

Mechanisms of action and therapeutic efficacies of the lipophilic antimycobacterial agents, clofazimine and bedaquiline

Moloko C. Cholo ^{1*}, Maborwa T Mothiba¹, Bernard Fourie² and Ronald Anderson³

¹*Department of Immunology, Faculty of Health Sciences, University of Pretoria, Pretoria 0001, South Africa*

²*Department of Medical Microbiology, Faculty of Health Sciences, University of Pretoria, Pretoria 0001, South Africa*

³*Institute for Cellular and Molecular Medicine, Department of Immunology, Faculty of Health Sciences, University of Pretoria, Pretoria 0001, South Africa*

Correspondence : Dr M. C. Cholo
Department of Immunology
P.O Box 2034
Pretoria 0001
South Africa

Tel. : +27 12 319 2162

Fax. : +27 12 323 0732

Email : moloko.cholo@up.ac.za

Running title: Clofazimine and bedaquiline

Summary

Drug-resistant (DR)-tuberculosis (TB) is the major challenge confronting the global tuberculosis (TB) control programme, necessitating treatment with second-line anti-TB drugs, often with limited therapeutic efficacy. This scenario has resulted in the inclusion of Group 5 antibiotics in various therapeutic regimens, two of which promise to impact significantly on the outcome of the therapy of DR-TB. These are the “re-purposed” riminophenazine, clofazimine, and the recently approved diarylquinoline, bedaquiline. Although they differ structurally, both of these lipophilic agents share cationic amphiphilic properties, which enable them to target and inactivate essential ion transporters in the outer membrane of *Mycobacterium tuberculosis*. In the case of bedaquiline, the primary target is the key respiratory chain enzyme, F₁/F₀-ATPase, while clofazimine is less selective, apparently inhibiting several targets, which may underpin the extremely low level of resistance to this agent. This review is focused on similarities and differences between clofazimine and bedaquiline, specifically in respect of molecular mechanisms of antimycobacterial action, targeting of quiescent and metabolically-active organisms, therapeutic efficacy in the clinical setting of DR-TB, resistance mechanisms, pharmacodynamics, pharmacokinetics, and adverse events.

Key words: Anti-inflammatory activity; bacterial sub-populations; diarylquinolines; early bactericidal activity; F₁/F₀-ATPase; potassium transporters; multidrug-resistance; *Mycobacterium tuberculosis*; resistance mechanisms; riminophenazines.

Introduction

Tuberculosis (TB) remains a major public health problem globally, killing more people than any other infectious disease.¹ In 2013, approximately nine million active TB patients and 1.5 million TB-related deaths were reported.²⁻⁴ The two major factors underpinning this global public health crisis include the ongoing TB pandemic driven by human immunodeficiency virus (HIV) coinfection and the associated alarming increase in drug-resistant (DR)-TB, increasing the transmission of *Mycobacterium tuberculosis* (*Mtb*) and mortality from the disease.^{2,5-7} In 2013, approximately 480 000 new multidrug-resistant (MDR)-TB cases and 210 000 deaths were reported worldwide with a very large proportion of these (60%) originating from Brazil, Russia, India, China and South Africa (BRICS).^{2,8}

Unlike drug-susceptible (DS)-TB, the treatment of DR-TB is complicated, often resulting in poor treatment outcomes, which vary according to the resistance profiles of the infecting strains to the constituent drugs in the various regimens. For instance, treatment of MDR-TB has been successful in less than 44% of patients,^{9,10} in comparison to the 95% success rate attained in the case of DS-TB cases.^{2,11} In the setting of complicated MDR-TB, such as patients infected with MDR-plus-, pre-extensively drug-resistant (pre-XDR)-, XDR-, or totally drug-resistant (TDR)-TB strains of *Mtb*, the recommended regimens yield even poorer results, achieving treatment success rates of around 11%, with associated high mortality rates of about 73%.^{6,11,12} Treatment outcome of TB/HIV coinfection is also poor, with an associated mortality rate of 70%.^{8,13} Notwithstanding, the drug resistance profiles of the infecting *Mtb* strains, the outcome of chemotherapy in this setting is dependent on additional factors including the severity of the two diseases, as well as drug-drug interactions

(DDIs) between the anti-infective agents used in the treatment of the two diseases.^{2,12}

The WHO regimen recommended for the treatment of DR-TB consists predominantly of several, under-researched and highly toxic, second-line antibiotics and is administered for a minimum of 18 months.¹⁴ The progression of treatment involves a 6 – 8-month intensive phase of administration of at least four second-line drugs, which include newer fluoroquinolones (levofloxacin and moxifloxacin), an injectable agent (kanamycin), prothionamide, and cycloserine or para-aminosalicylic acid, in addition to pyrazinamide, followed by a 12 month continuation phase with at least four oral drugs.^{7,11,15,16} Depending on the clinical and bacteriological responses, treatment duration can be extended if necessary.¹⁷ However, the duration of therapy and/or the composition of the drug regimen may have to be revised due to development of drug toxicity.

Questionable efficacy and/or unacceptably high toxicity of the recommended DR-TB drug regimen prompted the WHO to formulate an alternative strategy in an attempt to overcome treatment failure. This was based on the inclusion of Group 5 antibiotics in the regimens of those DR-TB patients who experience treatment failure. Group 5 antibiotics consist of “re-purposed” older agents such as clofazimine, linezolid, amoxicillin plus clavulanate, imipenem plus cilastatin, and clarithromycin, as well as the new drugs, bedaquiline and delamanid.^{9,10,16,18-20} However, as with the drugs which comprise the recommended WHO DR-TB regimen, Group 5 antimicrobial agents also have limitations as most have incomplete information in respect of their antimicrobial efficacies against mycobacterial subpopulations, DDIs,

safety, and mechanism(s) of action.¹⁹ The availability of such data is essential in formulating new, more effective treatment regimens, which should be less amenable to development of resistance by acting on multiple targets, including those located in the cell membrane.²¹

This review is focused on two Group 5 anti-TB agents, clofazimine and bedaquiline, that promise to impact significantly on improving the efficacy and shortening the duration of therapy of DR-TB. Both clofazimine and bedaquiline are the prototypes of different classes of lipophilic, antimycobacterial agents, having predicted logP (logarithm of partition (P) coefficient in Octanol/Pwater) lipophilicity values of 7.39 and 6.37, respectively.^{22,23} Both drugs are operative at the level of the cell membrane *Mtb*, targeting the proton motive force (PMF).²⁴ In addition to comparing and contrasting the mechanisms of antimycobacterial action of clofazimine and bedaquiline, their antimicrobial spectrums, and activities against mycobacterial subpopulations, other topics covered include: i) overviews of the efficacy of recently developed DR-TB regimens containing either bedaquiline or clofazimine; ii) mechanisms of development of drug resistance; iii) effects on eukaryotic cells; and iv) the pharmacokinetic, pharmacodynamic and adverse event (AE) profiles of each agent.

