

---

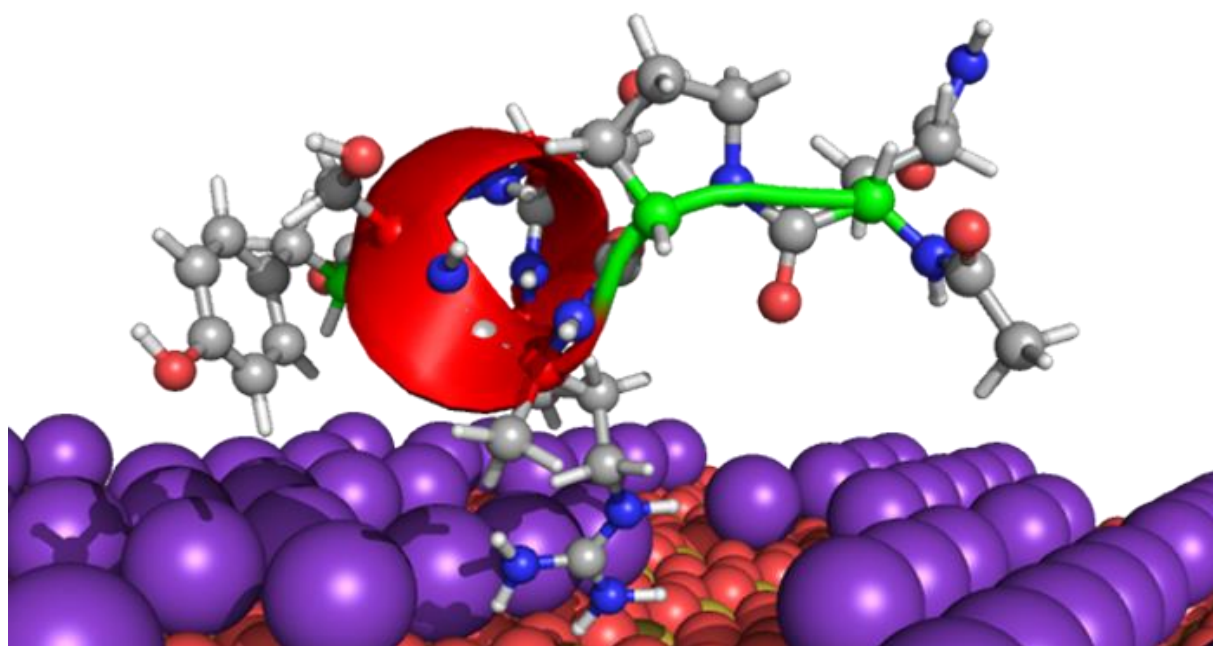
2<sup>nd</sup> International Conference on  
Peptide Materials for Biomedicine and  
Nanotechnology

**PepMat 2016**

Barcelona, 14-16 March 2016

**BOOK OF ABSTRACTS**

---



UNIVERSITAT POLITÈCNICA  
DE CATALUNYA  
BARCELONATECH



mecwins

**B|BRAUN**  
SHARING EXPERTISE



---

## INDEX

### ORAL PRESENTATIONS

OP-1	Activation or suppression of immune system with using peptide assemblies. S. Kimura	6
OP-2	Interactions between peptides and inorganic matter: from basic science to applications. M. Reches	7
OP-3	Peptide assemblies by design. V. Conticello	8
OP-4	$\beta$ -sheet forming peptides: from self-assembly to functional biomaterials. A. Saiani	9
OP-5	Folding and self-assembly of peptides using OPEP coarse-grained simulations. P. Derreumaux	10
OP-6	Peptide-based self-assembled hybrid materials. A. Moretto	11
OP-7	Self-assembly of phenylalanine oligopeptides. M.M. Pérez-Madrugal	12
OP-8	Peptide control of surface biomineralization. T. Weidner	13
OP-9	Structure-Function relationships in self-assembled peptide nanostructures: the case of charge transport. N. Ashkenasy	14
OP-10	Self-assembly of diphenylalanine backbone homologues and their combination with carbon nanotubes. D. Bhimareddy	15
OP-11	Hydrogen bonding and lipid bilayer mediated aggregation of various helix topologies. U. Diederichsen	16
OP-12	Peptides and peptide conjugates: from self-assembly towards applications in biomedicine. I.W. Hamley	17
OP-13	Self-assembling protein nanoparticles for targeting in colorectal cancer. A. Villaverde	18
OP-14	Short peptide supramolecular hydrogels for crystal growth. L. Álvarez de Cienfuegos	19

---

---

OP-15	Peptides as protein-surface interactors. E. Giralt	20
OP-16	Peptide nanofibers for diagnostic and therapeutic applications. G. Morelli	22
OP-17	Computational studies on peptides in drug design: insights from molecular dynamics simulations. A. Bidon-Chanal	23
OP-18	Intramolecular backbone...backbone hydrogen bonds in peptide conformations: the other way around. C. Toniolo	24
OP-19	Bioinspired visible fluorescent nanolabels for photonic and medical applications. G. Rosenman	25
OP-20	Mecwins: developing biomedicine applications using nanomechanical sensors. O. Ahumada	26
OP-21	Controlling multicomponent dipeptide hydrogels. D.J. Adams	28
OP-22	Drug delivery via cell membrane fusion using lipopeptide modified liposomes. A. Boyle	29
OP-23	Nanoscale molecular junctions incorporating peptides and peptidomimetics. S. Sek	30
OP-24	Perspectives for stimuli-responsive peptide: from self-assembly to biomolecular spring. G.M.L. Messina	31
OP-25	Ultrashort self-assembled peptides for biomaterial applications. G. Laverty	32
OP-26	Molecular dynamics simulations of intermolecular interactions involving peptides. G. Bocchinfuso	33
OP-27	Peptide aggregation in 2D and 3D. M. Venanzi	34
OP-28	Recombinamer-based injectable artificial ECM. J.C. Rodríguez-Cabello	35

---

---

## POSTER PRESENTATIONS

P-1	Crystal structures of the halogenated variants of the amyloidogenic core sequences DFNKF and KLVFF. G. Cavallo	37
P-2	Bacterial resistance to silver: the role of SilE protein. V. Chabert	38
P-3	Aqueous self-assembly of short hydrophobic peptides containing $\alpha,\alpha$ -disubstituted amino acids F. Clerici	39
P-4	Gel synthesis of iron oxides nanoparticles. R. Contreras-Montoya	40
P-5	Self-assembly of PEGylated tetra-phenylalanine derivatives: structural insights from solution and solid state studies. C. Diaferia	41
P-6	Aligning perylene bisimides for enhanced photoconductive properties. E. Draper	42
P-7	HIV-1 peptide inhibitor derived from the E2 protein of GB virus C conjugated to nanosystems. M.J. Gómara	43
P-8	Microarrays based on synthetic chimeric fibrin/filaggrin peptides for the diagnosis of rheumatoid arthritis. I. Haro	44
P-9	Structural chameleonic behavior in peptides derived from choline binding repeats. M.A. Jiménez	45
P-10	Halogenation promotes the self-assembly and gelation properties of DFNKF peptides L. Lascialfari	46
P-11	Protein nanomaterials from renewable sources. C. Lendel	47
P-12	Injectable peptide hydrogels for controlled-release. C. Martin	48
P-13	The evolution of surface-coating molecules in biomaterials science: integrin-binding peptidomimetics and multifunctional platforms. C. Mas-Moruno	49
P-14	Hierarchical surface-mediated self-assembly of triphenylalanine with N- and C- terminal fluorenylmethyl groups. E. Mayans	50
P-15	Halogen bonding: A new supramolecular tool in the molecular self-assembly of peptides P. Metrangolo	51

---

---

P-16	Ultrashort peptides as bifunctional nanomaterials. A.P. McCloskey	52
P-17	$\beta^{2,3}$ diaryl amino acids as molecular tools for the preparation of peptide nanotubes. S. Pellegrino	53
P-18	Elastin-like peptide nanoparticles for crossing the blood-brain barrier. J. Pille	54
P-19	Tuning Amyloid Peptide Self-assembly by Halogenation. A. Pizzi	55
P-20	Fibrin association at hybrid biointerfaces made of clot-binding peptides and polythiophene. A. Puiggali-Jou	56
P-21	Engineering of single-chain polypeptides to self-assemble as BBB-crossing protein nanoparticles. N. Serna	57
P-22	Electroactive polymer modified with RGD-derived peptide for cell adhesion. J. Triguero	59
P-23	Solution-state NMR to understand peptide gel fibre surface chemistry and gelation M. Wallace	60
P-24	Probing interactions in $\beta$ -sheet peptide / graphene derivatives hybrid hydrogels. J. Wychowaniec	61
P-25	Bifunctional Surfaces: A Peptide-Based Coating that Resists Fouling and Promotes Cell Adhesion. S. Yuran	62
P-26	Self-assembling peptide hydrogels cross-linked by disulfide bonds. A. Zamuner	63
P-27	Analysis of gold-tethered cationic RPAR peptide. A. Flores	64
P-28	Self-assembled monolayers of redox peptides. E.M. Cilli	65
P-29	Functional Peptide Assemblies on Surfaces. S. Nir	66
AUTHOR INDEX		67
PEOPLE ATTENDING THE MEETING		73

---

---

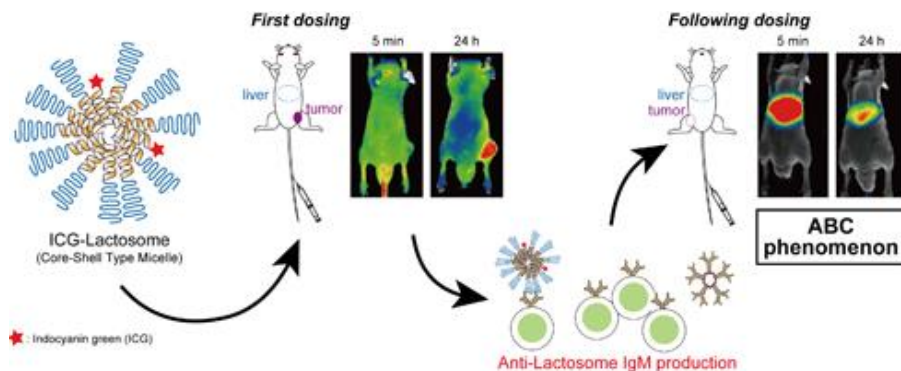
# Activation or Suppression of Immune System with Using Peptide Assemblies

Shunsaku Kimura

Department of Material Chemistry, Graduate School of Engineering, Kyoto University

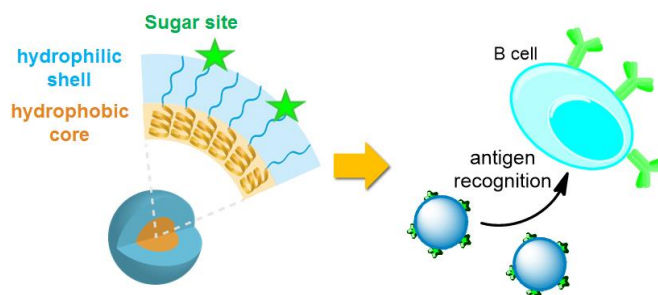
Nanoparticles are expected to be applicable for the theranostics medicine as a carrier vehicle of the diagnostic and therapeutic agents. We have prepared nanoparticles in a wide range of size, morphology, and surface character. As a result, we found that immune system is deeply involved in the *in vivo* disposition of nanoparticles upon multiple administration. The ideas how to suppress or activate the immune system with modifying nanoparticles will be shown<sup>1,2</sup>.

## Alteration of *in vivo* Disposition of Polymeric Micelle



Scheme 1 Schematic illustration of the accelerated blood clearance of polymeric micelles.

## Activation of Immune System with Raising anti-Lewis<sup>y</sup> IgM Production



Scheme 2 Schematic illustration of triggering the immune system against Lewis<sup>y</sup>.

<sup>1</sup> Hara E. et al., *J. Pept. Sci.* **2014**, 20, 570-577.

<sup>2</sup> Hara, E. et al., *ACS Med. Chem. Lett.* **2014**, 5, 873-877.



# Interactions between Peptides and Inorganic Matter: From Basic Science to Applications

*Meital Reches*

*The Institute of Chemistry and The Center for Nanoscience & Nanotechnology, The Hebrew University of Jerusalem*

Several natural processes are mediated by the interactions between proteins and peptides with inorganic materials. The immune response towards an implant is mediated by proteins. Composite materials are formed by the interactions of organic materials (usually proteins) and minerals. Biofouling, the process in which organisms attached to surfaces, is also mediated mainly by proteins. Moreover, the ability to anchor peptides to inorganic surfaces is important for many areas of research. This includes the development of biosensors, coatings and thin films. Understanding the nature of interactions between peptides and proteins with inorganic materials will bring to the development of improved implants, new composites, antifouling materials and functional surfaces.

Using single-molecule force spectroscopy with AFM, we study the underlying forces between individual biomolecules (either amino acid residues or short peptides) and inorganic surfaces in aqueous solution. Using this method, we were able to measure low adhesion forces and could clearly determine the strength of interactions between individual amino acid residues and inorganic substrates.<sup>1</sup> Our results with peptides also shed light on the factors that control the interactions at the organic-inorganic interface.<sup>2</sup>

Based on our knowledge from single molecule experiments, we designed a tripeptide that can spontaneously form a coating that resists biofilm formation. Our results demonstrate the formation of a coating on various surfaces (glass, titanium, silicon oxide, metals and polymers). In addition, we showed that this coating prevents the first step of antifouling, which involves the adsorption of bioorganic molecules to the substrate. Moreover, the coating significantly reduced the attachment of various organisms to surfaces. This includes pathogenic bacteria and fungi.<sup>3</sup>

---

<sup>1</sup> Razvag, Y., Gutkin, V. & Reches, M. (2013) *Langmuir* 29, 10102–10109

<sup>2</sup> Maity, S., Zanuy, D., Razvag, Y., Aleman, C. & Reches, M. (2015) *Phys. Chem. Chem. Phys.* 17, 15305-15315.

<sup>3</sup> Maity, S., Nir, S., Zada, T., Reches, M. (2014). *Chem. Commun.* 50, 11154-11157

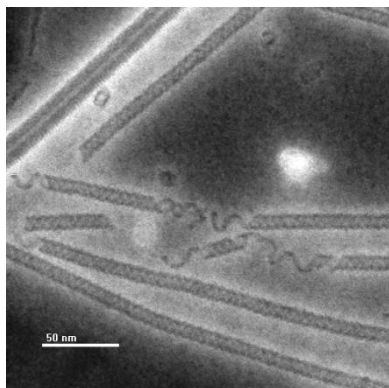
# Peptide Assemblies by Design

Charles Modlin, Spencer Hughes, Tao Jiang, Elizabeth Magnotti,

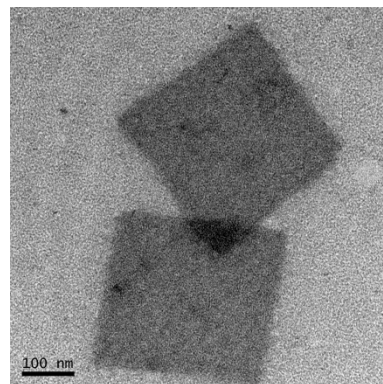
Vincent Conticello

Department of Chemistry, Emory University, Atlanta, GA 30322 U.S.A.

Structurally defined materials on the nanometer length-scale have been historically the most challenging to rationally construct and the most difficult to structurally analyze. Sequence-specific biomolecules, such as proteins, have advantages as design elements for construction of these types of nano-scale materials in that correlations can be drawn between sequence and higher order structure, potentially affording ordered assemblies in which functional properties can be controlled through the progression of structural hierarchy encoded at the molecular level. However, the predictable design of self-assembled structures requires precise structural control of the interfaces between peptide subunits (protomers). In contrast to the robustness of protein tertiary structure, quaternary structure has been postulated to be labile with respect to mutagenesis of residues located at the protein-protein interface. We have employed simple self-assembling peptide systems to interrogate the concept of designability of interfaces within the structural context of nanotubes and nanosheets (see below).<sup>1,2</sup> These peptide systems provide significant evidence that the designability of protein interfaces is a critical consideration for control of supramolecular structure in self-assembling systems.



**Figure 1.** STEM image of nanotubes derived from self-assembly of a *de novo* designed, bi-faceted coiled-coil peptide.



**Figure 2.** TEM image of two-dimensional nano-sheets derived from self-assembly of charge-complementary collagen peptides.

<sup>1</sup> Egelman, E.; Xu, C.; DiMaio, F.; Magnotti, E.; Modlin, C.; Yu, X.; Wright, E.R.; Baker, D.; Conticello, V.P. *Structure* **2015**, *23*, 280-9. <sup>2</sup> Jiang, T.; Xu, C.; Zuo, X.; Conticello, V.P. *Angew. Chemie Int. Ed. Engl.* **2014**, *53*, 8367-71





# **$\beta$ -sheet forming peptides: from self-assembly to functional biomaterials**

*Aline F. Miller<sup>1</sup> and Alberto Saiani<sup>2</sup>*

<sup>1</sup>*The University of Manchester, School of Chem. Eng. and Institute of Biotechnology*

<sup>1</sup>*The University of Manchester, School of Materials and Institute of Biotechnology*

*Polymers and Peptides Research Group*

[www.polymersandpeptides.co.uk](http://www.polymersandpeptides.co.uk)

The use of non-covalent self-assembly to construct materials has become a prominent strategy in material science offering practical routes for the construction of increasingly functional materials for a variety of applications ranging from electronic to biotechnology. A variety of molecular building blocks can be used for this purpose, one such block that has attracted considerable attention in the last 20 years is de-novo designed peptides. Peptides offer a number of advantages to the material scientists. The library of 20 natural amino acids offers the ability to play with the intrinsic properties of the peptide such as structure, hydrophobicity, charge and functionality allowing the design of materials with a wide range of properties. Synthetic peptides are chemically fully defined and easy to purify through standard processes. Being build from natural amino acids they result usually in low toxicity and low immune response when used in-vivo and can be degraded and metabolised by the body. In the past 10 years our group has focussed on the development of a technological platform for the design of novel biofunctional materials, in particular hydrogels, exploiting the self-assembly of  $\beta$ -sheet forming peptides. The  $\beta$ -sheet motif is of increasing interest as short peptides can be designed to form  $\beta$ -sheet rich fibres that entangle and consequently form hydrogels. These hydrogels can be functionalised using specific biological signals and can also be made responsive through the use of enzymatic catalysis and/or conjugation with responsive polymers. Through the fundamental understanding of the self-assembly of these peptides we have been able to design hydrogels with tailored properties for a range of applications including injectable systems for in-vivo cell delivery and sprayable systems for topical drug delivery.



# Folding and self-assembly of peptides using OPEP coarse-grained simulations

*P. Derreumaux*

*Laboratory of Theoretical Biochemistry, UPR9080 CNRS, University of Paris 7 – Sorbonne Cité Centre, PSL, IBPC, rue Pierre et Marie Curie, 75005, Paris, France*

I shall present where OPEP coarse-grained simulations coupled to advanced sampling techniques stand in the context of fast and accurate peptide structure determination, and the self-assembly of amyloid proteins associated with Alzheimer's disease.<sup>1,2</sup>

The understanding of protein/RNA behavior in cell-like environments from computer simulations is still a challenge because of the size and the spread of length- and time-scales characterizing the systems, and I will show where we are in this context.

- 
1. Nasica-Labouze J, Nguyen PH, Sterpone F, Berthoumieu O, Buchete NV, Coté S, De Simone A, Doig AJ, Faller P, Garcia A, Laio A, Li MS, Melchionna S, Mousseau N, Mu Y, Paravastu A, Pasquali S, Rosenman DJ, Strodel B, Tarus B, Viles JH, Zhang T, Wang C, **Derreumaux P**. Chem Rev. 2015 May 13;115(9):3518-63.
  2. Sterpone F, Melchionna S, Tuffery P, Pasquali S, Mousseau N, Cragolini T, Chebaro Y, St-Pierre JF, Kalimeri M, Barducci A, Laurin Y, Tek A, Baaden M, Nguyen PH, **Derreumaux P**. Chem Soc Rev. 2014 Jul 7;43(13):4871-93.

