

1 **A first approach of using ultrasound as an alternative for blanching in vacuum-packaged potato strips**

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17 **ABSTRACT:** The effect of ultrasound (US) (40 kHz, 200 W, 3 min), blanching (85 °C, 3.5 min) and the
18 combination of both methods was evaluated on the quality of vacuum packaged potato strips stored at 3±1 °C
19 for up to 10 days. For this study two cultivars of potatoes were assessed. For blanched Agata samples, the
20 lightness (L*) decreased over 12 % ($p < 0.05$). Moreover, their hue increased up to 100, obtaining lesser yellow
21 potato strips. In contrast, US did not affect the hue values. The losses of firmness of blanched potato strips were
22 notable (35 % for Agata and 51 % for Agria), whereas US did not change this property ($p < 0.05$). Nevertheless,
23 no significant differences were found in the total starch content at 10 days. Agata and Agria showed different
24 metabolic behavior of sucrose in the refrigerated storage. Therefore Agria cultivar retained better colour after
25 frying. These results suggest that US had less impact in colour and improve the firmness in vacuum-packaged
26 potato strips with no added chemicals.

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30 **Keywords:** Colour, firmness, starch content, reducing sugars, vacuum packaging

31 **Introduction**

32 Minimal processing operations of vegetables and tubers can induce undesirable changes in the colour and
33 appearance during storage, caused mainly by enzymatic browning. This can accelerate the loss of quality and
34 reduce the product's shelf life (Tomás-Barberán and Espín 2001; Cantos et al. 2002).

35 Blanching is a method commonly used in minimally processed potatoes to prevent enzymatic browning by
36 inactivating polyphenol oxidase. Blanching promoted in potato slices more uniform colour after frying and the
37 layer of gelatinized starch formed, limited oil absorption and improved texture Moreira et al. (1999). Severini et
38 al. (2003) achieved the inactivation of polyphenol oxidase in potatoes through blanching treatments and the
39 combination with calcium chloride, which promoted increased lightness (L^*), probably due to its chelating
40 properties. Meanwhile, Alvarez et al. (2000) observed that blanching led to a loss of firmness and of other
41 quality attributes of the product, such as nutrients, flavor and colour.

42 To prevent the undesirable effects of blanching, ultrasound may be a viable method for non-thermal processing.
43 Acoustic cavitation of ultrasound can accelerate chemical reactions, increase diffusion rates, disperse
44 aggregates, inhibit enzymes and destroy microorganisms (Sala et al. 1995; Knorr et al. 2002). Zuo et al. (2012)
45 studied the behavior of high intensity low-frequency ultrasound (20 kHz) on potato starch granules and observed
46 that treatment by ultrasound resulted in the damage to the starch surface and in some cases in the shattering of
47 the granules. Furthermore, the increase in the number of defect was found to be linear with the sonication
48 power. Comandini et al. (2013) studied the application of ultrasound produced through a 35 kHz sonotrode
49 during immersion freezing of potatoes and concluded that the treatment affected important freezing parameters,
50 like the anticipation of nucleation and the reduction of freezing time. Karizaki et al. (2013) investigated the
51 possibility of using ultrasound-assisted osmotic dehydration as a pretreatment prior to frying and its effects on
52 the quality of fried potatoes. The authors concluded that the association with ultrasound reached the goal by
53 reducing oil and moisture content as compared to that of untreated ones.

54 For the authors' knowledge, few applications have been described of the use of ultrasound in the fresh vegetable
55 industry to retain food quality. Color and texture changes have to be taken into account regarding the quality of
56 potato strips as they are the two parameters that define their appearance. Furthermore, the effects of ultrasound
57 on some specific food components have not been sufficiently studied, so the actual effect of degradation due to
58 the use of this treatment is not yet fully understood (Mizrach 2008). For this purpose, this study aimed to
59 evaluate the effect of ultrasound compared to conventional blanching on the colour, pH, firmness, sugars and

60 starch content of vacuum-packed potato strips stored at 3 ± 1 °C for up to 10 days. Colour and firmness of the
61 fried potato strips were also assessed.

