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Protocol of a prospective observational cohort study on clinical and biological predictors of disease
course and outcome in Guillain-Barré syndrome

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Abstract

Guillain-Barré syndrome (GBS) is an acute polyradiculoneuropathy with a highly variable clinical presentation, course, and outcome. The factors that determine the clinical variation of GBS are poorly understood which complicates the care and treatment of individual patients. The protocol of the ongoing International GBS Outcome Study (IGOS), a prospective, observational, multi-centre cohort study that aims to identify the clinical and biological determinants and predictors of disease onset, subtype, course and outcome of GBS is presented here. Patients fulfilling the diagnostic criteria for GBS, regardless of age, disease severity, variant forms, or treatment, can participate if included within two weeks after onset of weakness. Information about demography, preceding infections, clinical features, diagnostic findings, treatment, course and outcome is collected. In addition, cerebrospinal fluid and serial blood samples for serum and DNA is collected at standard time points. The original aim was to include at least 1000 patients with a follow-up of 1-3 years. Data are collected via a web-based data entry system and stored anonymously. IGOS started in May 2012 and by January 2017 included more than 1400 participants from 143 active centres in 19 countries across 5 continents. The IGOS data/biobank is available for research projects conducted by expertise groups focusing on specific topics including epidemiology, diagnostic criteria, clinimetrics, electrophysiology, antecedent events, antibodies, genetics, prognostic modelling, treatment effects and long-term outcome of GBS. The IGOS will help to standardize the international collection of data and biosamples for future research of GBS.

Keywords: Guillain-Barré syndrome, diagnosis, treatment, prognosis, outcome, biomarkers

ClinicalTrials.gov Identifier: NCT01582763

Introduction

Guillain-Barré syndrome (GBS) is an acute polyradiculoneuropathy clinically characterized by a rapidly progressive symmetrical flaccid weakness of the limbs (*van den Berg, et al., 2014; Willison, et al., 2016*). The clinical presentation, course and outcome of GBS is variable. Some patients have mild weakness of the lower legs only, while others develop complete tetraplegia and respiratory failure requiring mechanical ventilation (*van den Berg, et al., 2014; Willison, et al., 2016*). Patients may have variant forms of GBS, including Miller Fisher syndrome (MFS), pure motor, paraparetic or pharyngeal-cervical-brachial forms (*van den Berg, et al., 2014; Wakerley, et al., 2014; Willison, et al., 2016*). The electrophysiological findings are equally heterogeneous with subgroups of patients showing features of either demyelination or axonal degeneration (*Hadden, et al., 1998; Ho, et al., 1995; Uncini and Kuwabara, 2012*). The clinical recovery also varies: some patients recover spontaneously with no residual limitations, while others require mechanical ventilation for months and remain wheel chair bound for the rest of their lives or even die despite treatment (*van den Berg, et al., 2014; Willison, et al., 2016*). Independent of the GBS subtype, severity or predicted outcome, the standard treatment regimen for the past two decades has consisted of intravenous immunoglobulin (IVIg) or plasma exchange (PE) (*Hughes, et al., 2007; Hughes, et al., 2014; Raphael, et al., 2012*). Despite treatment, outcome of GBS is frequently poor since 2-5% of the patients die and 10-20% of patients remain severely disabled (*Fokke, et al., 2014; Hughes, et al., 2007; Willison, et al., 2016*). Outcome is even worse in low-income countries: in Bangladesh for example 85% of patients receive no treatment, 15% die, and 30% remain severely disabled (*Islam, et al., 2010*). These findings show the need for more effective, personalized and accessible treatments worldwide.

The factors that determine the heterogeneity of GBS and that could provide a basis to personalize treatment are largely unknown. Previous studies observed an association between the clinical course in patients and the acute phase clinical characteristics or biomarkers such as electrophysiological subtype, preceding infections, anti-ganglioside antibodies, serum levels of IgG and albumin and genetic polymorphisms (*Fokkink, et al., 2016; Geleijns, et al., 2006; Kuitwaard, et*

al., 2009; Kusunoki, *et al.*, 2008; van Koningsveld, *et al.*, 2007; Walgaard, *et al.*, 2010; Walgaard, *et al.*, 2011; Walgaard, *et al.*, 2016). However, most of these studies were retrospective, and the findings were derived from relatively small series or selected groups of patients with a short follow-up period using a limited set of suboptimal outcome measures. In addition, the variety in inclusion criteria and methods used in these studies complicates comparing or combining collected data. Moreover, GBS varies considerably between geographical regions (*Kuwabara and Yuki, 2013; Mori, et al., 2012; Willison, et al., 2016*). Solving these problems requires a prospective study design with standardized collection of clinical data and biomaterials from a large group of well-defined GBS patients with a long follow-up period. The International GBS Outcome Study (IGOS) was initiated to collect the required clinical data and biosamples. After several organizational meetings, a final study protocol was prepared, and local investigators were invited to participate. IGOS is being conducted in collaboration with the Peripheral Nerve Society and Inflammatory Neuropathy Consortium (INC) (www.pnsociety.com/inflammatory-neuropathy-consortium/), an international organization of clinical neurologists and scientists involved in the investigation and care of patients with GBS.