# Peptide-based Self-assembled Hybrid Materials

A. Moretto

University of Padova , Department of Chemical Science,  
35131 – Padova, Italy

Since J.M. Lehn's pioneering contributions, the bottom-up self-assembly strategy has emerged as one of the most applicable approaches to drive small molecule arrangements. Following a hierarchical process, under appropriate conditions the combination of a variety of non-covalent interactions may allow properly designed molecules to form stable, ordered aggregates that can evolve into nano-, micro- and macroscale materials characterized by discrete 3D-architectures with specific shapes, dimensions and functions, exploitable in areas ranging from chemistry to materials science and medicine. In this connection, peptides represent powerful building blocks on the nanoscale because of their modularity, molecular diversity, which can be expanded beyond the repertoire of genetically coded amino acids, and secondary structure variety. More specifically, nanotubes and nanofibers predominate as molecular architectures among the self-assemblies formed by short peptides, while vesicles-like structure are commonly found for high molecular weight peptides. Notably, such architectures may adsorb or entrap or covalently link other molecules, thus generating ordered hybrid materials with interesting applications. In particular, we made use of short or long peptides (Figure 1) to organize at molecular levels fullerenes,<sup>1</sup> carbon nanotubes,<sup>2</sup> carbon quantum dots,<sup>3</sup> gold nanoparticles,<sup>4</sup> azobenzene derivatives,<sup>5</sup> and several other chromophores, and we explored novel applications of these peptide-based, self-assembled hybrid materials.

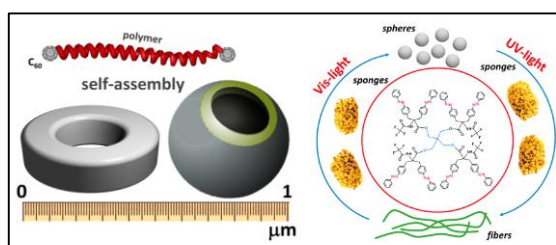


Figure 1. Two examples of self-assembled peptide-based nanostructure. Left: a  $C_{60}$ -poly-g-benzyl-L-glutamate hybrid polymer. Right: a reversible multistate hydrogel, based on a photo-responsive non-natural  $\alpha$ -amino acid.

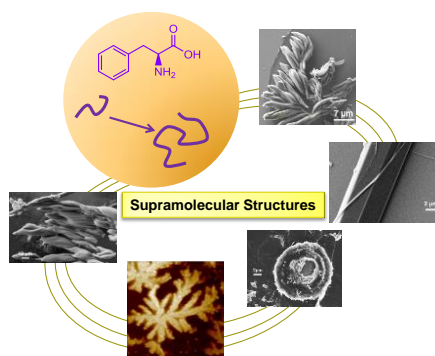
<sup>1</sup> D.Mazzier, M. Mba, M. Zerbetto, A. Moretto Chem. Commun. (2014), 50, 4571. <sup>2</sup> M. Mba, A. I. Jiménez, A. Moretto Chem. Eur. J. (2014), 20, 3888. <sup>3</sup> D. Mazzier, M. Favaro, S. Agnoli, G. Granozzi, S. Silvestrini, M. Maggini, A. Moretto Chem. Commun. (2014), 50, 6592. <sup>4</sup> D. Mazzier, F. Carraro, M. Crisma, M. Rancan, C.Toniolo, A. Moretto Soft Matter (2016), 12, 238. <sup>5</sup> D. Mazzier, M. Maran, O. Polo Perruchin, M. Crisma, M. Zerbetto, V. Causin, C.Toniolo, A. Moretto Macromolecules (2014), 47, 7272

## Self-assembly of phenylalanine oligopeptides

M. M. Pérez-Madrigal,<sup>a,b</sup> E. Mayans,<sup>a,b</sup> J. Casanovas,<sup>c</sup> L. J. del Valle,<sup>a,b</sup>  
C. Cativiela,<sup>d</sup> J. Puiggali<sup>a,b</sup> and C. Alemán<sup>a,b</sup>

<sup>a</sup>Departament d'Enginyeria Química, ETSEIB (UPC) - Avda. Diagonal 647, Barcelona E-08028, Spain; <sup>b</sup>Center for Research in Nano-Engineering (UPC)- c/Pasqual i Vila s/n, Barcelona E-08028, Spain; <sup>c</sup> Departament de Química, Escola Politècnica Superior, Universitat de Lleida, c/ Jaume II n° 69, Lleida E-25001, Spain; <sup>d</sup> Department of Organic Chemistry and Instituto de Síntesis Química y Catálisis Homogénea (ISQCH), University of Zaragoza–CSIC, 50009 Zaragoza, Spain

Self-assembled structures in the nano- and microscale have become essential for future technological applications in the nanoscience and biomaterials fields. Hence, since the pioneering work of Reches and Gazit in 2003,<sup>1</sup> in which diphenylalanine (FF) nanotubes were obtained in aqueous solution, relevant efforts have been oriented to develop a new generation of materials consisting in the self-assembly of aromatic peptides. However, research on self-assembled structures based on triphenylalanine (FFF) and tetraphenylalanine (FFFF) is very scarce. This contribution describes the self-assembly of a series of different phenylalanine-based oligopeptides that differ at the N- and C-termini. Hence, depending on environmental conditions such as solvent:co-solvent ratio, peptide concentration, and temperature, these oligopeptide derivatives self-organize into a wide variety of stable polymorphs. The overall of the results suggest that highly aromatic oligopeptides with four, or even more, phenylalanine residues should be considered as powerful building blocks for the fabrication of complex and relatively infrequent structures for advanced functional applications.



<sup>1</sup> Reches, M.; Gazit, E. *Science* **2003**, 300, 625-627



# Protein control of soft and hard matter

*Tobias Weidner*

*Max Planck Institute for Polymer Research, Mainz, Germany*

Proteins can act as Nature's engineers at interfaces and manipulate both hard and soft tissue – they shape biominerals, manipulate cell membranes and nucleate materials. Despite the apparent importance for physicists and chemists working in the fields of biomineralization, surface engineering, drug delivery, or diagnostics, the molecular mechanisms behind interfacial protein action have remained largely elusive. In order to understand protein function our goal is to probe the structure and structural dynamics of such active proteins – in action at the surface.

Mineral proteins have the ability to control and steer the growth of biogenic hard tissue by binding specific mineral facets. They control the intricate mineral morphologies found in diatom cell walls, mollusk nacre, but also human teeth and bone. Inspired by diatom silification we used amphiphilic peptides consisting of leucine and lysine (LK peptides) to investigate biomineralization at surfaces. These peptides can adopt helical or beta sheet structures at the air-water interface. Upon addition of a silica precursor we obtained freestanding peptide-silica hybrid sheets with thicknesses of  $\sim 4$  nm. We have followed the biomineral composition and interactions between peptides and silica at different early stages of biomineralization using a combination of surface spectroscopies and microscopies. Our experimental findings were complemented with molecular dynamics simulations. Our data shows that the peptide surface folding dictates the nanometer scale morphology of the prepared silica film.

A particularly fascinating example of protein driven nucleation and phase transitions are ice-nucleating proteins. These proteins are used by specific bacteria to attack plants and cause frost damage by growing ice crystals at temperatures that would otherwise not allow ice formation. A recent survey by the NASA found large amounts of biological ice nucleators in the troposphere where they may affect global precipitation patterns. We have followed the interaction of freeze proteins with surrounding water molecules – how specialized protein sites lock water molecules in place and manipulate the flow of energy within the surrounding layers of water.



# Structure-Function Relationships in Self-Assembled Peptide Nanostructures: The case of Charge transport

*Prof. Nurit Ashkenasy*

*The Ilse Katz Institute for Meso- and Nanoscale Science and Technology,  
Ben Gurion University of the Negev, Beer-Sheva, Israel*

The function of Proteins is determined by the spatial arrangement of amino acids, which in return depends on the sequence of amino acid in the proteins' chain. Inspired by these relationships, intensive effort is made to reveal the effect of peptides' sequence on their ability to fold and self-assemble into nanostructures. Naturally, the following challenge is to reveal the structure - function relationships of these assemblies. Such understanding will provide a predicting power that can be used for the design of bio-inspired functional materials. Our group focusses on understanding the structural factors that govern charge transport in self-assembled peptide nanostructures. In this talk, I will demonstrate that the sequence of the peptide affects charge transport both by controlling the morphology of the resulting nanostructures, and by direct involvement of the side chains in the charge transport process. The influence of these factors on both electron and proton transport will be demonstrated using several self-assembling peptide motifs. Finally, I will show how these understandings are used for the design of new sequences with improved electronic characteristics.

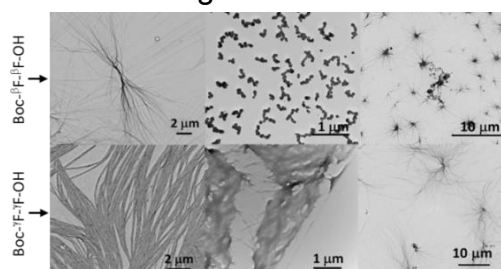
# Self-assembly of diphenylalanine backbone homologues and their combination with carbon nanotubes

*Bhimareddy Dinesh, Cécilia Ménard-Moyon, Alberto Bianco\**

*CNRS, Institut de Biologie Moléculaire et Cellulaire, Laboratoire d'Immunopathologie et Chimie Thérapeutique, 67000 Strasbourg, France*

The natural simple constituents of life - amino acids, sugars, lipids and nucleic acids - are intrinsically bioactive, biodegradable and biocompatible which makes them as perfect building blocks to generate new materials.<sup>1</sup> In particular, the research on peptide-based materials has intensified over the last few years, not only because of their synthetic versatility and their potential in biomedical applications, but also they are promising bio-based alternatives to synthetic materials.<sup>1</sup> The peptides composed of natural amino acids or in combination with other unnatural analogues are known to self-assemble or self-organise generating complex architectures, such as peptide nanowires,<sup>2</sup> nanotubes<sup>3</sup> and nanofibers.<sup>4</sup>

Herein, we studied the self-assembly of  $\beta$  and  $\gamma$  homologues of diphenylalanine peptides under different solvent and pH conditions. The backbone effected self-assemblies were shown to form nanofibers of different lengths and dimensions although possessing the same side chain. The pH variation transformed the nanofibers into spherical structures. Moreover, the co-assembly of  $\beta$  and  $\gamma$  peptides with carbon nanotubes (CNTs) covalently functionalized with the same peptide generated unique dendritic assemblies. This comparative study on self-assembly and co-assembly with CNT covalent conjugates is the first example exploring the capacity of  $\beta$  and  $\gamma$  peptides to adopt precise nanostructures, particularly in combination with CNTs (*Bhimareddy et.al. Nanoscale, 2015, 7, 15873*).



<sup>1</sup>Alemán, C; Bianco, A; Venanzi, M *Peptide Materials: From Nanostructures to Applications*, John Wiley & Sons, **2013**.

<sup>2</sup>Reches, M; Gazit, E *Science* **2003**, 300, 625-627.

<sup>3</sup>Scanlon, S; Aggeli, A; *Nano Today* **2008**, 3, 22-30.

<sup>4</sup>Renliang, H; Rongxin, S; Wei, Q; Jun, Z; Zhimin, H *Nanotechnology* **2011**, 22, 245609.

# Hydrogen bonding and lipid bilayer mediated aggregation of various peptide helix topologies

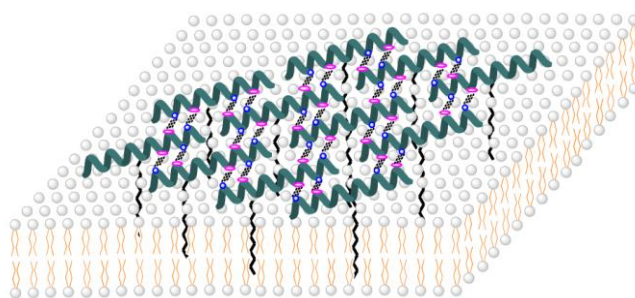
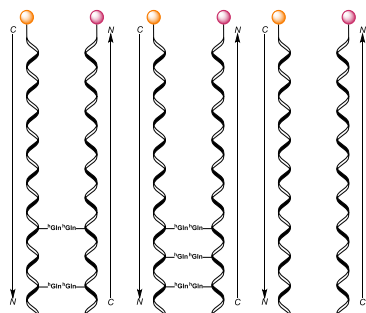
*Ulrike Rost, Gerialin Höger, Markus Wiegand, Ulf Diederichsen*

*Georg-August University Göttingen, Institute for Organic and Biomolecular Chemistry,  
Tammannstr. 2, D-37077 Göttingen, Germany, [udieder@gwdg.de](mailto:udieder@gwdg.de)*

The influence of lipid bilayers on the organization of peptide helices is investigated by various helical peptide topologies, especially focusing on  $\beta$ -peptide helices.  $\beta$ -Peptides were designed to be incorporated into membranes and thereby organized as transmembrane helices or to be aggregated on the membrane surface.

Peptide nucleic acids based on a  $\beta$ -peptide 14-helix backbone ( $\beta$ -PNA) are ideally suited to function as helix architectural unit that can be further organized in two or even three dimensional space to build up supramolecular assemblies.<sup>5,6</sup>  $\beta$ -PNAs are especially well suited as architectural elements because of the rigid  $\beta$ -peptide 14-helix scaffold, exactly three amino acids forming one helical turn, and the defined base pair hydrogen bonding complementarity.

$\beta$ -Peptide helices were designed as transmembrane helices and their incorporation in lipid bilayers and aggregation within the membrane will be presented depending on helix topology, lipid interaction, and hydrogen bonding recognition units (Figure left).



A second approach uses  $\beta$ -peptide aggregation on the membrane surface (Figure right) in order to provide a membrane supporting peptide layer with shape inducing properties. Alternatively, the aggregation of lipidated  $\beta$ -peptides on the membrane surface will be used to influence the membrane lipid composition, thereby, inducing phase separation and signaling.

<sup>5</sup> Brückner, A. M.; Chakraborty, P.; Gellman, S. H.; Diederichsen, U. *Angew. Chem. Int. Ed.* **2003**, *42*, 4395-4399.

<sup>6</sup> Chakraborty, P.; Diederichsen, U. *Chemistry Eur. J.* **2005**, *11*, 3207-3216.





# Peptides and Peptide Conjugates: From Self-assembly Towards Applications in Biomedicine

*Ian William Hamley*

*Department of Chemistry, University of Reading, United Kingdom*

Self-assembling peptides and their conjugates offer exceptional potential in nanomedicine. I will present some of our recent work on nanoscale assembled peptides and their conjugates, focussing on lipopeptides<sup>1,2</sup> and PEG-peptide conjugates.<sup>3</sup> PEGylation is an important technique in the development of conjugates for applications in therapeutics. It is found to greatly influence self-assembly of peptides and proteins - one example from our own work is a peptide which itself forms twisted fibrils but when PEG is attached, self-assembly of the conjugate leads to spherical micelles.<sup>4</sup> The conjugate can be enzymatically degraded using alpha-chymotrypsin, releasing the peptide. This nanocontainer delivery and release system could be useful in therapeutic applications. Thermoresponsive telechelic PEG/peptides with hydrophobic dipeptide end groups (di-tyrosine or di-phenylalanine) were developed, one of which shows a de-gelation transition near body temperature and which may be useful in bioresponsive delivery systems.<sup>5</sup> Examples from our recent work on self-assembling lipopeptides will also be outlined. Our focus is to investigate potential relationships between self-assembly and bioactivity, in particular in the fields of regenerative medicine,<sup>6-10</sup> antimicrobial systems<sup>11,12</sup> and immune therapies.<sup>13</sup>

---

[1] Dehsorkhi A, Castelletto V, Hamley IW. Self-Assembling Amphiphilic Peptides. *J. Pept. Sci.* 2014;20:453-67. [2] Hamley IW. Self-Assembly of Amphiphilic Peptides. *Soft Matter* 2011;7:4122-38. [3] Hamley IW. PEG-Peptide Conjugates. *Biomacromolecules* 2014;15:1543-59. [4] Castelletto V, McKendrick JME, Hamley IW, Cenker C, Olsson U. PEGylated Amyloid Peptide Nanocontainer Delivery and Release System. *Langmuir* 2010;26:11624-27. [5] Hamley IW, Cheng G, Castelletto V. Self-Assembly of Telechelic PEG End-capped with Hydrophobic Dipeptides. *Macromol. Biosci.* 2011;11:1068-78. [6] Jones RR, Castelletto V, Connon CJ, Hamley IW. Collagen Stimulating Effect of Peptide Amphiphile C<sub>16</sub>-KTTKS on Human Fibroblasts. *Mol. Pharm.* 2013;10:1063-69. [7] Castelletto V, Hamley IW, Whitehouse C, Matts P, Osborne R, Baker ES. Self-Assembly of Palmitoyl Lipopeptides Used in Skin Care Products. *Langmuir* 2013;29:9149-55. [8] Gouveia RJ, Castelletto V, Alcock SG, Hamley IW, Connon CJ. Bioactive films produced from self-assembling peptide amphiphiles as versatile substrates for tuning cell adhesion and tissue architecture in serum-free conditions. *Journal of Materials Chemistry B* 2013;1:6157-69. [9] Castelletto V, Gouveia RJ, Connon CJ, Hamley IW, Seitsonen J, Ruokolainen J, Longo E, Siligardi G. Influence of elastase on alanine-rich peptide hydrogels. *Biomaterials Science* 2014;2:867-74. [10] Gouveia RJ, Castelletto V, Connon CJ, Hamley IW. Submitted 2014. [11] Dehsorkhi A, Castelletto V, Hamley IW, Seitsonen J, Ruokolainen J. Interaction Between a Cationic Surfactant-Like Peptide and Lipid Vesicles and Its Relationship to Antimicrobial Activity. *Langmuir* 2013;29:14246-53. [12] Hamley IW, Dehsorkhi A, Castelletto V. Self-Assembled Arginine-Coated Peptide Nanosheets in Water. *Chem. Comm.* 2013;49:1850-52. [13] Hamley IW, Kirkham S, Dehsorkhi A, Castelletto V, Reza M, Ruokolainen J. Toll-like Receptor Agonist Lipopeptides Self-Assemble into Distinct Nanostructures. *Chem. Comm.* 2014; 50: 15948-51.

## Self-assembling protein nanoparticles for targeting in colorectal cancer

Fabián Rueda<sup>1,2,3</sup>, María Virtudes Céspedes<sup>3,4</sup>, Oscar Conchillo<sup>1</sup>,  
Alejandro Sanchez-Chardi<sup>5</sup>, Joaquin Seras-Franzoso<sup>1,2,3</sup>, Rafael  
Cubarsi, Alberto Gallardo, Mireia Pesarrodonna<sup>1,2,3</sup>, Neus Ferrer-Miralles<sup>1,2,3</sup>,  
Xavier Daura, Esther Vázquez<sup>1,2,3</sup>, Elena García-Fruitós<sup>1,2,3</sup>,  
Ramón Mangués<sup>3,4</sup>, Ugutz Unzueta<sup>3,4</sup>, Antonio Villaverde<sup>1,2,3</sup>

<sup>1</sup> Institut de Biotecnologia i de Biomedicina, Universitat Autònoma de Barcelona, Bellaterra, 08193 Cerdanyola del Vallès. <sup>2</sup> Dept. de Genètica i de Microbiologia, UAB. <sup>3</sup> CIBER de Bioingeniería, Biomateriales y Nanomedicina (CIBER-BBN). <sup>4</sup> Biomedical Research Institute Sant Pau (IIB-SantPau), Hospital de la Santa Creu i Sant Pau, Barcelon. <sup>5</sup> Servei de Microscòpia, UAB. <sup>6</sup> Dept. de Matemàtica Aplicada IV, Universitat Politècnica de Catalunya, Barcelona. <sup>7</sup> Catalan Institution for Research and Advanced Studies (ICREA), Barcelona, Spain

We have developed a protein engineering principle, based on the modular combination of cationic peptides and poly-histidines, to generate self-assembling building blocks of protein-only nanoparticles<sup>1-3</sup>. This principle, that exploits the cationic nature of several tumor homing peptides, has been applied to the *in vivo* targeting of primary tumor and metastatic foci in animal models of cancer<sup>4-6</sup>. We have recently found that nanoarchitecture and function of nanoparticles can be controlled at the cell factory level during biofabrication<sup>7</sup>. This fact impels the further development of this nano-architectonic principle towards the generation of true artificial viruses for highly specific and efficient drug delivery in colorectal cancer<sup>8</sup>.

<sup>1</sup> Vazquez,E. *et al.* Protein nanodisk assembling and intracellular trafficking powered by an arginine-rich (R9) peptide. (2010) *Nanomedicine. (Lond)* 5, 259-268.

<sup>2</sup> Unzueta,U. *et al.* Non-amyloidogenic peptide tags for the regulatable self-assembling of protein-only nanoparticles. (2012) *Biomaterials* 33, 8714-8722.

<sup>3</sup> Vazquez,E. *et al.* Internalization and kinetics of nuclear migration of protein-only, arginine-rich nanoparticles. (2010) *Biomaterials* 31, 9333-9339.

<sup>4</sup> Cespedes,M.V. *et al.* In Vivo Architectonic Stability of Fully de Novo Designed Protein-Only Nanoparticles. (2014) *ACS Nano*. 8, 4166-4176.

<sup>5</sup> Unzueta,U. *et al.* Intracellular CXCR4(+) cell targeting with T22-empowered protein-only nanoparticles. (2012) *Int. J. Nanomedicine*. 7, 4533-4544.