62 **Materials and Methods**

63 ***Materials***

64 Potatoes (*Solanum tuberosum* L.) of the Agata and Agria cultivars were acquired from Mercabarna (Mercados
65 de Abastecimientos de Barcelona SA), selected, washed in tap water to remove surface dirt, dried and stored in
66 the dark at 18 ± 2 °C overnight prior to processing. Agata presented values of dry matter of 18.40 ± 1.20 g·100g⁻¹
67 ¹ and Agria 20.95 ± 0.25 g·100g⁻¹.

68 ***Ultrasound equipment***

69 An ultrasound bath with 40 kHz of frequency and 200 W of generation power (JP-SELECTA 3000617,
70 Barcelona, Spain) was used to sonicate the potatoes. The machine was made of welded aluminum sheet, with a
71 capacity of 9 L, the dimensions were 12 cm x 46 cm x 12 cm (height x width x depth). The ultrasound bath had
72 four steel cone-shape transducers (45 mm/38 mm of diameter; 47 mm length). The experiment was conducted
73 in batch mode in non-refrigerated equipment. The operating conditions had previously been optimized and the
74 distribution of the ultrasound in the bath was uniform (Amaral et al. 2015). The specific heat and power of
75 dissipation were calculated and were 47234 kJ·kg⁻¹ and 157.45 W, respectively. The increase of the temperature
76 in the water was lesser than 1.5°C after 3 min of treatment.

77 ***Experimental procedure***

78 Potato tubers, free of defects, were hand-peeled, cut into rectangular strips with a cross-section of 10 x 10 mm
79 with a manual vegetable slicer and immediately rinsed in distilled water (1:4 w/w; potato:distilled water ratio).
80 The strips were then centrifuged in a manual centrifuge to eliminate excess water. For the treatment phase,
81 potato strips of each variety were separated into two portions: from the first portion, packs of 100 ± 5 g were
82 selected at random from the whole bunch and vacuum packed (-98 kPa, in a vacuum sealer VM-18 ORVED
83 S.p.A., Italy) in coextruded polyamide/high density polystyrene (Coex. PA/PEHD-70/150; thickness: 22 µm; O₂
84 transmission rate: 8 cm³/m² dbar at 25 °C) bags. The bags were then separated to receive either no further
85 treatment (control samples) or a 3-minute ultrasound treatment at 40 kHz. The second portion of the potato
86 strips was blanched in a non-agitated system (85 °C for 3.5 min, Pedreschi and Moyano 2005), samples were
87 manually shaken during blanching and the potato:water ratio was 1:4 (w/w). Blanched samples were cooled, for
88 five minutes at 22 ± 2 °C and centrifuged. They were also vacuum packed (-98 kPa) in PA/PEHD bags. Then,
89 the packages were divided into another two portions: one portion was subjected to sonication and the other

90 refrigerated. All samples were stored at 3 ± 1 °C for up to 10 days. Two replicates of 100 ± 5 g of vacuum-
91 packaged potatoes were assessed for each treatment and sampling date. For total starch content and total sugars,
92 samples were collected, immediately frozen at -20 °C and subsequently freeze-dried at -54 °C and 0.07 mbar
93 vacuums for 40 h by Telstar Cryodos -50 freeze dryer (1 KVA of potency, model 2G-6, Telstar, Barcelona,
94 Spain). Prior to frying, 1 L of olive oil (0.5°) was pre-heated in a deep-fat fryer. Samples were kept at 3 ± 1 °C
95 before frying. Then, 200 ± 10 g of each treatment of potato strips were fried at 180 °C (Taurus Professional
96 Compac2 fryer, Spain) for 5 minutes for Agata potatoes and 7 minutes for Agria potatoes. Frying times were
97 previously fixed according to the palatability of the fried strips. After frying, the strips were removed from the
98 oil, drained for one minute and then air-dried at 22 ± 2 °C for 10 minutes. The colour and firmness of the
99 samples were then assessed.