Study aim and objectives

The overall aim of IGOS is to determine the clinical and biological determinants and predictors of the clinical course and outcome of GBS. The objectives are: (1) to define the variation in clinical presentation, subtypes, progression and recovery of GBS in patients from a broad range of geographical areas, (2) to describe the current practice of diagnosis, treatment, and care for GBS, (3) to identify environmental and host factors that determine the disease onset and the variation in clinical course, treatment response and outcome, (4) to develop improved prognostic models to predict the clinical course and outcome in individual patients, and (5) to facilitate the collection of standardized and relevant data and biosamples for future studies of GBS.

Methods and Materials

Study design

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IGOS is an international, prospective, observational, multi-center cohort study. It uses a predefined protocol to collect data regarding baseline characteristics, clinical presentation and course, electrophysiology, diagnosis, treatment and outcome during a follow-up of 1 year with the possibility to extend the follow-up to 2 or 3 years (Figure 1). The protocol specifies the timing of the collection of biosamples, including blood for DNA and serum studies and cerebrospinal fluid (CSF). Predictive models for the clinical course and outcome will be based on results collected in the first two weeks. The first 1000 participants will constitute a derivation cohort. The next 500 or more patients will provide a validation cohort.

Inclusion and exclusion criteria for patients

Inclusion requires fulfilling the following criteria:

1. The diagnostic criteria for GBS of the National Institute of Neurological Disorders and Stroke (NINDS) (See supplement 1), or one of the variants of GBS, including the MFS and overlap syndromes (*Asbury and Cornblath, 1990; Sejvar, et al., 2011; Wakerley, et al., 2014*).
2. Entry within two weeks of onset of weakness (or other symptoms attributed to GBS).
3. Opportunity to continue a follow-up for at least one year.
4. Informed consent of the participant or, for children, the parents or legal guardians.

The aim is to enroll patients representing the full spectrum of GBS. There are no exclusion criteria, and all patients with GBS or its variants, including MFS and overlap forms, may participate, regardless of age, disease severity, or treatment.

IGOS data- and biobank

Baseline, clinical and treatment data

We collect baseline data about the patients' demography, co-morbidity, family history and antecedent events. In addition, we record the first clinical symptoms and signs of GBS, the timing and

key features of the diagnosis, hospital transfers, neurological examination findings and the clinical course. The severity and distribution of clinical manifestations of GBS will be documented in detail, including cranial nerve deficits, limb weakness using the MRC sum score (*Kleyweg, et al., 1991*) and Rasch-built MRC score (*Vanhoutte, et al., 2012*), sensory deficits, ataxia, limb tendon reflexes, GBS disability scale (*Hughes, et al., 1978*), pain, autonomic dysfunction, respiratory failure, and associated medical complications. In addition, we record the occurrence of treatment related fluctuations and transitions to chronic inflammatory demyelinating polyneuropathy (CIDP) (*Ruts, et al., 2010*). From 4 weeks onward, we collect the following clinical outcome measures: Overall Neuropathy Limitation Scale (ONLS) (*Graham and Hughes, 2006*), Rasch-built Overall Disability Score (R-ODS) (*van Nes, et al., 2011*), Fatigue Severity Scale (FSS) (and Rasch-built FSS) (*van Nes, et al., 2009*) and the EuroQol EQ-5D health questionnaire (*Brooks, 1996*) (Figure 1). A full neurological examination will be performed at study entry and after 1 week, 2 weeks, 4 weeks, 26 (± 2) weeks and 52 (± 4) weeks. Participants still in hospital at 8 (± 1) weeks and 13 (± 1) weeks will be examined but those who have been discharged have only a telephone assessment of the GBS disability scale, ONLS, R-ODS, FSS and EuroQoL EQ-5D. All patient reported outcome measures have been translated (and back translated) into the language of the participants. We collect detailed information about the treatment for GBS including the type, timing, regimen and side-effects of treatment, admission to intensive care, and start and end of mechanical ventilation. If a patient dies, we document the timing and cause of death.

Electrophysiology data

We collect the results of the routine diagnostic electrophysiological examinations, including the raw data and the local investigators' interpretation and classification of the subtype, including acute inflammatory demyelinating polyneuropathy (AIDP), acute motor axonal neuropathy (AMAN), acute motor-sensory axonal neuropathy (AMSAN), inexcitable nerves, and equivocal or normal results. The study protocol recommends performing electrophysiological studies according to a predefined standard format (see Supplement 2). Some clinics routinely perform a second diagnostic nerve

conduction study and we collect these results when available. We document locally used normative values and standard electrophysiology protocols.

Blood samples

We collect blood and CSF samples according to a predefined schedule (Figure 1). Blood samples provide serum and DNA. The protocol specifies that the first serum sample is to be collected before the start of treatment, although in some patients treatment is being initiated before study entry (e.g., when patients transfer to a center participating in IGOS after treatment at a local community hospital). Blood samples are frozen and stored at -80°C (or initially stored -20°C for a maximum of 6 months) at the local center or at the center of the country coordinator, and transported on dry ice to a central biobank, at Erasmus MC in Rotterdam, the Netherlands, with a reserve biobank at the University of Glasgow, Scotland, UK.

Cerebrospinal fluid samples

If participants have a routine lumbar puncture to examine CSF for diagnostic studies, we keep an aliquot for biomarker studies. In centers participating in an advanced proteomics study, an extra aliquot is sampled (*Teunissen, et al., 2009*). This study requires centrifugation of the CSF sample within one hour after the lumbar puncture. The supernatant (without pellet) is removed and immediately stored in a polystyrene tube at -20°C until being transferred to the country coordinating center and stored at -80°C . This optional research module requires additional informed consent.