<sup>6</sup> Pesarrodonna,M. *et al.* Intracellular targeting of CD44 cells with self-assembling, protein only nanoparticles. (2014) *Int. J. Pharm.* 473, 286-295.

<sup>7</sup> Rueda,F. *et al.* Bottom-Up Instructive Quality Control in the Biofabrication of Smart Protein Materials. (2015).*Adv. Mater.* doi: 10.1002/adma.201503676.

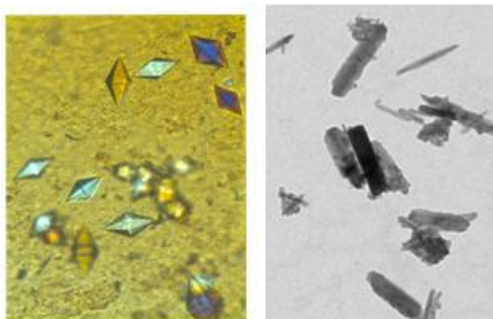
<sup>8</sup> Unzueta,U. *et al.* (2015) Towards protein-based viral mimetics for cancer therapies. *Trends Biotechnol.* 33, 253-258.

# Short peptide supramolecular hydrogels for crystal growth

Rafael Contreras-Montoya,<sup>1</sup> María Teresa Conejero-Muriel,<sup>2</sup> José A. Gavira,<sup>2</sup> and Luis Álvarez de Cienfuegos\*<sup>1</sup>

<sup>1</sup> Facultad de Ciencias, Departamento de Química Orgánica, Universidad de Granada, 18071 Granada, Spain. [lac@ugr.es](mailto:lac@ugr.es). <sup>2</sup> Laboratorio de Estudios Cristalográficos, Instituto Andaluz de Ciencias de la Tierra (Consejo Superior de Investigaciones Científicas-Universidad de Granada), Avenida de las Palmeras 4, 18100 Armilla, Granada, Spain.

Short-peptides supramolecular gels are a subgroup of low molecular weight gelators that have found useful biotechnological applications thank to their biocompatible and biodegradable nature.<sup>1</sup> In this work we have studied for the first time the behavior of these types of gels as a medium for protein crystallization. Results of the crystallization behavior and crystal quality of these proteins in the hydrogels will be presented and discussed. The influence of the chirality in protein crystallogenesis is also studied.<sup>2</sup> Additionally, preliminary results of the influence of these hydrogels in magnetite growth will be commented.



<sup>1</sup> Dasgupta, A.; Mondal, J. H.; Das, D. *RSC Adv.* **2013**, 3, 9117-9149.

<sup>2</sup> Conejero-Muriel, M.; et al. *Chem. Commun.* **2015**, 51, 3862-3865.



# Peptides as Protein-Surface Interactors

*Ernest Giralt*<sup>1,2</sup>

<sup>1</sup> *Institute for Research in Biomedicine (IRB Barcelona), Baldori Reixac 10, 08028, Barcelona, Spain, [ernest.giralt@irbbarcelona.org](mailto:ernest.giralt@irbbarcelona.org) ;*

<sup>2</sup> *Department of Organic Chemistry, University of Barcelona, Marti Franqués 1, 08028, Barcelona, Spain*

The breakthrough concept that proteins function as a contact network rather than as independent individuals is not only one of the most important advances in our comprehension of living systems, but also translates to a new era in drug discovery. The few reported examples of diseases caused by “impolite” protein social behavior certainly represent only the tip of the iceberg. Therapeutic intervention through molecules designed to selectively modulate the strength and specificity of protein-protein interactions (PPIs) is becoming a reality. This will not only feature molecules with inhibitory capacity: equally or even more interesting are those compounds which can rescue pre-established interactions or structures whose loss results in disease.

In this context, peptides are destined to play a major role as therapeutic agents. My laboratory is contributing to speeding up this process. On the one hand, we devote efforts to studying the molecular details and dynamics of the events that occur during molecular recognition at protein surfaces. We succeeded to design and synthesize peptides able to modulate these recognition events either permanently or in response to light. On the other hand, we are discovering and designing peptides able to cross biological barriers. Our aim is to use these peptides as shuttles for targeting therapeutic agents to organs, tissues, or cells, with a special emphasis on drug delivery to the brain.

PPIs are the result of an ensemble of exquisitely regulated molecular recognition events that take place at protein surfaces. Inspection of protein-protein interfaces allows distinguishing two categories of PPIs: domain-domain and peptide-mediated PPIs [1]. Relatively rigid peptides and peptidomimetics have proved to be very efficient inhibitors of this last class of interactions. In this presentation, recent results from our group related to the use of peptides to modulate PPIs will be discussed. This include, among others: *i)* the recent development of cell-permeable

photoswitchable PPI inhibitors, that opens the way to manipulating a specific PPI locally and in a time-controlled manner using illumination patterns [2,3]; *ii*) the application of the retro-enantio approach to obtain a peptide capable of overcoming the blood–brain barrier [3,4,5]; and *iii*) the use of peptides to modulate the dynamic behavior of prolyl oligopeptidase (POP), a large 80 kDa protease relevant as therapeutic target in schizophrenia [6].

- 
- [1] Nevola L, Giralt E. *Chem. Commun.*, **2015**, 45, 3302-3315.  
[2] Nevola L, Martin-Quiros A, Eckelt K, Camarero N, Tosi S, Llobet A, Giralt E, Gorostiza P. *Angew. Chem.Int. Ed.*, **2013**, 52, 7704–7708.  
[3] Prades R, Oller-Salvia B, Schwarzmaier SM, Selva J, Moros M, Balbi M, Grazú V, de La Fuente JM, Egea G, Plesnila N, Teixidó M, Giralt E. *Angew. Chem. Int. Ed. Engl.*, **2015**, 54, 3967-3972.  
[4] Arranz-Gibert P, Guixer B, Malakoutikhah M, Muttenthaler M, Guzman F, Teixidó M, Giralt E. *J. Am. Chem. Soc.*, **2015**, 137, 7357-7364.  
[5] Oller-Salvia B, Sánchez-Navarro M, Ciudad S, Guiu M, Arranz-Gibert P, Garcia C, Gomis RR, Cecchelli R, García J, Giralt E, Teixidó M. MiniAp-4: A Venom-inspired peptidomimetic for brain delivery. *Angew Chem Int Ed Engl.* **2015** Oct 23. doi: 10.1002/anie.201508445.  
[6] M. Kotev, D. Lecina, T. Tarrago, E. Giralt, V. Guallar. *Biophys J.*, **2015**, 108, 116-125.



# Peptide Nanofibers for Diagnostic and Therapeutic applications

*Antonella Accardo, Carlo Diaferia, Giancarlo Morelli*

*Department of Pharmacy, University of Naples "Federico II" Via Mezzocannone, 16  
80134 Napoli, Italy*

Aromatic peptides such as diphenylalanine (FF) or tetraphenylalanine (FFFF) are able to self-assemble into many ordered self assembled nanostructures from nanotubes to organogels. These nanostructures have been investigated for their mechanical, electrochemical and optical properties. Recently, few studies have been devoted to investigate their properties for biomedical and biotechnological applications, for example for regenerative nanomedicine, to fabricate stable drug-delivery systems with proteolytic resistance, to produce modern nanodevices, and to prepare ultrasensitive and selective sensors for toxin detection<sup>1</sup>. However, research of poly-phenylalanine based compounds as diagnostic tools, for example as MRI contrast agents, remains largely unexplored.

Magnetic Resonance Imaging (MRI) is one of the most powerful and non-invasive technique for medical diagnosis. Currently, stable Gd(III)-poly(aminocarboxylate) complexes are widely used as contrast agents (CAs) in MRI to produce high quality images.

In the present communication, novel peptide materials, based on poly-phenylalanine are discussed as a new class of supramolecular gadolinium based MRI contrast agents. These materials contain phenylalanine residues, Gd-DOTA or Gd-DTPA complexes and PEG spacers<sup>2</sup>. Their structural and relaxometric properties and their capacity to increase contrast in Magnetic Resonance Imaging are discussed.

---

<sup>1</sup> Mayans, E; Ballano, G; Casanovas, J; Diaz, A; Perez-Madriral, M; Estrany, J; Cativiela, C; Aleman, C; *Chem. Eur. J.* **2015**, 21, 16895-16905

<sup>2</sup> Diaferia, C; Gianolio, E; Palladino, P; Arena, F; Boffa, C; Morelli, G; Accardo, A; *Adv. Funct. Mat.* **2015**, 25, 7003-7016.



## Computational studies on peptides in drug design: insights from molecular dynamics simulations

*Axel Bidon-Chanal<sup>1</sup>, Rubí Zamudio<sup>2</sup>, Javier Ruiz-Rodríguez<sup>2</sup>, F. Javier Luque<sup>1</sup>, Francesc Mitjans<sup>2</sup>, Rodolfo Lavilla<sup>2</sup>, Fernando Albericio<sup>2</sup>*

*<sup>1</sup>Campus de l'Alimentació de Torribera, Universitat de Barcelona, <sup>2</sup>Parc Científic de Barcelona, Universitat de Barcelona*

The development of new peptides with therapeutic effects has gained interest within the pharmaceutical research during the last decade with around one hundred and forty peptides now being developed in clinical phase I, II or III<sup>1,2</sup>. Their high specificity along with their low toxicity profiles and the development of novel design and modification techniques are within the reasons that have promoted it, however, it is usually more difficult to fully understand their mechanism of action. Peptides are larger in size and more flexible compared to the common small molecules used as drugs, thus studying the way they interact with a receptor, for example, demands greater efforts than what usually is needed. With the implementation of supercomputers and the most recent development of algorithms and codes adapted to use GPUs, the use of computational techniques within the drug design process has received a major bump allowing for example the use of long timescale molecular dynamics simulations to study protein-ligand(peptide) complexes or the folding of small peptides or even small proteins. Here we will present two different cases where the use of molecular dynamics simulations helped in the development of peptides with therapeutic effects and to explain their possible mechanism of action<sup>3,4</sup>.

<sup>1</sup> Otvos, L.; Wade JD. *Frontiers in Chemistry* **2014**, 2, 62.

<sup>2</sup> Fosgerau, K.; Hoffmann, T. *Drug Discovery Today* **2015**, 1, 122-128.

<sup>3</sup> Zamudio-Vazquez R. *et al. Chemical Science* **2015**, 6, 4537-4549.

<sup>4</sup> Ruiz-Rodríguez J. *Chemical Science* **2014**, 5, 3929-3935.

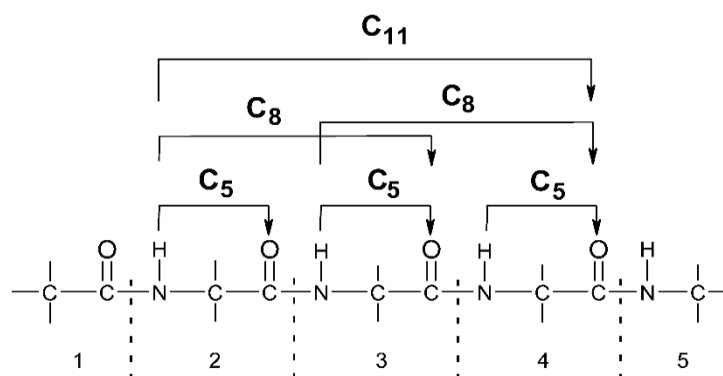
# Intramolecular Backbone···Backbone Hydrogen Bonds in Peptide Conformations: The Other Way Around

Claudio Toniolo

Department of Chemistry, University of Padova, 35131 Padova, Italy

Folded conformations stabilized by intramolecular H-bonds between a main-chain amide N-H donor and a main-chain amide C=O acceptor are extremely common observations in solution studies and crystal-state analyses of peptides and proteins. Moreover, fully-extended, intramolecularly H-bonded 3D-structural motifs have also been reported, although to a very limited extent.

In particular, in a system of *four* linked peptide units the possible conformations intramolecularly H-bonded “the other way around” (as compared to the traditional and extensively investigated C<sub>7</sub>, C<sub>10</sub>, and C<sub>13</sub> forms, where C stands for *cyclo* and the number indicates how many atoms are involved in the *pseudo*-ring closed by the intramolecular H-bond) are C<sub>5</sub>, C<sub>8</sub> and C<sub>11</sub> (see *Scheme* below).



In this lecture, I will examine the characteristic properties of: (i) the fully-extended (C<sub>5</sub>) conformation and its related 2.0<sub>5</sub>-helix, (ii) the C<sub>8</sub>- (also called δ-) turn, and (iii) the C<sub>11</sub>- (also called ε-) turn. In particular, the C<sub>8</sub> and C<sub>11</sub> forms, that are either sterically forced to or may include a *cis*-amide disposition, have been only occasionally mentioned and rarely authenticated experimentally. Specifically, results from X-ray diffraction analyses, conformational energy calculations, and solution investigations will be discussed.





# Bioinspired Visible Fluorescent Nanolabels for Photonic and Medical Applications

*Gil Rosenman*

*School of Electrical Engineering, Tel Aviv University, 69978 Tel Aviv, Israel*

Genetically encoded green fluorescent protein and its homologs opened the avenue for a wide applications of these intrinsic biological labels representing specific proteins. We report on a new visible blue/green fluorescent phenomenon found in  $\beta$ -sheet nanowires of ultrashort bioinspired peptides<sup>1</sup>.

Di- and tri-peptide nanostructures self-assembled from biomolecules of different compositions and origin such as aromatic and aliphatic (diphenylalanine, FF-; dileucine, LL-; triphenylalanine, FFF- monomers) have been studied. At elevated temperature 140-180°C these supramolecular structures of diverse morphologies undergone the same irreversible thermally-induced phase transformation. This reconstruction process is followed by deep modification at all levels: molecular, electronic, elementary symmetry, peptide secondary structure, displaying new nanowire morphology and completely new common physical properties. The thermally-induced nanowire supramolecular ensembles acquire  $\beta$ -sheet secondary structure. In this phase the  $\beta$ -sheet nanowires, irrespective of their native biomolecules origin, exhibit similar profound alteration of optoelectronic properties and visible (blue and green) fluorescence ascribed to hydrogen bonds. Observed visible fluorescent of peptide fiber nanostructures can be used as intrinsic optical labels in biomedicine as biocompatible markers and for a new generation of bio-nanophotonics such as bio-lasers and integrated optical devices.

---

<sup>1</sup> A.Handelman, N.Kuritz, A. Natan, Gil Rosenman, Reconstructive Phase Transition in Ultrashort Peptide Nanostructures and Induced Visible Fluorescence, Invited Feature Article, Langmuir, October 23, 2015



# Mecwins: developing biomedicine applications using nanomechanical sensors

*Oscar Ahumada*

*Mecwins S.A., Plaza de la Encina 10-11, Tres Cantos (Madrid), Spain*

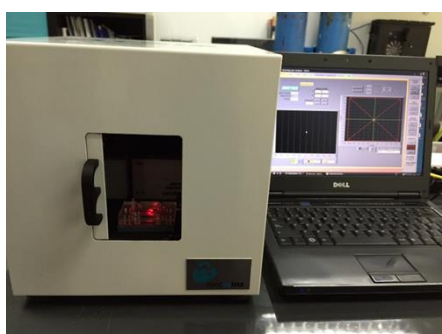
[oahumada@mecwins.com](mailto:oahumada@mecwins.com)

Mecwins was founded in 2008 by Dr. Javier Tamayo and Dr. Montserrat Calleja, Bionanomechanics group leaders from the Institute of Microelectronics of Madrid (IMM - CSIC) [1]. Since then, we have been developing cutting edge technology for nanomechanical sensing. The technology, based on detecting variations in the deflection and resonance frequency of nanomechanical sensors, was the groundwork for the technical improvements that led us to our new ultrasensitive detection device, SCALA

SCALA (SCAnning Laser Analyzer) [2] is a commercial platform with high potential for the analysis of biomolecule interactions in human whole blood samples for biosensing applications. Cantilever arrays have been extensively explored as high-sensitive nanomechanical biosensors [3]. The molecular recognition on the surface of a biofunctionalized cantilever results in a nanomechanical response, that produces cantilever bending of a few nanometers (static mode) or changes in cantilever resonance frequency (dynamic mode) where the added mass due to selective molecular recognition decreases resonance frequency value.

In the present study, SCALA has been used for the successful detection of different protein biomarkers in clinical diagnosis, focusing our preliminary experiments on oncology, cardiac and infectious diseases biomarkers. SCALA combines mechanical detection technique (microcantilevers resonance frequency analysis) with a new optical detection technique that increases current levels of sensitivity in clinical practice a million times [4] in comparison with techniques currently used in hospitals and central laboratories and without increasing current cost per sample. The adoption of ultrasensitive detection equipment will enable screening for early detection of a wide range of diseases with established diagnostic biomarkers from a droplet of blood.

The adoption of ultrasensitive detection equipment will enable screening for early detection of a wide range of diseases with established diagnostic biomarkers from a droplet of blood. Moreover, we have developed our technology to be portable, in order to be introduced in the market as a POC device (Figure 1) that allows the implementation of fast and efficient clinical procedures for diagnosis, monitoring and prognosis that will cut down costs in healthcare expenditure, insurance or laboratory work.



**Figure 1. SCALA Platform**

---

[1] Spanish Research Council: <http://www.imm-cnm.csic.es/bionano/es>

[2] <http://mecwins.com/>

[3] Arlett *et al.* Nat Nanotechnol 6(4),203-15 (2011)

[4] Kosaka *et al.* Nat Nanotechnol. 9(12),1047-53 (2014)



## Controlling Multicomponent Dipeptide Hydrogels

E.R. Draper,<sup>a</sup> E.G.B. Eden,<sup>a</sup> T.O. McDonald,<sup>a</sup> C. Colquhoun,<sup>a</sup> K. Morris,<sup>b</sup>  
L. C. Serpell<sup>b</sup> and Dave J. Adams<sup>a</sup>

<sup>a</sup> Department of Chemistry, University of Liverpool, Crown Street, Liverpool, UK

<sup>b</sup> School of Life Sciences, Chichester II Building, University of Sussex, Falmer BN1  
9QG, UK

Low molecular weight gelators (LMWG) self-assemble into one-dimensional fibrous structures; this self-assembly leads to the immobilisation of the solvent and the formation of a gel. These materials are attracting significant interest, for example in cell culturing, where the low LMWG concentration needed can be useful, as can the gel's reversibility as the cells grow and re-form their environment.

In the majority of cases, gels are formed from a single LMWG. Mixing different LMWG (all of which can form gels independently) can result in interesting new materials.<sup>1-3</sup> Depending on how these LMWG assemble, this method can adjust the properties of the final gels, or to prepare systems with higher information content by the selective positioning of specific functional groups in space. Here, we will describe a range of mixed dipeptide-based LMWG systems. We will show how fibrous structures form in these systems and show how we can control how different types of fibrous networks are built up in multicomponent systems. We describe both self-sorted and co-assembled networks and the effect of these different networks on the gel properties.

---

<sup>1</sup> Colquhoun, C.; Draper, E.R.; Eden, E.G.B.; Cattoz, B.N.; Morris, K.L.; Chen, L.; McDonald, T.O.; Terry, A.E.; Griffiths, P.C.; Serpell, L.C.; Adams, D.J. *Nanoscale* **2014**, *6*, 13719-13725

<sup>2</sup> Morris, K.L.; Chen, L.; Raeburn, J.; Sellick, O.R.; Cotanda, P.; Paul, A.; Griffiths, P.C.; King, S.M.; O'Reilly, R.K.; Serpell, L.C.; Adams, D.J. *Nature Commun.*, **2013**, *4*, 1480

<sup>3</sup> Draper, E.R.; Eden, E.G.B.; McDonald, T.O.; Adams, D.J. *Nature Chem.*, **2015**, *7*, 848-852.

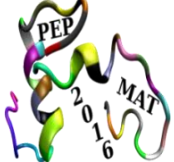


# Drug Delivery via Cell Membrane Fusion using Lipopeptide Modified Liposomes

A. Boyle, A. Kros

*Leiden Institute of Chemistry, Leiden University, The Netherlands*

Efficient delivery of drugs to living cells is still a major challenge. Currently, most methods rely on the endocytotic pathway resulting in low delivery efficiency due to limited endosomal escape and/or degradation in lysosomes. Here, we report a new method for direct drug delivery into the cytosol of live cells *in vitro* and *in vivo* utilizing targeted membrane fusion between liposomes and live cells. A pair of complementary coiled coil lipopeptides was embedded in the lipid bilayer of liposomes and cell membranes respectively, resulting in targeted membrane fusion with concomitant release of liposome encapsulated cargo including fluorescent dyes, high molecular weight dextran, cytochrome c. Next, the cytotoxic drug doxorubicin was delivered into cells thereby increasing its therapeutic efficacy and cytosolic delivery *in vivo* using zebrafish embryos was proven. Using a wide spectrum of endocytosis inhibitors and endosome trackers we demonstrate that the major site of cargo release is at the plasma membrane. This method thus allows for the quick and efficient delivery of drugs and is expected to have many applications both *in vitro* and *in vivo*.