Clofazimine

Background

Clofazimine is a riminophenazine antibiotic, originally developed for the treatment of TB.²⁵ Despite its impressive antimicrobial activity against *Mtb* isolates *in vitro*, clofazimine monotherapy was unsuccessful in earlier studies undertaken in higher primates and humans, while skin discolouration with associated mental disturbances, including depression, was also a deterrent to clinical application.²⁶ Poor treatment outcomes with clofazimine also coincided with the discovery of the first-line anti-TB agents, pyrazinamide and ethambutol in 1952 and 1961, respectively, both of which showed better therapeutic efficacy and fewer side-effects, surpassing clofazimine as preferred agents for the treatment of TB. Subsequently, rifampicin (1968) was discovered, which contributed significantly to shortened duration of treatment. These three agents, together with streptomycin and isoniazid, which were discovered prior to clofazimine in 1943 and 1945, respectively, were combined to form the earlier and current regimens used in the treatment of TB for the past 50 years. The efficacy of these regimens resulted in loss of interest in clofazimine as an anti-TB agent. However in 1981, clofazimine was recommended by the WHO for inclusion as a component of the multi-drug treatment of leprosy due to its beneficial combination of antimicrobial and anti-inflammatory properties.¹⁸

In the light of the growing XDR-TB epidemic, there has been a re-emergence of interest in clofazimine, which has become an important component of newer treatment regimens.^{27,28} One of these, referred to as the 9-month short-course

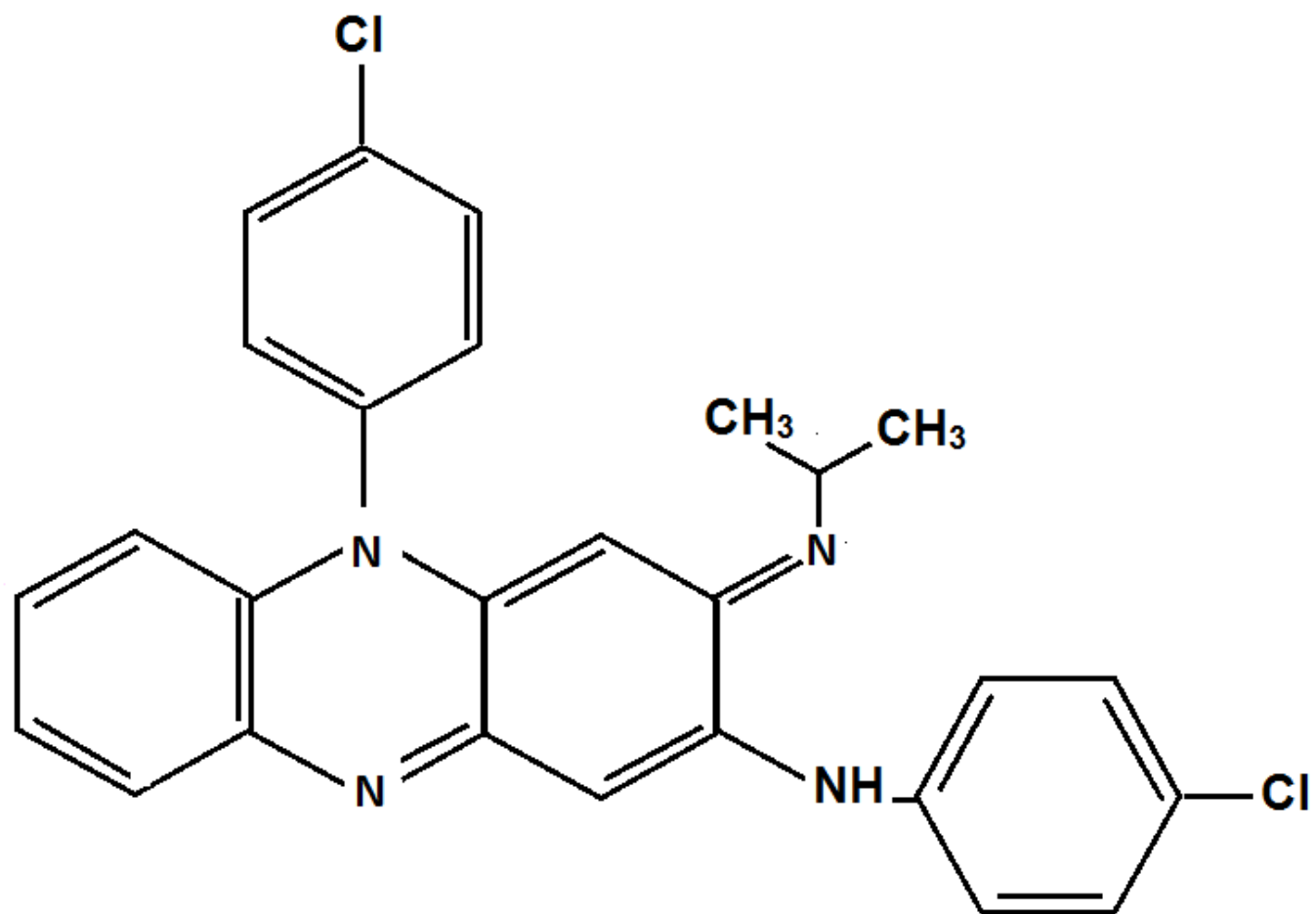
regimen, based on the outcome of a clinical trial conducted in Bangladesh, was found to be efficacious and is currently being evaluated for its possible application as a future standard regimen for the treatment of DR-TB patients.^{27,29,30}