Extended follow-up of two and three years

The time course of nerve regeneration and clinical recovery in GBS is unclear. Some patients continue to improve even after one year (*Dhar, et al., 2008; Rudolph, et al., 2008*). To determine the further recovery and long-term residual deficits, there is an optional long-term follow-up research module with a telephone assessment at 2 years (104 ± 4 weeks) and 3 years (156 ± 4 weeks) after onset. The data collected then include the GBS disability score, ONLS, R-ODS, FSS (and Rasch-FSS),

and EuroQoL EQ-5D. Severely affected patients may have more extensive evaluation including neurological examination at each visit. The long-term follow-up requires additional informed consent.

Children with GBS

Children with GBS differ from adult patients regarding their preceding infections, clinical features, GBS subtype, treatment and outcome (*Korinthenberg, et al., 2007; Roodbol, et al., 2011*). In children it is more difficult to obtain biosamples and the adult outcome measures have not all been validated. For this reason we are using an adapted protocol for children with age-dependent sampling of biomaterials and clinical assessment scales.

Data collection

We have developed a web-based data entry system that meets the standards of security and privacy of Erasmus MC, the host institution. The local investigators use this website to enter the data. The information stored is strictly anonymous. All participants have a unique code for use throughout the study. The quality of the collected data is controlled regularly by the IGOS Coordinating Center according to protocol and additional controls will be conducted by the IGOS Expertise Groups.

Sample size

The extent of the clinical variation of GBS in the world is currently unknown and limits the possibility to conduct a power calculation. There is only circumstantial evidence available to estimate the required size of the study. It is recommended for the development of predictive logistic regression models that the smallest outcome group should include at least 100 patients (*Vergouwe, et al., 2005*). Previous therapeutic trials with GBS defined poor outcome as a GBS disability score of >2 (not being able to walk unaided or worse) (*Hughes, et al., 1978*). In treatment trials about 10-15% of participants have had a poor outcome after one year (*Hughes, et al., 2007; van Koningsveld, et al.,*

2007). If we aim to include at least 100 participants with a poor outcome defined in this way, the whole study will need at least 1000 participants. Therefore this is the minimum size of the derivation cohort to identify new clinical and biological markers to predict poor outcome or evaluate previously described prognostic factors. In addition, we collect an independent validation cohort of at least 500 patients to validate any new clinical or biomarkers that emerge from the derivation cohort.

Statistical analyses

Descriptive statistics are used to analyze the clinical data. We will develop prognostic models to evaluate the use of biomarkers and clinical characteristics to predict outcome in clinical practice according to our previously published statistical approaches (*van Koningsveld, et al., 2007; Walgaard, et al., 2010; Walgaard, et al., 2011*). The association between putative prognostic factors and outcome variables will be analyzed using univariate and multivariable logistic regression models. If two similar variables are equally associated with outcome, the variable most easily obtainable in clinical practice will be selected. We will quantify model performance with respect to discrimination (area under receiver operating characteristic curve). Multivariable regression coefficients will be used to develop new prognostic models for GBS.

IGOS Consortium and IGOS Research Policy Agreement

IGOS is conducted by the IGOS Consortium which consists of (1) the members of the Steering Committee, (2) the staff of the Coordinating Center at Erasmus MC, Rotterdam, the Netherlands, (3) the Country Coordinators and (4) their networks of local investigators (see Appendix). To be able to participate in IGOS and to become a member of the IGOS Consortium all participants signed the IGOS Consortium Agreement which defined the conduct of the study. All members of the IGOS Consortium can apply to the Steering Committee to use the IGOS data/biobank to address specific research questions. Research projects are also being conducted by the Expertise Groups, consisting of members of the IGOS Consortium and additional researchers if external expertise is required.

Ethical regulations

The IGOS received approval from the Medical Ethics Review Committee of the Erasmus MC Medical University Rotterdam in 2012. Each participating center also had approval from their local Institutional Review Board (IRB). The IGOS is being conducted according to the principles of the Declaration of Helsinki (59th WMA General Assembly, Seoul, October 2008) and the Medical Research Involving Human Subjects (WMO). The procedures set out in this protocol were designed to ensure that the investigators abide by the principles of the GCP guidelines of the European Community (ICH topic E6, CPMP/ICH/135/95, Directive 2001/20/EC) in the conduct, evaluation and documentation of this study. Inclusion in IGOS requires informed consent from each participant or their legal representative. IGOS was registered before the start of the study at ClinicalTrials.gov (NCT01582763).

Discussion

IGOS started in May 2012 and in June 2015 IGOS had enrolled 1000 patients who will form the derivation cohort (Figure 2). By January 2017, more than 1400 patients with GBS had been included in IGOS by 143 active sites from 19 countries across five continents (Figure 3). We are continuing to recruit more patients to complete the independent validation cohort, and anticipate enrollment of 1500 patients by July 2017.