# ***Nanoscale Molecular Junctions Incorporating Peptides and Peptidomimetics***

***Slawomir Sek<sup>\*a</sup>, Karolina Pulka-Ziach<sup>b</sup>, Jan Pawlowski<sup>a</sup>***

*<sup>a</sup>Faculty of Chemistry, Biological and Chemical Research Centre, University of Warsaw, Zwirki i Wigury 101, 02-089 Warsaw, Poland*

*<sup>b</sup>Faculty of Chemistry, University of Warsaw, Pasteura 1, 02-093 Warsaw, Poland*

Peptides are involved in numerous biologically relevant processes and their broad range of functions results from ability to adopt variety of structural motifs. Therefore when suitably designed, peptides can provide desired properties that are useful in biosensing devices as well as in nanoscale electronic circuits. However, there are still numerous problems which need to be solved in order to design and build functional molecular devices such as molecular wires, logic gates, switches, or molecular-scale transistors. One of the crucial issues is to understand the fundamental mechanisms determining efficiency of charge transport through these molecules. Numerous experimental approaches can be utilized to probe electron transport efficiency through organic molecules. Among them, scanning probe microscopy (SPM) offers unique capability to investigate electric properties of individual molecules or molecular films at nanoscale. In general this method involves entrapment of the molecules between two metallic contacts established by metal support and SPM probe. The electric properties of the resulting metal–molecule–metal junction can be probed while the bias voltage is applied between the electrodes. The additional advantage of SPM-based method comes from the fact that it enables modulation of the efficiency of peptide mediated ET by controlling mechanical strain or stress of the molecules entrapped within the junction. Using such experimental approach, it is possible to probe electric properties of peptides as well as peptide-mimicking compounds. The latter can be represented by *N,N'*-substituted oligoureas and their derivatives. These compounds adopt very robust 2.5-helix in the solution and in the solid state. To the best of our knowledge, such unusual structural motif was not investigated so far in terms of the efficiency of electron transmission.



# Perspectives for Stimuli-responsive peptide: from self-assembly to biomolecular spring

*G.M.L. Messina and G. Marletta*

*Laboratory for Molecular Surfaces and Nanotechnology, Dept. of Chemical Sciences,  
University of Catania, Catania, I-95125, Italy*

Biomolecule-based materials that change properties in response to different local stimuli are increasingly being studied in the context of several applications, with particular attention to bioelectronics.

Peptides are ideally suited for this purpose because of the range of distinct physical properties available from the amino acids. This diversity allows several non-covalent interactions including electrostatic (acidic and basic amino acids), hydrophobic,  $\pi$ -stacking (aromatic amino acids), hydrogen bonding (polar amino acids) as well as covalent (disulfide) bonds and steric contributions (strand directing amino acids). Crucially, these interactions depend by different factors such as ionic strength, pH and temperature. The dynamic nature of these interactions then allows the molecular organisation to be altered in response to changes in the direct environment. Changing the nature of these contacts by introducing responsive amino acids can alter the stability of the conformation and provide a mechanism for control of dynamic materials. The use of acidic and basic amino acids that can be protonated or deprotonated by a change in pH allows dynamic control over the secondary structure of the peptides and can be used to control the assembly of coiled-coils.

This presentation will focus on the study of different peptides immobilized on surface and on their response by changing external stimuli. The loading and the conformational switching properties of the surface-bound peptides were investigated by means of several surface techniques. The self-organization process is shown to be severely affected by the type of peptide and surface properties, including specifically the charge state at a given pH. The results strongly support a self-assembly mechanism based onto a very specific organization processes at the liquid-solid interface, based onto the active role played by the surfaces to promote the orientation and organization of single molecules as the prime processes at nanometer scale, followed by the aggregation process at the mesoscopic scale.

Applications in the emerging field of the bio-inspired nanotechnology are suggested.



# Ultrashort Self-assembled Peptides for Biomaterial Applications

Dr Garry Laverty, Alice McCloskey


*Biofunctional Nanomaterials Group, School of Pharmacy, Queens University Belfast,  
N.Ireland*

Antimicrobial resistance is one of the greatest threats facing society worldwide. Biomaterial infections, commonly caused by multi-drug resistant pathogens, are responsible for the majority of healthcare associated infections. The implant surface provides an optimum environment for microbial attachment and growth. Pathogens form an extracellular polymeric biofilm matrix, providing protection against the host immune response and standard therapies. This results in failure of treatment, increased spread of resistant pathogens, device removal, morbidity and increased mortality. Despite their high value to healthcare and the economy, infections of biomaterials remain unsolved and an ongoing burden. Our group has developed a library of ultrashort, cost-effective, diphenylalanine-based peptides ( $X_1$ -FF- $X_2$ ) which target (viability reduced >90%) the most resistant biofilm pathogens implicated in biomaterial infection including: methicillin resistant and sensitive *Staphylococcus aureus* and *Staphylococcus epidermidis*; *Pseudomonas aeruginosa*, *Proteus mirabilis* and *Escherichia coli*. They demonstrate a reduced cell cytotoxic profile, possessing limited haemolysis.<sup>7</sup> These peptides respond form supramolecular hydrogel nanomaterials at low concentrations (~0.5%w/v) in response to subtle changes in pH associated with infection. Conjugation of variety of aromatic-based drugs at the  $X_1$  position, for example non-steroidal anti-inflammatories (NSAIDs), confer further pharmacological properties enhancing their therapeutic potential. *In vivo* studies using waxworms (*Galleria mellonella*) demonstrate the low toxicity and high antimicrobial activity of these low molecular weight gelators in animal models. This work shows biofunctional peptide-based nanomaterials hold great promise for future translation to patients as biomaterial platforms.<sup>2</sup>

<sup>7</sup> Laverty, G; McCloskey, A.P; Gilmore, B.F; Jones, D.S, Zhou, J; Xu, B. *Biomacromolecules* **2014**, 12, 6566-6596.

<sup>2</sup> McCloskey, A.P; Gilmore, B.F; Laverty, G. *Pathogens* **2014**, 3, 791-821.





# Molecular dynamics simulations of intermolecular interactions involving peptides

Gianfranco Bocchinfuso

*Dipartimento di Scienze e Tecnologie Chimiche – Università di Roma Tor Vergata  
(Italy)*

Peptides have a wide range of applications, going from drug design to innovative materials. This high versatility comes from their chemical and conformational flexibility. On the other hand, structural flexibility is also the main problem to face when studying this class of molecules. In this context, molecular dynamics (MD) simulations represent an essential tool to evaluate the structural and dynamic features of peptides in different environments. In the last years, we have applied a combined approach of MD and spectroscopic techniques for studying peptides under different conditions, by focusing both on topics of biological relevance and on aspects more related with a possible use in innovative materials.

In the former case, we have also investigated antimicrobial peptides (AMPs). AMPs are a promising class of antimicrobial drugs, which act as permeabilizing agents of the bacterial cell membrane. This nonspecific mechanism makes difficult for the pathogens to develop resistance. In this context, we have studied different structural features that influence the interaction with phospholipid bilayers.<sup>1</sup> Other biological problems involving peptides are related to their interactions with proteins. In this field, we have studied protein-peptide complexes involved in the onset of pathological conditions.<sup>2</sup>

With the aim to use peptides as building blocks for innovative materials, it is pivotal to understand the properties that influence the aggregation processes. In the last years, we have studied both self-aggregation processes of peptides and their ability to interact with ions.<sup>3</sup>

In the talk, an overview of these works will be presented, particularly focused on the contributions obtained by applying

<sup>1</sup> Farotti, A et al. *BBA Biomembrane*. **2015**, 1848, 581-592; Bobone, S et al. *BBA-Biomembrane*. **2013**: 1828, 1013–1024; Bocchinfuso, G et al. *Cell Mol Life Sci* **2011**, 68, 2281-2301; Bocchinfuso, G et al. *J Pept Sci* **2009**, 15, 550–558.

<sup>2</sup> Niceta, M et al. *Am J Hum Genet* **2015**, 96, 816-825

<sup>3</sup> Caruso, M et al. *J Phys Chem B* **2013**, 117, 5448–5459; Venanzi, M et al. *ChemBioChem* **2009**, 10, 91-97.



## Peptide aggregation in 2D and 3D

Mariano Venanzi

Dept. of Chemical Sciences and Technologies, University of Rome 'Tor Vergata',  
Rome (Italy)

Peptide-based materials have been finding application in very different fields, from molecular electronics to tissue engineering.<sup>1</sup>

In this contribution, we will discuss on a molecular basis how the aggregation propensity of different peptide secondary structures may affect the morphology of nano- and mesoscopic peptide architectures. This effect is the result of hierarchical self-assembly, *i.e.* the self-organizing aggregation of molecular building blocks.

Following this principle, a suitable modification of the peptide sequence may propagate its effect from the secondary structure to the formation of nanostructures of controlled size and shape, to the assembly of mesoscopic structures. The understanding of the basic interactions governing this process will help to fix the conditions leading to structures of the desired morphology and function.

The strategy to prosecute this program from the very elementary constituents is to use oligopeptides predominantly formed by C<sup>α,α</sup>-tetrasubstituted amino acids. The proper selection of these conformationally constrained residues allow to get oligopeptides attaining specific conformations, to be used as building blocks in the self-assembly process.

The conformational properties and physical interaction governing the formation of peptide aggregates in solution (fibrils, globules)<sup>2,3</sup> and as molecular films (self-assembled monolayers, Langmuir-Blodgett film) will be thoroughly discussed at the Conference.<sup>4,5</sup>

---

<sup>1</sup> Aleman, C.; Bianco, A.; Venanzi, M. Peptide Materials. From nanostructures to applications. John Wiley & Sons Ltd., United Kingdom (2013).

<sup>2</sup> Caruso, M.; Placidi, E.; Gatto, E.; Mazzuca, C.; Stella, L.; Bocchinfuso, G.; Palleschi, A.; Formaggio, F.; Toniolo, C.; Venanzi, M. J. Phys. Chem. B **2013**, 117, 5448-5459.

<sup>3</sup> Caruso, M.; Gatto, E.; Placidi, E.; Ballano, G.; Formaggio, F.; Toniolo, C.; Zanuy, D.; Aleman, C.; Venanzi, M. Soft Matter **2014**, 10, 2508-2519.

<sup>4</sup> Gatto, E.; Venanzi, M. Polymer J. **2013**, 1-13

<sup>5</sup> Longo, E.; Wright, K.; Caruso, M.; Gatto, E.; Palleschi, A.; Scarselli, M.; De Crescenzi, M.; Crisma, M.; Formaggio, F.; Toniolo, C.; Venanzi, M. Nanoscale **2015**, 7, 15495-15506.



# Recombinamer-based injectable artificial ECM

*J. Carlos Rodríguez-Cabello*

*Bioforge, University of Valladolid Ciber-BBN, Valladolid, Spain*

## Introduction

Mimicking the natural extracellular matrix (ECM) has become the Holy Grail in the development of new scaffolds for regenerative medicine. However, that statement of intent turns out to be very elusive due to the complexity of the natural ECM as well as the complex relationship existing between cells and ECM. Properties of the ECM that must be taken into consideration range from mechanical properties<sup>1</sup> and microstructure to much more specific bioactivities<sup>2</sup>. In addition, such scaffold will have to cope with potential rejection by immune system of the host. Finally, the adequacy of its injectability is highly relevant. Therefore, the system that can fulfill all those requirements must rely on powerful self-assembling processes that ideally drive both the gelation of the system as well as the development of the proper microstructure.

This task could be at reach by exploiting the potential of recombinant technology and the use of recombinant protein polymers as the source of complex materials that contains all the functions required, including capacity to self-assemble and form stable gels upon injection.

## Results

Among the different option within the area of recombinamers Elastin-like recombinamers (ELR) stand out<sup>3</sup>. In this work, different ELRs have been produced and used sowing different approaches to achieve injectable artificial ECM. The full range of ELRs designed and produced for this purpose include:

- Injectable two-component ELR for covalent cross-linking.
- Thermosensitive amphiphilic multiblocks for physical cross-links
- Advanced beta-sheet forming molecules for developing fibrous gels on setting.

The first group relies on catalyst-free click chemistry reactions. Those materials have proved exceptional performance in different applications, especially in the cardiovascular area. The second group relies on the thermal sensitiveness of the ELR. Due to their basic amphiphilic block nature and the ability to show a self-

aggregating effect induced by an increase in the temperature, those materials have the chance to be liquid below body temperature and make stable gels at BT. Those injectable scaffolds have proven their efficiency in musculoskeletal applications, particularly in the total regeneration of osteochondral injuries. That is also relevant its role in designing biomimicked mineralization templates and scaffolds. The last group is based on ELRs that, along with the elastin-like main contribution and all the epitopes rendering cell-adhesiveness, protease sensitiveness, etc., included epitopes taken from silk that endow the system with a capacity to self-organize in a regular microfibrillar structure resembling that of the natural ECM proteins such as collagen.

---

<sup>1</sup> J. Swift et al. *Science*, 341: 6149. 2013.

<sup>2</sup> Sudhir Khetan, et al. *Nature Materials* 12, 458–465, 2013

<sup>3</sup> Rnjak-Kovacina J, et al. *Comprehensive Biomaterials*, vol. 2 (Edit: P. Ducheyne, et al). Elsevier:329, 2011.

# Crystal Structures of the Halogenated Variants of the Amyloidogenic Core Sequences DFNKF and KLVFF

G. Cavallo<sup>1</sup>, A. Bertolani<sup>1</sup>, L. Gazzera<sup>1</sup>, A. Pizzi<sup>1</sup>, N. Demitri<sup>2</sup>,  
P. Metrangolo<sup>1</sup>, G. Resnati<sup>1</sup> and G. Terraneo<sup>1</sup>

<sup>1</sup>Laboratory of Nanostructured Fluorinated Materials (NFMLab), Dept. Chem., Mater., and Chem. Eng. "Giulio Natta", Politecnico di Milano, Milan, Italy.

<sup>2</sup>Elettra-Sincrotrone Trieste, 34149 Basovizza, Trieste, Italy.

Amyloid fibril self-assembly is linked to pathological disorders such as type II diabetes mellitus, Alzheimer's, Parkinson's, Creutzfeldt–Jakob, and Huntington's diseases.<sup>1</sup> While some general details about the overall features of amyloid fibrils are well known, there still remains a lack of knowledge about exact oligomer conformations and details of the molecular mechanisms of aggregation. These may be addressed by elucidating high-resolution structures of amyloid conformations, but due to low solubility and crystallinity of these structures this remains a real challenge. Here, we describe that halogenation in the *para*-position of a phenylalanine residue (Phe5) of DFNKF (Figure 1)<sup>2</sup> or KLVFF induces their crystallization allowing us to determine for the first time the single crystal X-ray structure of DFNKF(I) along with those of KLVFF(I) and KLVFF(Br). Moreover, we demonstrate by molecular dynamics simulations that wild-type DFNKF and the iodinated peptide DFNKF(I) populate similar conformations therefore confirming that the crystal structure of DFNKF(I) is representative of the structure of DFNKF itself.

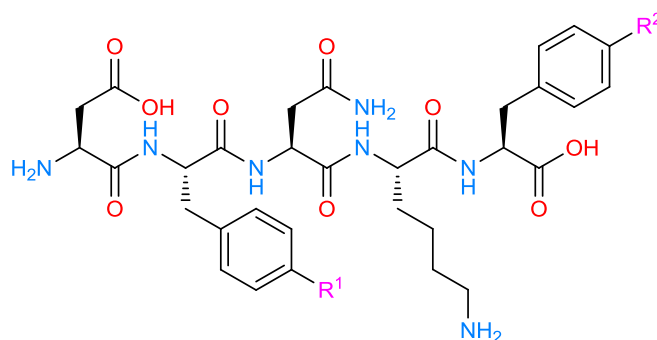


Figure 1: Chemical structure of the DFNKF (R1: H, R2: H) and DFNKF(I) (R1: H, R2:I) peptides.

<sup>1</sup> Dobson C.M. *Nature* **2003**, 426, 884-890.

<sup>1</sup> Bertolani A. *et al. Nat. Commun.* **2015**, 6, 7574, doi: 10.1038/ncomms857.

# Bacterial Resistance to Silver: The Role of SilE Protein

Valentin Chabert, Katharina M. Fromm\*

University of Fribourg, Switzerland

Silver has been used for hundreds of years for its antimicrobial properties. Since the emergence of many multi-resistant bacterial strains against classical antibiotics, the research of new silver compounds is now at its apogee. Nowadays, a lot of researches are focused on compounds with slow- and stimuli-responsive- release of  $\text{Ag}^+$ . While these drugs have been shown to be highly able to kill bacteria, some of these pathogens have developed a resistance to high concentrations of  $\text{Ag}^+$ .<sup>1</sup>

This resistance is provided by the plasmid pMG101, which encodes for eight proteins that act together in an efflux pump system to deal with silver ions. Among these, the SilE protein is the only one of which its mode of action is actually unknown.<sup>2</sup>

To identify the role of SilE in this bacterial machinery, two approaches have been intended in our group. While one way is to study the interaction of the whole protein with silver ions, the other is based on a bottom-up approach, investigating the interaction of silver ions with short peptide sequences of this protein.

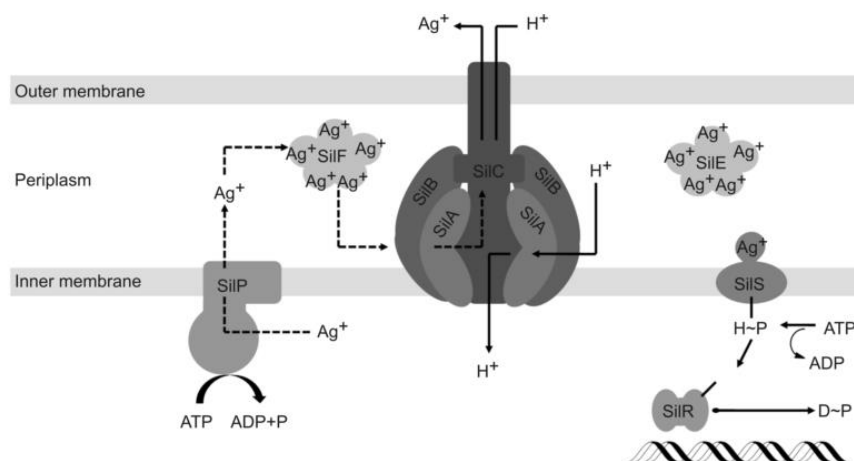


Figure 1: Proteins products of pMG101 silver resistance genes.

<sup>1</sup> Eckhardt, S. ; Brunetto, P. S. ; Gagnon, J. ; Priebe, M. ; Giese, B. ; Fromm, K. M. *Chem. Rev.* **2013**, 113, 4708-4754.

<sup>2</sup> Silver, S. ; Phung, L. T. ; Silver, G. *J. Ind. Microbiol. Biotechnol.* **2006**, 33, 627-634.

# Aqueous self-assembly of short hydrophobic peptides containing $\alpha,\alpha$ -disubstituted amino acids

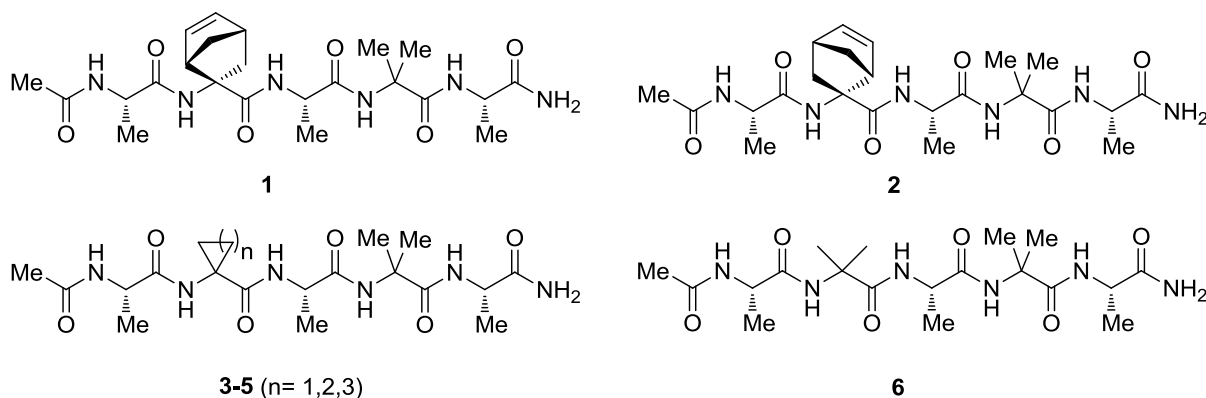
*F. Clerici, A. Ruffoni, S. Locarno, D. Nava*

*Università degli Studi di Milano, Dipartimento di Scienze Farmaceutiche, Milan, Italy*

Here we report on the preparation of a series of short hydrophobic peptides and on their different ability to stabilize the formation of supramolecular assemblies in water. Pentapeptides AcAla-**X**-Ala-Aib-AlaNH<sub>2</sub> **1-6** containing unnatural constrained amino acids **X**, are insoluble in organic solvents but completely soluble in water despite the presence of hydrophobic non polar scaffolds.

