

1 Genotype-specific evolution of hepatitis E virus

2

3 Adam B. Brayne,^a Bethany L. Dearlove,^a James S. Lester,^a Sergei L. Kosakovsky Pond,^b

4 Simon D. W. Frost^a #

5

6 University of Cambridge^a; Temple University^b

7

8

9 Running Head: Genotype-specific evolution of hepatitis E virus

10

11 #Address correspondence to Simon D.W. Frost, sdf22@cam.ac.uk

12

13

14

15

16

17

18

19 Abstract: 151 words. Other text: 5410 words

20

21 **Abstract**

22 Hepatitis E virus (HEV) is the most common cause of acute viral hepatitis globally.
23 HEV comprises four genotypes with different geographic distributions and host
24 ranges. We utilise this natural case-control study for investigating the evolution of
25 zoonotic viruses compared to single host viruses, using 244 near full length HEV
26 genomes. Genome wide estimates of dN/dS located a region of overlapping reading
27 frames, which is subject to positive selection in genotypes 3 and 4. The open reading
28 frames (ORFs) involved have functions related to host-pathogen interaction, so
29 genotype specific evolution of these regions may reflect their fitness. Bayesian
30 inference of evolutionary rates shows genotypes 3 and 4 have significantly elevated
31 rates relative to genotypes 1 across all ORFs. Reconstructing phylogenies of zoonotic
32 genotypes demonstrates significant intermingling of isolates between hosts. We
33 speculate that the genotype specific differences may result from cyclical adaptation
34 to different hosts in genotypes 3 and 4.

35 *Importance:*

36 Hepatitis E virus (HEV) is increasingly recognised as a pathogen which affects both
37 the developing, and the developed world. While most often clinically mild, HEV can
38 be severe or fatal in certain demographics, such as expectant mothers. Like many
39 other viral pathogens, HEV has been grouped into several distinct genotypes. We
40 show that most of the HEV genome is evolutionarily constrained. One locus of
41 positive selection is unusual as it encodes two distinct protein products. We are the

42 first to detect positive selection in this overlap region. Genotype 1, which only
43 infects humans, appears to be evolving differently to genotypes 3 and 4, which infect
44 multiple species, possibly because genotypes 3 and 4 are unable to achieve the same
45 fitness due to repeated host jumps.

46 **Introduction**

47 Hepatitis E virus (HEV) is a non-enveloped, single stranded, positive sense RNA
48 virus, which infects around 20 million people globally each year (1). It causes large
49 propagated epidemics of acute hepatitis in Asia and Africa, and low level, sporadic
50 food-associated infections in the developed world (2, 3). Pathogenicity varies from
51 acute liver failure and up to 20% mortality in some sub-populations (for example in
52 pregnant women), to apparently asymptomatic infections in others (4). Acquired via
53 the fecal-oral route, HEV is associated with poor hygiene and living conditions. It
54 can also be acquired by eating contaminated food, including infected artiodactyls
55 (swine, deer and boar) and shellfish (4–6).

56 Mammalian HEV exists in four internationally recognised genotypes (7). Genotyping
57 is based on nucleotide divergence of the capsid open reading frame (8), and whole
58 genome phylogenetic analysis (9). Genotypes differ at epidemiological (distribution,
59 hosts) and virological (pathogenicity, translation mechanisms) levels.

60 In terms of epidemiology, there is a striking global distribution of autochthonous
61 genotypes whose origins are obscure (10): Genotype 1 is found in Asia and North
62 Africa; genotype 2 in Mexico and Southern Africa; genotype 3 in North and South
63 America, Europe and Asia; and genotype 4 almost exclusively in Japan and China. All

64 four genotypes infect humans, but only genotypes 3 and 4 infect other animals such
65 as artiodactyls. In the developed world infections are sporadic and the genotype is
66 usually the same as that in the native swine population, suggesting zoonotic
67 transmission by food or contact (2). Most likely this involves the consumption of
68 undercooked pork. In contrast in developing countries infections can be epidemic as
69 well as sporadic, with human and swine strains most often different. A recombinant
70 vaccine against HEV exists, based on its capsid protein, which has passed phase III
71 trials (11, 12). The vaccine is based on genotype 1 strains, and appears to provide
72 cross protection against at least genotype 3 (12).

73 Pathogenicity and molecular mechanisms vary between genotypes. In developed
74 countries clinical disease is rare, and seroprevalence vastly outweighs documented
75 incidence (13–15). In developing countries, the clinical presentation of HEV
76 infection tends to be more symptomatic than in the developed world. Symptoms are
77 shared with many viral illnesses and include fever, gastro-intestinal upset and
78 malaise, and liver function tests may be deranged (15). The natural history also
79 varies by demographic, with a strikingly high mortality amongst pregnant women in
80 the developing world (10-25%) and also more disease in children compared to the
81 developing world where it is elderly men that are most often symptomatic (15, 16).
82 Primate models suggest these differences in pathogenicity are associated with the
83 genotypes, as genotypes 3 and 4 produce less clinical disease in comparison to
84 genotypes 1 and 2 in rhesus monkeys (17). There are few known differences in
85 molecular mechanisms between genotypes, however genotype 4 viruses do have a

86 distinct mechanism for the translation of open reading frame 3 (ORF3), due to a
87 frame-disrupting single nucleotide insertion (18).

88 HEV has a c. ~7200 nucleotide genome comprising three partially overlapping open
89 reading frames. ORF1 encodes a nonstructural region, and ORF2 encodes the capsid
90 protein (19). The function of ORF3, which almost entirely overlaps ORF2, is not
91 totally clear. Interestingly ORF3 is not necessary for *in vitro* infection (20), but is
92 necessary for *in vivo* infection of macaques (21). It is most likely multifunctional
93 (19) and involved in pathogenesis (22–25). Most of the coding region in the HEV
94 genome is under purifying selection (26, 27), *i.e.* selection against change in the
95 amino acid sequence. Areas with an excess of amino acid substitutions, a signal of
96 positive selection, have been found in the N-terminus of ORF2 and the C-terminus of
97 ORF3, with another in the RNA dependent RNA polymerase (RdRp) region in ORF1
98 (26). Purdy *et al.* (28) have described positive selection in the hypervariable region
99 (HVR) of ORF1; however, Smith *et al.* (27) failed to reproduce these results with a
100 broader selection of statistical tests.

101 Phylogenetic analyses of HEV may help to shed light on evolutionary differences
102 between genotypes, which underlie the epidemiological and clinical disparities. In a
103 previous study, Chen *et al.* (26) failed to discern any difference in selection
104 pressures between genotypes. Since 2012, the number of appropriate full genome
105 samples has increased by 150%. Using this expanded dataset, we revisit the
106 question of evolutionary differences between the genotypes of HEV, using state-of-
107 the-art methods. We focus on detecting natural selection, specifically investigating

108 regions of positive selection which stand out from a background of purifying
109 selection against non-synonymous substitutions. Our particular focus is on the
110 overlap region, making ours the first analysis of this region as a focus of positive
111 selection. We also carry out a detailed analysis of evolutionary rates, and link
112 phylogenetic findings to the virological characteristics of the genotypes.

113 **Methods**

114 **Sequence acquisition**

115 All available sequences of hepatitis E virus in Genbank (29) were obtained by
116 searching the NCBI Nucleotide Database using the taxonomic identifier (txid) 12461,
117 along with associated metadata on host, country, and date of sampling. As of August
118 6th, 2014 there were 10,041 sequences, of which 258 sequences were at least 7000
119 nucleotides long (i.e. near full length genomes).

120 **Sequence processing**

121 Open reading frames, corresponding to sequence regions between consecutive stop
122 codons, were identified for each sequence using `getorf`, part of the EMBOSS
123 package (30). ORFs 1, 2, and 3 for each sequence were identified by `blastp` (31),
124 with amino sequences of ORFs from the NCBI Reference Sequence NC_001434 as the
125 query, and translated ORF sequences as the reference. Multiple sequence alignments
126 (MSAs) for each ORF were generated using Clustal Omega (32), based on the
127 translated sequences. Nucleotide sequences were mapped on to the corresponding
128 aligned amino acid sequences using Seaview v. 4.5.0 (33). MSAs were trimmed,

129 based on the start and stop of ORFs in NC_001434, and checked manually. In order
130 to obtain a single in-frame sequence for the near-full length genome, we
131 concatenated ORFs 1 and 2. The alignments and associated inferred data are
132 available for download from github.com/veg/HEV-evolution-2015.

133 Sequences were screened for recombination using RDP4 (version 4.36 beta) (34),
134 using eight available methods; RDP (35), GENECONV (36), BootScan (37), MaxChi
135 (38), Chimaera (39), SiScan (40), PhylPro (41), LARD (42), and 3Seq (43), using
136 default settings. Following exploratory analyses to determine whether
137 recombination detection was simply an artifact of complex patterns of mutations, a
138 sequence was deemed recombinant if three or more methods had reported it as a
139 recombinant. Consistent with prior reports of recombination in HEV (26, 44, 45), we
140 identified 14 recombinant viruses, including novel recombinants (see Table 1).

141 Genotypes were assigned to each sequence by sequence similarity and phylogenetic
142 reconstruction. We used `tblastx` (from the BLAST 2.2.30+ software suite (31, 46))
143 to find the most similar sequences prototypical for each genotype; M73218
144 (genotype 1 (47)); M74506 (genotype 2 (48)); AF060668 (genotype 3 (49)); and
145 AJ272108 (genotype 4 (18)). Designations were further investigated by inspecting
146 phylogenetic reconstructions obtained using `FastTree v2.1.8` (50). Of the 258 near-
147 full length genomes, 127 were isolated from humans, and were selected for further
148 analysis. Two sequences were excluded on the basis that they were abnormally
149 divergent from the other sequences: M74506, which is a genotype 2 virus, and
150 JQ013793, which is similar to a strain of HEV isolated from rabbits (51). Genotype-

151 specific alignments were generated and merged into a single master alignment
152 using MACSE v.1.01b (52). Sequences with a 100% identity to other isolates were
153 removed, resulting in a final dataset of 113 unique near full-length genomes isolated
154 from humans, comprised of concatenated ORF1 and ORF2 regions, with 26 genotype
155 1 sequences, 42 genotype 3 sequences, and 45 genotype 4 sequences. We split the
156 alignment into ORF1 and ORF2 regions, extracted the overlapping part of ORF3
157 from ORF2, and split ORF2 into the region overlapping ORF3, and the non-
158 overlapping region. We also identified 56 unique HEV genomes isolated from swine.
159 The swine HEV sequence alignment was merged with the human HEV dataset using
160 profile alignment in codon space using MACSE.