100

101 ***Physicochemical analysis***

102 Analyses were carried out on days 1, 6 and 10 for the vacuum-packaged and fried potatoes. *pH* was measured
103 in triplicate according to AOAC 981.12. *Colour* was determined with a colorimeter (Konica Minolta CR-400,
104 Japan), measuring $L^*a^*b^*$ parameters in the CIE Lab scale using a D_{65} light source and 10° as the observed
105 standard; results are expressed in L^* , Chroma ($C^* = ((a^*)^2 + (b^*)^2)^{1/2}$) and hue ($H^* = \tan^{-1}(a^*/b^*)$). Ten readings
106 were taken on two sites on the surface of the potato strips for each treatment (Oner and Walker 2011). *Firmness*
107 was measured using the texture analyzer (TA.XT Plus, Stable Micro Systems Co. Ltd., UK) equipped with a 30
108 kg load cell and connected with a Warner-Bratzler blade set with a speed of $1 \text{ mm}\cdot\text{s}^{-1}$. Twenty potato strips of
109 each treatment and day were analysed (Yang et al. 2016). Firmness was measured as the maximum shear
110 strength values and expressed as maximum force (N). *Total starch* analyses were conducted using 100 ± 0.1 mg
111 of lyophilized sample according to AOAC 996.11 (amyloglucosidase/ α -amylase method) and AACC 76.13
112 procedures. D-glucose was oxidised to D-gluconate, which was quantitatively measured from the absorbance at
113 510 nm of a colorimetric reactant. The results were expressed by $\text{g}\cdot 100 \text{ g}^{-1}$ of lyophilized weight (LW). These
114 determinations were made in triplicate. *Sugars* were extracted and measured as described by (López-Hernández
115 et al. 1998) with slight modifications. 2g of lyophilized sample were extracted by refluxing for 30 min with 70%
116 ethanol. The extract was vacuum-filtered and filled to 25 mL with 70 % ethanol. A 5 mL aliquot of the solution
117 was passed through a Waters Sep-Pak C column, filtered ($0.45 \mu\text{m}$ pore size membrane), and injected into the
118 HPLC Hewlett Packard series 1100, equipped with a Beckman 110B injector and a Beckman Refraction Index
119 Detector (RID). The fructose, glucose and sucrose separation was performed using a Phenomenex Luna column

120 at 28 °C using acetonitrile-water (78:22 v/v). The average of the results of three replications were expressed by
121 $\text{g} \cdot 100\text{g}^{-1}$ of LW.

122 ***Statistical analysis***

123 The statistical study of variations after application of treatments and during storage was carried out using two-
124 way ANOVA using Minitab (v.16, MINITAB Inc, State College, PA) at a 95% confidence level. ANOVA was
125 carried out for each cultivar independently. The differences between samples were determined using Tukey's
126 least significant difference test.

127

128 **Results and Discussion**

129

130 ***Effect of the treatments on pH of the potato strips***

131 For Agata and Agria, the interaction between both factors (treatment and storage day) was significant for pH
132 values (data not shown). Both heat-treated Agata and Agria samples (blanched and blanched+US) had
133 significantly higher pH values (6.09-6.11 and 6.48-6.10, respectively) than the control (5.77-5.88) and US
134 samples (5.72 -5.89) at the beginning of the experiment ($p < 0.05$). In general, a statistically significant decrease
135 of pH (≤ 0.2 pH units) was observed in both cultivars and treatments at the end of 10 storage days at 3 ± 1 °C.
136 Only, the pH of the blanched treatment for Agata was not significant different than samples analysed at the
137 beginning of the experiment. Our previous study about the effect of different times of application of ultrasound
138 (40 kHz, 200 W) only observed alterations in pH after 5 minutes of sonication (Amaral et al. 2015).