NMR, CD, IR experiments were performed to gain insight on their secondary structure. Furthermore, the formation of supramolecular assemblies in water was studied by DLS.

In the case of **1** and **2**, containing the two enantiomers of the norbornene amino acid, DLS was performed either on the pure compounds or on their mixture.



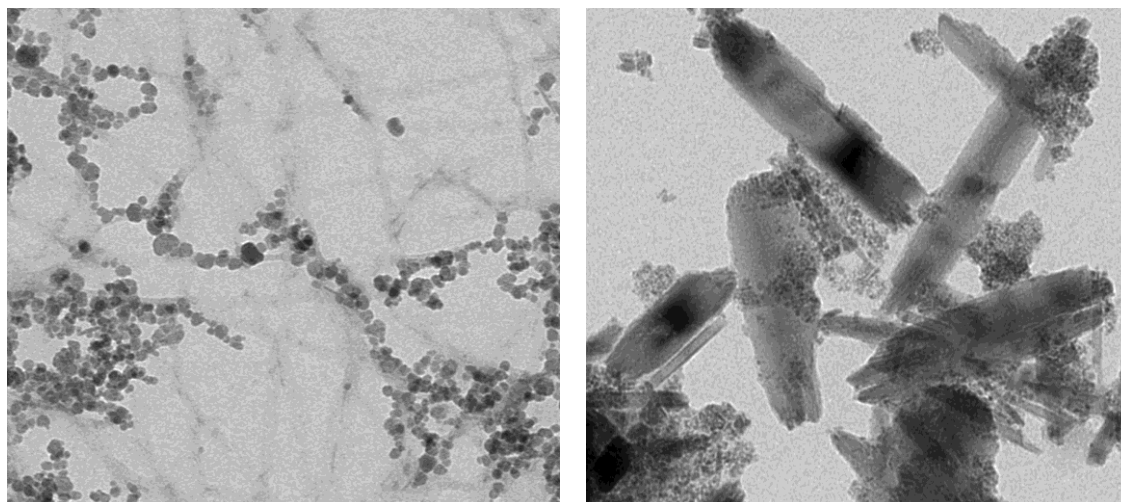
Interestingly, each couple of the diastereomeric mixture of the oligopeptide gave monodisperse structures similar to that obtained by the single diastereoisomer. This result is of strong value because allows the availability of a large amount of the peptide material useful for several applications. The stability of the oligopeptide assemblies in serum at 37°C was tested showing unmodified dimension opening the way to further studies as candidates for drug delivery.

## Gel synthesis of iron oxides nanoparticles

*Rafael Contreras-Montoya<sup>1</sup>, Laura Rodriguez-Arco,<sup>2</sup> Ana B. Bonhome-Espinosa,<sup>2</sup> Modesto T. Lopez-Lopez,<sup>2</sup> Juan M. Cuerva<sup>1</sup> and Luis Álvarez de Cienfuegos\*,<sup>1</sup>*

<sup>1</sup> *Facultad de Ciencias, Departamento de Química Orgánica, Universidad de Granada, 18071 Granada, Spain. [lac@ugr.es](mailto:lac@ugr.es).* <sup>2</sup> *Facultad de Ciencias, Departamento de Física Aplicada, Universidad de Granada, 18071 Granada, Spain.*

Short-peptides supramolecular gels are a subgroup of low molecular weight gelators that have found useful technological applications thank to their versatile nature.<sup>1</sup> In this work we have studied for the first time the behaviour of these types of gels as a medium to growth magnetite and goethite nanoparticles. The influence of the hydrogels and the protocols in the different kinds of structures obtained will be presented.



<sup>1</sup> Dasgupta, A.; Mondal, J. H.; Das, D. *RSC Adv.* **2013**, 3, 9117-9149.



# Self-assembly of PEGylated tetra-phenylalanine derivatives: structural insights from solution and solid state studies

C. Diaferia,<sup>1</sup> F.A. Mercurio,<sup>2</sup> C. Giannini,<sup>3</sup> T. Sibillano,<sup>3</sup> G. Morelli,<sup>1</sup> M. Leone,<sup>2</sup> A. Accardo<sup>1</sup>

<sup>1</sup>Department of Pharmacy and CIRPeB, University of Naples "Federico II", via Mezzocannone 16, 80134 Napoli (Italy) <sup>2</sup>Institute of Biostructure and Bioimaging (IBB), CNR, via Mezzocannone 16, 80134 Napoli (Italy) <sup>3</sup>Institute of Crystallography (IC), CNR, Via Amendola 122, 70126 Bari (Italy)

Peptide nanostructures containing diphenylalanines (FF), which constitutes the core motif of Alzheimer's  $\beta$ -amyloid, have been investigated for their mechanical, electrochemical and optical properties. More recently FF aggregates have been also proposed for applications in nanomedicine.<sup>1</sup> Despite the growing literature about FF, only few examples of aggregates based on tetraphenylalanines (F4) or their polymeric analogues, chemically modified with PEG or PEO chains at the N-terminus, have been reported until now.<sup>2,3</sup> We developed PEGylated tetra-phenylalanine conjugates (DOTA(Gd)-L<sub>6</sub>-F4 and DTPA(Gd)-L<sub>6</sub>-F4) as potential Magnetic Resonance Imaging (MRI) contrast agents.<sup>4</sup> In these F4-conjugates the aromatic framework is derivatized at N-terminus with a PEG chain and with DOTA or DTPA chelating agents for achieving gadolinium coordination. The replacement of the Gd(III) with radioactive metal ions (<sup>111</sup>In, <sup>67/68</sup>Ga) could open novel perspective in diagnostic field. As alternative, chelating agent could be replaced with active pharmaceutical ingredient (API). Here we report a physicochemical characterization of supramolecular aggregates of L<sub>6</sub>-F4 and its analogue containing DOTA chelating agent (DOTA-L<sub>6</sub>-F4) using several techniques (CD, IR, NMR, WAXS and SAXS). Results of this characterization open new knowledge on the comprehension of the molecular interactions at the bases of the hierarchical organization of molecules.

<sup>1</sup> Yan, X., Zhu, P, Li, J. *Chem. Soc. Rev.* **2010** 39(6), 1877-1890.

<sup>2</sup> Castelletto, V, Hamley, I. W. *Biophys. Chem.* **2009**, 141(2-3), 169-174.

<sup>3</sup> Tzokova, N., Fernyhough, C. M., Topham, P. D.; Sandon, N., Adams, D. J., Butler, M. F., Armes, S. P., Ryan, A. J. *Langmuir*, **2009**, 25(4), 2479-2785.

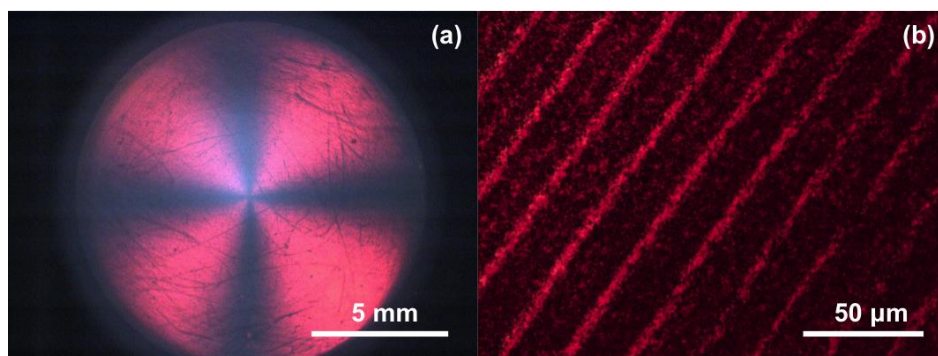
<sup>4</sup> Diaferia, C., Gianolio, E., Palladino, P., Arena, F., Boffa, C., Morelli, G., Accardo, A *Adv Funct. Mater.* **2015**, 25, 7003-7016.

# Aligning Perylene Bisimides for Enhanced Photoconductive Properties

*Emily R. Draper<sup>a</sup>, Matthew Wallace<sup>a</sup>, Oleksandr O. Mykhaylyk<sup>b</sup>, and Dave J. Adams<sup>a</sup>*

<sup>a</sup> University of Liverpool, UK. <sup>b</sup> University of Sheffield, UK.

We have shown that a series of amino acid functionalised perylene bisimides (PBI) form aggregates in solution that exhibit photoconductive behaviour upon irradiation exclusively with UV light.<sup>1</sup> PBIs show promise for use as n-type materials in organic electronics such as photovoltaic devices. The PBI molecules form fibre-like aggregates, these are formed by self-assembled  $\pi$ -stacking of the molecules in aqueous solution at high pH due to their hydrophobicity. By aligning these aggregates in solution before drying into a thin film, the pathway of the electron is shorter. A shorter path length shows a greater conductivity. We show by drying PBI solutions in the presence of a magnetic field, we are able to form well defined aligned structures compared to when allowed to dry in air. We also are able to align solutions and gels using shear to give reproducibly aligned films (Fig. 1a and b). These films show great directional dependence with larger currents along the alignment of the fibres than against the alignment of the fibres. This ability to reproducibly align the structures is essential for their use in application such as p-n heterojunctions.



**Figure 1.** Optical microscope images under cross-polarised light of (a) solution under shear and (b) a shear aligned dried film.

<sup>1</sup> E. R. Draper, J. J. Walsh, T. O. McDonald, M. A. Zwijnenburg, P. J. Cameron, A. J. Cowan and D. J. Adams, *J. Mat. Chem. C*, 2014, **2**, 5570-5575



## HIV-1 peptide inhibitor derived from the E2 protein of GB virus C conjugated to nanosystems

María J. Gómara<sup>1</sup>, Anna Paús<sup>1</sup>, Alberto Merino-Mansilla<sup>2</sup>, Víctor Sánchez-Merino<sup>2</sup>, Eloísa Yuste<sup>2</sup> and Isabel Haro<sup>1</sup>

(1) Unit of Synthesis and Biomedical Applications of Peptides,  
IQAC-CSIC, Barcelona

(2) AIDS Research Unit, Hospital Clinic, Barcelona

The use of nanocarriers for the targeted delivery of antiretroviral drugs significantly increases their therapeutic efficacy and safety, offering a large number of benefits such as protection of the drug against enzymatic degradation and their delivery to the specific site of action. They also provide conformational and physicochemical stability to the active molecule they transport, increase the cellular uptake of not very membrane-permeable drugs, reduce their cellular and tissue elimination, and maintain the prolonged release of active molecules at their site of action<sup>1</sup>.

This work involves the anti-retroviral study of a synthetic peptide derived from the N-terminal E2 protein of GB virus C which was previously defined as an entry HIV-1 inhibitor<sup>2</sup> incorporated on lipid vesicles and polymeric nanoparticles. These nanosystems were obtained following procedures previously described<sup>3,4</sup>. The lipid vesicles mimicking the lipid composition of HIV-1 membrane (Large Unilamellar Vesicles, LUVs composed of POPC/DPPC/SM/CHOL) rendered an increase in the peptide anti-HIV-1 activity about one order of magnitude.

These results demonstrate that the liposomes composed of a lipid raft-mimicking mixture facilitate targeted delivery of the inhibitor peptide to membrane subdomains where the process of entry of the HIV-1 virus in the cell takes place.

---

<sup>1</sup> Mallipeddi, R; Rohan, LC. . *Int. J. Nanomedicine* **2010**, 4, 533-547

<sup>2</sup> Eissman, K; Mueller, S; Stich, H; Jung, S; Zou, P; Jiang, S; Gross, A; Eichler, J; Fleckenstein, B; Reil, H. *PLoS One* **2013**, 8, e54452

<sup>3</sup> Vasconcelos, A; Vega, E; Pérez, Y; Gómara, MJ; García, ML; Haro, I. *Int. J. Nanomedicine* **2015**, 10, 609-631

<sup>4</sup> García, M; Alsina, MA; Reig, F; Haro I. *Vaccine* **2000**, 18, 276-283.



# Microarrays based on synthetic chimeric fibrin/filaggrin peptides for the diagnosis of Rheumatoid Arthritis

*M. José Gómara<sup>1</sup>, Javier Rodríguez<sup>1</sup>, Pablo Salvador<sup>2</sup>, M. José Bleda<sup>1</sup>, Raimon Sanmartí<sup>3</sup> and Isabel Haro<sup>1</sup>*

*(1) Unit of Synthesis and Biomedical Applications of Peptides, IQAC-CSIC, Barcelona*

*(2) Nanobiotechnology for Diagnostics Group, IQAC-CSIC, Barcelona*

*(3) Unit of Arthritis, Hospital Clínic, Barcelona*

Rheumatoid Arthritis (RA) is a chronic autoimmune disease that causes joint inflammation and extra-articular manifestations. To prevent progressive and irreversible structural damage, early diagnosis of RA is of paramount importance. We have recently obtained and highlighted the application of chimeric peptide bearing different citrullinated protein domains for the design of improved RA diagnosis/prognosis systems<sup>1-3</sup>. In the present work, we pursue the transfer of immunological assays from microtiter plates to microarray formats in order to allow the simultaneous analysis of several peptide sequences and the reduction in the volume of serum required from patients. Peptides were covalently immobilized on the microarray slides previously derivatized with epoxide groups. The immobilization of peptides or printing was conducted using a Miniarrayer (BioOdyssey™ Calligrapher™, BioRad) and the assay was carried out according to the protocol described previously for the ELISA plates<sup>1</sup>, using as secondary antibody anti-human IgG labelled with Alexa Fluor 647. Fluorescence of the spots was quantified in a scanner (ScanArray GX Plus, Perkin Elmer). A comparison of the results obtained by ELISA and Microarrays methodologies working with two cohorts of sera (96 from RA patients and 96 from healthy blood donors) will be presented.

<sup>1</sup> Pérez, ML; Gómara, MJ; Ercilla, G; Sanmartí, R; Haro, I. *J.Med.Chem.* **2007**, 50, 3573-3584.

<sup>2</sup> Sanmartí, R; Graell, E; Pérez, ML; Ercilla, G; Viñas, O; Gómez-Puerta, JA; Gratacós, J; Balsa, A; Gómara, MJ; Larrosa, M; Cañete, JD; Haro, I. *Arthritis Res. Ther.* 2009, 11(5):R135.

<sup>3</sup> Malakoutihak, M; Gómara, MJ; Gómez-Puerta, JA; Sanmartí, R; Haro, I. *J.Med.Chem.* **2011**, 54, 7486-7492.

## Structural chameleonic behaviour in peptides derived from choline binding repeats

Héctor Zamora-Carreras,<sup>a</sup> Beatriz Maestro,<sup>b</sup> Erik Strandberg,<sup>c</sup>  
Anne S. Ulrich,<sup>c</sup> Jesús M. Sanz,<sup>b</sup> & M. Angeles Jiménez<sup>a</sup>

<sup>a</sup> Instituto de Química Física Rocasolano (IQFR-CSIC), Madrid, Spain.

<sup>b</sup> Instituto de Biología Molecular y Celular, Universidad Miguel Hernández, Elche, Alicante, Spain. <sup>c</sup> Institute of Biological Interfaces (IBG-2), Karlsruhe Institute of Technology (KIT), Karlsruhe, Germany.

Choline-binding domains display a  $\beta\beta$ -solenoid structure formed by choline binding repeats (CBRs). Each CBR comprises a  $\beta$ -hairpin (approx. 14-residues) connected to the following repeat by a short linker sequence (about 6-residues). A choline molecule binds between consecutive CBRs. These domains are good models for understanding the folding and stability of repeat proteins, and also have applicability as affinity tags for protein purification and immobilisation. To find whether a minimal peptide comprising the  $\beta$ -hairpin core of a single CBR would conserve its native structure and its choline-binding capacity, we started the structural characterization of peptides derived from pneumococcal LytA autolysin. In these studies, we have found that a 14-mer peptide (LytA<sub>239-252</sub>) corresponding to the third CBR of LytA forms a very stable native-like  $\beta$ -hairpin in aqueous solution and also in the presence of trifluoroethanol, but it converts into a stable amphipathic  $\alpha$ -helix in the presence of detergent micelles.<sup>1</sup> Apart from its relevance in the fields of peptide design and protein folding, this result might be of potential applicability as a structure-switch biosensor. Hence, to get insights into the chemical-physical properties responsible for the unusual reversible  $\beta$ -hairpin to  $\alpha$ -helix transition exhibited by LytA<sub>239-252</sub>, we are analysing the effect of solvent conditions on the structural behaviour of designed LytA<sub>239-252</sub> variants, and of peptides corresponding to other CBR repeats of LytA by using CD and NMR.

<sup>1</sup>Zamora-Carreras, H.; Maestro, B.; Strandberg, E.; Ulrich, A. S.; Sanz, J. M.; Jiménez, M. Á. *Chem Eur J.* **2015**, *21*, 8076-8089.

## Halogenation promotes the self-assembly and gelation properties of DFNKF peptides

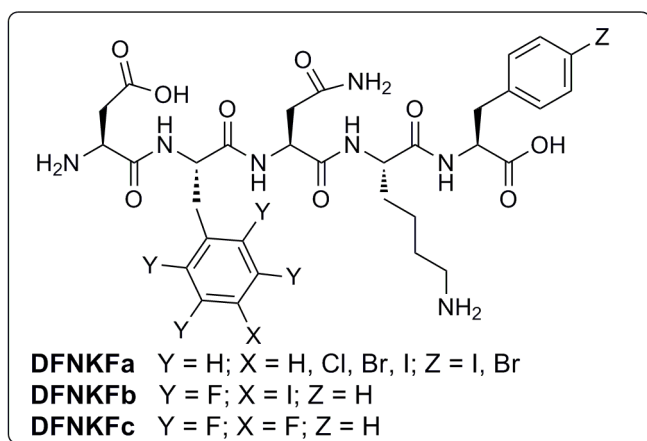
*Luisa Lasciari,<sup>a</sup> Arianna Bertolani,<sup>a</sup> Andrea Pizzi,<sup>a</sup> Alessandro Gori,<sup>a</sup> Nicola Demitri,<sup>b</sup> Giuseppe Resnati,<sup>a</sup> Pierangelo Metrangolo<sup>a,c</sup>*

*a) NFMLab, Department of Chemistry, Materials, and Chemical Engineering “Giulio Natta”, Politecnico di Milano, Via L. Mancinelli 7, 20131 Milano, Italy.*

*b) Elettra-Sincrotrone Trieste, 34149 Basovizza, Trieste, Italy.*

*c) VTT – Technical Research Centre of Finland, FI-02044 VTT Espoo, Finland.*

Amyloid fibrils, formed by the tight assembly of  $\beta$ -sheet-like structures, are widely studied because of their disease-related implications and because the robustness of their supramolecular architectures can be exploited in nanostructured bio-inspired materials.<sup>1</sup> It is at the same time true that, the halogenation of proteins is a consequence of oxidative-stress, which is often associated to misfolding and aggregation.<sup>2</sup> Choosing as a model fibrillation system the DFNKF core sequence of



hCT, we have demonstrated that halogenation strongly promoted fibrils, and thus, hydrogels formation.<sup>3</sup> Considering that DFNKF<sub>a</sub> aggregation performances increase as the halogen-bond donor (XB-d) character of the halogen increases, we then synthesised and studied the properties of DFNKF<sub>b</sub>, containing a

more effective XB-d aminoacid. Indeed, we found better gelation abilities of DFNKF<sub>b</sub> if compared to DFNKF<sub>a</sub>, while DFNKF<sub>c</sub>, lacking of XB-d residues, is a worse gelator. This point out a positive role of XB interactions in addition to common intermolecular interactions that are recognized as responsible in peptide systems aggregation.

<sup>1</sup> Mankar, S.; Anoop, A.; Sen, S.; Maji, S.K. *Nano Rev.* **2011**, 2, 6032-6043.

<sup>2</sup> Mazzulli, J.R.; Hodara, R.; Lind, S.; Ischiropoulos, H. *Prof. Rev.* **2006**, 4, 123-133.

<sup>3</sup> Bertolani, A.; Pirrie, L.; Stefan, L.; Metrangolo, P.; et al. *Nature Commun.* **2015**, 6, 7574.