161 **Genome-level selection analyses**

162 Selection analyses employed a suite of phylogenetic methods, as implemented in
163 HyPhy(53) and Datamonkey (54, 55) using default settings. FUBAR (56) was used to
164 characterize pervasive selective pressures, i.e., those aggregated over all branches in
165 the phylogeny. Both an alignment-wide distribution of synonymous and non-
166 synonymous substitution rates, and site-level estimates were obtained using
167 FUBAR. MEME (57) was applied to identify individual sites subject to episodic
168 positive selection (i.e. operating along a subset of tree branches). aBSREL (58)
169 allowed us to estimate the complexity of evolutionary processes along individual
170 tree branches, and to determine which branches in the tree were subject to positive
171 selection along a subset of sites in the alignment. Finally, RELAX (59) was employed

172 to formally test whether or not the evolutionary pressures were relaxed or
173 intensified for HEV infecting human hosts relative to those infecting swine hosts.
174 So that we could formally test whether or not selection was relaxed or intensified in
175 the overlapping region of ORF3 relative to ORF2, we modified the RELAX method
176 (60) to accept two gene alignments as input. Briefly, we fit a 3-rate random effects
177 branch-site class model (61) with three ω classes to accommodate the variation in
178 selective forces across sites and branches in an unrestricted fashion jointly to both
179 alignments, while endowing each with its own branch lengths, equilibrium codon
180 frequencies, and nucleotide substitution biases. The RELAX test enforces a
181 functional relationship between the ω ratios in reference (ORF2) and test (ORF3)
182 alignments: $\omega_{ORF3} = (\omega_{ORF2})^K$. The estimated value of K indicates whether
183 selection in the test frame is relaxed ($K < 1$) or intensified ($K > 1$) relative to the
184 reference frame. A likelihood ratio test of the null hypothesis ($K=1$), versus the
185 alternative hypothesis ($K \neq 1$) establishes statistical significance of relaxation (or
186 intensification).

187 **Codon substitution model for overlapping regions**

188 We fitted three codon substitution models that explicitly consider whether
189 mutations are synonymous in just one of ORF2 and ORF3, or both. These models,
190 which have been previously used to screen for biologically meaningful alternative
191 reading frames in mammalian genomes (62), generate estimates of rates R_{XY} , which
192 refer to the rates of substitutions which are synonymous ($X = 0$) or non-
193 synonymous ($X=1$) in the primary frame (ORF2), and synonymous ($Y = 0$) or non-

194 synonymous ($Y=1$) in the alternative frame (ORF3). R_{00} - the rate for substitutions
195 that are synonymous in both frames, is fixed at 1, and the other three rates are
196 estimated relative to R_{00} . Maximum likelihood parameter estimates and associated
197 95% confidence intervals (profile likelihood) were calculated for a model in which
198 R_{01} , R_{10} , and R_{11} were allowed to vary freely. We also performed likelihood ratio
199 tests comparing the full model with two null models. The first null model assumes
200 that R_{11} is greater than one or both of R_{01} and R_{10} ; the expectation is that R_{11}
201 (non-synonymous in both frames) should be less than either R_{01} or R_{10} , because
202 changing both frames should be evolutionary constrained. The second null model
203 assumes that $R_{01}=R_{10}$; rejection of this null hypothesis suggests that one frame is
204 more constrained than the other.

205 **Molecular clock analyses**

206 Sequences were annotated by year of sampling. In many cases, these data were
207 obtained from Genbank records. In other cases, the primary reference was used. In
208 the cases where neither source gave the sampling year, we used the submission date
209 to Genbank as an upper bound for the sampling date, with the lower bound set as
210 the earliest known sampling year (March 1990, from (63)). To estimate the
211 evolutionary rate for genotype specific alignments whilst accommodating the
212 uncertainty in sampling times, we used a Bayesian phylogenetic approach, as
213 implemented in MrBayes v3.2.2 (64). A general time reversible (GTR) model was
214 fitted, with rate variation modelled as a discrete gamma distribution with 4
215 categories. Base frequencies were fixed at their empirical values, and a uniform

216 prior placed on topologies. A relaxed clock model was used, assuming that
217 evolutionary rates were drawn independently from a gamma distribution. Default
218 priors were used, with the exception of the clock rate, which was set to $\text{lognorm}(-$
219 $9, 1)$. Two chains were run for 110 million generations with a burnin of 10 million,
220 thinned to give a sample of 1000 iterations. Results were processed using the coda
221 library (65) in R and the 95% upper and lower credible intervals were inferred from
222 the posterior distribution. Convergence was tested using manual inspection of
223 traces of parameter values, and calculation of the Gelman-Rubin statistic (66). The
224 `rv` library was used to generate credible intervals for the difference in clockrate
225 between genotypes in the same ORF. To validate the use of a relaxed clock we
226 analysed the parameter describing the variance of the rate distribution of the
227 relaxed clock, and found it to be distinct from zero with a median of 0.01914977
228 (95% credible interval=0.00115991-0.04887433), providing support for the use of a
229 relaxed clock over a strict clock. The `ggplot2` library (67) in R was used to create
230 rate plots.

231 **Host-specific patterns of evolution**

232 Human and swine HEV near full length genomes were split into genotype 3 and
233 genotype 4 alignments. Phylogenies for each genotype were reconstructed
234 separately using maximum likelihood with RAxML v.8 (68), assuming the GTR
235 model of nucleotide substitution with gamma distributed rate variation. Phylogenies
236 were rooted with `1sd` v.0.1 (69), using the median estimate of the sampling time for
237 each sequence. Terminal branches were classified as human or swine based on

238 which host they were isolated from. Interior branches were classified as
239 'human'/'swine' whenever all of their descendants were labelled as
240 'human'/'swine', following post-order tree traversal. Species-specific estimates of
241 the distribution of the ω ratio were obtained on the basis of the models
242 implemented in RELAX (59).

243 **Implementation**

244 Except where otherwise stated, selection analyses were performed using HyPhy
245 (53), using phylogenies of each region reconstructed using RAxML v.8 (68),
246 assuming the GTR model of nucleotide substitution with gamma distributed rate
247 variation, or the MG94xGTR model of codon substitution with analysis-defined
248 patterns of site-to-site and branch-to-branch rate variation. Tree visualisation was
249 carried out using the `phyloTree.js` widget implemented as an extension of the D3
250 (D3js.org) JavaScript visualisation library (<http://veg.github.io/hyphy-vision>).

251

252 **Results**

253 **Genome-wide patterns of selection**

254 To visually identify genomic regions under positive or purifying selection, we
255 estimated the number of non-synonymous (amino-acid changing, dN) and
256 synonymous (amino-acid preserving, dS) changes for each codon (Figure 1) using
257 the FUBAR method (56), which estimates these quantities for individual sites using
258 an Empirical Bayes procedure in the phylogenetic likelihood framework. Consistent

259 with previous findings, most of the genome was under purifying selection ($dN < dS$).
260 However, within each ORF, specific regions showed statistically significant evidence
261 of positive selection ($dN > dS$): the hypervariable region (HVR) in ORF1, the 5' end
262 of ORF2, and ORF3 (Figure 1). As the 5' end of ORF2 and ORF3 are overlapped, we
263 repeated FUBAR analysis of this area in each reading frame, finding a strong signal
264 of positive selection throughout the overlapped region of ORF2 and a weaker signal
265 in ORF3 (Figure 1).

266 **Rate variation amongst site and branches**

267 We fitted an adaptive branch-site model (58) to the alignment of 113 isolates with
268 near full length genomes. Overall, there was very strong evidence of variation in
269 selective pressure both over sites and lineages ($\Delta AIC = 1760$ in favour of the model
270 which allows such variation), with 54 (24%) of branches supporting site-to-site
271 variation, with 2 rate classes per branch. The remaining 169 branches could be
272 adequately explained by a model where all sites evolve at a single rate. Eleven
273 branches were subject to statistically significant ($p < 0.05$ after Holm-Bonferroni
274 multiple testing correction) positive selection. Of the eleven, one belonged to
275 genotype 1 (M94177), 3 to genotype 3 (KJ701409, AF060669, and AF060668), and 7
276 to genotype 4 (AB220977, AB291964, AB291959, AB220979, AB220976,
277 AB220978, and AJ272108). In all cases, 98% or more sites were under strong
278 purifying selection ($\omega < 0.05$), and the remainder were under very strong positive
279 selection ($\omega > 50$). Interestingly, despite the fact that the estimated distribution for
280 all interior branches separating the individual genotypes had a component with $\omega >$

281 1, none rose to the level of statistical significance for positive selection, after
282 multiple test correction.

283 **Selection on individual sites in the ORF2/ORF3 overlap region**

284 We performed selection analyses on each genotype separately. Consistent with the
285 whole genome FUBAR analysis, signals of positive selection were found in the
286 overlap region. Using multiple methods for detecting selection, positive selection
287 was found in both frames of genotypes 3 and 4, whilst neither reading frame of
288 genotype 1 exhibited any significantly positively selected sites (see Table 2). This
289 trend is shown in Figure 2, which renders the genotype-specific FUBAR distribution
290 estimates for each reading frame, representing the proportion of sites evolving at
291 different nonsynonymous and synonymous rates. These selective 'fingerprints'
292 demonstrate that there are sites subject to positive selection in both reading frames
293 in enzoonotic genotypes 3 and 4, but none in the human-only genotype 1.

294 In order to further disentangle selection on different reading frames, we fitted a
295 codon substitution model (see Table 3) that considers whether mutations are non-
296 synonymous in ORF2, ORF3, or both ORF2 and ORF3. In all three genotypes, the rate
297 of substitutions that were non-synonymous at a codon level in both frames was
298 significantly lower than the rate of non-synonymous mutations in either of the
299 specific frames. This finding is consistent with a dual-coding region where both
300 frames are under purifying selection for functional conservation, on average. The
301 point estimates derived from Genotype 1 are lower than for genotypes 3 and 4,
302 hinting at stronger conservation for the former. For genotypes 1 and 3, ORF2 and

303 ORF3 are evolving at significantly different rates, when considering non-
304 synonymous substitutions affecting only one of the frames, with ORF2 experiencing
305 more of the latter. For genotype 4, the rates are statistically indistinguishable.

306 To formally test whether ORF3 is evolving differently from the overlapping region of
307 ORF2, we modified the RELAX method(59) to accept two gene alignments as input.
308 The RELAX test enforces a functional relationship between the ω ratios in reference
309 (ORF2) and test (ORF3) alignments: $\omega_{\text{ORF3}} = (\omega_{\text{ORF2}})^K$. The estimated value of K
310 indicates whether selection in ORF3 is relaxed ($K < 1$) or intensified ($K > 1$) relative
311 to ORF2. A likelihood ratio test of the null hypothesis ($K=1$), versus the alternative
312 hypothesis ($K \neq 1$) establishes statistical significance of relaxation (or
313 intensification). The application of the RELAX procedure (Table 4) suggests strong
314 relaxation of selection in ORF3 (namely, through the elimination of the positively
315 selected component) in genotypes 1 and 3, and a weak (non-significant)
316 intensification of selection in ORF3 in genotype 4. This finding of ORF 2 apparently
317 driving the signal of positive selection reproduces, by different means, the findings
318 in Table 3.