Chemical structure of clofazimine

Clofazimine [3-(p-chloroanilino)-10-(p-chlorophenyl)-2,10-dihydro-2-(isopropylimino) phenazine] has a molecular formula of $C_{27}H_{22}Cl_2N_4$ and a molecular weight of 473.14 Daltons (Figure 1a).^{26,31} The key structural features of the riminophenazines are the phenazine nucleus, with an alkylimino (R-imino) group at the C-2 and phenyl substituents at the C-3 and N-10 positions of the phenazine nucleus.³² The basic nitrogen atom of the isopropylimino group at position C-2 of clofazimine contributes to the cationic amphiphilic properties of the molecule. Cationic amphiphilic drugs contain both a hydrophobic domain in the aromatic ring system and a hydrophilic domain in the ionisable amine functional group.^{33,34,35}

Modifications of the substituents at positions C-2, C-3 and N-10 of the clofazimine molecule have resulted in analogues that have demonstrated alterations in antimicrobial activity and pharmacological properties. The most promising of these are the tetramethylpiperidyl-substituted phenazines in which the isopropyl group at position C-2 of the phenazine nucleus is replaced by the tetramethylpiperidyl group,^{26,36-38} or by a methoxypyridylamino group at C-3.^{39,40} However, none of these, to our knowledge, are currently in clinical development.

(a)



(b)

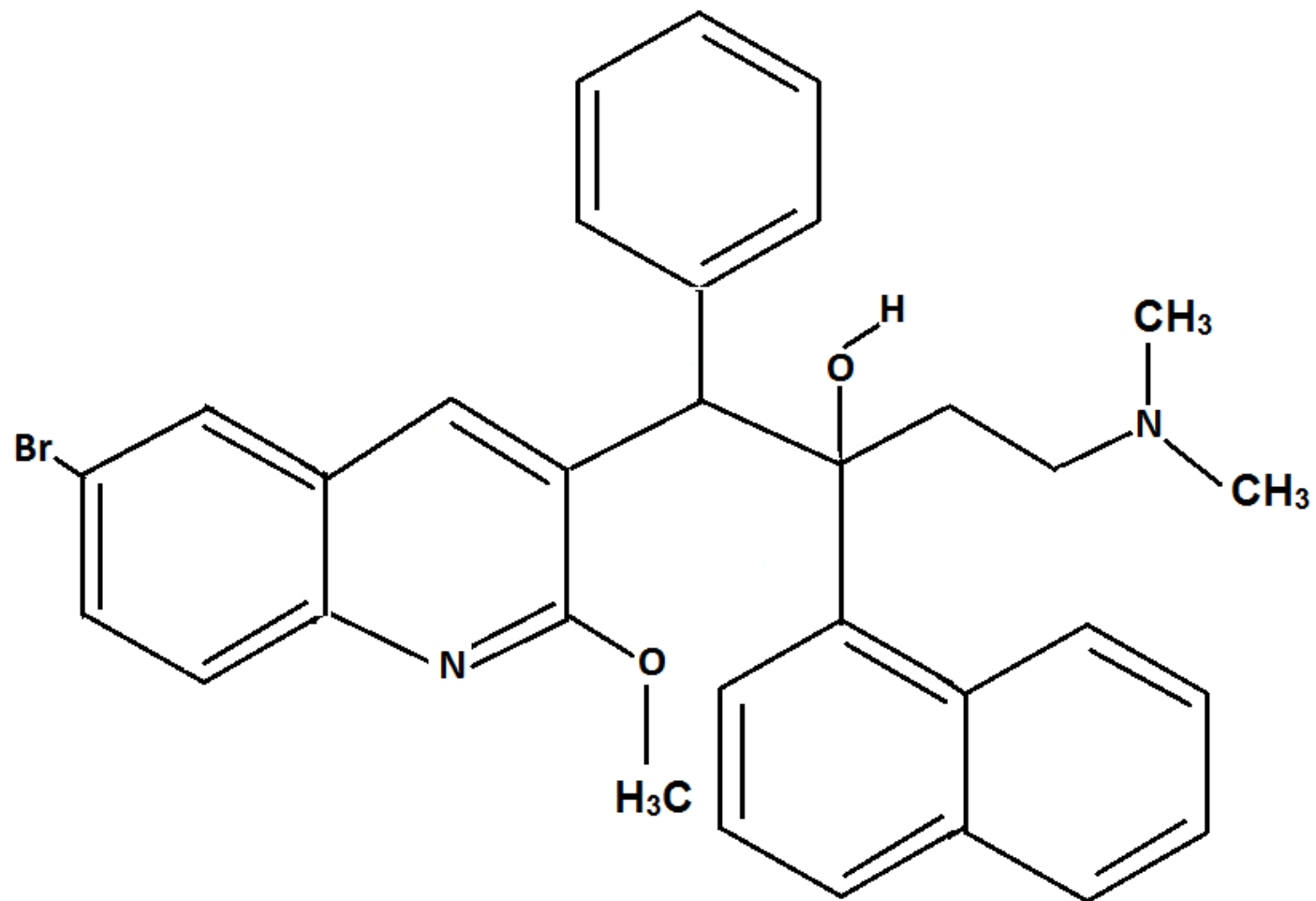


Figure 1. Molecular structures and systematic names of (a) clofazimine ([3-(p-chloroanilino)-10-(p-chlorophenyl)-2,10-dihydro-2-(isopropylimino) phenazine]),³² and (b) bedaquiline ([1-(6-bromo-2-methoxy-quinolin-3-yl)-4-dimethylamino-2-naphthalen-1-yl-1-phenyl-butan-2-ol]).¹³⁰

Various formulations of clofazimine, oral, intravenous and inhaled, have also been evaluated in the experimental setting, but none has yet undergone clinical evaluation.^{32,41,42}

Antimicrobial activity of clofazimine

Clofazimine has a broad spectrum of antimicrobial activity, acting against many types of microorganisms, including bacteria, parasites and fungi. Among the bacteria, clofazimine is active against Gram-positive organisms, while Gram-negative organisms are uniformly resistant.^{32,43-46} Parasites that are susceptible to clofazimine include *Plasmodium falciparum*,⁴⁷ *Leishmania donovani*,^{48,49} *Trypanosoma cruzi*,⁵⁰ *Babesia*, *Theileria*⁵¹ and *Schistosoma* species^{52,53} while the yeast, *Candida albicans*, is also susceptible.⁵³

