limb 189 190 to avoid focal tissue damage.

191

192 Mechanical threshold testing

A pneumatic actuator containing a 1 mm round-ended pin was secured with a brushing boot on the left clipped metacarpal area, held against the leg with an elasticated band secured with Velcro and connected to the control unit with non-distensible tubing (Taylor et al. 2016). The automatic control system increased the force of the pin pressing on the skin surface at 0.8 N second⁻¹. The stimulus was stopped when an aversive response was observed or the cut-out value of 20 N reached.

199

200 Abdominal auscultation

Intestinal sounds were assessed by auscultation over one minute in each of the four abdominal quadrants: right dorsal, right ventral, left dorsal and left ventral. The total motility score was the sum of the scores from each quadrant (Boscan et al. 2006) (Appendix B).

204

205 Statistical analysis

For each variable, normality was assessed by the Shapiro-Wilk test and graphical analysis. 206 Effects of time and treatment were analyzed with a mixed-model analysis of variance 207 (ANOVA) followed by Tukey's test for HHAG, cardiopulmonary variables, thermal stimulus 208 and intestinal motility (mean ± SD). Data from electrical and mechanical stimuli were not 209 normally distributed. Thus, logarithmic transformation was performed, and these data are 210 presented as geometric mean (back-transformed bounds of the 95% confidence interval). 211 Different covariance structures were tested. Data for ataxia, responses to tactile and 212 audiovisual stimuli and VAS, non-normally distributed, were analyzed by Kruskal-Wallis 213 with Dunn's tests and are presented as median (range). Significance level was set at 0.05. 214

215

216 **Results**

The HHAG was lower than baseline (sedation was greater) after DET (66 ± 26 versus 98 ± 10 cm) and MMD (77 ± 19 versus 99 ± 3 cm) for 30 minutes and after MHD (68 ± 26 versus 99 ± 9 cm) for 45 minutes (Fig. 1). Comparing treatments, HHAG with DET was lower than SAL, MET and MLD for 30 minutes. The HHAG with MMD was lower than SAL and MLD for 15 minutes and MET for 30 minutes, respectively. The HHAG with MHD was lower than SAL, MET, MLD for 45 minutes and between 15 and 45 compared with MMD. There were no differences between DET and MHD.

Visual analogue scores were significantly higher than baseline for 30 minutes only in
treatments DET, MMD and MHD (Table 1). Comparing treatments, MHD scores were not

different from MMD or DET at any time point, but were higher than MLD for 45 minutes,

226

and higher than SAL and MET for 60 minutes (Table 1). Scores in MMD were not different 227 from DET or MLD at any time point, but were higher than SAL and MET for 30 minutes. 228 229 The ataxia scores were higher than baseline [0 (0 - 0)] with DET [1 (0 - 1)], MMD [1(0-2)] and MHD [1 (0 - 3)] up to 15 minutes (p < 0.05). Comparing treatments, ataxia was 230 more pronounced with MHD [1 (0-3)] than with SAL [0 (0-0)] and MET [0 (0-0)] for 30 231 minutes (p < 0.05). Ataxia scores with MHD [2 (1 – 3)] were also higher than MLD [1 (0 – 232 3)] at 5 minutes (p < 0.05). 233 Scores for responses to tactile stimuli were lower than baseline [3(2-3)] with MMD 234 [1.5 (0-2)] for 5 minutes and up to 30 minutes in MHD [1 (0-2)] (p < 0.05). Comparing 235 treatments, scores were lower with MHD [2 (0 - 3)] than SAL [3 (3 - 3)] and MET [3 (3 - 3)] 236 3)] up to 45 minutes, lower than MLD at 15 [[0 (0-1)] versus [3 (1-3)]] and at 30 minutes 237 [[1 (0-2)] versus [3 (1-3)] and lower than DET at 15 [[0 (0-1)] versus [2.5 (1-3)]] and 238 45 minutes [[2 (0-3)] versus [3 (3-3)]] (p < 0.05). Scores were also lower with MMD [1.5] 239 (0-2)] than SAL [3 (3 – 3)] and MET [3 (2 – 3)] at 5 minutes (p < 0.05). 240 Scores of responses to audiovisual stimuli were lower than baseline [1 (1 - 2)] for 15 241 minutes in MHD [0 (0 - 1)] (p < 0.05). Comparing treatments, scores were lower with MHD 242 [0 (0 - 1)] than SAL [1 (1 - 2)] and MET [1 (1 - 2)] for up to 15 minutes and lower than 243 MLD at 5 minutes [0 (0-0)] versus [1 (1-2)] (p < 0.05). 244 HR were not significantly changed from baseline within all treatments (Table 2). 245 Statistically significant changes in SAP and $f_{\rm R}$ were recorded in some treatments (Table 2). 246 Electrical thresholds in treatments MHD, MMD, MLD and DET were significantly 247 higher than baseline for 30, 15, 5 and 15 minutes, respectively (Table 3). Comparing 248 treatments, MHD thresholds were higher than MMD at 15 and 30 minutes only, but were 249

250 higher than DET, MLD and SAL for 30 minutes and MET for 45 minutes. After MMD,

thresholds were higher than MET and SAL only for 15 minutes. Thresholds at, and after 60minutes were not different from baseline and between treatments.