319 **Estimates of time-scaled synonymous and nonsynonymous substitution rates**

320 Differences in the rate of evolution between different genotypes could arise due to
321 different selection pressures on the genotypes (i.e. different ratios of
322 nonsynonymous to synonymous substitution), as suggested by the selection
323 pressure analyses, or could simply be due to differences in the substitution rate (i.e.
324 differences in synonymous rates), independent of selection pressure. To address

325 this question, we derived time-scaled estimates of synonymous and non-
326 synonymous rates, using the procedure described in (70). Briefly, a Maximum Clade
327 Credibility tree obtained using MrBayes was used as input to a codon analysis in
328 HyPhy, using the Muse-Gaut codon-substitution model with branch-specific α
329 (synonymous) and β (non-synonymous) rate parameters, which were used to
330 partition the fixed branch length into synonymous and non-synonymous
331 components. The conversion from expected substitutions per site to expected
332 substitutions / site / year was carried out under the assumption of a strict
333 molecular clock. The results are summarized in Table 5, and demonstrate that the
334 synonymous substitution rate of genotype 1 is approximately half that of genotypes
335 3 and 4. Whilst this is an important confounding factor, this effect merely adds to an
336 extant signal of positive selection in genotype 1 sequences, because lower dS would
337 work to elevate dN/dS for genotype 1 (for example, results in Table 3 are robust to
338 this confounding factor), it has not created the effect *de novo*.

339 **Analysis of evolutionary rates**

340 We estimated the evolutionary rate of each genotype, including information on the
341 estimated time of sampling (Table 6, Figure 3). The mean evolutionary rate was
342 similar across ORFs at approximately 0.003-0.005 substitutions per site per year.
343 Evolutionary rates of genotype 1 were significantly lower than those of genotypes 3
344 and 4 across all ORFs. Genotypes 3 and 4 demonstrate remarkably similar profiles,
345 with differences non significant across all ORFs. Table 5 shows that this is likely due
346 to both lower synonymous and non-synonymous substitution rates. Unsurprisingly,

347 the overlap region of ORF 2 appears very similar to ORF 3, as they overlap
348 extensively. More surprisingly the non-overlap region of ORF 2 has a similar
349 evolutionary rate profile to ORF 1, with which it does not overlap at all.

350 **Host-specific differences in evolution**

351 We investigated whether patterns of evolution differ not only by genotype but also
352 species of isolation. We constructed phylogenies of the human and swine lineages
353 for genotypes 3 and 4, and assigned branches as either human, swine or
354 indeterminate. For both genotypes 3 and 4, there was notable intermingling of
355 lineages (Figure 4), representing a continuous zoonotic process. Genome-wide
356 analyses of selective pressures using the null and alternative models in the RELAX
357 suite, found a slight, but statistically significant intensification of selection along
358 human branches relative to swine branches. For genotype 3, RELAX inferred the
359 intensification coefficient of $K = 1.09$ ($p = 0.013$ when compared to the null
360 hypothesis of $K = 1$). For genotype 4, the inferred values were $K = 1.12$ and $p <$
361 0.001 . In brief, this test establishes that ω estimates on human branches are more
362 extreme (further away from $\omega = 1$, i.e., neutrality), than on swine branches. For
363 these analyses, indeterminate branches were endowed with their own ω
364 distribution and branch-level relaxation/intensification coefficients, treated here as
365 nuisance parameters. For genotype 3, 91.5% of the bootstrapped trees supported p-
366 value of ≤ 0.05 or less (count = 211, median p value = 0.019 (4E-5-0.0699), median
367 $K = 1.14987$ (1.0324-1.2159)). For G4 every single p-value for RELAX was < 0.05
368 (count = 352, median p value = 9E4 (7E-11-0.0051), median $K = 1.4059$ (1.0900-
369 1.8709)).

370

371 **Discussion**

372 We have demonstrated differences in the evolution of hepatitis E virus (HEV)
373 between the three open reading frames, and quantified how evolutionary patterns
374 differ between genotypes. Using a high quality alignment comprising all available
375 near full length genomes, our analyses have identified and focused in on the main
376 genomic region of interest: the ORF2/ORF3 overlap region (Figure 1). Selection
377 analysis of the overlap region revealed multiple sites/regions undergoing positive
378 selection in genotypes 3 and 4, but a much weaker signal in genotype 1 (Figure 2).
379 This pattern is the same as that found in evolutionary rates, with significantly
380 reduced evolutionary rates in both ORF2 overlap and ORF3 of genotype 1 (Figure 3),
381 driven by differences in both synonymous and nonsynonymous rates. A genome-
382 wide analysis of genotype 3 and 4 isolates revealed a slight but statistically
383 significant intensification of selective pressures in human lineages compared to
384 swine lineages. We speculate, as genotype 1 viruses only infect one host and
385 genotypes 3 and 4 are enzoonotic, that genes in genotype 1 are subject to reduced
386 diversifying or balancing selection pressure as they have fine-tuned fitness by
387 specializing to their single host species. This functional constraint on amino acid
388 changes is particularly pertinent as this effect was found in both ORF2 and ORF3,
389 which are both believed to be important in the pathogen-host response as the
390 capsid protein and an immunomodulatory phosphoprotein, respectively (19). ORF1,
391 in contrast, contains housekeeping genes, which are less likely to be host-specific.

392 Cyclical host jumps seen in arboviruses, e.g., West-Nile virus, are associated with
393 purifying selection (71, 72). The concept behind this is that only substitutions
394 conferring a selection advantage in both hosts are preserved. However this
395 paradigm may not be globally applicable. *In silico* models of evolution under varying
396 selection pressure show that the rate of evolution and dN/dS can be either
397 suppressed or increased depending on how the timescale of the environmental
398 change compares to that of adaptation to the new environment (73, 74). In an
399 environment with very slow environmental fluctuations, each substitution will
400 either fix or go extinct during the epoch in which it arose, whilst in faster
401 oscillations a substitution will have the opportunity to be selected in both
402 environments (74). Therefore it may not be the case that all cyclical environments
403 induce stronger purifying selection. HEV may be an instance where the interaction
404 of oscillation period and time taken to reach a particular fitness level interact in
405 such a way as to promote diversity and a signal of positive selection. We therefore
406 postulate the signal of positive selection in those genomic regions which interact
407 with the host (ORF2 and ORF3, ORF1 contains housekeeping genes) represents a
408 cyclical but ultimately futile selection process in each species which results in a
409 phenotype which is sub-optimally fit in both. Although, interestingly, our host-
410 specific analysis provides evidence that the scales are currently tipped towards
411 optimizing for the human host.

412 Overlapping reading frames are not uncommon in RNA viruses (75), and have been
413 suggested as a mechanism of packing more genes in a limited genomic space (76).

414 Whilst other studies have found a scattering of positively selected codons in this

415 region (26), none have investigated the overlap region as a locus for positive
416 selection. Overlapping coding regions are often constrained as substitutions impact
417 two protein products instead of one, causing a reduction in evolutionary rates (77).
418 However, there is a precedent for rapid evolution in overlapped regions in both
419 viruses (e.g. PB1-F2 and PA-X in Influenza A virus (78, 79)) and mammalian
420 genomes (80), and statistical techniques designed for mammalian overlapping
421 regions (62) helped us to shed light on what is driving evolution in the overlap
422 region. Investigating selection in a region of overlapping reading frames requires
423 reading-frame aware models. Apart from a currently computationally infeasible full
424 Bayesian treatment of co-dependent evolution in multiple reading frames (e.g. see
425 (81)), two approximate approaches have been used in practice. Firstly, the
426 overlapping reading frames can be treated entirely independently, and analysed
427 using standard methods (e.g. (82)). When this approach is taken to estimate
428 synonymous and non-synonymous rates and carry out tests of selection, the
429 interpretation of results becomes difficult (e.g., how valid is the concept of a frame-
430 specific synonymous rate in this context?), and can lead to false positive results (83).
431 Secondly, codon-substitution models which correct for the "expected" context of a
432 codon in the alternative reading frame have been proposed (62, 84, 85). The benefit
433 of these models is that, while remaining computationally tractable, they directly
434 estimate frame-aware rates of synonymous and non-synonymous mutations. Such
435 models have been successfully used to perform genome-wide screens of ORFs with
436 multiple overlapping reading frames for functional constraint (62), and the
437 evolution of overlapping reading frames in Influenza A virus (84). Our analysis of

438 the overlapping reading frames shows a significant difference between the rate of
439 substitutions that were non-synonymous in both frames compared to only one
440 (Table 3), indicative of positive selection. Applying this model also allowed us to
441 find out which reading frame (and therefore likely gene product) was driving the
442 positive signal in this region. Although both reading frames are subject to positive
443 selection, the ORF2 overlap region appears to be driving selection (at least in
444 genotypes 1 and 3).

445 Evolutionary rates showed significant differences between the anthroptropic and
446 zoonotic genotypes. Across all genomic regions genotype 1 had significantly lower
447 evolutionary rates than genotypes 3 and 4, whilst genotypes 3 and 4 had
448 remarkably similar values. Evolutionary rates inferred from the posterior
449 distribution were typical of an RNA virus (86) and related viruses e.g. norovirus
450 (87). We, like Nakano *et. al.* (2012) and Purdy *et. al.* (2012), found a relaxed clock
451 most appropriate to reflect the variation in substitution rates between branches in
452 HEV, although our estimates of evolutionary rate are higher than those reported
453 previously (88–90). It should be noted that apparent evolutionary rates show time
454 dependency, with an elevation towards the present due to transient unfixed
455 substitutions, and apparent reduction in the past due to saturation (91).

456 Interestingly the ORF 2 overlap region has a very distinct profile of evolutionary
457 rates across all genotypes when compared with the non-overlap region. The non-
458 overlap region is strikingly similar to the ORF 1 profile, which is believed to contain
459 housekeeping genes.

460 Our analyses of the host specific patterns of evolution are important in showing that
461 the differences described above are largely genotype, not host species, dependent.
462 For genotypes 3 and 4, we detected a slight increase in selection intensity in human-
463 associated viral lineages compared to swine associated viral lineages, in contrast to
464 the large differences between genotypes. The construction of phylogenies
465 demonstrated intermingling of swine and human lineages, and suggest a high rate of
466 host jumps indicative of the frequent transmission between swine and humans and
467 back again. The transmission of HEV from humans to swine has been demonstrated
468 extensively in laboratory settings (92, 93), however its frequency and mechanism in
469 the wild remain unclear (94). As phylogenies do not contain independent
470 information on the direction of transmission, it is hard to demonstrate such 'reverse
471 zoonoses' from sequence data alone.