IGOS has important advantages over previous observational studies of GBS. First, IGOS already includes the largest number of prospectively collected patients with a confirmed diagnosis of GBS. Second, there was no selection of patients based on age, clinical subtype or severity, aiming to investigate the full range of variants within the spectrum of GBS. Third, we collected data from patients from various geographical regions, including high- and low-income countries using the same protocol and will be able to compare and contrast various attributes of the condition worldwide. Fourth, IGOS has a long follow-up period and uses several well-defined and validated outcome measures to assess the long-term outcome of GBS. Fifth, we are collecting biosamples prospectively according to the protocol at recruitment and follow-up visits coinciding with clinical assessments. We will use these samples to study preceding infections, antibodies to peripheral nerves and other immunological factors, pharmacokinetics, genetic factors, and other potential biomarkers for correlation with clinical features. Because of these advantages, IGOS will provide the most extensive research data- and biobank of GBS patients collected so far. This will enhance our understanding of the pathogenesis and the individual clinical course, prognosis, treatment response and outcome. The overall aim is to develop more effective personalized treatment regimens based on a better understanding of the variation of the disease.

IGOS has already enhanced international collaboration in research into the cause and treatment of GBS by strengthening international networks. Expertise Groups will be formed on various topics including (1) emerging preceding events related to GBS, (2) diagnostic criteria and protocols for GBS, (3) electrophysiological subtypes of GBS, (4) biological determinants including genetic

polymorphisms, preceding infections and serum antibodies in the pathogenesis of GBS and its subtype, (5) biomarkers for monitoring treatment, pharmacokinetics and disease activity, (6) validating and improving outcome measures, (7) long-term outcome, (8) prediction of prognosis, and (9) improving treatment.

By providing an infrastructure for standardized collection of data and biomaterials, IGOS will facilitate international research projects on emerging infections associated with GBS, such as the recent outbreak of Zika virus infection (*Cao-Lormeau, et al., 2016; Parra, et al., 2016*). This infrastructure will also help to record other emerging preceding events that have been previously related to the development of GBS, including vaccinations. Our aim is also to use the international expertise involved in IGOS and the collected materials to compare and standardize assays for relevant biomarkers in GBS including preceding infections, antibodies, genetic polymorphisms and pharmacokinetic analysis. The extensive recording of the clinical course and outcome at serial visits during long-term follow-up provides a unique opportunity for international validation of outcome measures that so far have been developed in limited regions only. The IGOS data- and biobank provide an easily accessed source of control natural history data for modelling studies and comparison with patients treated with novel treatment regimens. One ongoing study is already comparing one with two courses of IVIg in patients with a poor predicted outcome (International Second IVIg Dose GBS Study). A second observational study within IGOS is comparing IVIg treatment with supportive care alone for patients with initially mild GBS. IGOS provides an exciting opportunity to support future clinical trials. The consortium is open to collaboration with other academic partners and with pharmaceutical companies interested in improving the treatment of GBS.

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specific countries or projects was received from gain (<http://www.gaincharity.org.uk/>), Erasmus MC, Rotterdam, The Netherlands, University of Glasgow, Glasgow, United Kingdom, Grifols, CSL Behring, Shire Pharmaceuticals and Annexon. IGOS is scientifically independent, and the funding agencies have no influence on the study and infrastructure design of IGOS, nor on the collection, statistical analysis and interpretation of the data collected in IGOS, nor on the writing, publication of manuscripts or other presentations based on these data.