Thermal thresholds in treatments MHD, MMD, MLD and DET were significantly 253 higher than baseline for 45, 30, 15 and 15 minutes, respectively (Table 3). Comparing 254 treatments, MHD thresholds were higher than MMD only at 30 minutes, higher than MLD 255 and DET from 15-45 minutes and higher than SAL and MET at all time points up to 45 256 minutes. Thermal thresholds in MMD were higher than DET only at 30 minutes, and higher 257 than SAL and MET for 30 minutes. Thresholds in MLD and DET were higher than MET for 258 5 minutes and SAL for 15 minutes. Thresholds at, and after 60 minutes were not different 259 from baseline and between treatments. 260

Mechanical thresholds in MHD, MMD, MLD and DET were significantly higher than baseline for 60, 30, 15 and 5 minutes, respectively (Table 3). Comparing treatments, MHD thresholds were higher than MLD only at 30 minutes, higher than DET at 15-60 minutes, higher than MET for 45 minutes, and higher than SAL for up to 60 minutes. Mechanical thresholds in MMD were higher than DET only at 15 minutes, and higher than MET and SAL for 15 minutes. Thresholds at, and after 75 minutes were not different from baseline and between treatments.

Intestinal motility was below baseline in MET, DET, MLD and MMD for 30 minutes and up to 75 minutes in MHD (Fig. 2). Comparing treatments, MET, DET, MMD and MLD were lower than SAL for 30 minutes, and MHD was lower for 60 minutes. MHD was lower than MET and MMD for 30 minutes and lower than MLD and DET for 45 minutes.

272 Methadone alone (MET) resulted in mild behavioral effects in the first 5-15 minutes: 273 two horses were restless, two flicked their lower lips, one made minor attempts to move and 274 one was sedated. No adverse effects were observed in the remaining two horses. No abnormal

behavioral effects were observed after any other treatments except in MLD where one horseflicked the lower lips and one briefly became very alert to the environment.

One horse became deeply sedated in response to both drugs, either alone or in 277 combination. In this horse only, some superficial skin lesions became evident a few hours 278 after application of thermal stimuli in the third treatment, when thresholds close to the cut-out 279 temperature were reached. The lesions started as edematous wheals mimicking the footprint 280 of the probe. The outer surface of the skin was lost after two to three days, followed 281 subsequently by corneal flaking which resolved in 5 days. The lesions were initially slightly 282 painful to touch and were treated with wet cold towels, dried and followed by an ointment 283 containing methyl salicylate, Peru balsam (exudation of Myroxilon peruiferum) and zinc 284 oxide (Balsamex; Chemitec Agro-Veterinária Ltda, SP, Brazil). These injured local sites were 285 not used again for further thermal stimulation for the remaining treatments. 286

287

288 Discussion

Detomidine (10 μ g kg⁻¹) with methadone produced the most intense and prolonged 289 antinociception. When the detomidine dose was reduced to $5 \ \mu g \ kg^{-1}$, antinociception was of 290 similar intensity but of shorter duration, the horses were less sedated and intestinal motility 291 better preserved. This observation confirms that methadone potentiates detomidine-induced 292 antinociception, not only at the high detomidine dose of 10 μ g kg⁻¹ (Oliveira et al. 2014; 293 Lopes et al. 2016), but also at 5 μ g kg⁻¹, while sedative effects are minimized. Moreover, the 294 lowest dose of detomidine (2.5 μ g kg⁻¹) combined with methadone produced antinociception 295 similar to that after detomidine alone (5 μ g kg⁻¹), yet without producing measurable sedation. 296 No conclusion as to potentiation can be drawn regarding this combination as treatment with 297 detomidine alone at 2.5 μ g kg⁻¹ was not studied. 298

299 Nociceptive electrical, thermal and mechanical stimulation on the thoracic limbs is easy to apply and interpret, is reliable, sensitive and specific, and has been validated in horses 300 (Luna et al. 2015). Electrical stimuli activate not only the nociceptive A δ and C fibres, but 301 also large diameter AB fibres not directly involved in antinociception (Le Bars et al. 2001). 302 Thermal stimulation with a slow heating rate of 0.8 °C second⁻¹ predominantly activates C 303 fibres (Yeomans & Proudfit 1996; Lopes et al. 2016) and mechanical stimuli activate both 304 nociceptive A\delta and C fibres (Le Bars et al. 2001). Since opioids and α_2 -agonists have 305 different mechanisms of action, the use of a variety of nociceptive stimuli improves 306 sensitivity and specificity and therefore produces more accurate data (Luna et al. 2015; Lopes 307 et al. 2016). All three nociceptive testing modalities were sufficiently sensitive to detect 308 different degrees of antinociception between treatments, and, in general, followed a similar 309 310 trend.