472 Our study represents the most comprehensive HEV sequence analysis to date. It is
473 important, however, to note the limitations in the publicly available data. Genotypes
474 3 and 4 dominate in the developed world, whilst genotypes 1 and 2 are found in the
475 developing world (4). This global differential distribution of genotypes may be an
476 important confounder, as in fact the virus is not interacting with a single
477 homogeneous human host, but rather different clades of virus are interacting with
478 specific groups of human hosts. These groups are likely to differ significantly, e.g. in
479 the population composition of Human Leukocyte Antigen alleles (95), which in turn
480 imposes differential selective pressures on the pathogen as part of host-pathogen
481 interactions. As is the case for most pathogens, sampling is heavily biased by
482 location. There are many samples from Europe and East Asia, but few from

483 Australasia and Africa, and there are many countries for which there are no
484 sequence data. Furthermore, little is known about genotype 2, with too few full
485 length viral genomes publicly available to build a reliable alignment, so studies
486 either omit it (26), or have low statistical power (96).

487 Hepatitis E virus is of increasing interest to public health officials and clinicians.
488 Attention in the developed world to date has been limited, partly due to the acute
489 nature of the infection in healthy individuals and the apparently asymptomatic
490 nature of infection in swine. However, the emergence of new strains of HEV, such as
491 one recently documented in the U.K. (97), emphasise the need for continuing
492 surveillance and characterisation of this pathogen.

493

494 **Acknowledgements**

495 This work was funded in part by a Medical Research Council Methodology Research
496 Programme grant (MR/J013862/1) to SDWF, as well as by the Department of
497 Pathology at the University of Cambridge. SLKP and HyPhy development was
498 supported by the US National Institutes of Health (grants GM093939 and
499 GM110749). Many thanks to Prof. Ian Goodfellow (University of Cambridge) for
500 providing a genomic map of HEV.

501 AB performed data collection, annotation and interpretation, ran analyses, created
502 figures and drafted the initial paper. SP devised overlapping reading frame analysis,
503 ran further analyses, created figures, and revised the draft paper. BD and JL

504 provided technical assistance, including generation of figures, and revised the draft
505 paper. SF devised and initiated the collaborative project, designed the data
506 collection and analysis strategies and revised the draft paper.

507

508 **References**

509 1. **Rein DB, Stevens GA, Theaker J, Wittenborn JS, Wiersma ST.** 2012. The global
510 burden of hepatitis e virus genotypes 1 and 2 in 2005. *Hepatology* **55**:988–997.

511 2. **Meng X-J.** 2013. Zoonotic and foodborne transmission of hepatitis E virus. *Semin*
512 *Liver Dis* **33**:41–49.

513 3. **Berto A, Martelli F, Grierson S, Banks M.** 2012. Hepatitis E virus in pork food
514 chain, united kingdom, 2009-2010. *Emerg Infect Dis* **18**:1358–1360.

515 4. **Teshale EH, Hu DJ, Holmberg SD.** 2010. The two faces of hepatitis E virus. *Clin*
516 *Infect Dis* **51**:328–334.

517 5. **Lewis H, Wichmann O, Duizer E.** 2010. Transmission routes and risk factors for
518 autochthonous hepatitis E virus infection in europe: a systematic review. *Epidemiol*
519 *Infect* **138**:145–166.

520 6. **Banks M, Bendall R, Grierson S, Heath G, Mitchell J, Dalton H.** 2004. Human
521 and porcine hepatitis E virus strains, United Kingdom. *Emerg Infect Dis* **10**:953–955.

522 7. **Emerson S, Anderson D, Arankalle A.** 2004. VIIIth report of the ICTV. Report.

- 523 8. **Worm HC, Poel WHM van der, Brandstatter G.** 2002. Hepatitis E: an overview.
524 *Microbes Infect* **4**:657–666.
- 525 9. **Lu L, Li CH, Hagedorn CH.** 2006. Phylogenetic analysis of global hepatitis E virus
526 sequences: genetic diversity, subtypes and zoonosis. *Rev Med Virol* **16**:5–36.
- 527 10. **Schlauder GG, Mushahwar IK.** 2001. Genetic heterogeneity of hepatitis E virus.
528 *J Med Virol* **65**:282–292.
- 529 11. **Shrestha MP, Scott RM, Joshi DM, Mammen Jr MP, Thapa GB, Thapa N, Myint**
530 **KSA, Fourneau M, Kuschner RA, Shrestha SK, others.** 2007. Safety and efficacy of
531 a recombinant hepatitis e vaccine. *New England Journal of Medicine* **356**:895–903.
- 532 12. **Zhu F-C, Zhang J, Zhang X-F, Zhou C, Wang Z-Z, Huang S-J, Wang H, Yang C-L,**
533 **Jiang H-M, Cai J-P, others.** 2010. Efficacy and safety of a recombinant hepatitis e
534 vaccine in healthy adults: a large-scale, randomised, double-blind placebo-
535 controlled, phase 3 trial. *The Lancet* **376**:895–902.
- 536 13. **Kuniholm MH, Purcell RH, McQuillan GM, Engle RE, Wasley A, Nelson KE.**
537 2009. Epidemiology of hepatitis e virus in the united states: results from the third
538 national health and nutrition examination survey, 1988–1994. *Journal of Infectious*
539 *Diseases* **200**:48–56.
- 540 14. **Bendall R, Ellis V, Ijaz S, Ali R, Dalton H.** 2010. A comparison of two
541 commercially available anti-hEV igG kits and a re-evaluation of anti-hEV igG
542 seroprevalence data in developed countries. *Journal of medical virology* **82**:799–
543 805.

- 544 15. **Kamar N, Bendall R, Legrand-Abravanel F, Xia N-S, Ijaz S, Izopet J, Dalton**
545 **HR.** 2012. Hepatitis e. *The Lancet* **379**:2477–2488.
- 546 16. **Dalton HR, Stableforth W, Thurairajah P, Hazeldine S, Remnarace R, Usama**
547 **W, Farrington L, Hamad N, Sieberhagen C, Ellis V, others.** 2008. Autochthonous
548 hepatitis e in southwest england: natural history, complications and seasonal
549 variation, and hepatitis e virus igG seroprevalence in blood donors, the elderly and
550 patients with chronic liver disease. *European journal of gastroenterology &*
551 *hepatology* **20**:784–790.
- 552 17. **Purcell RH, Engle RE, Govindarajan S, Herbert R, St Claire M, Elkins WR,**
553 **Cook A, Shaver C, Beauregard S Michelle, Emerson S.** 2013. Pathobiology of
554 hepatitis e: lessons learned from primate models. *Emerging Microbes & Infections*
555 **2**:e9.
- 556 18. **Wang Y, Zhang H, Ling R, Li H, Harrison TJ.** 2000. The complete sequence of
557 hepatitis E virus genotype 4 reveals an alternative strategy for translation of open
558 reading frames 2 and 3. *J Gen Virol* **81**:1675–1686.
- 559 19. **Cao D, Meng X-J.** 2012. Molecular biology and replication of hepatitis e virus.
560 *Emerging microbes & infections* **1**:e17.
- 561 20. **Emerson SU, Nguyen H, Torian U, Purcell RH.** 2006. ORF3 protein of hepatitis
562 e virus is not required for replication, virion assembly, or infection of hepatoma cells
563 in vitro. *Journal of virology* **80**:10457–10464.

- 564 21. **Graff J, Nguyen H, Yu C, Elkins WR, Claire MS, Purcell RH, Emerson SU.** 2005.
565 The open reading frame 3 gene of hepatitis e virus contains a cis-reactive element
566 and encodes a protein required for infection of macaques. *Journal of virology*
567 **79**:6680–6689.
- 568 22. **Chandra V, Kar-Roy A, Kumari S, Mayor S, Jameel S.** 2008. The hepatitis E
569 virus ORF3 protein modulates epidermal growth factor receptor trafficking, STAT3
570 translocation, and the acute-phase response. *J Virol* **82**:7100–7110.
- 571 23. **Tyagi S, Korkaya H, Zafrullah M, Jameel S, Lal SK.** 2002. The phosphorylated
572 form of the oRF3 protein of hepatitis e virus interacts with its non-glycosylated form
573 of the major capsid protein, oRF2. *Journal of Biological Chemistry* **277**:22759–
574 22767.
- 575 24. **Tyagi S, Surjit M, Lal SK.** 2005. The 41-amino-acid c-terminal region of the
576 hepatitis e virus oRF3 protein interacts with bikunin, a kunitz-type serine protease
577 inhibitor. *Journal of virology* **79**:12081–12087.
- 578 25. **Tyagi S, Surjit M, Roy AK, Jameel S, Lal SK.** 2004. The oRF3 protein of hepatitis
579 e virus interacts with liver-specific α 1-microglobulin and its precursor α 1-
580 microglobulin/bikunin precursor (aMBP) and expedites their export from the
581 hepatocyte. *Journal of Biological Chemistry* **279**:29308–29319.
- 582 26. **Chen X, Zhang Q, He C, Zhang L, Li J, Zhang W, Cao W, Lv Y-G, Liu Z, Zhang J-X,**
583 **Shao Z-J.** 2012. Recombination and natural selection in hepatitis E virus genotypes. *J*
584 *Med Virol* **84**:1396–1407.

- 585 27. **Smith DB, Vanek J, Ramalingam S, Johannessen I, Templeton K, Simmonds**
586 **P.** 2012. Evolution of the hepatitis E virus hypervariable region. *J Gen Virol*
587 **93**:2408–2418.
- 588 28. **Purdy MA, Lara J, Khudyakov YE.** 2012. The hepatitis E virus polyproline
589 region is involved in viral adaptation. *PLoS One* **7**:e35974–e35974.
- 590 29. **Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J,**
591 **Sayers EW.** 2013. GenBank. *Nucleic Acids Res* **41**:D36–D42.
- 592 30. **Rice P, Longden I, Bleasby A.** 2000. EMBOSS: the european molecular biology
593 open software suite. *Trends in genetics* **16**:276–277.
- 594 31. **Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K,**
595 **Madden TL.** 2009. BLAST+: architecture and applications. *BMC Bioinformatics*
596 **10**:421.
- 597 32. **Sievers F, Higgins DG.** 2014. Clustal omega, accurate alignment of very large
598 numbers of sequences. *Methods Mol Biol* **1079**:105–116.
- 599 33. **Gouy M, Guindon S, Gascuel O.** 2010. SeaView version 4: A multiplatform
600 graphical user interface for sequence alignment and phylogenetic tree building. *Mol*
601 *Biol Evol* **27**:221–224.
- 602 34. **Martin DP, Murrell B, Golden M, Khoosal A, Muhire B.** 2015. RDP4: Detection
603 and analysis of recombination patterns in virus genomes. *Virus Evol* **1**:vev003.