---

# Protein nanomaterials from renewable resources

*Christofer Lendel*

*Dept. of Chemistry, KTH Royal Institute of Technology, Stockholm, Sweden*

Proteins are Nature's own high-performance materials providing both extraordinary mechanical properties (e.g. muscles, silks) and sophisticated functionality (e.g. adhesion, biological signalling). The key to copy these properties to man-made materials lies in controlling the hierarchical material structure, from molecular- to macro level. From this perspective, the self-assembly of proteins and peptides into ordered nanofibers is of utmost interest. The use of protein nanofibers to create new materials is an expanding field of research. Much of the efforts in this field of research have, however, been made on recombinantly produced and highly purified starting material. For large-scale applications, methods for production of nanofibers and macroscopic materials from crude starting materials are needed. We explore the potential of producing the nanocomposites based on proteins from renewable source, e.g. plant proteins. The questions we address are:

- 1) How can we produce nanofibers from various crude starting materials?
- 2) How can we control the hierarchical assembly of nanofibers into macroscopic materials?
- 3) How can the nanofibers be used to create improved or new functionalities in bioplastics?

## Injectable peptide hydrogels for controlled-release

C. Martin,<sup>1</sup> E. Oyen,<sup>1</sup> M. Bibian,<sup>1</sup> J. Gardiner,<sup>2</sup> B. Van Mele,<sup>3</sup> A. Madder,<sup>4</sup>  
R. Hoogenboom,<sup>5</sup> M. Spetea<sup>6</sup> and S. Ballet<sup>1</sup>

<sup>1</sup>Research Group of Organic Chemistry, Vrije Universiteit Brussel, Belgium. <sup>2</sup>CSIRO Materials Science & Engineering, Australia. <sup>3</sup>Vrije Universiteit Brussel, Physical Chemistry and Polymer Science, Belgium. <sup>4</sup>Organic and Biomimetic Chemistry Research Group, Ghent University, Belgium. <sup>5</sup>Supramolecular Chemistry Group, Ghent University, Belgium. <sup>6</sup>Institute of Pharmacy, University of Innsbruck, Austria.

Currently, most drugs are directly administered into patients orally or systemically, via parenteral routes. Therefore, to get the desired therapeutic effect, high doses are required due to substantial biodegradation of the drug prior to interaction with the biological target, resulting in the appearance of adverse effects. To overcome this, hydrogels have been reported as suitable controlled drug-delivery systems. These systems present several advantages such as the protection of the drug against the enzymatic degradation by encapsulation in the hydrogel network, while maintaining the therapeutic plasma drug concentration over a long period of time. Consequently, lower dosage and frequency of administration are possible and result in an improvement of the drug efficacy while reducing the risk of side effects.

In this work, a new family of hydrogel-forming peptides was designed starting from a short, tunable and amphipathic hexapeptide hydrogelator.<sup>1</sup> After characterization, and in order to study their eventual therapeutic potential, the hydrogels have been used for entrapment and sustained release of opioid drugs. The *in vitro* drug release properties and hydrogel toxicity (cell viability experiments) were also determined. Based on the best physicochemical, mechanical, and noncytotoxic properties, selected hydrogels were investigated for *in vivo* release of opioids. Opioid administration by subcutaneous injection and subsequent testing in the tail-flick assay (acute pain model), showed sustained antinociceptive effects over longer periods of time, as compared to drug injections in saline solutions.<sup>2</sup>

<sup>1</sup> Bibian, M.; Mangelschots, J.; Gardiner, J.; Waddington, L.; Acevedo, M. M. D.; De Geest, B. G.; Van Mele, B.; Madder, A.; Hoogenboom, R. and Ballet, S. *J. Mater. Chem. B* **2015**, 3, 759-765.

<sup>2</sup> Martin, C.; Oyen, O.; Mangelschots J.; Bibian, M.; Ben Haddou, T.; Andrade, J.; Gardiner, J.; Van Mele, B.; Madder, A.; Hoogenboom, R.; Spetea, M. and Ballet, S. *Med. Chem. Com.* DOI:10.1039/c5md00440c.



# The evolution of surface-coating molecules in biomaterials science: integrin-binding peptidomimetics and multifunctional platforms

R. Fraioli,<sup>1</sup> F. Rechenmacher,<sup>2</sup> S. Neubauer,<sup>2</sup> J. M. Manero,<sup>1</sup> H. Kessler,<sup>2</sup>  
F. J. Gil,<sup>1</sup> and C. Mas-Moruno<sup>1\*</sup>

1. Group of Biomaterials, Biomechanics and Tissue Engineering & Centre for Research in NanoEngineering, Technical University of Catalonia, Barcelona, Spain
2. Institute for Advanced Study and Center for Integrated Protein Science, Technical University of Munich, Garching, Germany

**E-mail:** carles.mas.moruno@upc.edu

The biofunctionalization of implant materials with integrin-binding peptides and proteins derived from the extracellular matrix has long been investigated to improve cell-material interactions. However, classical biomimetic strategies present some limitations and may prove insufficient to elicit the biological signals required in the process of tissue regeneration.<sup>1</sup>

To overcome these limitations, we have focused on the development of innovative biological coatings with enhanced integrin-specificity and/or multiple biological potential, aiming at designing cell-instructive and versatile biomaterials for tissue regeneration.

In this work two novel strategies will be presented: i) the use of two RGD peptidomimetics with high activity and selectivity for integrins  $\alpha\beta3$  or  $\alpha5\beta1$ ;<sup>2</sup> and ii) a peptide-based platform with the capacity to simultaneously present two distinct bioactive motifs in a chemically controlled fashion.<sup>3</sup>


The potential of these biomolecular tools to engineer biomaterials with bone regenerative properties has been proven *in vitro* using bone-like and mesenchymal stem cells, and *in vivo* in a calvarial defect model in rats.

---

<sup>1</sup> Williams, D.F. *Biomaterials* **2011**, 32, 4195-4197.

<sup>2</sup> a) Rechenmacher, F.; et al. *Angew. Chem. Int. Ed.* **2013**, 52, 1572-157; b) Fraioli, R.; et al. *Colloid Surf. B-Biointerfaces* **2015**, 128, 191-200.

<sup>3</sup> Mas-Moruno, C.; et al. *ACS Appl. Mater. Interfaces* **2014**, 6, 6525-6536.



# Hierarchical surface-mediated self-assembly of triphenylalanine with N- and C-terminal fluorenylmethyl groups

E. Mayans<sup>a</sup>, G. Fabregat<sup>a</sup>, J. Casanovas<sup>b</sup>,  
C. Cativiela<sup>c,\*</sup>, J. Puiggali<sup>a,\*</sup> and C. Alemán<sup>a,\*</sup>

<sup>[a]</sup> *Departament d'Enginyeria Química, ETSEIB, Universitat Politècnica de Catalunya,  
Diagonal 647, Barcelona E-08028, Spain*

<sup>[b]</sup> *Departament de Química, EPS, Universitat de Lleida, c/ Jaume II nº 69, Lleida E-25001,  
Spain*

<sup>[c]</sup> *Department of Organic Chemistry and Instituto de Síntesis Química y Catalisis  
Homogenea (ISQCH), University of Zaragoza-CSIC, 50009 Zaragoza, Spain*

The 9-fluorenylmethoxycarbonyl (Fmoc) N-capped oligophenylalanine derivatives (F<sub>n</sub>, n=2,3,4) form a wide range of micro- and nano-assemblies that include fibrils, nanoplates and hydrogels depending on the preparation and incubation conditions. Particularly, morphological differences in the surface self-assembly are tested for Fmoc-FF by varying peptide concentration and the pH of the aqueous media.<sup>1</sup>

In this work we have investigated the influence of the substrate in the formation of supramolecular assemblies by a highly aromatic FFF derivative capped with N- and C-terminal fluorenylmethyl groups. Surface-induced assemblies in 9 substrates that differ in its chemical nature were examined. Dendritic-like morphologies were obtained for glass and plasma treated polystyrene. Micrometric jellyfish-like structures were obtained onto silanized glass. Crystalline assemblies and birefringent spherulites were observed for mica, whereas structures grown onto steel AISI 316, silicon wafer and evaporated carbon coatings consist on spherical aggregates arranged in necklaces. The reported structures point out that come from a complex hierarchical assembly of densely packed nano-assemblies.

---

<sup>1</sup> Liu, Yun; Xu, Xiao-Ding; Chen, Jing-Xiao; Cheng, Han; Zhang, Xian-Zheng; Zhuo, Ren-Xi; *Colloids Surf., B* **2011**, 87, 192-197.

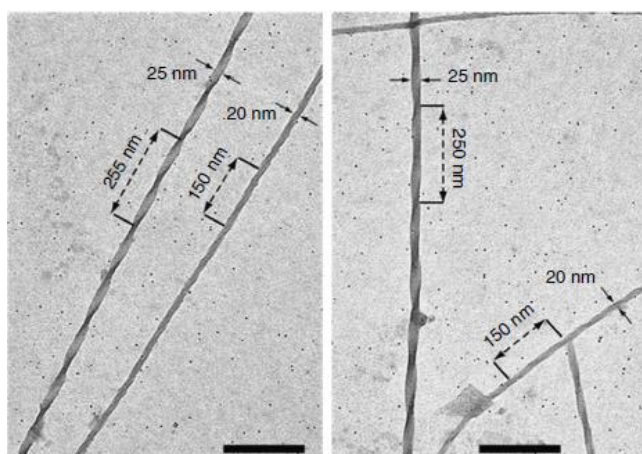
## Halogen bonding: A new supramolecular tool in the molecular self-assembly of peptides

Pierangelo Metrangolo,<sup>a,b</sup> Luisa Lascialfari,<sup>a</sup> Arianna Bertolani,<sup>a</sup> Andrea Pizzi,<sup>a</sup> L oic Stefan,<sup>a</sup> Alessandro Gori,<sup>a</sup> Giuseppe Resnati<sup>a</sup>

a) NFMLab, Department of Chemistry, Materials, and Chemical Engineering "Giulio Natta", Politecnico di Milano, Via L. Mancinelli 7, 20131 Milano, Italy.

b) VTT – Technical Research Centre of Finland, FI-02044 VTT Espoo, Finland.

A halogen bond occurs when there is evidence of a net attractive interaction between an electrophilic region associated with a halogen atom in a molecular entity and a nucleophilic region in another, or the same, molecular entity.<sup>1</sup>



Although many modifications of amyloidogenic sequences have been utilized to tune their self-assembly behaviour, halogenation has rarely been pursued. The advantage of a strategy based on the introduction of halogen atoms on peptide motifs lies in the fact that halogenation is a minimal structural modification, which,

on the other hand, may induce a large difference in the peptide supramolecular behaviour as a consequence of the rich variety of noncovalent interactions given by halogen atoms.<sup>2</sup> In this presentation we show how the halogen bond can be used to promote the molecular self-assembly of peptides. We have applied this new supramolecular concept to the augmented fibrillation of amyloidogenic peptides and proteins, such as DFNKF,<sup>3</sup> KLVFF, and hCT. Implications of oxidative stress-induced halogen of proteins are discussed in terms of biomarkers of diseases such as Parkinson and Alzheimer's. The obtainment of a novel unnatural amino acid functioning as strong halogen-bond donor may pave the way to totally new design principles in peptide-based molecular self-assembly.

<sup>1</sup> Metrangolo, P.; *et al. Chem. Rev.* **2016**, DOI: 10.1021/acs.chemrev.5b00484.

<sup>2</sup> Metrangolo, P.; *et al. Acc. Chem. Res.* **2013**, *46*, 2686.

<sup>3</sup> Metrangolo, P.; *et al. Nature Commun.* **2015**, *6*, 7574.



## Ultrashort peptides as bifunctional nanomaterials

*McCloskey A.P, Gilmore S.M, Gilmore B.F, Laverty G.*

*Bifunctional Nanomaterials Research Group, School of Pharmacy, Queen's University Belfast*

Advances in modern medicine have resulted in increased life expectancy and quality of life for patients. Biomaterials have contributed greatly to these advances in healthcare however, despite their many uses in replacing/ facilitating normal physiological functions, they provide an ideal surface for bacterial adhesion and subsequent development of biofilm infections.<sup>1</sup>

The presence of and sometimes trauma caused by insertion of a biomaterial, stimulates the host immune response system to upregulate inflammatory mediators. These can lead to destruction or impairment of the biomaterial. This in turn, prolongs illness and delays healing impacting negatively on the patient's recovery.<sup>2</sup>

Peptides are ideal candidates to help overcome these associated challenges due to their vast bioavailability in nature, biocompatibility and biodegradability. Ultrashort peptides are of increasing interest in novel research and development due to their economic advantages over longer counterparts benefiting their potential transition from the lab bench to the clinic.<sup>3</sup>

In this research an ultrashort diphenylalanine- dilysine peptide sequence has been conjugated to the clinically used non-steroidal anti-inflammatory drugs (NSAIDs)- naproxen, indomethacin and ibuprofen. These peptides have inherent dual antimicrobial and anti-inflammatory activity and at suitable concentrations NSAID conjugated peptides have been shown to possess the ability to self-assemble.<sup>4</sup>

Broad-spectrum antimicrobial activity was demonstrated against a range of Gram positive and Gram negative bacteria. Anti-inflammatory activity was tested in terms of inhibition of two key enzymes, cyclooxygenase-1 and -2 and conjugation of the NSAID to the peptide did not affect inhibition of these. An ability to form hydrogels was assessed in terms of a vial inversion assay, rheological analysis and cryo-SEM and TEM imaging to determine the presence of nanofibrous networks. Cytotoxicity and hemolytic activity assays showed that the peptide-NSAID conjugates were more selective for bacterial cells.

The findings of this research are very promising and indicate that peptide- drug conjugates such as these have great potential as bifunctional materials for biomedical applications.

<sup>1</sup> Karp, J; Langer, R *Current Opinion in biotechnology*, **2007**, 18.5, 454-459.

<sup>2</sup> Morais,J; Papadimitrakopoulos,F; Burges,D *The AAPS journal*, **2012**, 12.2, 188-196.

<sup>3</sup> Laverty,G; McCloskey,A; Gilmore,B; Jones,D; Zhou,J; Xu,B, *Biomacromolecules*, **2014**, 15(9), 3429-3439.

<sup>4</sup> Li,J; Kuang,Y; Shi,J; Gao,Y; Zhou,J; Xu,B, *Beilstein journal of organic chemistry*, **2013**, 9(1), 908-917.

# $\beta^{2,3}$ diaryl amino acids as molecular tools for the preparation of peptide nanotubes

S. Pellegrino,<sup>a</sup> R. Bucci,<sup>a</sup> F. Meneghetti,<sup>a</sup> N. Ferri,<sup>b</sup> M. L. Gelmi,<sup>a</sup> M. Reches<sup>c</sup>

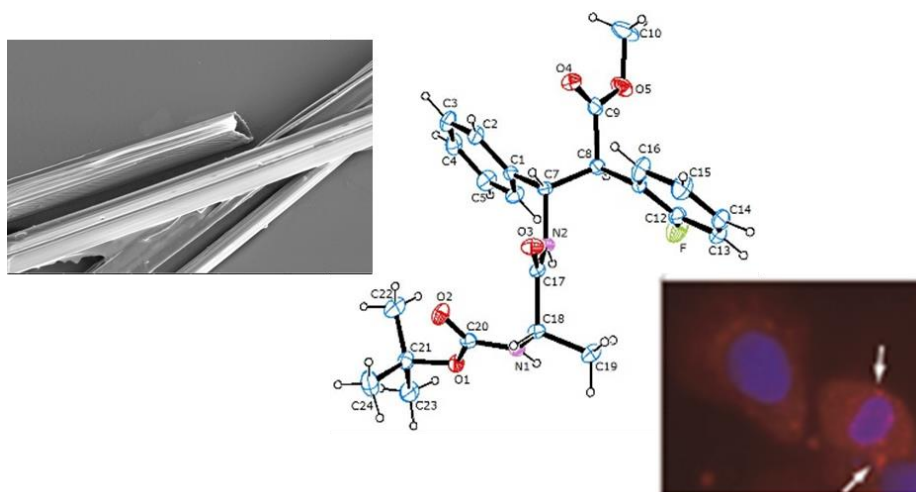
<sup>a</sup> Università degli Studi di Milano, Dipartimento di Scienze Farmaceutiche, Milan, Italy

<sup>b</sup> Università degli Studi di Padova, Dipartimento di Scienze del Farmaco, Padua, Italy

<sup>c</sup> The Hebrew University of Jerusalem, Institute of Chemistry, Jerusalem, Israel

Peptide nanotubes are a class of organic nanotubes attracting great interest due to their potential applications in nanotechnology and biomedicine. On the other hand, one drawback of using peptides is their high susceptibility toward proteases and their short half-lives.

Here, we present the preparation of proteolytic stable nanotubes containing fluorine-substituted (S,S)- $\beta^{2,3}$ -diaryl-amino acid and (L)-alanine. In the solid state, the supramolecular architecture is strengthened by intermolecular N-H...O hydrogen bonds, C $\pi$ -H...O, C $\pi$ -H...F and van der Waals interactions.<sup>1</sup> Furthermore, the nanotubes were able to incorporate small organic molecules, to enter the cell and to locate in the cytoplasmic/perinuclear region. Therefore they represent interesting candidates for biomedical applications.



<sup>1</sup> Bonetti, A. ; Pellegrino, S.; Das, P.; Yuran, S.; Bucci, R.; Ferri, N.; Meneghetti, F.; Castellano, C.; Reches, M.; Gelmi, M.L. *Org. Lett.*, **2015**, 17, 4468–4471

# Elastin-like peptide nanoparticles for crossing the blood-brain barrier

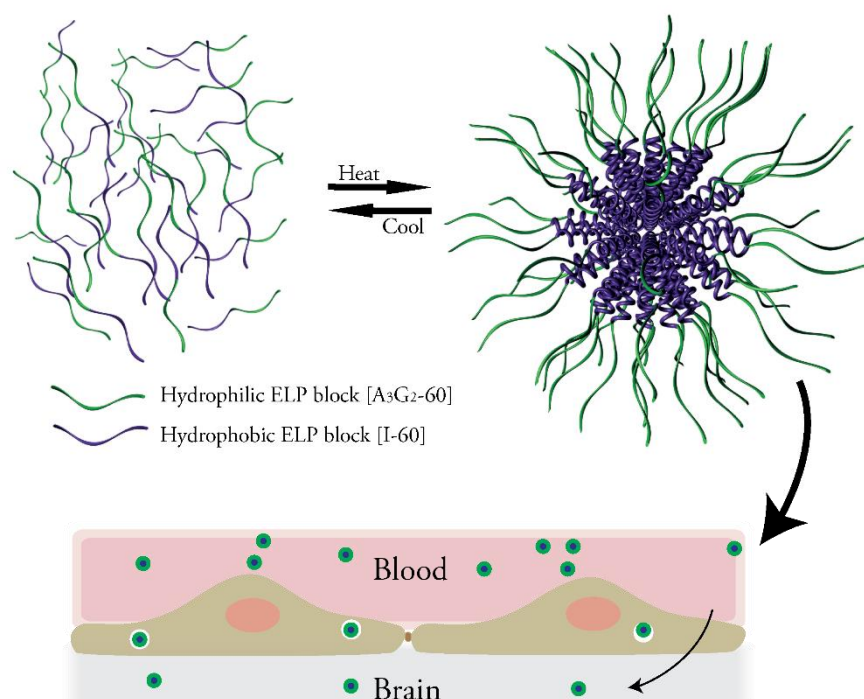
Jan Pille<sup>1</sup>, L. van Oppen<sup>2,3</sup>, J.Smeitink<sup>3</sup>, J.C.M van Hest<sup>1</sup>

<sup>1</sup>Bio-Organic Chemistry, Institute for Molecules and Materials, Radboud University

<sup>2</sup>Department of Biochemistry, Radboud Institute for Molecular Life Sciences, Radboud University Medical Center

<sup>3</sup>Department of Pediatrics, Radboud Institute for Molecular Life Sciences, Radboud University Medical Center

Treating neurological disorders demands overcoming the blood-brain barrier. Hijacking the active transport mechanisms of this tight, selective endothelial cell layer shows great promise in administering therapeutic agents to the brain. A potential carrier system needs to be well-defined, bio-compatible and able to display functional moieties for transcytosis. Here we develop elastin-like peptide (ELP) nanoparticles that are highly monodisperse, stable down to nanomolar concentrations and are able to display a range of functional peptides. The GM1b-binding peptide G23<sup>1</sup> is used to induce uptake and transcytosis of ELP nanoparticles by brain endothelial cells.