The equipment employed for application of nociceptive stimuli was the same as 311 previously reported (Luna et al. 2015; Lopes et al. 2016). Additional features were included 312 for the electrical stimulus: before application, a strict protocol of clipping, washing, 313 degreasing and cleaning was followed. Since current intensity is equal to voltage divided by 314 resistance (Ohm's law), increases in skin resistance reduce intensity. Studies where resistance 315 was not measured showed heterogeneous, less accurate data (Lopes et al. 2016). Thus, proper 316 control of resistance is mandatory, not only by preparing the area and maintaining the same 317 distance between electrodes (7-8 cm on the coronary band), but also by keeping resistance 318 values below $3 k\Omega$ (Hopster et al. 2014; Risberg et al. 2014). 319

The drug doses used in this study were chosen to identify a combination that would produce adequate antinociception with minimal adverse effects. Detomidine (10-20 μ g kg⁻¹) and methadone (0.1-0.2 mg kg⁻¹) are commonly used clinical doses. In the present study, methadone alone (0.2 mg kg⁻¹) administered IV resulted in only transitory, minor behavioral

324 effects and no antinociception. Similar results were reported by Oliveira et al. (2014), where mild ataxia and head shaking were observed for 30 minutes, without consistent 325 antinociception. However, the addition of methadone (0.2 mg kg⁻¹) to detomidine (5 μ g kg⁻¹) 326 as described here, and to $10 \ \mu g \ kg^{-1}$ as reported by Oliveira et al. (2014) and Lopes et al. 327 (2016), prolonged the detomidine-induced antinociception and abated the methadone-induced 328 behavioral effects. Combination with detomidine (2.5 µg kg⁻¹) also appeared to reduce the 329 adverse effects of methadone, resulting in minimal adverse reactions in only two of the 330 horses. 331

Antinociception was observed in all treatments that included detomidine. In general, thresholds followed a similar pattern in the two treatments DET and MLD and in MMD and MHD, with longer durations of antinociception when methadone was combined with 5 or 10 μ g kg⁻¹ detomidine. The results indicate that the combination of methadone (0.2 mg kg⁻¹) with detomidine (5 μ g kg⁻¹) could provide useful antinociceptive effects for some clinical procedures lasting around 30 minutes.

Excessively deep sedation and severe ataxia may be considered adverse effects under 338 clinical conditions. In the study presented here, a reduction in the detomidine dose from 10 in 339 MHD to 5 μ g kg⁻¹ in MMD resulted in a similar degree of antinociception, but less sedation 340 (HHAG). Sedation in DET (5 μ g kg⁻¹) was comparable to MHD, but of shorter duration. The 341 lowest detomidine dose combined with methadone (MLD) produced similar antinociception 342 to detomidine alone (DET), without apparent sedation. The HHAG variable has been used to 343 measure the degree of sedation induced by administration of α_2 -agonists. Previous authors 344 have defined 'sufficient' sedation as when the horse's head position is equal to or lower than 345 50% of the awake position (Ringer et al. 2012; Marly et al. 2014). Using this definition, 346 adequate sedation was achieved only at 15 minutes after detomidine alone and for 30 minutes 347 in MHD, whereas sufficient sedation was not achieved with the other treatments. However, 348

349 the term 'sufficient' should be used carefully when an opioid is added to low doses of α_2 -350 agonists, as opioid-linked behavioral effects may overcome the sedative effects of the α_2 -351 agonist, depending on the relative dose ratio of the drugs.

Contrary to HHAG, VAS was similar in DET, MHD and MMD for the first 30 minutes. Moreover, the VAS detected more differences in sedation quality than the NRS used to score ataxia and responses to tactile and audiovisual stimuli. As two of these three tactile stimuli were performed on the limbs, it might be expected that methadone would increase locomotor activity (Oliveira et al. 2014) and, potentially increase the bias of these responses. However, the significant ataxia seen in treatments with detomidine at doses equal to or higher than 5 μ g kg⁻¹ may have reduced the responses to the tactile stimuli..

Throughout the experiments, the values for HR, SAP and $f_{\rm R}$ remained within 359 clinically acceptable ranges. As reported by Wagner et al. (1991) with a detomidine dose of 360 10 µg kg⁻¹, initial increases in arterial pressure, followed by a decrease, was observed only 361 with the combinations including the highest dose of detomidine. Reducing this dose limited 362 the biphasic effect on arterial pressure. $f_{\rm R}$ decreased below baseline for 60 minutes only in 363 MHD, but this was not statistically different from the other treatments. It is possible that 364 detomidine and its combination with methadone may cause respiratory depression, especially 365 at the highest dose, but arterial blood gases were not measured in this study. 366

367 Invasive arterial pressure measurements would have produced more accurate 368 measurements. However, a noninvasive method was used because evaluation of the 369 cardiopulmonary effects of the treatments was not the main aim of the study, and arterial 370 catheterization may have stimulated the horses and affected subsequent sedation and 371 antinociception.

372 Duration and intensity of intestinal hypomotility was detomidine dose-dependent. The 373 reduction in intestinal motility and its possible consequences are well reported after both α_{2} -

agonists (Daunt & Steffey 2002; Valverde 2010) and opioids (Bennett & Steffey 2002;
Boscan et al. 2006; Clutton 2010; Schauvliege 2014).