- 604 35. **Martin D, Rybicki E.** 2000. RDP: detection of recombination amongst aligned
605 sequences. *Bioinformatics* **16**:562–563.
- 606 36. **Padidam M, Sawyer S, Fauquet CM.** 1999. Possible emergence of new
607 geminiviruses by frequent recombination. *Virology* **265**:218–225.
- 608 37. **Martin DP, Posada D, Crandall KA, Williamson C.** 2005. A modified bootscan
609 algorithm for automated identification of recombinant sequences and
610 recombination breakpoints. *AIDS Res Hum Retroviruses* **21**:98–102.
- 611 38. **Maynard Smith J.** 1992. Analyzing the mosaic structure of genes. *J Mol Evol*
612 **34**:126–129.
- 613 39. **Posada D, Crandall KA.** 2001. Evaluation of methods for detecting
614 recombination from DNA sequences: computer simulations. *Proc Natl Acad Sci U S A*
615 **98**:13757–13762.
- 616 40. **Gibbs MJ, Armstrong JS, Gibbs AJ.** 2000. Sister-scanning: a monte carlo
617 procedure for assessing signals in recombinant sequences. *Bioinformatics* **16**:573–
618 582.
- 619 41. **Weiller GF.** 1998. Phylogenetic profiles: a graphical method for detecting
620 genetic recombinations in homologous sequences. *Mol Biol Evol* **15**:326–335.
- 621 42. **Holmes EC, Worobey M, Rambaut A.** 1999. Phylogenetic evidence for
622 recombination in dengue virus. *Mol Biol Evol* **16**:405–409.

- 623 43. **Boni MF, Posada D, Feldman MW.** 2007. An exact nonparametric method for
624 inferring mosaic structure in sequence triplets. *Genetics* **176**:1035–1047.
- 625 44. **Cuyck H van, Fan J, Robertson DL, Roques P.** 2005. Evidence of recombination
626 between divergent hepatitis E viruses. *J Virol* **79**:9306–9314.
- 627 45. **Wang H, Zhang W, Ni B, Shen H, Song Y, Wang X, Shao S, Hua X, Cui L.** 2010.
628 Recombination analysis reveals a double recombination event in hepatitis E virus.
629 *Virol J* **7**.
- 630 46. **Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ.** 1990. Basic local
631 alignment search tool. *J Mol Biol* **215**:403–410.
- 632 47. **Tam AW, Smith MM, Guerra ME, Huang CC, Bradley DW, Fry KE, Reyes GR.**
633 1991. Hepatitis e virus (hEV): molecular cloning and sequencing of the full-length
634 viral genome. *Virology* **185**:120–131.
- 635 48. **Huang CC, Nguyen D, Fernandez J, Yun KY, Fry KE, Bradley DW, Tam AW,**
636 **Reyes GR.** 1992. Molecular cloning and sequencing of the mexico isolate of hepatitis
637 E virus (HEV). *Virology* **191**:550–558.
- 638 49. **Schlauder GG, Dawson GJ, Erker JC, Kwo PY, Knigge MF, Smalley DL,**
639 **Rosenblatt JE, Desai SM, Mushahwar IK.** 1998. The sequence and phylogenetic
640 analysis of a novel hepatitis E virus isolated from a patient with acute hepatitis
641 reported in the united states. *J Gen Virol* **79 (Pt 3)**:447–456.
- 642 50. **Price MN, Dehal PS, Arkin AP.** 2010. FastTree 2–approximately maximum-
643 likelihood trees for large alignments. *PLoS One* **5**:e9490.

644 51. **Izopet J, Dubois M, Bertagnoli S, Lhomme S, Marchandean S, Boucher S,**
645 **Kamar N, Abravanel F, Guérin J-L.** 2012. Hepatitis E virus strains in rabbits and
646 evidence of a closely related strain in humans, France. *Emerg Infect Dis* **18**:1274–
647 1281.

648 52. **Ranwez V, Harispe S, Delsuc F, Douzery EJ.** 2011. MACSE: Multiple alignment
649 of coding SEquences accounting for frameshifts and stop codons. *PLoS One*
650 **6**:e22594.

651 53. **Pond SLK, Frost SDW, Muse SV.** 2005. HyPhy: hypothesis testing using
652 phylogenies. *Bioinformatics* **21**:676–679.

653 54. **Pond SLK, Frost SDW.** 2005. Datamonkey: rapid detection of selective pressure
654 on individual sites of codon alignments. *Bioinformatics* **21**:2531–2533.

655 55. **Delpont W, Poon AFY, Frost SDW, Pond SLK.** 2010. Datamonkey 2010: a suite
656 of phylogenetic analysis tools for evolutionary biology. *Bioinformatics* **26**:2455–
657 2457.

658 56. **Murrell B, Moola S, Mabona A, Weighill T, Sheward D, Kosakovsky Pond SL,**
659 **Scheffler K.** 2013. FUBAR: a fast, unconstrained bayesian approximation for
660 inferring selection. *Mol Biol Evol* **30**:1196–205.

661 57. **Murrell B, Wertheim JO, Moola S, Weighill T, Scheffler K, Pond SK.** 2012.
662 Detecting individual sites subject to episodic diversifying selection. *PLoS Genetics*
663 **8**:e1002764–e1002764.

664 58. **Smith MD, Wertheim JO, Weaver S, Murrell B, Scheffler K, Kosakovsky Pond**
665 **SL.** 2015. Less is more: an adaptive branch-site random effects model for efficient
666 detection of episodic diversifying selection. *Mol Biol Evol* **32**:1342–53.

667 59. **Wertheim JO, Murrell B, Smith MD, Kosakovsky Pond SL, Scheffler K.** 2015.
668 RELAX: detecting relaxed selection in a phylogenetic framework. *Mol Biol Evol*
669 **32**:820–32.

670 60. **Wertheim JO, Murrell B, Smith MD, Pond SLK, Scheffler K.** 2015. RELAX:
671 detecting relaxed selection in a phylogenetic framework. *Molecular biology and*
672 *evolution* **32**:820–832.

673 61. **Pond SLK, Murrell B, Fourment M, Frost SD, Delport W, Scheffler K.** 2011. A
674 random effects branch-site model for detecting episodic diversifying selection.
675 *Molecular biology and evolution* msr125.

676 62. **Chung W-Y, Wadhawan S, Szklarczyk R, Pond SK, Nekrutenko A.** 2007. A first
677 look at ARFome: Dual-coding genes in mammalian genomes. *PLoS Comput Biol*
678 **3**:855–861.

679 63. **Reyes GR, Purdy MA, Kim JP, Luk KC, Young LM, Fry KE, Bradley DW.** 1990.
680 Isolation of a cDNA from the virus responsible for enterically transmitted non-A,
681 non-B hepatitis. *Science* **247**:1335–1339.

682 64. **Ronquist F, Teslenko M, Mark P van der, Ayres DL, Darling A, Hohna S,**
683 **Larget B, Liu L, Suchard MA, Huelsenbeck JP.** 2012. MrBayes 3.2: Efficient

684 bayesian phylogenetic inference and model choice across a large model space. Syst
685 Biol **61**:539–542.

686 65. **Plummer M, Best N, Cowles K, Vines K**. 2006. CODA: Convergence diagnosis
687 and output analysis for mCMC. R News **6**:7–11.

688 66. **Gelman A, Goegebeur Y, Tuerlinckx F, Van Mechelen I**. 2000. Diagnostic
689 checks for discrete data regression models using posterior predictive simulations. J
690 R Stat Soc Ser C Appl Stat **49**:247–268.

691 67. **Wickham H**. 2009. ggplot2: elegant graphics for data analysis. Springer New
692 York.

693 68. **Stamatakis A**. 2014. RAxML version 8: a tool for phylogenetic analysis and post-
694 analysis of large phylogenies. Bioinformatics **30**:1312–1313.

695 69. **To T-H, Jung M, Lycett S, Gascuel O**. 2015. Fast dating using least-squares
696 criteria and algorithms. Systematic biology syv068.

697 70. **Lemey P, Pond SLK, Drummond AJ, Pybus OG, Shapiro B, Barroso H, Taveira
698 N, Rambaut A**. 2007. Synonymous substitution rates predict HIV disease
699 progression as a result of underlying replication dynamics. PLoS Comput Biol **3**:e29.

700 71. **Coffey LL, Forrester N, Tsetsarkin K, Vasilakis N, Weaver SC**. 2013. Factors
701 shaping the adaptive landscape for arboviruses: implications for the emergence of
702 disease. Future microbiology **8**:155–176.

703 72. **Parameswaran P, Charlebois P, Tellez Y, Nunez A, Ryan EM, Malboeuf CM,**
704 **Levin JZ, Lennon NJ, Balmaseda A, Harris E, others.** 2012. Genome-wide patterns
705 of intrahuman dengue virus diversity reveal associations with viral phylogenetic
706 clade and interhost diversity. *Journal of virology* **86**:8546–8558.

707 73. **Kashtan N, Noor E, Alon U.** 2007. Varying environments can speed up
708 evolution. *Proceedings of the National Academy of Sciences* **104**:13711–13716.

709 74. **Cvijović I, Good BH, Jerison ER, Desai MM.** 2015. Fate of a mutation in a
710 fluctuating environment. *Proc Natl Acad Sci U S A* **112**:E5021–E5028.

711 75. **Neuhaus K, Oelke D, Fürst D, Scherer S, Keim DA.** 2010. Towards automatic
712 detecting of overlapping genes-clustered bLAST analysis of viral genomes. Springer.

713 76. **Chirico N, Vianelli A, Belshaw R.** 2010. Why genes overlap in viruses.
714 *Proceedings of the Royal Society of London B: Biological Sciences* **277**:3809–3817.

715 77. **Simon-Loriere E, Holmes EC, Pagan I.** 2013. The effect of gene overlapping on
716 the rate of RNA virus evol. *Mol Biol Evol* **30**:1916–1928.

717 78. **Suzuki Y.** 2006. Natural selection on the influenza virus genome. *Molecular*
718 *biology and evolution* **23**:1902–1911.

719 79. **Jagger B, Wise H, Kash J, Walters K-A, Wills N, Xiao Y-L, Dunfee R,**
720 **Schwartzman L, Ozinsky A, Bell G, others.** 2012. An overlapping protein-coding
721 region in influenza a virus segment 3 modulates the host response. *Science*
722 **337**:199–204.

- 723 80. **Szklarczyk R, Heringa J, Pond SK, Nekrutenko A.** 2007. Rapid asymmetric
724 evolution of a dual-coding tumor suppressor INK4a/ARF locus contradicts its
725 function. *Proc Natl Acad Sci U S A* **104**:12807–12812.
- 726 81. **Pedersen A-MK, Jensen JL.** 2001. A dependent-rates model and an mCMC-
727 based methodology for the maximum-likelihood analysis of sequences with
728 overlapping reading frames. *Molecular Biology and Evolution* **18**:763–776.
- 729 82. **Obenauer JC, Denson J, Mehta PK, Su X, Mukatira S, Finkelstein DB, Xu X,**
730 **Wang J, Ma J, Fan Y, others.** 2006. Large-scale sequence analysis of avian influenza
731 isolates. *Science* **311**:1576–1580.
- 732 83. **Holmes EC, Lipman DJ, Zamarin D, Yewdell JW.** 2006. Comment on“ large-
733 Scale sequence analysis of avian influenza isolates”. *Science* **313**:1573–1573.
- 734 84. **Sabath N, Landan G, Graur D.** 2008. A method for the simultaneous estimation
735 of selection intensities in overlapping genes. *PLoS One* **3**:e3996.
- 736 85. **Mir K, Schober S.** 2014. Selection pressure in alternative reading frames. *PloS*
737 *one* **9**.
- 738 86. **Jenkins GM, Rambaut A, Pybus OG, Holmes EC.** 2002. Rates of molecular
739 evolution in RNA viruses: A quantitative phylogenetic analysis. *J Mol Evol* **54**:156–
740 165.
- 741 87. **Cotten M, Petrova V, Phan MV, Rabaa MA, Watson SJ, Ong SH, Kellam P,**
742 **Baker S.** 2014. Deep sequencing of norovirus genomes defines evolutionary
743 patterns in an urban tropical setting. *J Virol* **88**:11056–11069.