Clofazimine has demonstrated impressive activity against various mycobacterial species,^{54,55} including rapidly- (*M. abscessus*, *M. fortuitum* and *M. smegmatis*) and slow-growing bacilli [*Mtb*, *M. avium intracellulare* complex (MAC) and *M. leprae*].^{54,56-59} In addition, clofazimine acts synergistically with other antimicrobial agents, such as amikacin and clarithromycin, against several mycobacterial species, including *M. avium* and *M. abscessus in vitro*.⁵⁷ In the case of *Mtb*, clofazimine, at low concentrations, is active against both DS- and DR-TB strains *in vitro* and *in vivo*, exhibiting differential activities against *Mtb* populations according to the stage of growth.^{32,37,60} Clofazimine has demonstrated impressive bacteriostatic activity, but poor bactericidal activity, against actively-replicating bacilli, *in vitro* and *in vivo*, the former achievable at minimum inhibitory concentration (MIC) values ranging

from 0.06 - 2 mg/L.^{32,61,62} Clofazimine also acts synergistically in combination with the primary anti-TB agents, ethambutol and pyrazinamide,^{63,64} as well as with the second-line agents, linezolid,⁶⁵ bedaquiline and moxifloxacin, against *Mtb* isolates.⁶⁶ Clofazimine monotherapy has been reported to reduce the bacterial load in BALB/c mice infected with the H37Rv strain of *Mtb*, reducing it from 6log₁₀ to 4log₁₀ within 8 weeks.⁶⁷ In another murine model of experimental chemotherapy, combining clofazimine with the primary anti-TB drugs, rifampicin, isoniazid and pyrazinamide, shortened the treatment time to achieve cure from 6 months to 4 months in comparison with mice treated with a primary anti-TB drug regimen alone (rifampicin, isoniazid and pyrazinamide).⁶⁸ In humans, however clofazimine fails to kill this actively-growing microbial population during the first 14 days of therapy, which is attributable to its lack of early bactericidal activity (EBA). *In vitro*, clofazimine also failed to demonstrate EBA during the first 10 days of treatment when used at concentrations of 0.3 - 2.5 mg/L. However, an EBA was achievable when higher concentrations of clofazimine (5 - 20 mg/L) were used.⁶²

Among the different *Mtb* subpopulations, clofazimine has demonstrated highest activity against slow-replicating bacilli. The MIC and minimum bactericidal concentration (MBC) values against this bacterial subpopulation *in vitro* were reported to be 0.06 and 0.3 mg/L, respectively,⁶² while reducing the slow-replicating *Mtb* bacterial load in C3HeB/FeJ infected mice by 5.8log₁₀ cfu/mL.⁶⁷ Slow-replicating bacilli are responsible for the formation of biofilm *in vitro* and granuloma *in vivo*, both of which are attenuated by clofazimine, possibly facilitating exposure of the bacteria to other antimicrobial agents.^{62,67}

In the case of non-replicating bacilli, only those cultured in an aerated, streptomycin-starved (SS18b) model of dormancy *in vitro* were found to be susceptible to the lethal activity of clofazimine,⁶⁹ while those residing in non-aerobic, enclosed environments, such as preformed mycobacterial biofilm cultures were not affected.⁶² Likewise, in a C3HeB/FeJ murine model of experimental TB, organisms contained in the matured granuloma lesions in the lungs were only slightly reduced by 1.6log₁₀ following treatment with clofazimine.⁶⁷ These observations seemingly support the requirements for the availability of oxygen and/or accessibility of the bacteria to the antibiotic to achieve a mycobactericidal effect on dormant bacilli.^{62,67,70,71} The preferential microbicidal action of clofazimine on non-replicating bacilli, may explain the lack of EBA, while contributing to shortening of the duration of chemotherapy via late bactericidal activity (LBA).

Mechanisms of action of clofazimine

Effect of clofazimine on microbial cells

The Irish group which discovered clofazimine suggested that the antimycobacterial activity of this agent was attributable to two unusual properties, these being its high lipophilicity, enabling efficient transmembrane penetration, together with a redox potential of -0.18V at pH7,²⁵ favouring intracellular redox cycling.⁴³ Intracellular oxidation of reduced clofazimine was proposed to result in the generation of antimicrobial reactive oxygen species (ROS).²⁵ However, convincing evidence for the existence of such a mechanism was provided only 50 years later by Yano *et al.*⁷⁰ These authors, using isolated membrane fractions from *M. smegmatis*,

demonstrated that clofazimine appears to compete for electrons with menaquinone, the substrate for type 2 nicotinic adenine dinucleotide hydrogen (NADH):quinone oxidoreductase, which is the initial event in the mycobacterial respiratory chain.⁷⁰ Reduced clofazimine generated by this mechanism was proposed to undergo spontaneous oxidation, resulting in the generation of antimicrobial ROS such as superoxide and hydrogen peroxide.^{70,72} This putative mechanism of antimicrobial activity is supported by a more recent study which reported that supplementation of the bacterial growth medium with high concentrations of menaquinone antagonized the antimycobacterial activity of clofazimine.⁷³ In addition, and seemingly consistent with an inhibitory effect on bacterial respiration, selective inactivation of the cytochrome *bd*-type quinol oxidase of the branched respiratory chain operative in mycobacteria was found to increase the susceptibility of *M. smegmatis* to clofazimine.⁷⁴ The authors speculated that the protective action of cytochrome *bd* is achieved via neutralization or inhibition of clofazimine-generated ROS.⁷⁴

Although redox cycling as described by Yano *et al.*⁷⁰ appears to contribute to the antimycobacterial activity of clofazimine, others believe that this is unlikely to be the only mechanism, favouring the existence of a multifaceted mechanism of antimicrobial activity. If correct, this may explain the remarkably low level of resistance to clofazimine in both the clinical and experimental settings. Evidence in support of this contention originates from several sources. Firstly, although clofazimine is assumed to compete with menaquinone for electrons generated via the activity of type 2 NADH:quinone oxidoreductase, the existence of such a mechanism remains to be conclusively established.⁷³ In addition, menaquinone possesses secondary membrane-stabilizing properties,⁷⁵ which may counteract the

disruptive effect of clofazimine on the mycobacterial membrane. Secondly, in an earlier study, Van Rensburg *et al.*⁴³ reported that exposure of a single strain each of *Staphylococcus aureus* and *Streptococcus pyogenes* to clofazimine under anaerobic conditions actually increased the susceptibility of these microorganisms to clofazimine.⁴³ More recently, Lu *et al.*⁷⁶ using a low oxygen recovery assay (LORA), reported that exposure of *M. tuberculosis* to clofazimine at very low oxygen concentrations (<0.16%) resulted in only moderate loss of antimycobacterial activity of the antibiotic.⁷⁶ These authors proposed that different clofazimine-mediated antimycobacterial mechanisms may be operative under different environmental conditions.⁷⁶ Thirdly, the susceptibility of Gram-negative bacteria to the antimicrobial actions of ROS is not entirely consistent with their relative lack of susceptibility to clofazimine.^{32,43}