References

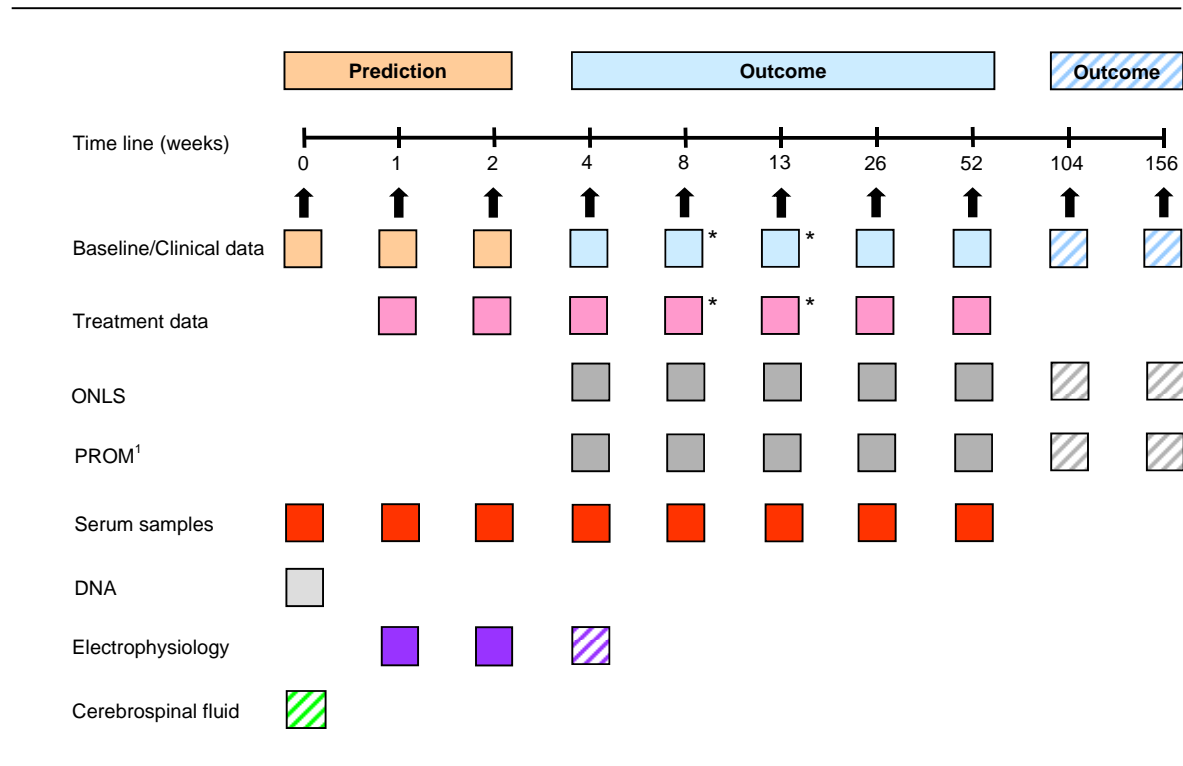
- Asbury AK, Cornblath DR (1990). Assessment of current diagnostic criteria for Guillain-Barre syndrome. *Ann Neurol* 27 Suppl:S21-24.
- Brooks R (1996). EuroQol: the current state of play. *Health Policy* 37:53-72.
- Cao-Lormeau VM, Blake A, Mons S, Lastere S, Roche C, Vanhomwegen J, Dub T, Baudouin L, Teissier A, Larre P, Vial AL, Decam C, Choumet V, Halstead SK, Willison HJ, Musset L, Manuguerra JC, Despres P, Fournier E, Mallet HP, Musso D, Fontanet A, Neil J, Ghawche F (2016). Guillain-Barre Syndrome outbreak associated with Zika virus infection in French Polynesia: a case-control study. *Lancet* 387:1531-1539.
- Dhar R, Stitt L, Hahn AF (2008). The morbidity and outcome of patients with Guillain-Barre syndrome admitted to the intensive care unit. *J Neurol Sci* 264:121-128.
- Fokke C, van den Berg B, Drenthen J, Walgaard C, van Doorn PA, Jacobs BC (2014). Diagnosis of Guillain-Barre syndrome and validation of Brighton criteria. *Brain* 137:33-43.
- Fokkink WR, Walgaard C, Kuitwaard K, Tio-Gillen AP, van Doorn PA, Jacobs BC (2016). Association of Albumin Levels With Outcome in Intravenous Immunoglobulin-Treated Guillain-Barre Syndrome. *JAMA Neurol*.
- Geleijns K, Roos A, Houwing-Duistermaat JJ, van Rijs W, Tio-Gillen AP, Laman JD, van Doorn PA, Jacobs BC (2006). Mannose-binding lectin contributes to the severity of Guillain-Barre syndrome. *J Immunol* 177:4211-4217.
- Graham RC, Hughes RA (2006). A modified peripheral neuropathy scale: the Overall Neuropathy Limitations Scale. *J Neurol Neurosurg Psychiatry* 77:973-976.
- Hadden RD, Cornblath DR, Hughes RA, Zielasek J, Hartung HP, Toyka KV, Swan AV (1998). Electrophysiological classification of Guillain-Barre syndrome: clinical associations and outcome. Plasma Exchange/Sandoglobulin Guillain-Barre Syndrome Trial Group. *Ann Neurol* 44:780-788.
- Ho TW, Mishu B, Li CY, Gao CY, Cornblath DR, Griffin JW, Asbury AK, Blaser MJ, McKhann GM (1995). Guillain-Barre syndrome in northern China. Relationship to *Campylobacter jejuni* infection and anti-glycolipid antibodies. *Brain* 118 (Pt 3):597-605.
- Hughes RA, Newsom-Davis JM, Perkin GD, Pierce JM (1978). Controlled trial prednisolone in acute polyneuropathy. *Lancet* 2:750-753.
- Hughes RA, Swan AV, Raphael JC, Annane D, van Koningsveld R, van Doorn PA (2007). Immunotherapy for Guillain-Barre syndrome: a systematic review. *Brain* 130:2245-2257.
- Hughes RA, Swan AV, van Doorn PA (2014). Intravenous immunoglobulin for Guillain-Barre syndrome. *Cochrane Database Syst Rev*:CD002063.
- Islam Z, Jacobs BC, van Belkum A, Mohammad QD, Islam MB, Herbrink P, Diorditsa S, Luby SP, Talukder KA, Endtz HP (2010). Axonal variant of Guillain-Barre syndrome associated with *Campylobacter* infection in Bangladesh. *Neurology* 74:581-587.
- Kleyweg RP, van der Meche FG, Schmitz PI (1991). Interobserver agreement in the assessment of muscle strength and functional abilities in Guillain-Barre syndrome. *Muscle Nerve* 14:1103-1109.
- Korinthenberg R, Schessl J, Kirschner J (2007). Clinical presentation and course of childhood Guillain-Barre syndrome: a prospective multicentre study. *Neuropediatrics* 38:10-17.
- Kuitwaard K, de Gelder J, Tio-Gillen AP, Hop WC, van Gelder T, van Toorenenbergen AW, van Doorn PA, Jacobs BC (2009). Pharmacokinetics of intravenous immunoglobulin and outcome in Guillain-Barre syndrome. *Ann Neurol* 66:597-603.
- Kusunoki S, Kaida K, Ueda M (2008). Antibodies against gangliosides and ganglioside complexes in Guillain-Barre syndrome: new aspects of research. *Biochim Biophys Acta* 1780:441-444.
- Kuwabara S, Yuki N (2013). Axonal Guillain-Barre syndrome: concepts and controversies. *Lancet Neurol* 12:1180-1188.
- Mori M, Kuwabara S, Yuki N (2012). Fisher syndrome: clinical features, immunopathogenesis and management. *Expert Rev Neurother* 12:39-51.
- Parra B, Lizarazo J, Jimenez-Arango JA, Zea-Vera AF, Gonzalez-Manrique G, Vargas J, Angarita JA, Zuniga G, Lopez-Gonzalez R, Beltran CL, Rizcala KH, Morales MT, Pacheco O, Ospina ML, Kumar A,