One horse developed small lesions at the site of thermal stimulation. Development of 376 skin lesions was unexpected as the cut-out was set at 60 °C, based on previous similar 377 studies, where only occasional superficial, local lesions were seen, usually at a thoracic site 378 and not on the limbs (Luna et al. 2015). A 60 °C cut-out may not be suitable for all conditions 379 as some individuals may be more sensitive to thermal stimulation. A lower cut-out such as 55 380 °C might be appropriate for different environments, sites and breeds to avoid tissue damage. 381 This horse became deeply sedated after all treatments, which may have influenced the results. 382 However, its exclusion would not be justified as this horse could represent a real effect seen 383 in a certain proportion of the population. It is also possible that allodynia developed after the 384 thermal injury in this horse, changing responses in subsequent experiments. Nonetheless, 385 separate statistical analysis performed without this horse's data did not significantly modify 386 the nociceptive results. 387

388

389 Conclusions

Methadone (0.2 mg kg⁻¹) potentiated the antinociception produced by detomidine at 5 μ g kg⁻³⁹¹ ¹. However, side effects such as deep sedation, ataxia and prolonged intestinal hypomotiliy were diminished when methadone was combined with 5 μ g kg⁻¹ detomidine when compared with the combination with 10 μ g kg⁻¹ detomidine. Further studies are justified to evaluate the value of this combination under clinical conditions.

395

396 Acknowledgements

This work was supported by Fundação de Amparo à Pesquisa de São Paulo (FAPESP),
Scholarships grant numbers 2010/08967-0 and 2014/00474-5.

The authors thank the pharmaceutical company Ourofino for donating the injectable form of detomidine. The authors also thank Altamiro Rosam for the special care and dedication with the horses used in this study and to Evaldo Mamedes Cândido de Queiroz for his unvaluable assistance, both at the São Paulo State University (UNESP), Botucatu, SP, Brazil.

404

405 Authors' contributions

MGM: study design, acquisition, management and interpretation of data, writing and critical review of the manuscript. SPLL: study design, interpretation of data, review of the manuscript. NC: study design, data acquisition, review of the manuscript. JNPF: data acquisition. FSP: statistical analysis and interpretation of data. LP: study design, management and interpretation of data, review of the manuscript. PMT: study design, interpretation of data, review of the manuscript.

412

413 **Conflict of interest**

414 PM Taylor is director of Topcat Metrology Ltd.

415

417 **References**

- Bennett RC, Steffey EP (2002) Use of opioids for pain and anesthetic management in horses.
 Vet Clin North Am Equine Pract 18, 47-60.
- 420 Boscan P, Van Hoogmoed LM, Farver TB, Snyder JR (2006) Evaluation of the effects of the
- 421 opioid agonist morphine on gastrointestinal tract function in horses. Am J Vet Res 67,
 422 992-997.
- Bryant CE, England GC, Clarke KW (1991) Comparison of the sedative effects of
 medetomidine and xylazine in horses. Vet Rec 129, 421-423.
- 425 Clutton RE (2010) Opioid analgesia in horses. Vet Clin North Am Equine Pract 26, 493-514.
- Combie J, Dougherty J, Nugent E et al. (1979) The pharmacology of narcotic analgesics in
 the horse. IV. Dose- and time-response relationships for behavioural responses to
 morphine, meperidine, pentazocine, anileridine, methadone, and hydromorphone. J
 Equine Med Surg 3, 377-385.
- 430 Daunt DA, Steffey EP (2002) Alpha-2 adrenergic agonists as analgesics in horses. Vet Clin
 431 North Am Equine Pract 18, 39-46.
- 432 Dixon MJ, Taylor PM, Slingsby LC, Murrell JC (2016) Refinement of a thermal threshold
 433 probe to prevent burns. Lab Anim 50, 54-62.
- Hopster K, Müller C, Hopster-Iversen C et al. (2014) Effects of dexmedetomidine and
 xylazine on cardiovascular function during total intravenous anaesthesia with midazolam
 and ketamine and recovery quality and duration in horses. Vet Anaesth Analg 41, 25-35.
- 437 Le Bars D, Gozariu M, Cadden SW (2001) Animal models of nociception. Pharmacol Rev
 438 53, 597-652.
- Linardi RL, Stokes AM, Keowen ML et al. (2012) Bioavailability and pharmacokinetics of
 oral and injectable formulations of methadone after intravenous, oral, and intragastric
 administration in horses. Am J Vet Res 73, 290-295.