744 88. **Takahashi K, Toyota J, Karino Y, Kang JH, Maekubo H, Abe N, Mishiro S.**
745 2004. Estimation of the mutation rate of hepatitis E virus based on a set of closely
746 related 7.5-year-apart isolates from sapporo, japan. *Hepatol Res* **29**:212–215.

747 89. **Purdy MA, Khudyakov YE.** 2010. Evolutionary history and population dynamics
748 of hepatitis E virus. *Plos One* **5**:9.

749 90. **Nakano T, Takahashi K, Pybus OG, Hashimoto N, Kato H, Okano H,**
750 **Kobayashi M, Fujita N, Shiraki K, Takei Y, Ayada M, Arai M, Okamoto H, Mishiro**
751 **S.** 2012. New findings regarding the epidemic history and population dynamics of
752 japan-indigenous genotype 3 hepatitis E virus inferred by molecular evolution. *Liver*
753 *Int* **32**:675–688.

754 91. **Ho SY, Shapiro B, Phillips MJ, Cooper A, Drummond AJ.** 2007. Evidence for
755 time dependency of molecular rate estimates. *Systematic biology* **56**:515–522.

756 92. **Meng X-J, Halbur PG, Shapiro MS, Govindarajan S, Bruna JD, Mushahwar IK,**
757 **Purcell RH, Emerson SU.** 1998. Genetic and experimental evidence for cross-
758 species infection by swine hepatitis E virus. *J Virol* **72**:9714–9721.

759 93. **Feagins A, Opriessnig T, Huang Y, Halbur P, Meng X.** 2008. Cross-species
760 infection of specific-pathogen-free pigs by a genotype 4 strain of human hepatitis E
761 virus. *J Med Virol* **80**:1379.

762 94. **Messenger AM, Barnes AN, Gray GC.** 2014. Reverse zoonotic disease
763 transmission (zooanthroponosis): a systematic review of seldom-documented
764 human biological threats to animals. *PloS one* **9**:e89055.

765 95. **Buhler S, Sanchez-Mazas A.** 2011. HLA DNA sequence variation among human
766 populations: molecular signatures of demographic and selective events. PLoS One
767 6:e14643.

768 96. **Okamoto H.** 2007. Genetic variability and evolution of hepatitis E virus. Virus
769 Res 127:216–228.

770 97. **Ijaz S, Said B, Boxall E, Smit E, Morgan D, Tedder RS.** 2014. Indigenous
771 hepatitis E in england and wales from 2003 to 2012: evidence of an emerging novel
772 phylotype of viruses. J Infect Dis 209:1212–1218.

773

774 **Tables**775 **Recombination analysis**

Accession	Recombination reference	Genotype	Host	Major parent	Minor parent
AB097811	Wang et al. (2010)	3	Swine	AB193177	AB481227
AB291954	NONE	3	Human	AB443626	AB291953
D11093	van Cuyck et al. (2005)	NA	Human	D11092	D10330
DQ450072	Wang et al. (2010)	4	Swine	JF915746	GU188851;AB091394
EU723513	NONE	3	Swine	EU723512	EU723515
FJ426404	NONE	3	Swine	Unknown	FJ426403
FJ457024	NONE	NA	Human	JF443725	AF459438
HM439284	NONE	4	Human	JQ993308	JX855794; EU676172
JF443720	NONE	1	Human	AF459438	JF443725
JN564006	NONE	3	Human	AB089824	JQ679014
JQ655735	NONE	4	Human	GU188851	JQ655733
JX565469	NONE	Rabbit	Rabbit	AB740222	GU937805
KJ013414	NONE	Rabbit	Rabbit	Unknown	JQ768461;JX121233
KJ013415	NONE	Rabbit	Rabbit	Unknown	JQ768461;JX121233

776 Table 1: Details of recombinants found. 14 recombinant HEV sequences were
777 identified in the 258 near full length genomes, generated by concatenating ORF1
778 and ORF2, by screening with RDP4 (version 4.36 beta) (34). With the exception of
779 KJ013414 and KJ013415, which shared a recombinant structure, all recombinants
780 were unique. Three recombinants had been previously described (see

781 Recombination reference column). The table also shows the genotype of the
782 recombinant, the host it was isolated from, and the putative major and minor
783 parents.

784

Genotype	Reading frame (ORF)	FUBAR (posterior ≥ 0.95)	MEME ($p \leq 0.05$)
1	2	0	0
1	3	0	0
3	2	7	4 (4)
3	3	2	4 (2)
4	2	6	7 (6)
4	3	1	6 (1)

785 Table 2: The number of positively selected codon sites in each reading frame of each
786 genotype of the overlap region (numbers in parentheses show how many sites were
787 shared between MEME and FUBAR sets). Genotype 1 lacks any positively selected
788 sites, meanwhile genotypes 3 and 4 produce a consistent signal of positively
789 selected sites in both reading frames. Note that MEME is generally more sensitive,
790 because it can detect selection on a subset of viral lineages, whilst FUBAR pools the
791 signal of selection from all branches.

792

Genotype	ORF2	ORF3	Both	Both < ORF2/ORF3 LRT p-value	ORF2 ≠ ORF3 LRT p- value
1	0.056	0.032	0.012	0.005	0.039
95% CI	(0.036,0.083)	(0.020,0.048)	(0.005,0.024)		
3	0.167	0.082	0.011	<0.001	<0.001
95% CI	(0.138,0.201)	(0.066,0.099)	(0.005, 0.018)		
4	0.113	0.092	0.015	<0.001	0.12
95% CI	(0.090,0.140)	(0.076,0.110)	(0.009, 0.024)		

793 Table 3. Estimates of substitution rates that result in non-synonymous changes in at
794 least one frame, relative to the rate of substitutions that are synonymous in both
795 frames. A dimensionless metric, based on the model from Chung *et al.* (62). The last
796 two columns show LRT-based p-values for rejecting the corresponding null
797 hypotheses. Genotypes 3 and 4 demonstrate highly significant reading frame
798 specific positive selection with ORF 2 convincingly driving the signal in genotype 3
799 but not 4 (rejection of null hypothesis). Genotype 1 has a lower background rate of
800 non synonymous substitutions although does achieve significance, with the ORF 2
801 rate again significantly higher than ORF 3 rate.

802

Genotype	Relaxation parameter (K)	RELAX test p-value
1	< 0.0001	0.002
3	< 0.0001	< 0.0001
4	1.42	0.16

803 Table 4. Application of the RELAX procedure suggests strong relaxation of selection
804 (namely, through the elimination of the positive selected component) in ORF3 of
805 genotypes 1 and 3 relative to ORF2, and a weak, non-significant intensification of
806 selection in genotype 4 of ORF3 relative to ORF2. This suggests that ORF2 and ORF3
807 are evolving differently, and ORF 2 is more responsible than ORF 3 for the signal of
808 positive selection in genotypes 1 and 3.

809

Region	Genotype	Expected Synonymous substitutions / site / year	Expected non-synonymous substitutions / site / year
ORF 1	1	0.0022	0.00034
ORF 1	3	0.0051	0.00039
ORF 1	4	0.0053	0.00054
ORF 2 (non-overlap)	1	0.0030	0.00022
ORF 2 (non-overlap)	3	0.0063	0.00022
ORF 2 (non-overlap)	4	0.0051	0.00024
ORF 2 (overlap)	1	0.0022	0.00027
ORF 2 (overlap)	3	0.0040	0.00029
ORF 2 (overlap)	4	0.0053	0.00054
ORF 3 (overlap)	1	0.00067	0.00016
ORF 3 (overlap)	3	0.0017	0.00092
ORF 3 (overlap)	4	0.0011	0.00075

810 Table 5. Estimation of genotype specific synonymous substitution rates and non-
811 synonymous substitution rates performed after Lemey *et. al.* (70). Synonymous
812 substitution rate of genotype 1 is approximately half that of genotypes 3 and 4,
813 which contributes to, but does not constitute, the signal of genotype specific positive
814 selection. The rate of substitutions in ORF 3 is consistently elevated in comparison
815 to other ORFs.

816

ORF	Genotype	Genotype	2.5% CredibleInterval	97.5% CredibleInterval	Significance
1	1	3	-0.004568928	-0.0004705684	*
1	1	4	-0.004663462	-0.0003018292	*
1	3	4	-0.002376133	+0.0023648283	
2nonoverlap	1	3	-0.004737947	-0.0002768805	*
2nonoverlap	1	4	-0.004861239	-0.0001427095	*
2nonoverlap	3	4	-0.002609059	+0.0025491087	
2overlap	1	3	-0.003233479	-0.0005518198	*
2overlap	1	4	-0.005747935	-0.0014066999	*
2overlap	3	4	-0.004201915	+0.0006952047	
3overlap	1	3	-0.003521166	-0.0007273096	*
3overlap	1	4	-0.005043289	-0.0010736460	*
3overlap	3	4	-0.003172870	+0.0013850900	

817 Table 6. Assessing significance in differences in clockrates between genotypes for
818 each ORF. The credible intervals are significant if they do not include zero. This
819 shows genotype 1 has a significantly different clockrate from genotypes 3 & 4 across
820 all ORFs. This supports the clockrate data in Figure 3.