- Cornblath DR, Munoz LS, Osorio L, Barreras P, Pardo CA (2016). Guillain-Barre Syndrome Associated with Zika Virus Infection in Colombia. *N Engl J Med* 375:1513-1523.
- Raphael JC, Chevret S, Hughes RA, Annane D (2012). Plasma exchange for Guillain-Barre syndrome. *Cochrane Database Syst Rev* 7:CD001798.
- Roodbol J, de Wit MC, Walgaard C, de Hoog M, Catsman-Berrevoets CE, Jacobs BC (2011). Recognizing Guillain-Barre syndrome in preschool children. *Neurology* 76:807-810.
- Rudolph T, Larsen JP, Farbu E (2008). The long-term functional status in patients with Guillain-Barre syndrome. *Eur J Neurol* 15:1332-1337.
- Ruts L, Drenthen J, Jacobs BC, van Doorn PA, Dutch GBSSG (2010). Distinguishing acute-onset CIDP from fluctuating Guillain-Barre syndrome: a prospective study. *Neurology* 74:1680-1686.
- Sejvar JJ, Kohl KS, Gidudu J, Amato A, Bakshi N, Baxter R, Burwen DR, Cornblath DR, Cleerhout J, Edwards KM, Heininger U, Hughes R, Khuri-Bulos N, Korinthenberg R, Law BJ, Munro U, Maltezos HC, Nell P, Oleske J, Sparks R, Velentgas P, Vermeer P, Wiznitzer M, Brighton Collaboration GBSWG (2011). Guillain-Barre syndrome and Fisher syndrome: case definitions and guidelines for collection, analysis, and presentation of immunization safety data. *Vaccine* 29:599-612.
- Teunissen CE, Petzold A, Bennett JL, Berven FS, Brundin L, Comabella M, Franciotta D, Frederiksen JL, Fleming JO, Furlan R, Hintzen RQ, Hughes SG, Johnson MH, Krasulova E, Kuhle J, Magnone MC, Rajda C, Rejda K, Schmidt HK, van Pesch V, Waubant E, Wolf C, Giovannoni G, Hemmer B, Tumani H, Deisenhammer F (2009). A consensus protocol for the standardization of cerebrospinal fluid collection and biobanking. *Neurology* 73:1914-1922.
- Uncini A, Kuwabara S (2012). Electrodiagnostic criteria for Guillain-Barre syndrome: a critical revision and the need for an update. *Clin Neurophysiol* 123:1487-1495.
- van den Berg B, Walgaard C, Drenthen J, Fokke C, Jacobs BC, van Doorn PA (2014). Guillain-Barre syndrome: pathogenesis, diagnosis, treatment and prognosis. *Nat Rev Neurol* 10:469-482.
- van Koningsveld R, Steyerberg EW, Hughes RA, Swan AV, van Doorn PA, Jacobs BC (2007). A clinical prognostic scoring system for Guillain-Barre syndrome. *Lancet Neurol* 6:589-594.
- van Nes SI, Vanhoutte EK, Faber CG, Garssen M, van Doorn PA, Merkies IS, PeriNom SSG (2009). Improving fatigue assessment in immune-mediated neuropathies: the modified Rasch-built fatigue severity scale. *J Peripher Nerv Syst* 14:268-278.
- van Nes SI, Vanhoutte EK, van Doorn PA, Hermans M, Bakkers M, Kuitwaard K, Faber CG, Merkies IS (2011). Rasch-built Overall Disability Scale (R-ODS) for immune-mediated peripheral neuropathies. *Neurology* 76:337-345.
- Vanhoutte EK, Faber CG, van Nes SI, Jacobs BC, van Doorn PA, van Koningsveld R, Cornblath DR, van der Kooij AJ, Cats EA, van den Berg LH, Notermans NC, van der Pol WL, Hermans MC, van der Beek NA, Gorson KC, Eurlings M, Engelsman J, Boot H, Meijer RJ, Lauria G, Tennant A, Merkies IS, PeriNom SSG (2012). Modifying the Medical Research Council grading system through Rasch analyses. *Brain* 135:1639-1649.
- Vergouwe Y, Steyerberg EW, Eijkemans MJ, Habbema JD (2005). Substantial effective sample sizes were required for external validation studies of predictive logistic regression models. *J Clin Epidemiol* 58:475-483.
- Wakerley BR, Uncini A, Yuki N, Group GBSC, Group GBSC (2014). Guillain-Barre and Miller Fisher syndromes--new diagnostic classification. *Nat Rev Neurol* 10:537-544.
- Walgaard C, Lingsma HF, Ruts L, Drenthen J, van Koningsveld R, Garssen MJ, van Doorn PA, Steyerberg EW, Jacobs BC (2010). Prediction of respiratory insufficiency in Guillain-Barre syndrome. *Ann Neurol* 67:781-787.
- Walgaard C, Lingsma HF, Ruts L, van Doorn PA, Steyerberg EW, Jacobs BC (2011). Early recognition of poor prognosis in Guillain-Barre syndrome. *Neurology* 76:968-975.
- Walgaard C, Lingsma HF, van Doorn PA, van der Jagt M, Steyerberg EW, Jacobs BC (2016). Tracheostomy or Not: Prediction of Prolonged Mechanical Ventilation in Guillain-Barre Syndrome. *Neurocrit Care*.
- Willison HJ, Jacobs BC, van Doorn PA (2016). Guillain-Barre syndrome. *Lancet*.