139

140 ***Effect of treatment on carbohydrates: sugars and starch content of potato strips***

141 ***Total starch***

142 The content of total starch was $60.65 \pm 2.77 \text{ g} \cdot 100\text{g}^{-1}$ LW and $65.46 \pm 1.60 \text{ g} \cdot 100\text{g}^{-1}$ LW for Agata and Agria
143 cultivar, respectively. No significant differences were observed in the total starch content in either blanched or
144 control samples ($p > 0.05$) after one day of storage at 3 ± 1 °C in the two cultivars studied (data not shown).
145 Wang, Zhang and Mujumdar (2010) did not find significant differences either between unprocessed and
146 blanched potatoes (in boiling water for 5 min); although a slight decrease was noticed due to leaching losses.
147 Ultrasound affected the total starch content of the potato strips and promoted its reduction, but no significant
148 differences were found compared to the control and blanched samples at the end of storage. According to
149 Jambrak et al. (2010), ultrasound causes changes in the starch granule, such as a decrease in size and,

150 consequently, alterations in the physical-chemical properties of starch. Ultrasound treatment fosters the damage
151 and ruptures of the starch granules and distorts the crystalline region prior to a reversible hydration of the
152 amorphous phase. The samples treated with the combination of blanching and ultrasound had a slightly higher
153 content of total starch, but no significant differences were detected with the control and blanched potato strips
154 ($p > 0.05$). At 10 days, for both of the cultivars assessed, the potato strips did not show changes in their major
155 nutrient.

156 *Sugars*

157 Agata showed higher concentrations of fructose, glucose and sucrose (Figure 1) compared with Agria. This
158 information differs than that found by Uri et al. (2014) that observed the French fry cultivars have greater
159 amounts of fructose and glucose. In the refrigerated storage the results suggest a different metabolic behaviour
160 of sucrose for each cultivar, since Agata accumulates sucrose while the two monosaccharide from which it is
161 composed, reduced their quantities in contrast to Agria that presented reduction of the content of sucrose.
162 Folgado et al. (2014) pointed that the different behaviour of sucrose metabolism is due to the cold-acclimation
163 of each species of tuber. Although Park et al. (2009) stated that sucrose is a substrate for starch formation; in
164 this work there was no connection with the accumulation of this sugar and the total starch content.

165 The application of treatments decreased significantly ($p < 0.05$) the content of fructose and glucose of Agata
166 potato strips the first day of storage (Figure 1a and 1b). After 10 days, there was a decrease of the content of the
167 three sugars analysed for blanched samples ($p < 0.05$). In contrast, samples of the other treatments increased
168 significantly the content of glucose and fructose. For Agria samples it was observed a slightly increase of the
169 three sugars analysed ($p < 0.05$) at the beginning of the experiment (Figure 1d, 1e and 1f). Moreover, a negative
170 correlation between sucrose and day of storage ($r = -0.615$; $p < 0.01$; $n=12$) and positive correlation between
171 fructose and glucose ($r = 0.956$; $p < 0.01$; $n=12$) were found.

172

173 ***Effect of the treatments on Colour and firmness on vacuum packaged and fried potato strips***

174 *Colour*

175 For Agata vacuum-packaged potato strips, the interaction between both factors (treatment and storage day) was
176 significant for all parameters ($p < 0.05$). On the other hand, for Agria the interaction between treatment and
177 storage day was not significant for Chroma and Hue. In fried strips, the effects of treatment and time of storage
178 were significant for all parameters and the two varieties analysed, excepted for the parameter Chroma (data not
179 shown). For this parameter there were significant differences only for time of storage in Agria ($p < 0.05$).

180 The treated Agata potato strips showed a significant decrease ($p < 0.05$) in *lightness* (L^*), at one day of storage
181 but Agria potato strips were not affected (Figure 2a and 2c) It is important to remember that in potato strips, the
182 higher the L^* value, the lighter the colour (Cantos et al. 2002). Furthermore, both blanched samples showed
183 significantly lower values than the ultrasound-treated ones. These results are in agreement with Oner and
184 Walker (2011), who noted that unprocessed potato strips were lighter in colour than blanched and fresh-cut
185 strips. Enzymatic oxidation occurring before the denaturation of the polyphenol oxidase (PPO) in the blanched
186 samples may be the cause of their darker colour. According to our results, at the end of storage the blanching
187 operation leads to increase browning ($p > 0.05$) in potato strips, but in Agria this effect is lesser than Agata.
188 Nevertheless, the blanched Agria potato strips were darker than the ultrasound-treated and control potato strips
189 after 10 days of storage ($p < 0.05$). The control and ultrasound samples had lower *hue* (H^*) values for both of
190 the cultivars assessed (Figure 2b and 2d) ($p < 0.05$). The Agata samples had a higher H^* value than the Agria
191 samples after one day of storage at 3 ± 1 °C (100.12, 97.82, respectively) and this trend is maintained during
192 storage time for blanching treatments. Cabezas-Serrano et al. (2009) also found lower L^* values for Agata, but
193 the H^* value for this cultivar were higher than in Marabel, Agria and Almera. Our results are consistent with
194 those findings; a higher H^* value indicates that the potato flesh is less yellow. The H^* values of the blanched
195 Agata samples increased significantly after 10 days of storage, whereas the Agria samples did not change. This
196 may indicate that the colour of the blanched Agata potato became less attractive over time.