<sup>1</sup> Georgieva, Julia V.; Rene P. Brinkhuis *et al.* *Angew Chem Int Ed Engl* **2012** 51(33): 8339-8342



## Tuning amyloid peptide self-assembly by halogenation

*Andrea Pizzi,<sup>a</sup> Lisa Pirrie,<sup>b</sup> Arianna Bertolani,<sup>a</sup> Nicola Demitri,<sup>c</sup>*

*Giancarlo Terraneo,<sup>a</sup> Giuseppe Resnati,<sup>a</sup> Pierangelo Metrangolo<sup>a,b</sup>*

*a) NFMLab, Department of Chemistry, Materials, and Chemical Engineering “Giulio Natta”, Politecnico di Milano, Via L. Mancinelli 7, 20131 Milano, Italy.*

*b) VTT – Technical Research Centre of Finland, FI-02044 VTT Espoo, Finland.*

*c) Elettra-Sincrotrone Trieste, 34149 Basovizza, Trieste, Italy.*

Alzheimer’s disease (AD) is a pathology associated with the formation of  $\beta$ -sheet containing fibrils. The peptide associated with this disease, constituted by 40-42 aminoacids, is called Amyloid beta ( $A\beta$ ). Different groups have investigated the fibrillation of several fragments of  $A\beta$ . The results suggest that the sequence  $A\beta$  (16-20) -KLVFF- (Lys-Leu-Val-Phe-Phe) is critical for fibrillation<sup>1</sup>. There is no current evidence of halogenation in AD; however, there is substantial evidence of oxidative stress – which may result in a halogenated biomolecule – playing an important role in this disease<sup>2</sup>.

In this contribution we report on a study of various halogen-modified derivatives of KLVFF. We demonstrated by different techniques like AFM, TEM, and Congo Red staining that varying the halogen position in the sequence confers very different structural properties, as previously reported for another amyloidogenic sequence<sup>3</sup>. In general, the presence of a halogen atom in the 4-position (internal phenylalanine) of the peptide sequence increases the rate of fibrillation, since we obtained amyloid hydrogels using concentrations four time lower compared to the minimal gelation concentration of the wild type sequence. An unexpected behavior characterizes the fragments having just a halogen in the 5-position (terminal phenylalanine), since they form crystals rather than fibrils. Finally, di-iodination confers polymorphism to the sequence by varying concentration and pH.

<sup>1</sup> Krysmann, J. M.; Castelletto, V.; Kelarakis, A.; Hamley, I. W.; Hule, R. A. *Biochem.* **2008**, *47*, 4597-4605.

<sup>2</sup> Smith, D. G.; Cappai, R.; Barnham, K. J. “The redox chemistry of the Alzheimer’s disease amyloid  $\beta$  peptide” *Biochim. Biophys. Acta* **2007**, *1768*, 1976-1990.

<sup>3</sup> Bertolani, A.; Pirrie, L.; Stefan, L.; Houbenov, N.; Haataja, J. S.; Catalano, L.; Terraneo, G.; Giancane, G.; Valli, L.; Milani, R.; Ikkala, O.; Resnati, G.; Metrangolo, P. *Nat. Commun.* **2015**, *6*:7574

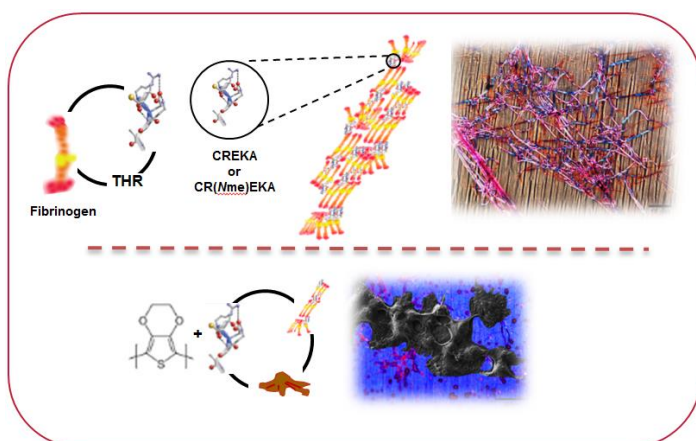
# Fibrin Association at Hybrid Biointerfaces Made of Clot-Binding Peptides and Polythiophene

*Anna Puiggali-Jou,<sup>1,2</sup> Luis J. del Valle,<sup>1,2,\*</sup> Elaine Armelin,<sup>1,2</sup> Joan Torras,<sup>3</sup> and Carlos Alemán<sup>1,2,\*</sup>*

<sup>1</sup> *Departament d'Enginyeria Química, E. T. S. d'Enginyeria Industrial de Barcelona, Universitat Politècnica de Catalunya, Diagonal 647, Barcelona E-08028, Spain*

<sup>2</sup> *Centre for Research in Nano-Engineering, Universitat Politècnica de Catalunya, Edifici C', C/Pasqual i Vila s/n, Barcelona E-08028, Spain*

<sup>2</sup> *Departament d'Enginyeria Química, EEI, Universitat Politècnica de Catalunya, Av. Pla de la Massa, 8, 08700 Igualada, Spain*



**Scheme 1.** Illustration of how peptides influenced fibrin polymerization and facilitate cell adhesion.

PEDOT-CREKA than for PEDOT-CR(NMe)EKA. This feature suggests that CREKA molecules enter into the PEDOT matrix individually or forming small groups (dimers or trimers), whereas entrapped CR(NMe)EKA forms bigger aggregates. Furthermore, the electrochemical and electrical behaviors of PEDOT improve upon the incorporation of the peptides. Both CREKA and CR(NMe)EKA affect fibrinogen thrombin-catalyzed polymerization causing the immediate formation of fibrin, whereas in absence of thrombin this is only observed for CR(NMe)EKA. The fibrin-adsorption capacity of fibrin decreases as follows: PEDOT-CR(NMe)EKA > PEDOT-CREKA  $\approx$  bare steel > PEDOT. PEDOT-peptide films coated with fibrin promote the attachment of metastatic cells with respect to normal cells. However, this clearly manifested selectivity is lost in absence of surface adsorbed fibrin.

Electroactive conducting polymer-peptide biocomposites formed by poly(3,4- ethylenedioxythiophene) (PEDOT) and CREKA or CR(NMe)EKA (where Glu has been replaced by *N*-methyl-Glu) have been prepared and their properties compared (**Scheme 1**).

The surface topology is considerably more leveled for





## Engineering of single-chain polypeptides to self-assemble as BBB-crossing protein nanoparticles

*Naroa Serna*<sup>1, 2, 3</sup>, *María Virtudes Céspedes*<sup>3, 4</sup>, *Paolo Saccardo*<sup>1, 2, 3</sup>,  
*Zhikun Xu*<sup>1, 2, 3</sup>, *Ugutx Unzueta*<sup>1, 2, 3</sup>, *Patricia Álamo*<sup>3, 4</sup>, *Mireia*  
*Pesarrodona*<sup>1, 2, 3</sup>, *Ramón Mangués*<sup>3, 4</sup>, *Neus Ferrer-Miralles*<sup>1, 2, 3</sup>,  
*Esther Vazquez*<sup>1, 2, 3</sup>, *Antonio Villaverde*<sup>1, 2, 3</sup>

<sup>1</sup> *Institut de Biotecnologia i de Biomedicina, Universitat Autònoma de Barcelona, Bellaterra, 08193 Barcelona, Spain*

<sup>2</sup> *Departament de Genètica i de Microbiologia, Universitat Autònoma de Barcelona, Bellaterra, 08193 Barcelona, Spain*

<sup>3</sup> *CIBER de Bioingeniería, Biomateriales y Nanomedicina (CIBER-BBN), Bellaterra, 08193 Barcelona, Spain*

<sup>4</sup> *Oncogenesis and Antitumor Drug Group, Biomedical Research Institute Sant Pau (IIB-SantPau), Hospital de la Sant Creu i Sant Pau, C/ Sant Antoni Maria Claret, 167, 08025 Barcelona, Spain*

The major challenge for the treatment of brain disorders is to develop optimal carriers to deliver therapeutic molecules, as chemicals or nucleic acids, to the Central Nervous System (CNS). Apart from the renal clearance that affects the retention of small molecules in the blood stream, the main obstacle to the targeted delivery is the blood–brain barrier (BBB) that acts as a filter between the bloodstream and the brain.

The aim of this work is the design, production and characterization of self-assembling protein nanoparticles directed to CNS, and to assess their biodistribution. Likewise, we have explored the potential influence of a supramolecular protein organization in BBB specific targeting.

Briefly, two different brain cell binding peptide domains, SEQ1 and Angiopep, were both engineered in order to increase their positive electrostatic charge to promote protein nanoparticle assembly. Each of these modified domains, fused to His-tagged GFP protein were produced in *E.coli* expression system and further characterized.

---

SEQ1charge8 peptide generates nanoparticles of about 30-40 nm. The nanoparticulate presentation dramatically enhances in vitro, LDLR-dependent cell penetrability of the protein compared to the parental monomeric version, but the assembled protein does not show any enhanced brain targeting upon systemic administration.

The presentation of protein chemicals in form of nanoparticles is advantageous when addressing non brain metastatic cancers, preventing renal clearance and ensuring re-circulation until cell binding and uptake. However, our data demonstrate that this principle might not apply to brain targeting, as the BBB imposes steric constraints different from those in systemic circulation. Irrespective of this fact, the fine engineering of proteins electrostatic profiles is revealed here as a powerful tool for the rational functional design of building blocks and of protein-only nanoparticles.

- 
1. Unzueta, U.; Ferrer-Miralles, N.; Cedano, J.; Zikung, X.; Pesarrodon, M.; Saccardo, P.; Garcia-Fruitos, E.; Domingo-Espin, J.; Kumar, P.; Gupta, K. C.; et al. Non-Amyloidogenic Peptide Tags for the Regulatable Self-Assembling of Protein-Only Nanoparticles. *Biomaterials* 2012, 33, 8714–8722.
  2. Vazquez, E.; Roldan, M.; Diez-Gil, C.; Unzueta, U.; Domingo-Espin, J.; Cedano, J.; Conchillo, O.; Ratera, I.; Veciana, J.; Daura, X.; et al. Protein Nanodisk Assembling and Intracellular Trafficking Powered by an Arginine-Rich (R9) Peptide. *Nanomedicine (London, U.K.)* 2010, 5, 259–268.
  3. Unzueta, U.; Cespedes, M. V.; Ferrer-Miralles, N.; Casanova, I.; Cedano, J. A.; Corchero, J. L.; Domingo-Espin, J.; Villaverde, A.; Mangues, R.; Vazquez, E. Intracellular CXCR4+ Cell Targeting with T22-Empowered Protein-Only Nanoparticles. *Int. J. Nanomed.* 2012, 7, 4533–4544.
  4. Ferrer-Miralles, N.; Rodriguez-Carmona, E.; Corchero, J. L.; Garcia-Fruitos, E.; Vazquez, E.; Villaverde, A. Engineering Protein Self-Assembling in Protein-Based Nanomedicines for Drug Delivery and Gene Therapy. *Crit. Rev. Biotechnol.* [Epub ahead of print]. DOI: 10.3109/07388551.2013.833163. Published online Oct 9, 2013.

## Electroactive polymer modified with RGD-derivated peptide for cell adhesion

*Jordi Triguero,<sup>a</sup> Silvana Maione,<sup>a,b</sup> Ana M. Gil,<sup>c</sup> Georgina Fabregat,<sup>a,b</sup> Lluís J. del Valle,<sup>a,b</sup> David Zanuy,<sup>a</sup> Oscar Bertran,<sup>d</sup> Carlos Cativiela<sup>\*c</sup> and Carlos Alemán<sup>\*a,b</sup>*

*a) Departament d'Enginyeria Química, ETSEIB, Universitat Politècnica de Catalunya, Avda. Diagonal 647, Barcelona E-08028, Spain*

*b) Center for Research in Nano-Engineering, Universitat Politècnica de Catalunya, Avda. Diagonal 647, Barcelona E-08028, Spain*

*c) Departamento de Química Orgánica, ISQCH, Universidad de Zaragoza - CSIC, 50009 Zaragoza, Spain*

*d) Departament de Física Aplicada, Escola d'Enginyeria d'Igualada, Universitat Politècnica de Catalunya, Pça Rei 15, Igualada 08700, Spain*

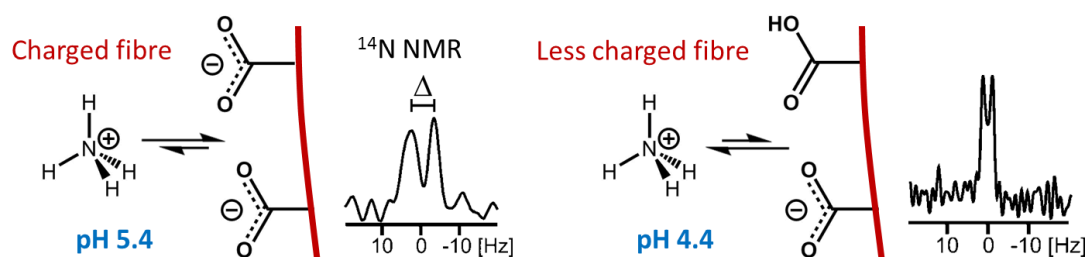
Electroactive polymer–peptide conjugates have been synthesized by combining poly(3,4-ethylenedioxythiophene), a polythiophene derivative with outstanding properties, and an Arg-Gly-Asp (RGD)-based peptide in which Gly has been replaced by an exotic amino acid bearing a 3,4-ethylenedioxythiophene ring in the side chain. The incorporation of the peptide at the ends of preformed PEDOT chains has been corroborated by both FTIR and X-ray photoelectron spectroscopy. The effect of the incorporation of the peptide at the ends of PEDOT chains on the surface and electrochemical properties have been characterized. Density functional theory calculations have been used to ascertain the conformational preferences of the peptide. Electroactive surfaces prepared using the conjugates displayed the higher bioactivities in terms of cell adhesion, with the relative viabilities being dependent on the roughness, wettability and electrochemical activity of the conjugate. In addition to the influence of the peptide fragment in the initial cell attachment and subsequent cell spreading and survival, the results indicate that PEDOT promotes the exchange of ions at the conjugate–cell interface.

# Solution-state NMR to understand peptide gel fibre surface chemistry and gelation

*Matthew Wallace, Jonathan A. Iggo and Dave J. Adams*

*Department of Chemistry, University of Liverpool, Crown Street, Liverpool, L69 7ZD, UK.*

Supramolecular hydrogels can be formed *via* the self-assembly in solution of low molecular weight peptide-based gelators and have a range of applications as diverse as cell culturing and catalysis. A number of well-established analytical techniques exist for the study of such hydrogels at the molecular level, the focus being on how the gelator molecules are packed together in the self-assembled gel fibres.<sup>1</sup> Hydrogels, however, are hierarchical systems with the primary gel fibres further associating with one another to form larger-scale structures that constitute the bulk macroscopic gels. Techniques for probing the surface chemistry of the gel fibres are thus required in order to build a better understanding of both the final hydrogels and the processes by which they are formed; however, such techniques remain relatively underdeveloped. To this end, we are developing a range of techniques based on solution-state NMR spectroscopy to probe the surface charge, hydrophobicity and ion binding dynamics of hydrogel fibres. The fibres themselves are invisible by conventional solution-state NMR, but much valuable information can be obtained by studying instead their selective interaction with probe molecules dissolved in the bulk solution of the gel. For example, by studying the relative residual quadrupolar couplings, ( $\Delta$ ), of  $\text{NH}_4^+$  and isopropanol- $\text{d}_8$  as the pH of the bulk solution is decreased, we are able to follow the loss of charge from the gel fibres which leads to a stiffening and contraction of the bulk gel.<sup>2</sup> In other systems, we are also able to observe the selective displacement of  $\text{Na}^+$  ions by  $\text{Mg}^{2+}$  or  $\text{Ca}^{2+}$  from a peptide fibre.



<sup>1</sup> Yu, G.; Yan, C.; and Huang, F., *Chem. Soc. Rev.*, **2013**, 42, 6697-6722.

<sup>2</sup> Wallace, M.; Iggo, J.A.; and Adams, D.J., *Soft Matter*, **2015**, 11, 7739-7747.

# Probing interactions in $\beta$ -sheet peptide / graphene derivatives hybrid hydrogels

Jacek Wychowaniec<sup>1\*</sup>; Maria Iliut<sup>2</sup>; Jonathan Moffat<sup>3</sup>; Aravind Vijayaraghavan<sup>2</sup> and Alberto Saiani<sup>1</sup>

<sup>1</sup>School of Materials and Manchester Institute of Biotechnology, University of Manchester. UK; <sup>2</sup>School of Materials and National Graphene Institute, University of Manchester. UK; <sup>3</sup>Asylum Research, Oxford Instruments, High Wycombe, UK

The exploitation of the self-assembling properties of  $\beta$ -sheet forming peptides has become a prominent strategy in material science for the construction of soft material (i.e.: hydrogels)<sup>1,2</sup>. The incorporation of nanofillers such as carbon nanotubes and graphene derivatives offers an effective route for the construction of increasingly complex functional materials for a variety of applications ranging from electronics to biomedical and tissue engineering<sup>3,4</sup>. Understanding the interactions between the  $\beta$ -sheet fibres and the nanofillers is key for the design of the materials. In this study the co-assembly of three peptides, FEFKFEFK (F8), FEFEFKFE (FE), VEVKVEVK (V8) (V: valine; F: phenylalanine; K: lysine; E: glutamic acid) with varying designs and charge states and graphene derivatives nanofillers (graphene oxide and reduced graphene oxide) was investigated using a variety of techniques including: oscillatory rheology, Raman spectroscopy and atomic force microscopy (AFM). Processes for the homogeneous dispersion of graphene oxide in the peptide hydrogels and its subsequent reduction to graphene were developed. Strong interactions between the graphene flakes and the peptides were observed that affected the overall mechanical properties of the hydrogels. In particular electrostatic interactions and  $\pi$ - $\pi$  stacking, when phenylalanine residues are present, were shown to play a key role in determining the final properties of the material.

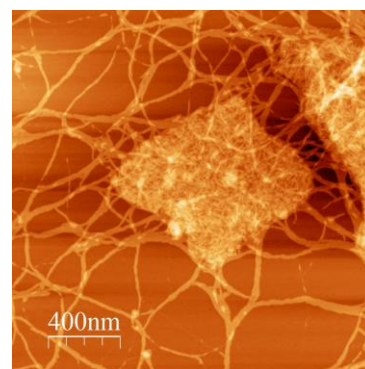


Figure 2. Network of F8 peptide fibres with Graphene Oxide flake.

<sup>1</sup> A. Mujeeb, et al., *Acta Biomaterialia*, **2013**, 9, 4609-4617.

<sup>2</sup> D. Roberts et al., *Langmuir*, **2012**, 28, 16196-16206.

<sup>3</sup> M. Sheikholeslam et al., *Carbon*, **2014**, 71, 284-293.

<sup>4</sup> J. Ramon-Azcon et al., *Advanced Materials*, **2013**, 25, 4028-4034.



# Bifunctional Surfaces: A Peptide-Based Coating that Resists Fouling and Promotes Cell Adhesion

*Sivan Yuran, Alona Dolid and Meital Reches*

*Institute of Chemistry & the Center for Nanoscience and Nanotechnology, The Hebrew University of Jerusalem, Israel*

The interface between organic and inorganic matter is of high importance when designing an implant or a biomaterial. On one hand, since these materials should be integrated into the human body, their interface with tissues should encourage cell attachment. On the other hand, these materials can serve as a surface for bacterial colonization, termed biofilm formation, which can lead to an infection. Despite the preventive measures against infections, such as sterilization, bacterial infection at the site of implants is still of major concern for the health care system. The infection causes discomfort for the patient and may result in the removal of the implant and even patient death. Different approaches attempt to prevent biofilm formation; however, there are disadvantages to each approach. This includes low stability, high cost, complexity of the synthesis or fabrication method and the limitation to a specific substrate.

Recently, our lab designed a tripeptide that self-assembles into a coating that resists fouling. It contains: (i) two aromatic residues that direct the peptide self-assembly into an ordered coating, (ii) the amino acid DOPA which promotes the adsorption of the peptide onto different substrates and (iii) fluorine atoms as substituent on the phenyl groups that mimic the properties of Teflon<sup>®</sup> as a nonstick surface and therefore provide the antifouling activity. Our results demonstrated the formation of a coating which prevents protein adsorption and significantly reduces the number of bacteria adsorbed on the substrate.<sup>1</sup> Based on this tripeptide, we designed and synthesized a bi-functional peptide that both acts as an anti-biofilm coating and encourages cell attachment. We study the formation of coating by several derivatives of this peptide, their antifouling activity and their ability to attract cells. These peptides can be used as a coating on implants and potentially reduce the risk of infection and simultaneously increase the probability of the implant to well integrate in the tissue.

---

<sup>1</sup> Maity S., Nir S., Zada T. & Reches M., *Chemical Communications* **2014**, 50.76 11154-11157

## Self-assembling peptide hydrogels cross-linked by disulfide bonds

*Zamuner A.<sup>§</sup>, Brun P.<sup>°</sup>, Messina M.G.L.\* , Gloria A.<sup>#</sup>, Calvanese L.<sup>^</sup>,  
Castagliuolo I.<sup>°</sup>, Marletta G.\* , Dettin M.<sup>§</sup>*

<sup>§</sup>Department of Industrial Engineering, University of Padova, Padova, 35131, Italy.