Additional mechanisms of antimicrobial activity, unrelated to redox cycling, are likely to result from the cationic amphiphilic, membrane disruptive properties of clofazimine alluded to above. Mycobacteria and Gram-positive bacteria are particularly susceptible to the membrane disruptive actions of cationic amphiphiles and other types of membrane disruptive agent.^{32,43,46,77-79} In this context, it is noteworthy that ion-transporting adenosine triphosphatases (ATPases), are particularly vulnerable to inhibition by cationic amphiphiles, which appear to induce conformational changes in protein molecular structure and loss of function.⁸⁰⁻⁸³ We have previously reported that interference with cation uptake, specifically potassium (K⁺), is one of the earliest occurring changes during exposure of *Mtb* to clofazimine at MIC concentrations, and is followed by depletion of ATP and inhibition of growth.³² Although we have previously proposed that selective targeting of mycobacterial K⁺

active transporters may underpin the antimycobacterial activity of clofazimine, a non-specific membrane disruptive mechanism, resulting in loss of activity of several different ion transporters, appears more likely.^{24,32,37,61}

Taken together, the currently available evidence is consistent with the existence of at least two mechanisms of clofazimine-mediated antimycobacterial activity *viz.* intracellular redox cycling and membrane disruption. As mentioned above, the relative contributions of these mechanisms may vary according to environmental conditions and may also explain the very low level of resistance to clofazimine encountered in the therapeutic setting. It is, however, noteworthy that the membrane disruptive antimycobacterial mechanism related to the cationic amphiphilic properties of clofazimine may not be effective in regions of the granuloma which are slightly alkaline, thereby neutralising the positive charge on the molecule.^{34,35}

Effect of clofazimine on eukaryotic cells

The effects of clofazimine on eukaryotic cells have been reviewed by us previously and are considered only briefly here.³² Not surprisingly, these include inhibition of the plasma membrane K⁺ transporters, sodium (Na⁺), K⁺-ATPase,⁸⁴ and the Kv1.3 potassium channel,^{85,86} both of which are electrogenic and essential for the activation and proliferation of T-lymphocytes. Clofazimine-mediated interference with T-cell activation via inhibition of transmembrane K⁺ fluxes is likely to underpin the reported benefit of this agent in the treatment of autoimmune and other chronic inflammatory disorders, as well as in controlling immune-mediated tissue damage during mycobacterial infection.^{32,84-86} Additional mechanisms of clofazimine-mediated

immunosuppressive activity include increased production of both prostaglandin E₂, and the interleukin (IL)-1 receptor antagonist by immune, inflammatory and other cell types.^{32,87-89}

As proposed previously, the immunosuppressive properties of clofazimine may be either beneficial or detrimental depending on the timing of administration of this agent.³² If administered at the outset of therapy, immunosuppressive activity may compromise the antimycobacterial efficacy of clofazimine, possibly contributing to the lack of EBA, as well as that of other agents in the drug regimen. Administration later in the course of therapy may contribute to the eradication of slow-growing persisters in the setting of controlled recovery of *Mtb*-specific immune reactivity.

Clinical efficacy of clofazimine

As with other anti-TB therapeutic agents, clofazimine is used in multidrug regimens to prevent the emergence of drug resistance. However, due to lack of a standardised regimen for clofazimine, several different regimens have been evaluated for efficacy in the treatment of DR-TB, most frequently at daily dosages ranging from 50-100 mg, with few reaching to 300 mg.^{17,18,90}

One of the most effective clofazimine-containing regimens evaluated to date has been the 9-month short-course regimen, based on a clinical trial conducted in Bangladesh from 1997 - 2007.²⁹ This regimen consisted of gatifloxacin, clofazimine, ethambutol and pyrazinamide throughout the treatment period of nine months, supplemented with prothionamide, kanamycin and high-dose isoniazid during the 4-

month intensive phase, followed by a continuation phase of five months with gatifloxacin, ethambutol, pyrazinamide and clofazimine. This regimen administered to treatment-naive MDR-TB patients resulted in a reduction in the duration of treatment from the 20-month period when using the WHO-recommended regimen to nine months. It was also associated with impressive relapse-free cure rates in patients who were followed for 24 months.²⁹ The efficacy of this regimen was confirmed in a follow-up clinical trial undertaken in Bangladesh⁹¹ and has since been adopted by many low-income countries, including Cameroon⁹² and Niger.⁹³ These studies, summarised in Table 1, have also demonstrated impressive treatment outcomes, with success rates ranging from 84 - 89%.⁹¹ A number of patients recruited to some of these studies harboured isolates resistant to at least six anti-TB agents, and also included those who were coinfecting with HIV. The outcomes of therapy reported from these trials appear to confirm the efficacy of the 9-month short-course regimen, irrespective of disease severity and socio-economic status of the patients. This regimen is currently being evaluated in several other West African countries and preliminary data has demonstrated sputum-culture conversion within four months of the intensive phase of therapy in 96% of patients.³⁰

The shortening of treatment associated with the Bangladesh-based regimen may be related to targeting of slow/non-replicating bacterial populations by clofazimine as described in experimental settings both *in vitro* and *in vivo*.^{60,62,67} The sustained relapse-free cure rates have been attributed to the prolonged half-life (70 days), high tissue accumulation,^{60,94} and/ or the long post-antibiotic effect (PAE) of clofazimine. The latter effect was recently demonstrated in the experimental setting when *Mtb*-infected BALB/c mice treated with clofazimine alone and as a constituent

Table 1. 9-month clofazimine-based short-course treatment regimen for DR-TB patients in different countries