197 In general, all of the samples of the Agata cultivar displayed a sharper depletion in L^* values (47.9% for control
198 potato strips) after frying than the Agria samples (Figures 2e, 2g). Ultrasound-treated Agria potato strips had the
199 highest L^* value (71.68), but no significant differences were found between blanched and control samples ($p >$
200 0.05). For the Agata cultivar, the lightness of the control and ultrasound-treated samples improved over time (p
201 < 0.05), whereas blanched samples became slightly darker at the end of storage ($p > 0.05$). Scores under this
202 value indicate a darker, brownish colour for fried potato strips (Oner and Walker 2011). Consequently, the
203 Agata cultivar seems highly suitable for use as a fresh-cut product, but the colour after frying may be considered
204 less attractive to consumers. Its darker colour after frying may be due to the compounds obtained after the
205 Maillard reaction, which is determined by the superficial reducing sugar and amino acid content. However, it is
206 important to note that for the Agata cultivar, the lightness of the control and ultrasound-treated samples
207 improved over time ($p < 0.05$). The H^* values follow the same pattern as the lightness values for the Agata
208 cultivar and confirm their brownish colour (Figure 2f). At the end of storage, blanched samples had the lowest
209 value ($p < 0.05$). As is shown in Figure 2h, the treatments did not have an impact on the H^* angle of the fried

210 Agria potato strips at the beginning of the experiment ($p > 0.05$). Despite the H^* value of the vacuum-packaged
211 potato strips, after frying, values were around 90°(yellow) for all the treatments as well as the control.

212 *Firmness*

213 There was a significant decrease in *firmness* in both of the blanched potato strips: 35% for Agata and over 51%
214 for Agria, indicating a remarkable softening of the product due to the heat treatment (Table 1). Starch
215 gelatinization weakens the structure of potato parenchyma which occurs when strips are blanched at high
216 temperatures (over 70 °C); meanwhile, the intensity of softening depends on both temperature and time (Liu and
217 Scanlon 2007). Pedreschi and Moyano (2005) reported that blanching induced a reduction of almost 29% in the
218 maximum force of raw slices of potato. Ultrasound treatment did not change the firmness of either the Agata or
219 the Agria potato strips compared to the control over time.

220 There were no significant differences in maximum force on fried potato strips on day 1 ($p < 0.05$) but,
221 comparing with the vacuum-packaged samples, the control and sonicated potato strips underwent the greatest
222 losses in firmness after frying (>70%). Nevertheless, the Agria fried potato strips were firmer than the Agata
223 ones (Table 1). According to Nourian and Ramaswamy (2003), within five minutes of frying time, potato strips
224 lose more than 80% of their original texture. This is due to the combination effect of cell rupture, solubilisation
225 of the middle lamellae and the gelatinization of starch. Oner and Walker (2011) obtained a greater loss of
226 firmness in unprocessed potato strips (84%) than in blanched ones (35-47%) for the Russet Burbank cultivar.
227 This trend was similar to both potato cultivars assessed in this work. In contrast, Pedreschi et al. (2009) did not
228 find a significant effect on the texture of fried potatoes that had previously been blanched. During refrigerated
229 storage, the control and blanched samples of the Agata cultivar had significantly higher values than the
230 ultrasound-treated and blanched and sonicated samples. The greatest firmness was found on day 10 for the
231 blanched potato strips, which increased significantly over time. In the Agria fried samples, the values were
232 higher for the sonicated potato strips at the end of storage at 3 ± 1 °C.