- 442 Lopes C, Luna SP, Rosa AC et al. (2016) Antinociceptive effects of methadone combined
 443 with detomidine or acepromazine in horses. Equine Vet J 48, 613-618.
- Luna SPL, Lopes C, Rosa AC et al. (2015) Validation of mechanical, electrical and thermal
 nociceptive stimulation methods in horses. Equine Vet J 47, 609-614.
- Marly C, Bettschart-Wolfensberger R, Nussbaumer P et al. (2014) Evaluation of a romifidine
 constant rate infusion protocol with or without butorphanol for dentistry and
 ophthalmologic procedures in standing horses. Vet Anaesth Analg 41, 491-497.
- 449 Oliveira FA, Pignaton WA, Teixeira-Neto FJ et al. (2014) Antinociceptive and behavioural
- effects of methadone alone or in combination with detomidine in conscious horses. J
 Equine Vet Sci 34, 380-386.
- 452 Pippi NL, Lumb WV (1979) Objective tests of analgesic drugs in ponies. Am J Vet Res 40,
 453 1082-1086.
- 454 Pollock AB, Tegeler ML, Morgan V, Baumrucker SJ (2011) Morphine to methadone
 455 conversion: an interpretation of published data. Am J Hosp Palliat Care 28, 135-140.
- 456 Ringer SK, Portier KG, Fourel I et al. (2012) Development of a xylazine constant rate
 457 infusion with or without butorphanol for standing sedation of horses. Vet Anaesth Analg
 458 39, 1-11.
- Ringer SK, Portier K, Torgerson PR et al. (2013) The effects of a loading dose followed by
 constant rate infusion of xylazine compared with romifidine on sedation, ataxia and
 response to stimuli in horses. Vet Anaesth Analg 40, 157-165.
- 462 Risberg A, Spadavecchia C, Ranheim B et al. (2014) Antinociceptive effects of three
 463 escalating dexmedetomidine and lignocaine constant rate infusions in conscious horses.
 464 Vet J 202, 489-497.
- 465 Schauvliege S (2014) Opioids for field procedures in equine practice. Vet Rec 175, 621-622.
- 466 Taylor P, Coumbe K, Henson F et al. (2014) Evaluation of sedation for standing clinical

- 467 procedures in horses using detomidine combined with buprenorphine. Vet Anaesth Analg468 41, 14-24.
- 469 Taylor PM, Crosignani N, Lopes C et al. (2016) Mechanical nociceptive thresholds using
 470 four probe configurations in horses. Vet Anaesth Analg 43, 99-108.
- 471 Valverde A (2010) Alpha-2 agonists as pain therapy in horses. Vet Clin North Am Equine
 472 Pract 26, 515-532.
- Wagner AE, Muir WW 3rd, Hinchcliff KW (1991) Cardiovascular effects of xylazine and
 detomidine in horses. Am J Vet Res 52, 651-657.
- 475 Yamashita K, Tsubakishita S, Futaok S et al. (2000) Cardiovascular effects of medetomidine,
- detomidine and xylazine in horses. J Vet Med Sci 62, 1025-1032.
- 477 Yeomans DC, Proudfit HK (1996) Nociceptive responses to high and low rates of noxious
 478 cutaneous heating are mediated by different nociceptors in the rat: electrophysiological
- 479 evidence. Pain 68, 141-150.
- 480
- 481

- 482 Appendix A Numerical rating scale (NRS) to assess ataxia and responses to tactile and audiovisual stimulation. Adapted from Bryant et al.
- 483 (1991) and Ringer et al. (2013).

	Degree of ataxia	Tactile stimuli	Audiovisual stimuli
	Evaluated visually and by	Touching an ear with a pen, then pressing	Acoustic stimulus was hand clapping
	pushing the horse to detect any	the coronary band of thoracic and pelvic	behind the horse and visual stimulus was
	swaying	limbs with the pen	Shaking a towel in front of the horse
NRS			
0	No ataxia	No response, even with strong pressing	No response, no signs of noise recognition
			or visual arousal
1	Stable but swaying slightly	Mild response, slightly diminished response	Mild response, minimal movement of ears
		to normal or strong pressing	and elevates head slightly
2	Unstable, swaying markedly	Intermediate response, animal elevates the	Intermediate response, subdued reactions
		limb after normal pressing	and movements, turning slowly

	3	Severe ataxia, risk of falling down	Fast response, movement of the limb after	Fast response, animal abruptly turns or
			touching, with mild pressing	moves away
484				
485			~	O_{λ}
			Y	

486 Appendix B Scoring of intestinal sounds (Boscan et al. 2006).

Score Criteria	487
0 No intestinal sounds auscultated	
1 Low-pitched crepitant sounds, frequency ≤ 1 minute ⁻¹	
2 Low-pitched crepitant sounds, frequency > 1 minute ⁻¹	
3 Long, loud, gurgling sounds, frequency 1 minute ⁻¹	
4 Long, loud, gurgling sounds, frequency 2 to 4 minute ⁻¹	
5 Long, loud, gurgling sounds, frequency > 4 minute ⁻¹	Ć
R	

491 **Table 1** Sedation quality evaluated by visual analogue scale (VAS) [median (range)]. 0 indicates no sedation and no ataxia and 10 indicates maximum

sedation and ataxia, in eight standing horses after six treatments administered intravenously: 10 mL saline (SAL), 5 µg kg⁻¹ detomidine (DET), 0.2 mg

493 kg^{-1} methadone alone (MET), or combined with 10 (MHD), 5 (MMD) or 2.5 (MLD) $\mu g kg^{-1}$ detomidine before (time 0) and after (times 5-180

494 minutes) drug administration.