Study name^a	1	2	3	4	5
Study site	Bangladesh	Bangladesh	Niger	Cameroon	STREAM stage 1 ^b
Period (years)	1997-2007 (10)	2005-2011 (6)	2008-2010 (2)	2008-2011 (3)	2012-2015 (3)
No of patients	206	515	65	150	424
Treatment regimen	4 (KCGEHZP)	4 (KCGEHZP)	4 (KCGEHZP)	4 (KCGEHZP)	4 (KCMEHZP)
Months (drugs)	5 (GEZC)	5 (GEZC)	5 (GEZC)	8 (GEZCP)	5 (MEZC)
Treatment duration (months)	9-12	9-12	12	12-13.6	9-11
Follow-up period (months)	24	24	24	24	27
Successful treatment outcome	182/206 (87.9%)	435/515 (84.4%)	58/65 (89%)	134/150 (89%)	NYA
Relapse-free cure rate	90%	82.3%	49/49 (100%)	100%	NYA
Treatment failure	1/206 (0.5%)	7 (1.4%)	1/65 (1.5%)	1/150 (0.7%)	NYA
Default	12 (5.8%)	40/515 (7.8%)	1/65 (1.5%)	5/150 (3.3%)	NYA
Death	11 (5.3%)	29/515 (5.6%)	6 (9.2%)	10/150 (5%)	NYA
FQ/KM resistance	20/206 (10%)	51/515 (10%)	1/65 (1.5%)	0	NYA

HIV-positive	0	0	1/58 (1.7%)	30/150 (20%)	NYA
Side effects	Vomiting (21.4%) Hearing impairment (6.3%) Ataxia (3.9%) Dysglycemia (3.9%)	Vomiting (21.6%)	Vomiting (26%) Hearing impairment (20%) Hyperglycaemia (9.2%) Gastritis (7.7%) Arthralgia (6.2%) Peripheral neuropathy (4.6%) Skin pigmentation (3.1%)	Hearing loss (43%) Vomiting Retrobulbar neuritis Transaminases (0.7%)	NYA

K, kanamycin; C, clofazimine; G, gatifloxacin; E, ethambutol; H, isoniazid; Z, pyrazinamide; P, prothionamide; M, moxifloxacin; NYA, not yet available.

^aStudies numbered according to authors' names; 1, 2, 3, 4 and 5 represent studies conducted by van Deun *et al.*, 2010²⁹; Aung *et al.*, 2014⁹¹; Piubello *et al.*, 2014⁹³; Kuaban *et al.*, 2015⁹² and Nunn *et al.*, 2015²⁷, respectively.

^bDifferent sites included in Stream stage 1 trial were Ethiopia, South Africa, Vietnam and Mongolia.

of a multi-drug regimen showed a delay in bacterial regrowth six weeks after treatment cessation, while those treated with an alternative clofazimine-free regimen showed regrowth immediately after treatment cessation.⁹⁵

Despite its success, several caveats exist in relation to widespread implementation of the 9-month short-course regimen. Most importantly, the efficacy of this regimen may be affected by the drug resistance profiles of the isolates. In Bangladesh, patients with *Mtb* strains with high-level fluoroquinolone (gatifloxacin) and pyrazinamide resistance had a poor treatment outcome.^{28,91} In this setting, cessation of the 9-month short-course regimen and replacement with a bedaquiline-containing regimen is recommended.⁹⁶

In addition to the ongoing West Africa study, the efficacy of the Bangladesh regimen is currently being evaluated in the STREAM trial, encompassing several countries including South Africa, Ethiopia, Mongolia and Vietnam, comparing its efficacy and safety with that of the WHO regimen as a necessary prerequisite prior to possible recommendation as a standard regimen for the treatment of MDR-TB patients. In this study, 424 patients, including those who are HIV-positive, have been enrolled during the period 2012 - 2015, while patients infected with fluoroquinolone- and injectable agent-resistant DR-TB isolates have been excluded. Modifications to the 9-month treatment regimen include replacement of gatifloxacin with moxifloxacin due to the association of the former with dysglycaemia.²⁸ Patients are being monitored for 27 months post-treatment. The trial is ongoing and the expected completion date is early 2018.^{27,28}

Based on the successful implementation of the STREAM trial, also referred to as STREAM stage 1, a stage 2 trial was planned and will be initiated during 2016 continuing for three years thereafter, with the results expected in 2021.²⁸ In this trial, the two major objectives are the formulation of: i) a less toxic fully oral 9-month regimen; and ii) a shortened 6-month regimen. With respect to the first objective, a fully oral 9-month regimen consisting of isoniazid, prothionamide, bedaquiline, levofloxacin, ethambutol, clofazimine and pyrazinamide during a 4-month intensive phase, and bedaquiline, levofloxacin, ethambutol, clofazimine and pyrazinamide during a 5-month continuation phase is under investigation. In this fully oral 9-month regimen, bedaquiline replaces the injectable kanamycin, while in both regimens levofloxacin is used instead of moxifloxacin to reduce the risk of QT prolongation that occurs during coadministration of bedaquiline and moxifloxacin. In the case of the second objective, a 6-month regimen consisting of bedaquiline, clofazimine, pyrazinamide, levofloxacin, isoniazid and kanamycin is administered during a 2-month intensive phase, followed by bedaquiline, levofloxacin, clofazimine and pyrazinamide during a 4-month continuation phase. On a cautionary note, however, several investigators have expressed concern about coadministration of clofazimine and bedaquiline, due to shared efflux pump-based mechanisms of drug resistance,^{6,97} high risk for increased QT prolongation^{98,99} and a possible increased occurrence of DDIs due to metabolism of both antibiotics by CYP3A4,^{2,100,101} necessitating high-level vigilance.