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Figures

Figure 1. Time schedule of research protocol for the IGOS database and biobank.

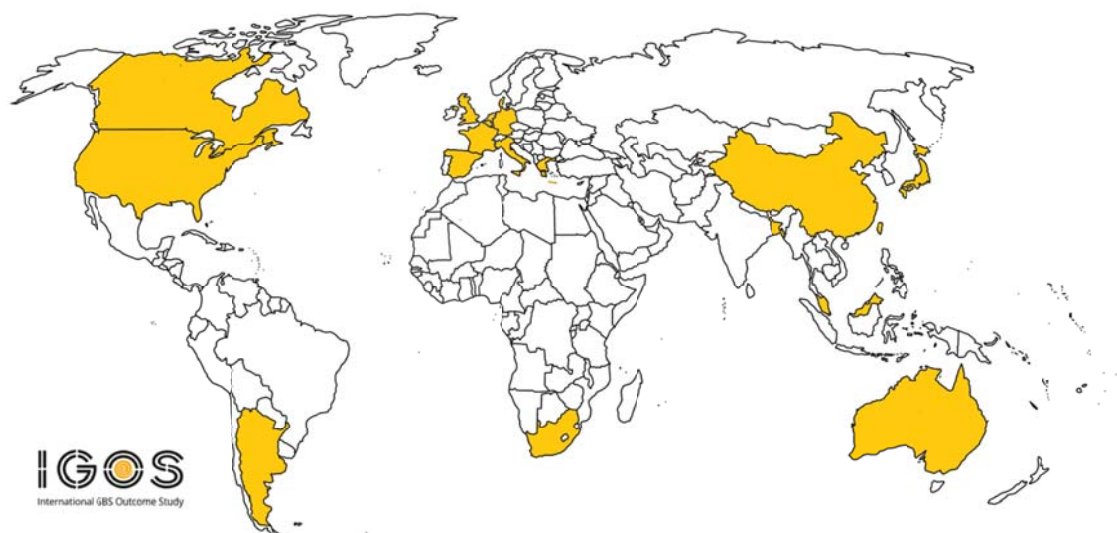


Legend figure 1.

The filled blocks refer to obligatory studies, the striped to optional substudies. The timeline represents the follow-up period after study entry in weeks. The first 2 weeks focus on collecting data and biomaterials to predict the clinical course and outcome in the period after 2 weeks. Blood samples are obtained as indicated for serial serological studies and for DNA extraction. Routine diagnostic electrophysiology will be conducted in the first or second week, and as an optional study at 4 weeks. * At 8 weeks and 13 weeks patients admitted at the hospital will have a full examination and serum sampling but discharged patients will have telephone assessment only and no serum sampling.

¹ Patient reported outcome measures (PROM) used are R-ODS, FSS, Rasch-FSS, EuroQol EQ-5D.

Figure 2. Countries with hospitals participating in IGOS.



Legend figure 2

Worldwide representation of IGOS in 19 countries (indicated in orange) including Argentina, Australia, Bangladesh, Belgium, Canada, China, Denmark, France, Germany, Greece, Italy, Japan, Malaysia, Netherlands, South Africa, Spain, Taiwan, United Kingdom and United States of America.

Figure 3. Quarterly number of patients included in IGOS.