Time	Treatment				O'	
(minutes)	SAL	MET	DET	MLD	MMD	MHD
0	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)
5	$0(0-0)^{a}$	0 (0 - 1.3) ^{a}	5.3 (2.1 - 6.4) ^{*abc}	0 (0 - 8.7) ^{ab}	6.3 (4.9 - 8.8) ^{*bc}	7.8 (6.4 -9.9) [*] c
15	$0(0-0)^{a}$	0 (0 - 0.9) ^{ab}	6.2 (2.9 - 7.5) ^{*bcd}	0 (0 - 8.9) ^{abc}	6.2 (4.0 - 8.1) ^{*cd}	9.1 (6.1 - 9.6) ^{*d}
30	$0(0-0)^{a}$	$0(0-0)^{a}$	4.6 (0 - 6.4) ^{*abc}	0 (0 - 7.1) ^{ab}	3.3 (1.4 - 7.9) ^{*bc}	6.7 (3.9 - 8.6) ^{*c}
45	$0(0-0)^{a}$	$0(0-0)^{a}$	1.0 (0 - 4.5) ^{ab}	$0(0-4.5)^{a}$	0.5 (0 - 6.2) ^{ab}	3.4 (1.3 - 7.8) ^b
60	$0(0-0)^{a}$	$0(0-0)^{a}$	0 (0 - 1.4) ^{ab}	0 (0 - 1.7) ^{ab}	0 (0 - 2.2) ^{ab}	1.4 (0 - 5.2) ^b
75	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 3)
90	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 2.1)
120	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)

	180	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	0 (0 - 0)	
495						<u> </u>		
496	*Signif	icantly different	from baseline (tir	me 0) ($p < 0.05$). ^{abc}	Different superscript	letters indicate sign	ificant differences among	g treatments at that time
497	point (p	o < 0.05).						
498								
499						5		
					(P)			
					A.			
				C				
				Y.				

500 **Table 2** Heart rate (HR), systolic arterial blood pressure (SAP) and respiratory rate (f_R) (mean \pm standard deviation) in eight standing horses

501 administered intravenous saline (SAL), 5 μg kg⁻¹ detomidine (DET), 0.2 mg kg⁻¹ methadone alone (MET), or combined with 10 (MHD), 5 (MMD) or

502 2.5 (MLD) μ g kg⁻¹ detomidine.

		Treatments				£	
	Time	SAL	MET	DET	MLD	MMD	MHD
Variable	(minutes)				S		
HR					$\overline{\mathbf{z}}$		
(beats minute ⁻¹)	0	37 ± 11^a	32 ± 4^{b}	34 ± 6^{bc}	36 ± 6^a	33 ± 5^{ac}	35 ± 5^{b}
	5	37 ± 14^a	33 ± 5^{b}	30 ± 5^{bc}	36 ± 10^{a}	33 ± 8^{ac}	29 ± 5^{b}
	15	37 ± 12^{a}	35 ± 3^{b}	34 ± 10^{bc}	36 ± 9^a	35 ± 8^{ac}	31 ± 5^{b}
	30	38 ± 13^a	31 ± 4^{b}	34 ± 8^{bc}	38 ± 11^a	35 ± 7^{ac}	30 ± 5^{b}
	45	35 ± 12^{a}	31 ± 3^{b}	35 ± 7^{bc}	38 ± 12^{a}	35 ± 7^{ac}	31 ± 4^{b}
	60	37 ± 12^{a}	33 ± 4^{b}	33 ± 6^{bc}	34 ± 6^a	35 ± 9^{ac}	32 ± 4^b
	75	37 ± 13^{a}	33 ± 3^{b}	34 ± 7^{bc}	37 ± 11^{a}	37 ± 9^{ac}	32 ± 5^b
	90	38 ± 13^a	32 ± 4^{b}	32 ± 6^{bc}	36 ± 6^{a}	40 ± 8^{ac}	33 ± 4^{b}
	120	37 ± 12^{a}	31 ± 3^{b}	36 ± 8^{bc}	37 ± 10^{a}	35 ± 7^{ac}	33 ± 5^{b}