233

234 **Conclusions**

235 The use of ultrasound indicated that the firmness, colour and pH of sonicated vacuum-packaged potato strips
236 were better than blanched. The combination of both techniques (blanched + US) didn't improve the studied
237 parameters. The colour of the fried Agria potato strips is better than the Agata, however, the use of ultrasound
238 led to the improvement in the lightness of fried Agata potato strips during refrigerated storage. Sugar content
239 showed different results after exposition to ultrasound for both cultivars analysed. Therefore, ultrasound can be

240 considered by the potato industry as an option to reduce the impact in colour and the loss of firmness of
241 vacuum-packaged potato strips with no added chemicals, although further research is needed to understand other
242 important aspects such as the maximum shelf life of this product and the consequences on composition and
243 sensory quality of the fried product.

244

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311

312 **Figure caption**

313 **Fig 1** Fructose, Glucose and saccharose contents for vacuum-packaged Agata (a, b and c) and Agria (d, e and f)
314 potato strips stored at 3 ± 1 °C. Values are the average \pm standard deviation (n=3). Different capital letters in the
315 same parameter and day of storage indicate significant differences. Different lower case letters in the same
316 parameter and treatment indicate significant differences ($p < 0.05$)

317 (□) control, (▤) blanching, (▥) ultrasound and (▦) blanching + ultrasound

318

319 **Fig 2** Changes in L^* and H^* for vacuum-packaged Agata (a, b) and Agria (c, d) and fried Agata (e, f) and Agria
320 (g, h) potato strips stored at 3 ± 1 °C. Values are the average \pm standard deviation (n=20). Different capital letters
321 in the same parameter and day of storage indicate significant differences. Different lower case letters in the
322 same parameter and treatment indicate significant differences ($p < 0.05$)

323 (□) control, (▤) blanching, (▥) ultrasound and (▦) blanching + ultrasound

324

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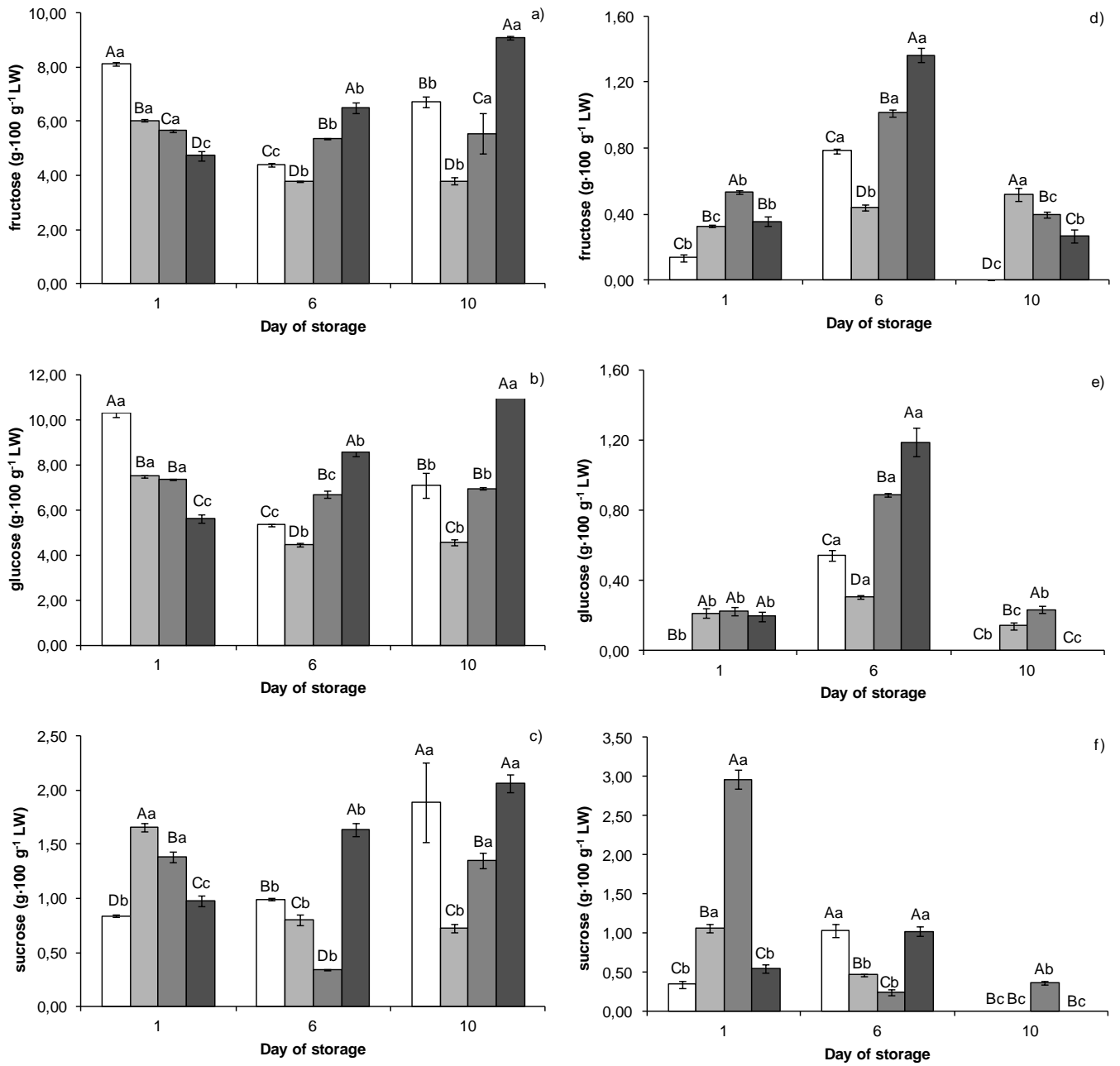