Other clofazimine-containing regimens, although resulting in improved treatment outcomes, have been less successful than the 9-month short-course regimen.^{29,102,103} These trials were undertaken in Benin, South Africa, Ukraine, Brazil

and Sri Lanka with reported cure rates ranging from 60 - 66%. Interestingly and importantly, comparable successful treatment outcomes of MDR- (65%) and XDR-TB (66%) cases were reported,^{90,104} illustrating that the efficacy of clofazimine is independent of the resistance of *Mtb* to the other antimicrobial agents in the regimens, especially the primary anti-TB agents. In most of these trials treatment duration ranged from 12 - 18 months.^{17,104}

Inclusion of clofazimine in the DR-TB drug regimens of TB/HIV-coinfected patients has also been reported to result in improved treatment success rates, increasing from 28.6% to 50%.^{13,17,18} However, treatment success in these patients is affected by the DDI effect of the anti-TB and antiretroviral (ARV) agents.¹⁰⁰

Mechanisms of resistance to clofazimine

Currently, no primary clofazimine-resistant *Mtb* clinical isolate, has been described, probably due to an extremely low mutation rate, necessitating exposure at a high bacterial density in a clinical lesion or culture ($1/10^{26}$ cfu/mL) for selection of resistance traits.^{6,105-108} Alternatively, the existence of multiple targets may underpin the low level of development of resistance to clofazimine.^{24,32}

In the absence of resistant clinical isolates, the mechanism of clofazimine resistance has been investigated using clofazimine-resistant mutant strains developed *in vitro*. Ninety-seven percent of these laboratory mutant strains had mutations at the *rv0678* gene, encoding the Rv0678 protein drug efflux pump.^{6,107,109} The Rv0678 protein is the transcriptional regulator, which represses expression of

mmpS5-mmpL5, the gene encoding the multi-substrate MmpS5-MmpL5 efflux pump.^{99,107} Interestingly, the *rv0678-mmpS5-mmpL5* locus is absent in *M. leprae*, which is also highly susceptible to clofazimine, and for which no clofazimine-resistant mutant has been isolated to date.¹⁰⁷ Mutations at the *rv0678* gene also lead to cross-resistance to bedaquiline in *Mtb* isolates.^{6,97} Although other genes associated with clofazimine resistance in *Mtb* (*rv1979c* and *rv2535c*) have been identified, their mechanisms of resistance have not been determined.¹⁰⁹

Pharmacokinetic and pharmacodynamic properties of clofazimine

Clofazimine is highly lipophilic leading to high accumulation in fat-tissues and relatively low serum concentrations (0.7 - 1 mg/L).^{32,39,94,104} The fat-tissues include macrophage-rich organs such as the lungs, livers, spleen, brain¹¹⁰ and the bone marrow.¹¹¹⁻¹¹³ As mentioned above, the drug has a long half-life of approximately 70 days, which contributes to skin discolouration, its most frequent AE.⁷⁵

During its long-term tissue accumulation, clofazimine undergoes xenobiotic sequestration resulting in the formation of crystal-like drug inclusions (CLDI) in the cytoplasm of tissue macrophages.¹¹⁴⁻¹¹⁷ These bodies are formed through an intracellular chloride transport mechanism within the cells and are composed of several layers 5-15 nanometers in thickness.^{115,118} Intracellularly, they do not destabilise mitochondria, neither do they induce oxidative damage as shown *in vitro* cultures.^{111,117}

In spite of its high lipophilicity, clofazimine is unable to penetrate and accumulate in caseous granulomas. Examination of granuloma lesions in patients, using matrix-assisted laser desorption/ionization (MALDI) imaging mass spectrometry has shown a strong accumulation of the drug in the highly cellular peripheral zone of the granuloma, consisting of macrophages, epithelioid macrophages, and lymphocytes, to levels considerably higher than in the enclosed necrotic core. Due to its high lipophilicity, it is internalised in the macrophages through endocytosis and lysosomal trapping, but fails to diffuse passively through the aqueous necrotic region.⁹⁴ The poor accumulation of clofazimine in this region of the granuloma may explain its failure to act on the non-replicating bacilli in the granuloma core of C3HeB/FeJ mice as described above.^{32,94}

Administration of clofazimine to DR-TB patients has been associated with occasional, usually manageable, AEs, with frequencies comparable to those of the standard first-line anti-TB agents.⁹⁰ The most common are transient discolouration of the skin and mucous membranes, and gastrointestinal tract (GIT) discomfort,^{32,90} such as vomiting and nausea.^{18,55,93,102} While skin discolouration can lead to mental disturbance, the GIT AEs may be, albeit infrequently, severe and occasionally fatal.³⁰ In a meta-analysis of cohort studies involving clofazimine therapy, the pooled proportion of all of these AEs attributable to clofazimine has been shown to be 21.9%, although those that required withdrawal or discontinuation was 0.1% and the frequency of all AEs was 5.1%.⁹⁰ In the study by Xu *et al.*¹⁰² 39 of 44 patients (87%) were reported to have experienced AEs after starting clofazimine, predominantly skin discolouration.⁹⁴ It has, however, been noted that administration of clofazimine in the 9-month short-course regimen, was associated with a relatively low frequency (3%)

of skin pigmentation (Table 1). Clofazimine therapy has also been reported to be associated with an increased QT interval.⁹⁹

Mortality associated with clofazimine therapy is uncommon.^{29,55,68} In several studies, mortality rates ranged from 1 - 63.4%, although none of the deaths was apparently attributable to clofazimine use,¹⁰⁴ but rather to respiratory failure complications, haemoptysis and cerebral malaria.

Bedaquiline

Background

Bedaquiline belongs to a new class of antimicrobial agents known as the diarylquinolines.^{105,119,120} It is the first drug that has been approved for the treatment of TB in over 40 years by the US FDA. Approval was based on the outcome of two phase IIb studies demonstrating improved efficacy in the clinical setting of DR-TB with cure rates of 62% and 44% in patients who received the bedaquiline/background regimen (BR) and placebo/BR, respectively, with corresponding reductions in the duration of therapy (83 vs 125 days). The preferred BR consisted of five antibiotics, ethionamide, pyrazinamide, ofloxacin, kanamycin and cycloserine.¹²¹

Despite the improved therapeutic efficacy of bedaquiline-containing regimens, safety issues remain a concern due to incomplete safety data based only on phase II clinical trials in the absence of phase III and IV trial data.^{12,119,122,123} Of concern, the

phase II studies revealed more deaths and AEs in the bedaquiline/BR-treated group than in the placebo/BR group. These safety issues led the WHO to recommend only conditional use of bedaquiline, limited to patients with pulmonary MDR-TB for whom no suitable alternatives are available.^{10,20,124} The WHO has also emphasized the necessity for timely completion of phase III and phase IV clinical trials.¹²⁵