	180	37 ± 12^{a}	32 ± 3^{b}	33 ± 6^{bc}	38 ± 12^{a}	37 ± 8^{ac}	33 ± 4^{b}
SAP (mmHg)	0	139 ± 15	161 ± 11	143 ± 12	122 ± 9	125 ± 6	127 ± 6
	5	$133 \pm 9^{\mathbf{a}}$	$143 \pm 13^{\mathbf{a}}$	149 ± 16^{a}	$148 \pm 11^{\mathbf{a}}$	$148 \pm 8^{\mathbf{a}}$	$193 \pm 13^{\textbf{*b}}$
	15	$153 \pm 22^{\mathbf{ab}}$	$156\pm22^{\mathbf{a}}$	$110\pm6^{*\mathbf{c}}$	137 ± 12^{abc}	127 ± 11^{bc}	138 ± 7^{ab}
	30	$154 \pm 11^{\text{ab}}$	$165\pm5^{\mathbf{a}}$	$149\pm7^{\textbf{ab}}$	131 ± 10 ^b	$141 \pm 15^{\mathbf{ab}}$	$134\pm13^{\textbf{b}}$
	45	132 ± 12	$130 \pm 4^*$	141 ± 6	143 ± 8	144 ± 7	138 ± 4
	60	$168 \pm 10^{*\mathbf{a}}$	$136\pm29^{\text{bc}}$	$151\pm41^{\text{ab}}$	137 ± 5^{bc}	114 ± 5^{c}	$143\pm17^{\textbf{ab}}$
	75	$134 \pm 6^{\mathbf{a}}$	$131 \pm 4^{*b}$	143 ± 25^{ab}	$126 \pm 21^{\mathbf{a}}$	$143 \pm 11^{\text{ab}}$	$168 \pm 9^{*b}$
	90	141 ± 23	145 ± 13	126 ± 6	137 ± 12	140 ± 12	132 ± 7
	120	141 ± 13	$127 \pm 11^*$	133 ± 25	127 ± 8	124 ± 9	138 ± 21
	180	127 ± 6^{ab}	126 ± 4^{ab}	$116 \pm 9^{\mathbf{a}}$	$149\pm7^{\textbf{b}}$	$139\pm7^{\textbf{ab}}$	$123\pm8^{\bm{ab}}$
$f_{ m R}$							
(breaths minute ⁻¹)	0	16 ± 4	16 ± 4	16 ± 4	15 ± 3	15 ± 2	17 ± 5
	5	$16 \pm 4^{\mathbf{a}}$	16 ± 6^{ab}	$10 \pm 2^{\mathbf{ab}}$	12 ± 6^{ab}	10 ± 3^{ab}	$8\pm2^{*b}$
	15	$16 \pm 3^{\mathbf{a}}$	17 ± 7 ^b	$10 \pm 2^{\mathbf{ab}}$	13 ± 6^{ab}	9 ± 2^{ab}	$8 \pm 1^{*b}$
	30	$16 \pm 4^{\mathbf{ab}}$	$19 \pm 6^{\mathbf{a}}$	$10 \pm 2^{\mathbf{bc}}$	13 ± 5^{abc}	$9\pm 2^{\mathbf{bc}}$	$8\pm1^{*c}$

45	17 ± 4^{ab}	$18 \pm 6^{\mathbf{a}}$	$11 \pm 2^{\mathbf{bc}}$	14 ± 6^{abc}	11 ± 3^{bc}	$10 \pm 2^{*c}$
60	$17\pm4^{\bm{ab}}$	19 ± 7^{a}	$11\pm 2^{\textbf{b}}$	14 ± 6^{ab}	$12 \pm 2^{\mathbf{b}}$	$10 \pm 2^{*b}$
75	17 ± 5	18 ± 7	12 ± 2	15 ± 4	12 ± 3	11 ± 1
90	17 ± 4	18 ± 6	13 ± 2	15 ± 5	14 ± 2	12 ± 2
120	18 ± 4	18 ± 5	14 ± 3	15 ± 3	14 ± 2	12 ± 2
180	18 ± 4	18 ± 5	14 ± 2	15 ± 3	15 ± 2	13 ± 2

503 *Significantly different from baseline (time 0) within a treatment (p < 0.05).

^{abc}Different superscript letters indicate significant differences among treatments at that time point (p < 0.05).

CERTE

505

Table 3 Electrical (upper safety limit 20 V), thermal (upper safety limit 60 °C) and mechanical (upper safety limit 20 N) thresholds in eight standing horses treated intravenously with 10 mL saline (SAL), 5 μ g kg⁻¹ detomidine (DET), 0.2 mg kg⁻¹ methadone (MET), or methadone combined with 10 (MHD), 5 (MMD) or 2.5 (MLD) μ g kg⁻¹ detomidine. Thermal threshold data are mean \pm standard deviation; electrical and mechanical threshold data are geometric means (back-transformed bounds of the 95% confidence interval).

Stimulus	Time	Treatment		S	2		
	(minutes)	SAL	MET	DET	MLD	MMD	MHD
Electrical (V)	0	1.7 (1.5, 2.1)	1.6 (1.3, 1.8)	1.4 (1.2, 1.7)	1.7 (1.4, 2.0)	1.7 (1.4, 2.0)	1.8 (1.5, 2.2)
	5	1.7 (1.4, 2.1) ^a	1.7 (1.3, 2.2) ^a	2.9 (2.0, 4.1) ^{*ab}	3.0 (1.5, 6.1) ^{*ab}	4.7 (3.4, 6.6) ^{*bc}	6.1 (3.4, 10.9) ^{*c}
	15	1.9 (1.6, 2.3) ^a	1.9 (1.5, 2.4) ^a	3.1 (2.2, 4.4) ^{*ab}	2.8 (1.8, 4.5) ^{ab}	4.0 (2.7, 5.9) ^{*b}	8.4 (6.0, 11.7) ^{*c}
	30	1.9 (1.6, 2.3) ^a	1.7 (1.4, 2.0) ^a	$1.8 (1.5, 2.2)^{a}$	2.4 (1.6, 3.6) ^a	2.9 (1.9, 4.5) ^a	6.2 (3.6, 10.8) ^{*b}
	45	1.7 (1.3, 2.2) ^{ab}	1.7 (1.4, 2.0) ^a	1.8 (1.4, 2.3) ^{ab}	2.0 (1.4, 2.9) ^{ab}	2.3 (1.6, 3.2) ^{ab}	3.0 (2.7, 3.4) ^b
Thermal (°C)	0	44 ± 4	44 ± 3	44 ± 3	44 ± 3	44 ± 4	44 ± 2
	5	$44 \pm 5^{\mathbf{a}}$	$44 \pm 3^{\mathbf{a}}$	$54\pm5^{*b}$	$53 \pm 6^{*b}$	$59 \pm 2^{*b}$	$60 \pm 0^{*b}$
	15	$43 \pm 3^{\mathbf{a}}$	47 ± 6^{ab}	$51\pm5^{*bc}$	$52\pm7^{*bc}$	$56 \pm 5^{*cd}$	$60 \pm 0^{*d}$