Fig. 1

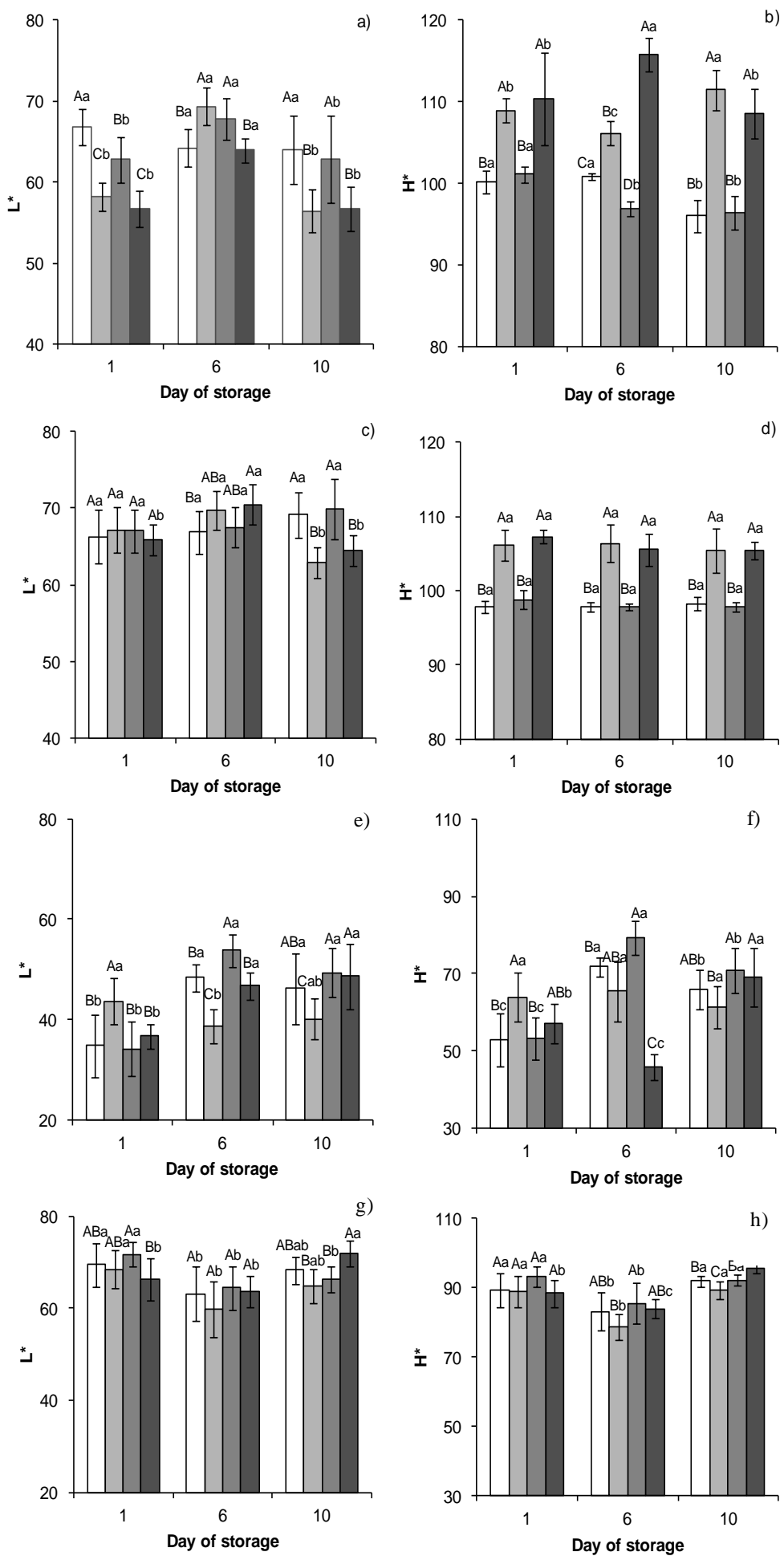


Fig. 2

Table 1. Firmness (N) of vacuum-packaged and fried Agata and Agria potato strips stored at 3 ± 1 °C

Treatment		Storage time (day)		
		1	6	10
Vacuum-packaged	Control	16.02 ± 2.89 Aa	16.45 ± 1.85 Ba	16.95 ± 1.95 Aa
	Blanching	10.44 ± 2.79 Ba	10.35 ± 2.86 Ca	11.02 ± 3.48 Ba
	US	15.21 ± 2.12 Ab	23.31 ± 5.80 Aa	17.40 ± 1.68 Ab
	Blanching + US	9.26 ± 2.86 Ba	9.20 ± 3.30 Ca	10.44 ± 3.85 Ba
Agata	Control	21.21 ± 3.37 Aa	23.59 ± 2.18 Aa	20.43 ± 4.73 Aa
	Blanching	10.33 ± 2.31 Ba	12.09 ± 1.94 Ca	10.90 ± 4.20 Ba
	US	23.54 ± 3.88 Aa	20.34 ± 2.43 Bb	22.74 ± 2.29 Aab
	Blanching + US	12.43 ± 4.00 Ba	10.46 ± 1.62 Ca	10.68 ± 2.00 Ba
Fried	Control	3.67 ± 1.20 Ab	4.95 ± 1.23 Aa	4.99 ± 1.16 ABa
	Blanching	4.13 ± 1.16 Ab	4.72 ± 0.98 Ab	6.55 ± 1.13 Aa
	US	3.82 ± 1.15 Aa	3.50 ± 1.04 Ba	4.68 ± 1.85 Ba
	Blanching + US	3.65 ± 0.99 Ab	3.37 ± 0.88 Bb	4.82 ± 1.20 Ba
Agria	Control	5.41 ± 1.95 Ab	6.59 ± 1.53 Bab	8.08 ± 2.51 Ba
	Blanching	5.51 ± 0.85 Ab	8.14 ± 2.27 ABa	6.74 ± 1.23 BCab
	US	5.15 ± 1.85 Ab	10.58 ± 4.00 Aa	10.40 ± 3.30 Aa
	Blanching + US	5.91 ± 1.68 Aa	6.39 ± 1.74 Ba	5.31 ± 1.86 Ca

Values are the average ± standard deviation (n=20). Different capital letters in the same parameter and day of storage indicate significant differences. Different lower case letters in the same parameter and treatment indicate significant differences ($p < 0.05$).