821

822 **Figure Legends**

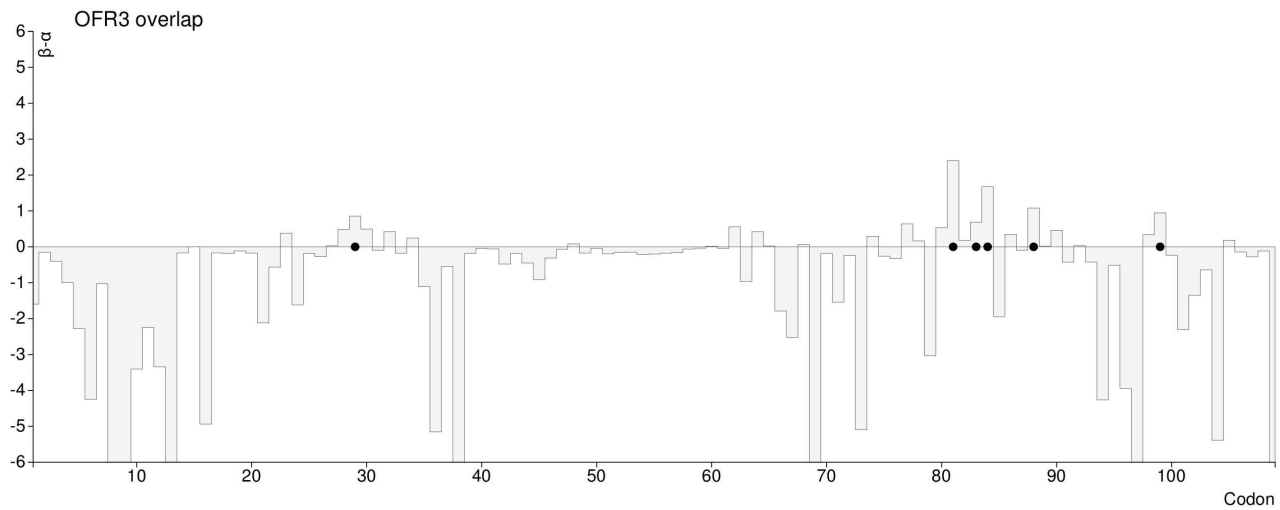
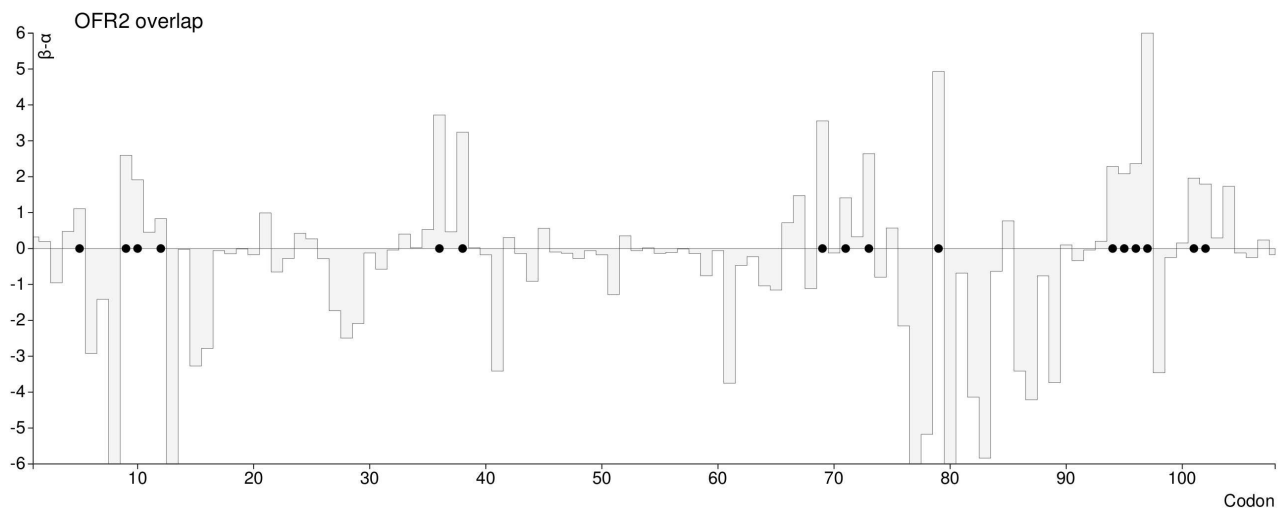
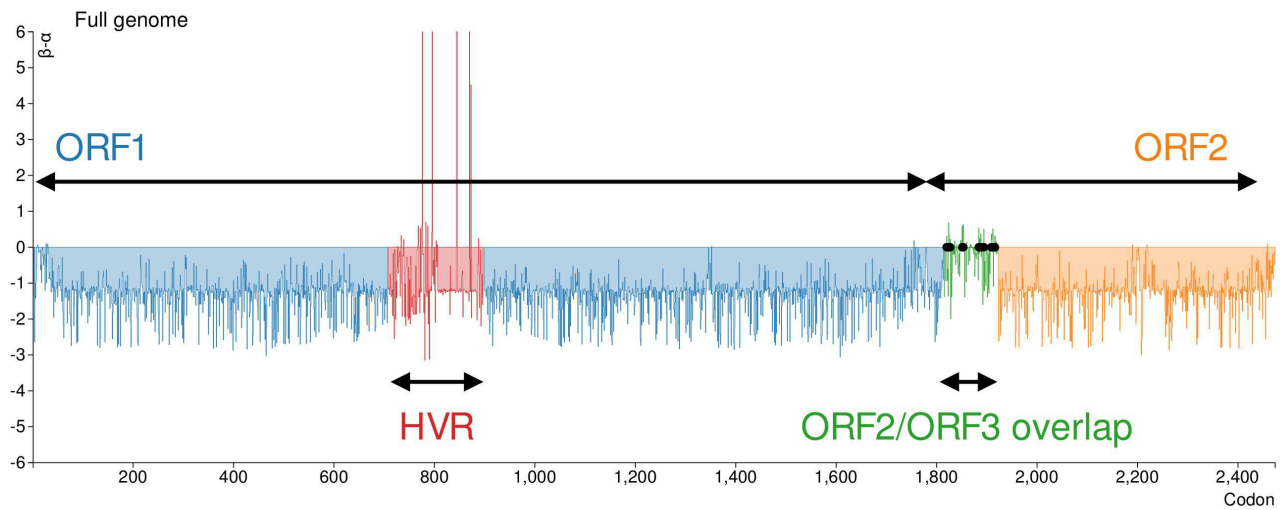
823 Figure 1. FUBAR analysis of concatenated ORF1 and ORF2 sequences isolated from
824 humans (n=113). Genome-wide patterns of non-synonymous (β) and synonymous
825 (α) substitutions per site show that HEV has a background of purifying selection
826 with two discrete regions of elevated diversity corresponding to the hypervariable
827 region (HVR) and the overlap region between ORF2 and ORF3, as shown on the
828 genomic map. Sites subject to significant pervasive positive selection (FUBAR
829 posterior probability ≥ 0.95) are shown as black circles on the x-axis. FUBAR
830 analysis of the ORF2/3 overlap regions in their respective reading frames, showing
831 positive selection in both frames, but with ORF2 demonstrating a stronger signal
832 than ORF3, both in terms of the number of positively selected sites, and the
833 magnitude of β - α .

834 Figure 2. FUBAR Rate analysis of the ORF2/3 overlap region showing conserved
835 patterns of groups of selected sites across genotypes. The x axis represents
836 synonymous rates (α), while the y axis represents non-synonymous rates (β). As
837 labelled, all sites above the $\alpha=\beta$ line positively selected, and those below are
838 negatively selected. The plane is coloured by the weight assigned to each area by the
839 FUBAR algorithm. All six plots use the same colouring scale, so they are directly
840 comparable. Genotype 1 is unusual in having a very low proportion of positively
841 selected sites. In genotypes 1 and 3 both the codon substitution model (Table 3) and
842 RELAX procedure (Table 4) estimate that ORF 2 has significantly a stronger signal of
843 positive selection.

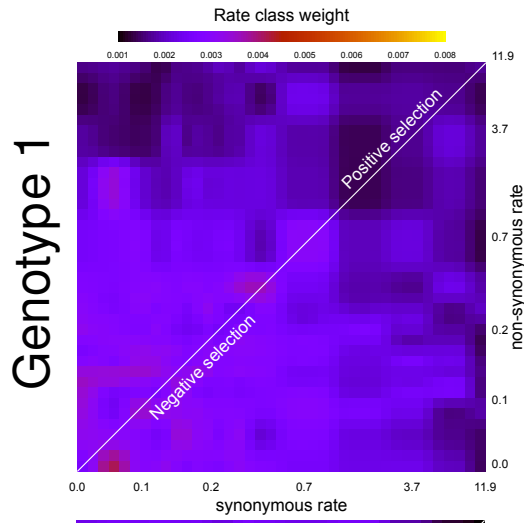
844 Figure 3. Estimates of evolutionary rates of HEV based on different genomic
845 regions. Anthropotropic genotype 1 has significantly reduced relative non-
846 synonymous evolutionary rates compared to their zoonotic counterparts across all
847 ORFs. Genotypes 3 and 4 demonstrate similar profiles, with non significant
848 differences across all ORFs. The overlap region of ORF 2 appears very similar to ORF
849 3, as they overlap extensively. Notably the non-overlap region of ORF 2 has a similar
850 evolutionary rate profile to ORF 1, with which it does not overlap at all. Asterisks
851 denote significance.

852 Figure 4. Maximum likelihood phylogenies of near-full-length sequences of HEV
853 isolated from humans and swine. Branch lengths are in expected substitutions per
854 nucleotide site estimated under the RELAX (59) general exploratory model. Swine
855 isolates are labelled using muted text, and all branches labelled as 'human' are
856 plotted using thicker lines. The k coefficients measures relaxation ($k < 1$) or
857 intensification ($k > 1$) of positive selective pressure relative to the phylogeny-wide
858 baseline (mean of k is constrained to be 1), represented by shades of grey. For G3,
859 91.5% of the bootstrapped trees supported p-value of ≤ 0.05 or less. For G4 every
860 single p-value of the bootstrapped trees supported p-value of < 0.05 .