		MAN		ODIT	
Λ ((E P	$\Lambda \Lambda \Lambda$		(\mathbf{R})	2
			NUD	$\sim n$	

	30	$44 \pm 4^{\mathbf{a}}$	$44 \pm 5^{\mathbf{a}}$	$45 \pm 2^{\mathbf{a}}$	$49 \pm 5^{\mathbf{ab}}$	$52\pm6^{*b}$	$60 \pm 1^{*c}$
	45	$42 \pm 4^{\mathbf{a}}$	$44 \pm 4^{\mathbf{a}}$	$45 \pm 3^{\mathbf{a}}$	$45 \pm 3^{\mathbf{a}}$	49 ± 5^{ab}	$55 \pm 5^{*b}$
Mechanical					R		
(N)	0	1.1 (0.6, 2.2)	0.9 (0.6, 1.3)	0.9 (0.6, 1.2)	0.8 (0.6, 1.2)	1.0 (0.6, 1.7)	0.9 (0.5, 1.5)
	5	1.8 (0.9, 3.5) ^a	$1.5 (0.5, 4.0)^{a}$	4.4 (2.1, 9.5) ^{*ab}	4.7 (1.8, 12.0) ^{*ab}	$17.4 (12.9, 20.0^{\dagger})^{*b}$	20.0 (20.0, 20.0) ^{*b}
	15	1.3 (0.7, 2.6) ^a	1.6 (0.6, 4.0) ^{a}	2.5 (0.9, 6.7) ^a	4.8 (2.0, 11.4) ^{*ab}	15.1 (9.9, 20.0†) [*]	20.0 (20.0, 20.0) ^{*b}
	30	1.5 (0.8, 2.9) ^a	1.7 (0.7, 4.0) ^{a}	2.4 (1.4, 4.0) ^a	3.1 (1.2, 7.9) ^a	6.5 (2.0, 20.0 ⁺) ^{*ab}	$19.2 (17.4, 20.0^{\dagger})^{*b}$
	45	1.5 (0.8, 2.7) ^a	1.5 (0.6, 3.4) ^a	1.4 (0.9, 2.2) ^a	3.2 (1.2, 8.1) ^{ab}	3.8 (1.6, 8.7) ^{ab}	13.1 (8.7, 19.7) ^{*b}
	60	1.3 (0.8, 2.2) ^a	1.7 (0.9, 3.2) ^{ab}	1.2 (0.7, 2.1) ^a	1.5 (0.8, 2.8) ^{ab}	1.9 (0.8, 4.5) ^{ab}	6.4 (3.2, 12.6) ^{*b}

*Significantly increased above baseline (time 0) (p < 0.05). Different lower case letters at each time point for each variable indicate significant

513 differences among treatments (p < 0.05).

⁵¹⁴ †Indicates that the upper limit of the confidence interval was higher than 20, although mechanical thresholds were censored at 20N.

515

517	Figure	legends

Figure 1 Height of head above the ground (HHAG) in eight standing horses administered intravenously saline (SAL), 5 μ g kg⁻¹ detomidine (DET), 0.2 mg kg⁻¹ methadone alone (MET), or combined with 10 (MHD), 5 (MMD) or 2.5 (MLD) µg kg⁻¹ detomidine. Data are mean \pm standard deviation. *Significantly lower from time 0 and lower than MET (p < 0.05). \pm Significantly lower from time 0 and lower than SAL, MET and MLD (p < 0.05). \pm Significantly lower from time 0 and lower than SAL, MET, MLD and MMD (p < 0.05). Figure 2 Intestinal motility scores (mean ± standard deviation) in eight standing horses administered intravenously saline (SAL), 5 µg kg⁻¹ detomidine (DET), 0.2 mg kg⁻¹ methadone alone (MET), or combined with 10 (MHD), 5 (MMD) or 2.5 (MLD) µg kg⁻¹ detomidine. *Significantly lower from time 0 (p < 0.05). †Significantly higher than all other treatments (p< 0.05). \pm Significantly higher than MHD (p < 0.05). \pm Significantly lower than MET, DET, MLD, MMD (p < 0.05). ¶Significantly lower than DET and MLD (p < 0.05).