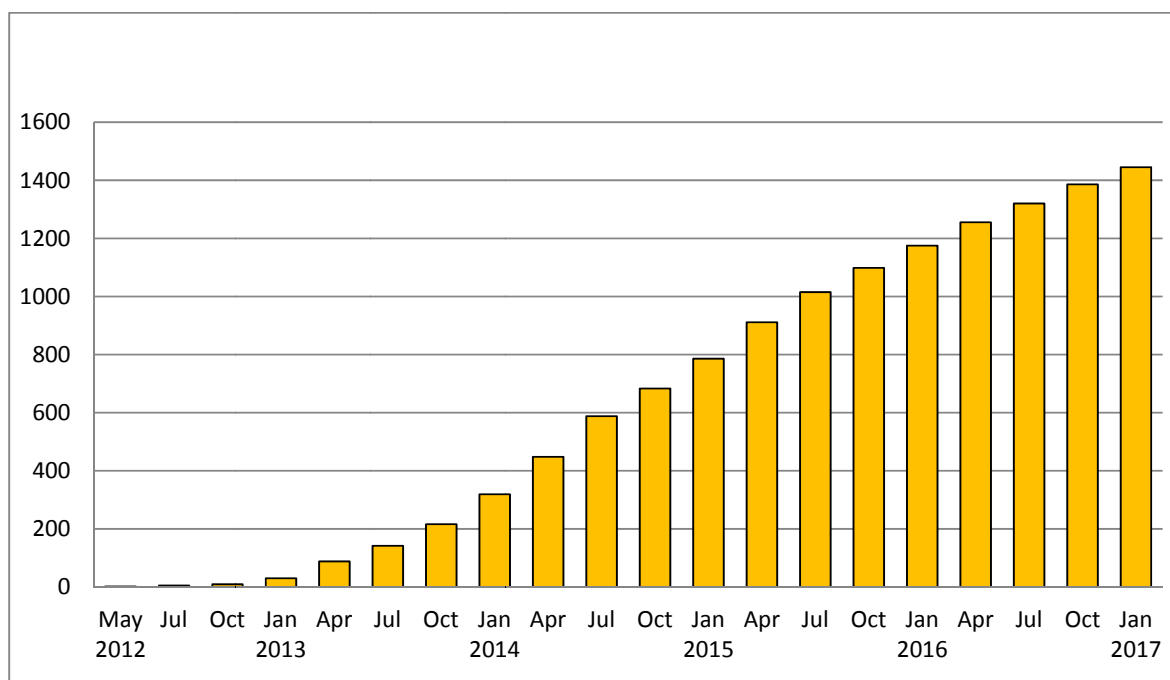


Figure 4. Logo for the International GBS Outcome Study (IGOS).



Appendix: Persons involved in IGOS Consortium

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Supplement Table 1: Diagnostic criteria for Guillain-Barré syndrome (GBS)

Features required for diagnosis

Progressive weakness in both arms and legs (might start with weakness only in the legs)
Areflexia (or decreased tendon reflexes)

Features that strongly support diagnosis

Progression of symptoms over days to 4 weeks
Relative symmetry of symptoms
Mild sensory symptoms or signs
Cranial nerve involvement, especially bilateral weakness of facial muscles
Autonomic dysfunction
Pain (often present)
High concentration of protein in CSF
Typical electrodiagnostic features

Features that should raise doubt about the diagnosis

Severe pulmonary dysfunction with limited limb weakness at onset
Severe sensory signs with limited weakness at onset
Bladder or bowel dysfunction at onset
Fever at onset
Sharp sensory level
Slow progression with limited weakness without respiratory involvement (consider subacute inflammatory demyelinating polyneuropathy or CIDP)
Marked persistent asymmetry of weakness
Persistent bladder or bowel dysfunction
Increased number of mononuclear cells in CSF ($>50 \times 10^6/L$)
Polymorphonuclear cells in CSF

Legend supplement Table 1

Adapted from Asbury and Cornblath et al. (*Asbury and Cornblath, 1990*)

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Motor nerve conductions studies

		Responsive (yes/no)	DML (ms)	Distal CMAP amplitude (baseline-peak) (mV)	Prox. CMAP ampl. I (baseline-peak) (mV)	Prox. CMAP ampl. II (baseline-peak) (mV)	NCV I (m/s)	NCV II (m/s)	Minimal F-wave latency(>10)(ms)	H-M lat (ms)
1.	Median nerve –APB (nd)	X	X	X	X		X		X	
2.	Ulnar nerve- ADM (nd)	X	X	X	X	X	X	X	X	
3.	Peroneal nerve-EDB (nd))	X	X	X	X	X	X	X	X	
4.	Other nerve*	X	X	X	X	X	X	X	X	
5.	H-reflex m. soleus (nd)	X								X

Sensory nerve conductions studies

		Responsive (yes/no)	DSL (ms)	Distal SNAP amplitude (baseline-peak) (μ V)	Prox. SNAP ampl. I (baseline-peak) (μ V)	Prox. SNAP ampl. II (baseline-peak)(μ V)	NCV I (m/s)	NCV II (m/s)	NCV III (m/s)
1.	Median nerve- dig II (nd)	X	X	X	X		X	X	
2.	Ulnar nerve- dig V (nd)	X	X	X	X	X	X	X	X
3.	Sural nerve (nd)	X	X	X			X	X	
4.	Optional radial nerve(nd)	X	X	X	X		X	X	

Optional needle EMG

		Fibrillations (yes/no)	Positive sharp waves (yes/no)	Polyphasic MU (yes/no)	Increased size MU (yes/no)	Decreased MU recruitment (yes/no)
1.	Dorsal Interosseus I muscle					
2.	Anterior tibial muscle					
3.	Proximal arm muscle (deltoid or biceps muscle)					
4.	Proximal leg muscle (vastus lateralis muscle)					

Legend supplement Table 2.

Local investigators are free to conduct the nerve conduction studies according to their local routine standards but recommended in IGOS is to perform a complete electrophysiological examination at two separate time points: the first within 7 days of admission or registration in IGOS, and the second at four weeks after admission or registration in IGOS. The collected data should include sensory studies in legs and arms (3-4 nerves), motor studies with F waves (3-4 nerves), tibial H-reflexes and EMG of a proximal and distal muscle in an arm and leg. Normative data and pictures of the waveforms will also be included in the report.

nd = non-dominant side

* = other nerve (median, ulnar or peroneal nerve on dominant-side or tibial nerve on dominant side) (*Uncini and Kuwabara, 2012*)