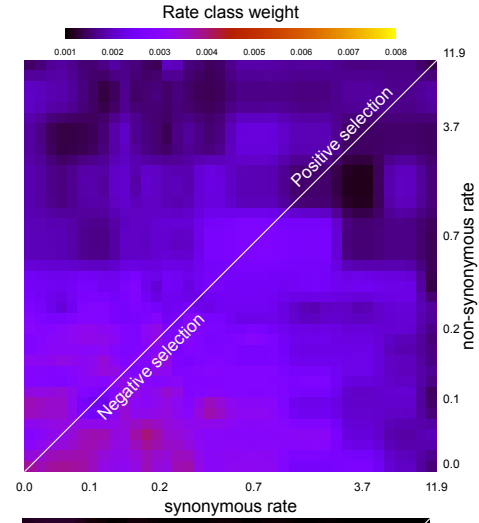
861



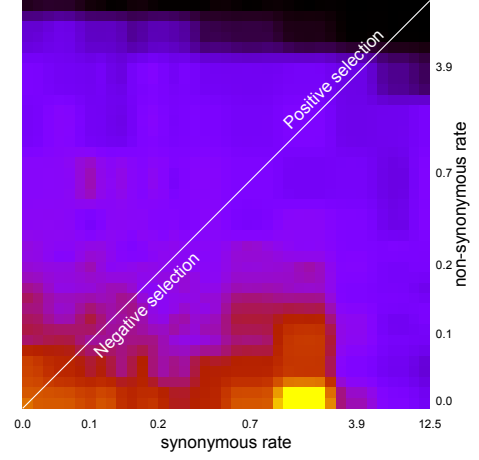
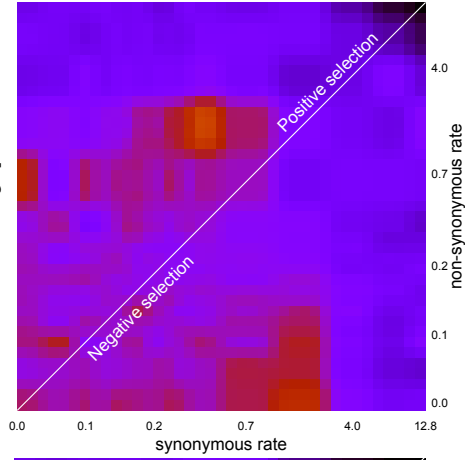
ORF2



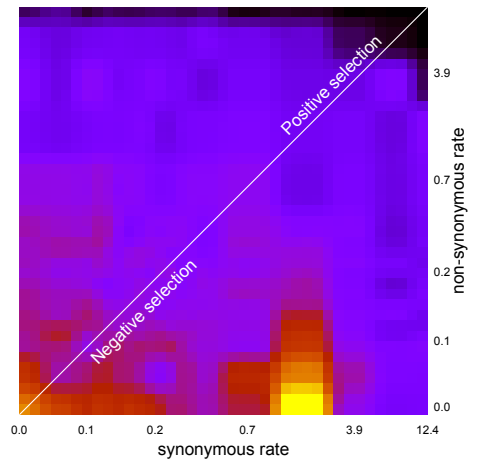
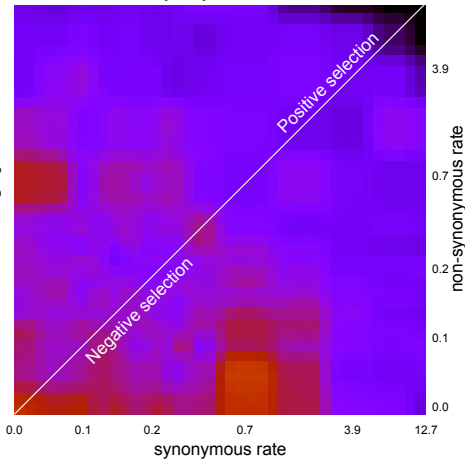
ORF3

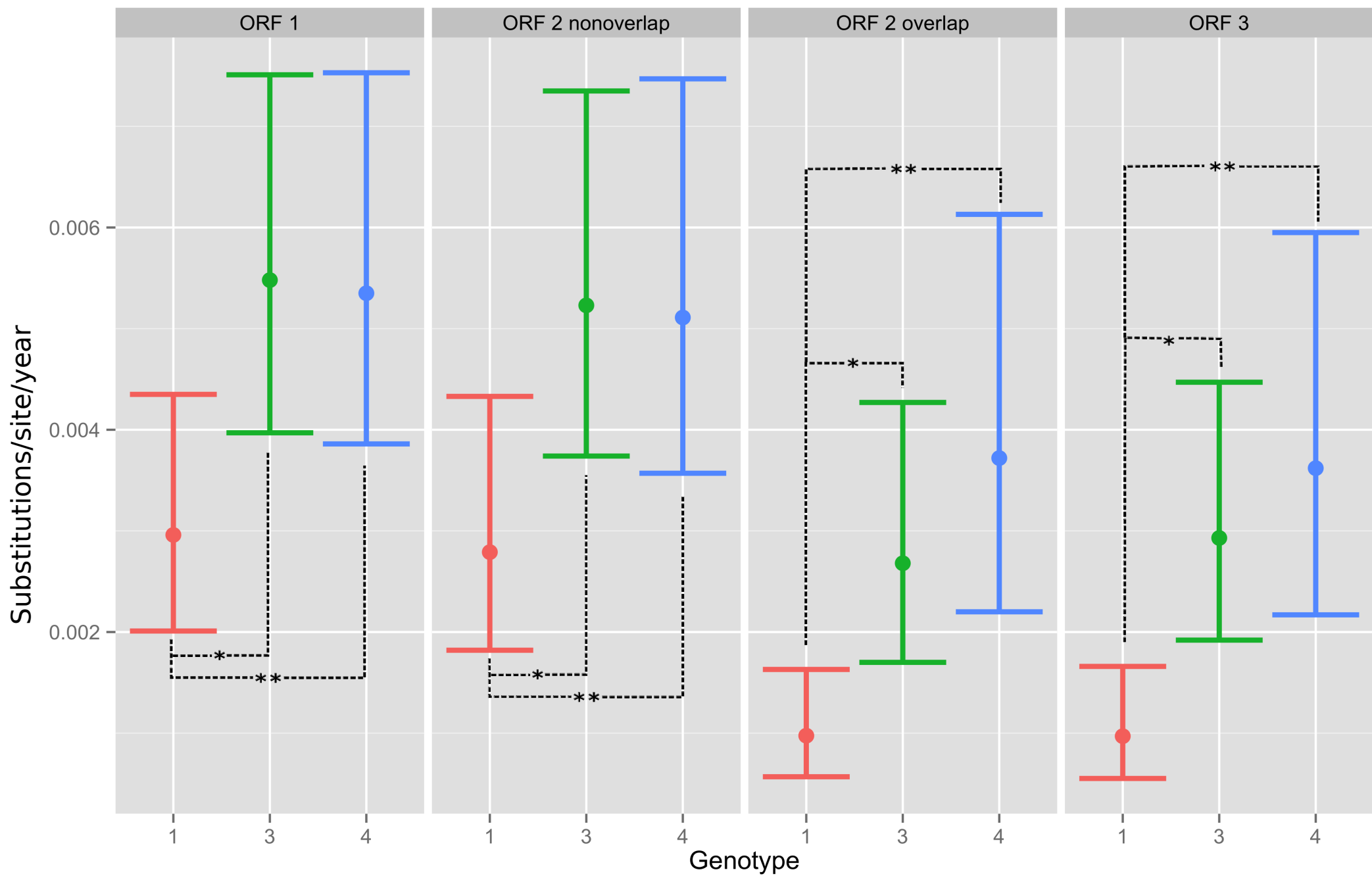


Genotype 3

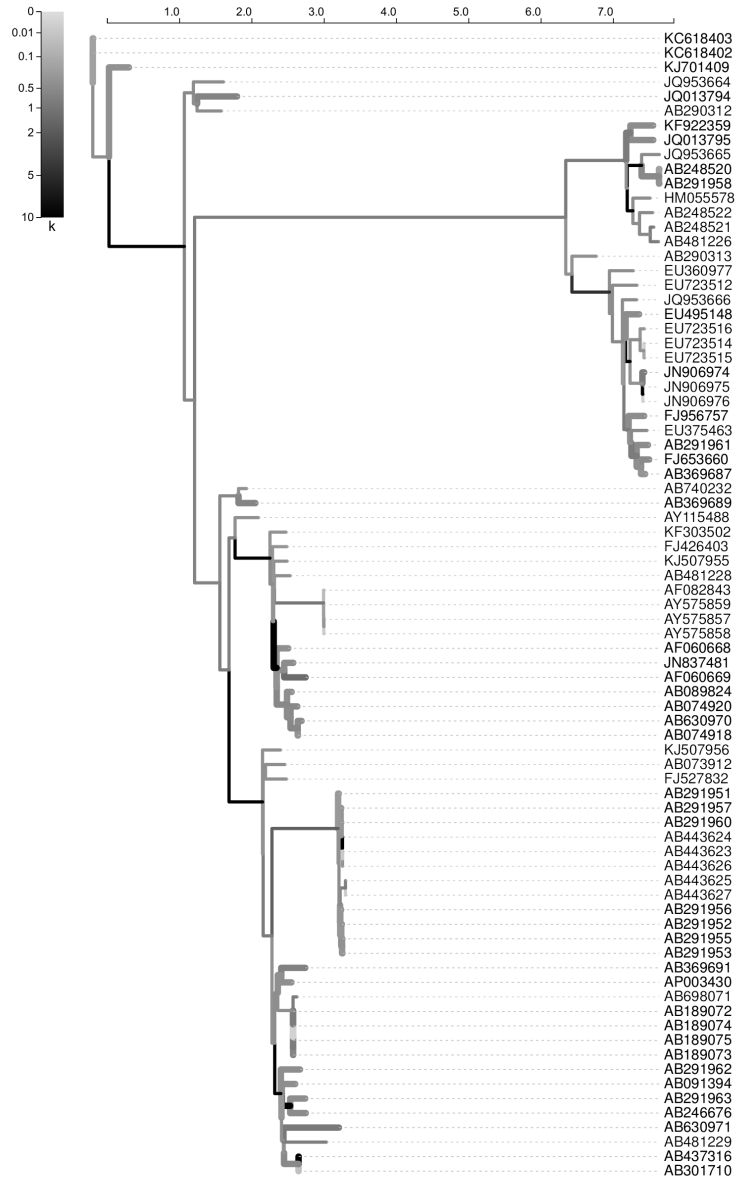


Genotype 4

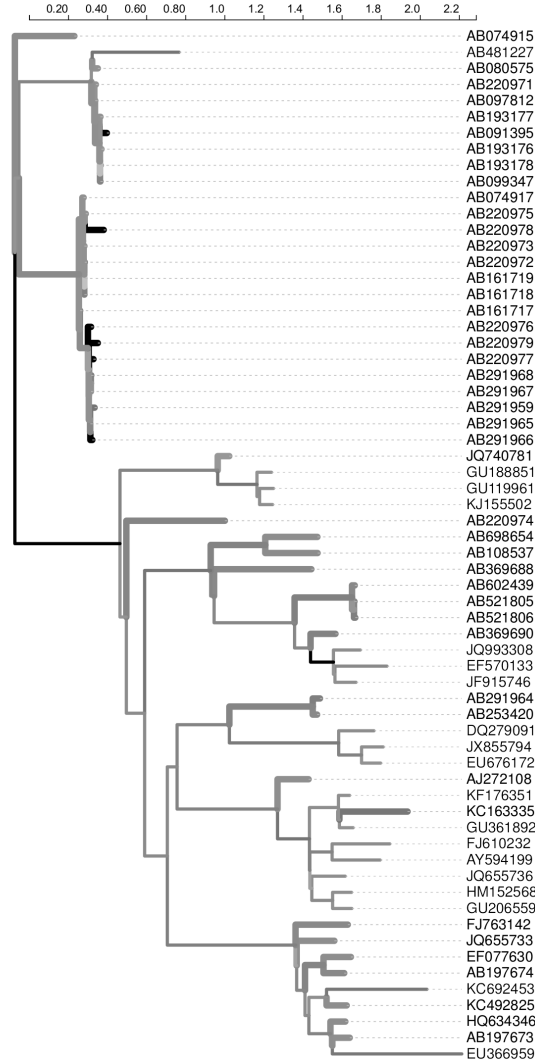




Genotype 3



Genotype 4



human isolates swine isolates