<sup>°</sup>Department of Molecular Medicine, University of Padova, Padova, 35121, Italy.

\*Department of Chemical Sciences, University of Catania, Catania, 95125, Italy.

<sup>#</sup>Institute of Polymers, Composites and Biomaterials-National Research Council of Italy, Naples, 80125, Italy.

<sup>^</sup>CIRPeB, University of Naples Federico II, Naples, 80134, Italy.

Self-assembling ionic complementary peptides (SAPs) have been receiving much interest from the scientific community as promising scaffolds for tissue-engineering. In order to investigate the effects of cross-linking on hydrogels characteristics, we have designed four SAP sequences: Cys-EAK (CAEAEAKAKAEAEAKAK), Cys-EAK-Cys (CAEAEAKAKAEAEAKAKC), Cys-EAK<sub>mod</sub> (AEAEAKAKCEAEAKAK) and EAK (AEAEAKAKAEAEAKAK) as reference<sup>1</sup>. Atomic Force Microscopy analysis confirmed the presence of nanofibers that were longer and more interconnected in the oxidized Cys-EAK hydrogel. Small amplitude shear tests showed a typical gel-like behavior for all samples, oxidized or not. In the case of Cys-EAK hydrogel, the oxidation produced a stiffer hydrogel but this trend was not observed for Cys-EAK-Cys. All SAPs hydrogels (oxidized or not) were good scaffolds for mouse fibroblasts and osteoblasts at 24 h. Interestingly at 72, immunohistochemical analysis showed apoptotic round-shape cells in not oxidized Cys-EAK and Cys-EAK-Cys scaffolds (Figure 1).

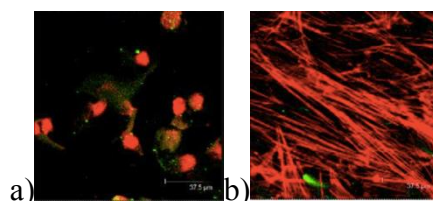


Figure 1. Immunohistochemical analysis of mouse fibroblasts at 72 h of not oxidized Cys-EAK (a) and oxidized Cys-EAK (b).

<sup>1</sup>Gambaretto, R; Tonin, L; Biopolymers, **2008**, 89 (11), 906-915



## **Analysis of Gold-Tethered cationic RPAR Peptide**

*A. Flores<sup>1</sup>, A. Villanueva-Montellano<sup>1</sup>, J. Flores-Garay<sup>1</sup>, J. Trigueros-Enguidanos<sup>2</sup>, D. Zanuy<sup>2</sup> & C. Alemán<sup>2</sup>*

*<sup>1</sup>Instituto de Ingeniería y Tecnología. Universidad Autónoma de Ciudad Juárez, Ave. del Charro 610 Nte., Chihuahua, México 32310.*

*<sup>2</sup>Departament d'Enginyeria Química, Universitat Politècnica de Catalunya, Diagonal 647, Barcelona 08028*

The ability of the peptides to recognize organic surfaces has been renewed attention due to the interaction of proteins with inorganic surfaces, this being very useful in fields such as biomedicine and the development of new composite materials. Blood vessels are generally impermeable to macromolecules and supramolecular complexes.

The increased permeability of blood vessels is associated with angiogenesis and carcinogenic processes. The RPAR peptide interacts with neuropilin (NRP-1), which is a receptor that plays an essential role in these processes. Molecular dynamics simulations have been used to study the conformational preferences of RPAR in solution and tethered to biocompatible inorganic surfaces.



## Self-assembled monolayers of redox peptides

*Piccoli, J.P.<sup>1</sup>; Santos, A.<sup>2</sup>; Bueno, P.R.<sup>2</sup>; Cilli, E.M.<sup>1</sup>*

*Institute of Chemistry, UNESP - Univ Estadual Paulista, <sup>1</sup>Department of Biochemistry and Technological Chemistry, <sup>2</sup>Department of Physical Chemistry, Nanobionics research group ([www.nanobionics.pro.br](http://www.nanobionics.pro.br)) São Paulo, Brazil.*

Biosensors based in protein-protein interaction plays an important role in the early diagnosis diseases. The development of redox capacitive biosensors, coupled with the electroanalytical techniques, has recently been introduced with great potential of low cost and fast method. However, there are disadvantages because of the high cost and low stability of the molecular recognition component. Peptides have proved to be a promising tool improve the affinity, specificity, and stability of molecular recognition components for the development of self-assembled monolayers. Herein, we described synthesis of redox-tagged peptides with self-assembling capability in electrochemically active capacitive surfaces, for the C Reactive Protein detection (biomarker of inflammation and cardiovascular disease). Peptides containing ferrocene (fc), redox group, was synthesized by solid phase peptide synthesis (SPPS), with the sequences: Fc-E-A-A-C-NH<sub>2</sub> and Fc-K-A-A-C-NH<sub>2</sub>. To obtain the electrochemically capacitive interface, the side chain of Cys was covalently bound to the gold electrode (sulfur group) and the N-terminus group was used to attach the ferrocene in the peptide chain. The anti-CRP was attached to the peptide using the side chain of glutamate and lysine, respectively. The self-assembly and redox capability was characterized by cyclic voltammetry and electrochemical impedance-based capacitance spectroscopy techniques. Although, the affinity constant and the surface coverage of both peptide SAMs were similar, the limits of detection were different: 0.11 nmol L<sup>-1</sup> and 0.31 nmol L<sup>-1</sup> for the peptides Fc-E-A-A-C-NH<sub>2</sub> and Fc-K-A-A-C-NH<sub>2</sub>, respectively. Results showed that, the bound of the anti-CRP to glutamate improved the detection. In addition, the design of redox active peptides self-assembly is predictably useful in the development of biosensor devices, making use of the electrochemical capacitance signal intrinsically existing in redox-active monolayers.<sup>1</sup>

<sup>1</sup> Santos, A. et al; Piccoli, J.P.; et al *Biosensors and Bioelectronics* **2014**, 68, 281-287.



## Functional Peptide Assemblies on Surfaces

*Sivan Nir, Tal Zada and Meital Reches*

*Institute of Chemistry and the Center for Nanoscience and Nanotechnology, The Hebrew University of Jerusalem, Israel*

Biofouling is an undesirable process in which a surface becomes encrusted with organisms and their by-products. This unwanted colonization has a serious impact on marine devices, as it leads to deterioration of the surfaces and can alter fluid flow rates leading to significant increase in cost of marine transportation. In the healthcare system, the attachment of bacteria and biofilm formation on medical devices may lead to a severe infection and consequently death. In the US alone, the American Centre for Disease Control and Prevention (CDC) reported that healthcare-associated infections account for an estimated 1.7 million infections and 100,000 deaths annually.

Many approaches to prevent biofouling have been suggested, however, they suffer from drawbacks such as release of toxic materials to the surroundings, low stability that limits their long-term application or complex and expensive synthesis.

We have recently designed a tripeptide that under different conditions self-assembles in an organized manner to form coatings that interfere with the attachment of organisms to the substrate and therefore act as an antifouling agent.<sup>1</sup>

The peptide contains three elements that enable i) its self-assembly, ii) its adsorption onto any substrate and iii) its antifouling activity. The peptide-based coating completely prevented the first stage of biofouling and abolished the adsorption of proteins to a substrate. Moreover, the coating significantly reduced the amount of different bacterial strains adsorbed on the substrate.

---

<sup>1</sup> Maity S., Nir S., Zada T. and Reches M. *ChemCommun*, **2014**, 50, 11154-11157.

## AUTHOR INDEX

---

Accardo, A.	OP-16, P-5
Adams, D.J.	OP-21, P-6, P-23
Ahumada, O.	OP-20
Álamo, P.	P-21
Albericio, F.	OP-17
Alemán, C.	OP-7, P-14, P-20, P-22, P-27
Álvarez de Cienfuegos, L.	OP-14, P-4
Armelin, E.	P-20
Ashkenasy, N.	OP-9
Ballet, S.	P-12
Bertolani, A.	P-1, P-10, P-15, P-19
Bertran, O.	P-22
Bhimareddy, D.	OP-10
Bianco, A.	OP-10
Bibian, M.	P-12
Bidon-Chanal, A.	OP-17
Bleda, M.J.	P-8
Bocchinfuso, G.	OP-26
Bonhome-Espinosa, A.B.	P-4
Boyle, A.	OP-22
Brun, P.	P-26
Bucci, R.	P-17
Bueno, P.R.	P-28
Calvanese, L.	P-26
Casanovas, J.	OP-7, P-14
Castagliuolo, I.	P-26
Cativiela, C.	OP-7, P-14, P-22
Cavallo, G.	P-1
Céspedes, M.V.	OP-13, P-21
Chabert, V	P-2
Cilli, E.M.	P-28
Clerici, F.	P-3

---

---

Colquhoun, C.	OP-21
Conchillo, O.	OP-13
Conejero-Muriel, M.T.	OP-14
Conticello, V.	OP-3
Contreras-Montoya, R.	OP-14, P-4
Cubarsi, R.	OP-13
Cuerva, J.M.	P-4
Daura, X.	OP-13
del Valle, L.J.	OP-7, P-20, P-22
Demitri, N.	P-1, P-10, P-19
Derreumaux, P.	OP-5
Detin, M.	P-26
Diaferia, C.	OP-16, P-5
Diederichsen, U.	OP-11
Dolid, A.	P-25
Draper, E.R.	OP-21, P-6
Eden, E.G.B.	OP-21
Fabregat, G.	P-14, P-22
Ferrer-Miralles, N.	OP-13, P-21
Ferri, N	P-17
Flores, A.	P-27
Flores-Garay, J.	P-27
Fraioli, R.	P-13
Fromm, K.M.	P-2
Gallardo, A.	OP-13
García-Fruitós, E.	OP-13
Gardiner, J.	P-12
Gavira, J. A.	OP-14
Gazzera, L.	P-1
Gelmi, M.L.	P-17
Giannini, C.	P-5
Gil, A.M.	P-22
Gil, F.J.	P-13

---

Gilmore B.F.	P-16
Gilmore, S.M.	P-16
Giralt, E.	OP-15
Gloria, A.	P-26
Gómara, M.J.	P-7, P-8
Gori, A.	P-10, P-15
Hamley, I.W.	OP-12
Haro, I.	P-7, P-8
Höger, G	OP-11
Hoogenboom, R.	P-12
Hughes, S.	OP-3
Iggo, J.A.	P-23
Iliut, M.	P-24
Jiang, T.	OP-3
Jiménez, M.A.	P-9
Kessler, H.	P-13
Kimura, S.	OP-1
Kros, A.	OP-22
Lascialfari, L.	P-10, P-15
Laverty, G.	OP-25, P-16
Lavilla, R.	OP-17
Lendel, C.	P-11
Leone, M.	P-5
Locarno, S.	P-3
López-López, M.T.	P-4
Luque, F.J.	OP-17
Madder, A.	P-12
Maestro, B.	P-9
Magnotti, E.	OP-3
Maione, S.	P-22
Manero, J.M.	P-13
Mangues, R.	OP-13, P-21
Marletta, G.	OP-24, P-26

Martin, C.	P-12
Mas-Moruno, C.	P-13
Mayans, E.	OP-7, P-14
McCloskey , A.P.	OP-25, P-16
McDonald, T.O.	OP-21
Ménard-Moyon, C.	OP-10
Meneghetti, F.	P-17
Mercurio, F.A.	P-5
Merino-Mansilla, A.	P-7
Messina, M.G.L.	OP-24, P-26
Metrangolo, P.	P-1, P-10, P-15, P-19
Miller, A.F.	OP-4
Mitjans, F.	OP-17
Modlin, C.	OP-3
Moffat, J.	P-24
Morelli, G.	OP-16, P-5
Moretto, A.	OP-6
Morris, K.	OP-21
Mykhaylyk, O.O.	P-6
Nava, D.	P-3
Neubauer, S.	P-13
Nir, S.	P-29
Oyen, E.	P-12
Paús, A.	P-7
Pawlowski, J.	OP-23
Pellegrino, S.	P-17
Pérez-Madrigal, M.M.	OP-7
Pesarrodona, M.	OP-13, P-21
Piccoli, J.P.	P-28
Pille, J.	P-18
Pirrie, L.	P-19
Pizzi, A.	P-1, P-10, P-15, P-19
Puiggalí, J.	OP-7, P-14

Puiggali-Jou, A.	P-20
Pulka-Ziach, K.	OP-23
Rechenmacher, F.	P-13
Reches, M.	OP-2, P-17, P-25, P-29
Resnati, G.	P-1, P-10, P-15, P-19
Rodríguez, J.	P-8
Rodríguez-Arco, L.	P-4
Rodríguez-Cabello, J.C.	OP-28
Rosenman, G.	OP-19
Rost, U.	OP-11
Rueda, F.	OP-13
Ruffoni, A.	P-3
Ruiz-Rodríguez, J.	OP-17
Saccardo, P.	P-21
Saiani, A.	OP-4, P-24
Salvador P.	P-8
Sanchez-Chardi, A.	OP-13
Sánchez-Merino, V.	P-7
Sanmartí, R.	P-8
Santos, A.	P-28
Sanz, J.M.	P-9
Sek, S.	OP-23
Seras-Franzoso, J.	OP-13
Serna, N.	P-21
Serpell, L.C.	OP-21
Sibillano, T.	P-5
Smeitink, J.	P-18
Spetea, M.	P-12
Stefan, L.	P-15
Stranderg, E.	P-9
Terraneo, G.	P-1, P-19
Toniolo, C.	OP-18
Torras, J.	P-20

Triguero, J.	P-22
Trigueros-Enguidanos, J.	P-27
Ulrich, A.S.	P-9
Unzueta, U.	OP-13, P-21
van Hest, J.C.M.	P-18
van Mele, B.	P-12
van Oppen, L.	P-18
Vázquez, E.	OP-13, P-21
Venanzi, M.	OP-27
Vijayaraghavan, A.	P-24
Villanueva-Montellano, A.	P-27
Villaverde, A.	OP-13, P-21
Wallace, M.	P-6, P-23
Weidner, T.	OP-8
Wiegand, M.	OP-11
Wychowaniec, J.	P-24
Xu, Z.	P-21
Yuran, S.	P-25
Yuste, E.	P-7
Zada, T.	P-29
Zamora-Carreras, H.	P-9
Zamudio, R.	OP-17
Zamuner, A.	P-26
Zanuy, D.	P-22, P-27



PEOPLE ATTENDING THE MEETING

Accardo, A.	Univ. of Naples	antonella.accardo@unina.it
Adams, D.J.	Univ. of Liverpool	d.j.adams@liverpool.ac.uk
Aguilar, M.A.	Univ. Peruana de Ciencias Aplicadas	ma23aguilar@gmail.com
Ahumada, O.	Mecwins	oahumada@mecwins.com
Alemán, C.	Univ. Politècnica de Catalunya	carlos.aleman@upc.edu
Álvarez de Cienfuegos, L.	Univ. de Granada	lac@ugr.es
Ashkenasy, N.	Ben Gurion Univ.	nurita@bgu.ac.il
Bertran, O.	Univ. Politècnica de Catalunya	oscar.bertran@upc.edu
Bhimareddy, D.	Univ. of Strasbourg	b.dinesh@ibmc-cnrs.unistras.fr
Bidon-Chanal, A.	Univ. de Barcelona	abidonchanalb@ub.edu
Bocchinfuso, G.	Univ. of Rome "Tor Vergata"	bccgfr00@uniroma2.it
Boyle, A.	Leiden Univ.	a.l.boyle@chem.leidenuniv.nl
Calleja, R.	Mecwins	rcalleja@mecwins.com
Casanovas, J.	Univ. de Lleida	jasanovas@quimica.udl.cat
Cavallo, G.	Politecnico di Milano	gabriella.cavallo@polimi.it
Chabert, V.	Univ. of Fribourg	valentin.chabert@unifr.ch
Cilli, E.	Univ. Estadual Paulista	eduardocilli@gmail.com
Clerici, F.	Univ. of Milan	francesca.clerici@unimi.it
Conticello, V.	Emory Univ.	vcontic@emory.edu
Contreras-Montoya, R.	Univ. de Granada	contrerasmontoya.r@gmail.com
del Valle, L.J.	Univ. Politècnica de Catalunya	luis.javier.del.valle@upc.edu
Derreumaux, P.	CNRS	philippe.derreumaux@ibpc.fr
Dettin, M.	Univ. of Padova	monica.dettin@unipd.it
Diaferia, C.	Univ. of Naples "Federico II"	carlo.diaferia@unina.it
Diederichsen, U.	Georg-August Univ. Göttingen	udieder@gwdg.de
Draper, E.	Univ. of Liverpool	Emily.Draper@liverpool.ac.uk
Flores, A.	Univ. Autónoma de Ciudad Juárez	ale.flores@uacj.mx

Gelmi, M.L.	Univ. di Milano	marialuisa.gelmi@unimi.it
Giralt, E.	Inst. for Research in Biomedicine	ernest.giralt@irbbarcelona.org
Gómara, M.J.	CSIC	mariajose.gomara@iqac.csic.es
Hamley, I.W.	Univ. of Reading	i.w.hamley@reading.ac.uk
Haro, I.	CSIC	isabel.haro@iqac.csic.es
Jiménez, M.A.	CSIC	majimenez@iqfr.csic.es
Kimura, S.	Kyoto Univ.	shun@scl.kyoto-u.ac.jp
Lascialfari, L.	Politecnico di Milano	luisa.lascialfari@polimi.it
Laverty, G.	Queen's Univ. Belfast	garry.laverty@qub.ac.uk
Lendel, C.	KTH Royal Inst. of Technology	lendel@kth.se
Martin, C.	Vrije Universiteit Brussel	charlotte3martin@gmail.com
Mas Moruno, C.	Univ. Politècnica de Catalunya	carles.mas.moruno@upc.edu
Matera, C.	IBEC, Parc Científic BCN	cmatera@ibecbarcelona.eu
Mayans, E.	Univ. Politècnica de Catalunya	enric.mayans@gmail.com
McCloskey, A.P.	Queen's Univ. Belfast	amccloskey16@qub.ac.uk
Messina, G.M.L.	Univ. of Catania	grmessi@unict.it
Metrangolo, P.	Politecnico di Milano	pierangelo.metrangolo@polimi.it
Morelli, G.	Univ. of Naples "Federico II"	gmorelli@unina.it
Moretto, A.	Univ. of Padova	alessandro.moretto.1@unipd.it
Nir, S	The Hebrew Univ. of Jerusalem	sivanvanir@gmail.com
Pellegrino, S.	Univ. degli Study di Milano	sara.pellegrino@unimi.it
Pérez-Madrigal, M.M.	Univ. Politècnica de Catalunya	m.mar.perez@upc.edu
Pille, J.	Radboud Univ. Nijmegen	J. Pille@science.ru.nl
Pizzi, A.	Politecnico di Milano	andrea.pizzi@polimi.it
Porter, S.	Queen's Univ. Belfast	sporter13@qub.ac.uk
Prischich, D.	Univ. de Barcelona	dprischich@ibecbarcelona.eu
Puiggali-Jou, A.	Univ. Politècnica de Catalunya	anna.puiggali@upc.edu
Reches, M.	The Hebrew Univ.	meital.reches@mail.huji.ac.il
Rodríguez-Cabello, J.C.	Univ. de Valladolid	roca@bioforge.uva.es

Rosenman, G.	Tel Aviv Univ.	gilr@eng.tau.ac.il
Saiani, A.	Univ. of Manchester	a.saiani@manchester.ac.uk
Sek, S.	Univ. of Warsaw	slasek@chem.uw.edu.pl
Serna, N.	Univ. Autònoma de Barcelona	srnaroa@gmail.com
Toniolo, C.	Univ. of Padova	claudio.toniolo@unipd.it
Torras, J.	Univ. Politècnica de Catalunya	joan.torras@upc.edu
Triguero, J.	Univ. Politècnica de Catalunya	antidot18@gmail.com
Venanzi, M.	Univ. of Rome "Tor Vergata"	venanzi@uniroma2.it
Villaverde, A.	Univ. Autònoma de Barcelona	antoni.villaverde@uab.cat
Wallace, M.	Univ. of Liverpool	m.wallace1@liverpool.ac.uk
Weidner, T.	Max Planck Inst.	weidner@mpip-mainz.mpg.de
Wychowaniec, J.	Univ. of Manchester	jacek.wychowaniec@postgrad.manchester.ac.uk
Yuran, S.	The Hebrew Univ.	sivan.yuran@mail.huji.ac.il
Zamuner, A.	Univ. of Padova	annj.zamuner@studenti.unipd.it
Zanuy, D.	Univ. Politècnica de Catalunya	david.zanuy@upc.edu