Nevertheless, despite limited availability of safety data, usage of bedaquiline in MDR-TB chemotherapy through compassionate use programmes has been beneficial, resulting in the survival of many MDR-TB patients with more than 1258 patients worldwide having experienced clinical benefit as of October 2015.^{123,126-129} In addition, the US Agency for International Development has decided to make bedaquiline available free-of-charge to >100 global fund-eligible countries for a 4-year period.^{12,99,126}

Chemical structure of bedaquiline

Bedaquiline [1-(6-bromo-2-methoxy-quinolin-3-yl)-4-dimethylamino-2-naphthalen-1-yl-1-phenyl-butan-2-ol] has a molecular formula of $C_{32}H_{31}BrN_2O_2$ and a molecular weight of 555.51 Daltons (Figure 1b).¹³⁰ Its alternative names are R207910 and TMC207.¹³⁰ Bedaquiline contains planar hydrophobic moieties and hydrogen-bonding acceptor or donor groups.¹³¹ The hydrophobic moieties, which include hydroxyl and N,N dimethyl (-N(CH₃)₂) groups, play an important role in binding to the mycobacterial target F₁/F₀-ATP synthase, interacting with amino acid residues Arg-186 and Glu-61 at the A and C subunits, respectively, while the hydrogen bonding provides stability.^{5,132} The diarylquinoline ring has also been shown to play a role in

the anti-TB activity of bedaquiline, although it is not indispensable for this activity. Other structural moieties that play a role in anti-TB activity include the hydroxyl group, the side chain with the N,N-dimethyl amino terminus and/or the naphthalene moiety.^{5,133}

Several bedaquiline analogues have been synthesised with the aim of improving the antimicrobial activity and spectrum of bedaquiline, as well as the pharmacological properties of the antibiotic, however, none of these has progressed to clinical evaluation.^{134,135}

Antimicrobial activity of bedaquiline

Unlike clofazimine, which has a broader spectrum of antimicrobial activity, bedaquiline has demonstrated weak activity against Gram-positive and Gram-negative bacteria, generally exhibiting MIC values >32 mg/L.^{2,120,130} Activity against other types of microorganisms, such as parasites and fungi, has not yet been reported. Importantly, bedaquiline has demonstrated selective activity against a wide variety of pathogenic mycobacteria, such as *Mtb*, *M. leprae*¹³⁶ and *M. avium*,¹³⁷ as well as non-pathogenic organisms including *M. smegmatis*, with MICs ranging from 0.003 - 0.5 mg/L.^{119,120,138} *Mtb*, and *M. smegmatis*, are the most susceptible mycobacterial species to bedaquiline with equivalent MIC values of 0.003 mg/L.^{2,119,120,138} On the other hand, several mycobacterial species are naturally resistant to bedaquiline with high MIC values (>8 mg/L) reported for *M. novocastrense*, *M. shimoidei* and *M. xenopi in vitro*. Resistance of these organisms to bedaquiline has been associated with phenotypic variations in the F₁/F₀-ATP

synthase enzyme, which, as described below, is the target for bedaquiline in susceptible mycobacterial species.^{2,139}

Like clofazimine, bedaquiline has showed varying activity against the different bacterial subpopulations within DS and DR strains of *Mtb*. It is highly bacteriostatic against actively-replicating organisms,¹⁴⁰ achieving MIC values ranging from 0.003 - 0.12 mg/L *in vitro*.^{2,119,120,138} It also acts synergistically when used in combination with other anti-TB agents, including the primary anti-TB drugs rifampicin, ethambutol and pyrazinamide, as well as second-line drugs such as AZD5847, tedizolid, oxazolidinone, rifapentine, linezolid, clofazimine,¹⁴¹⁻¹⁴³ BTZ043 and PBTZ169.¹⁴⁴ Other agents such as sutezolid and SQ109 have demonstrated additive interactions,¹⁴⁵ while others like pretonamid, interact antagonistically with bedaquiline.¹⁴⁶

Bactericidal activity of bedaquiline against actively-replicating *Mtb* organisms has also been demonstrated *in vivo*, resulting in accelerated sterilizing activity in a murine model of experimental chemotherapy as shown by earlier culture conversion.¹⁴⁵ During two months of chemotherapy, approximately 20% of mice treated with bedaquiline monotherapy demonstrated culture conversion to negativity, which was not observed in those treated with individual primary anti-TB drugs. The sterilizing activity of bedaquiline improved when this agent was added to the primary anti-TB drug regimen, resulting in culture conversion in 70 - 100% of infected mice.^{145,147} In humans, bedaquiline, like clofazimine, has demonstrated poor EBA during the first 14 days of chemotherapy. Like clofazimine, earlier onset of

bactericidal activity of bedaquiline can, however, be achieved by increasing the dosage of the drug.^{148,149}

Importantly, and similarly to clofazimine, bedaquiline possesses bactericidal activity against dormant/non-replicating *Mtb* bacilli at low, therapeutically attainable MBCs.² This highly beneficial property of bedaquiline on the dormant bacilli, has been attributed to its LBA,^{150,151} which is also consistent with shortening of chemotherapy. However, like clofazimine, it fails to act on mycobacterial populations residing in caseous granuloma lesions of C3HeB/FeJ mice due to its poor penetration of the caseous core.¹⁵²

Mechanisms of antimycobacterial action of bedaquiline

Bedaquiline, unlike clofazimine, is highly selective against *Mtb*, including MDR strains of the pathogen, as well as against other types of mycobacteria as mentioned above.¹³⁰ The drug selectively targets and inactivates, the F₁/F₀-ATP synthase of the pathogen,¹³⁰ but importantly, has no inhibitory effect on mammalian F₁/F₀-ATP synthase.¹⁴² F₁/F₀-ATP synthase is a highly conserved and key enzyme in the process of oxidative phosphorylation, which utilises the kinetic mechanical energy of PMF to drive ATP production.^{153,154} Protons generated by the bacterial, membrane-associated electron-transport chains coupled to oxidative phosphorylation are captured by, and funnelled into the membrane-embedded, F₀ proton channel of the enzyme and transported to the catalytic F₁ component, which undergoes a conformational change resulting in synthesis of ATP.^{2,154,155}

Computer-based molecular modelling/docking, mutational analyses and other approaches have identified the lipophilic F_0 component of F_1/F_0 ATP synthase, specifically the transmembrane, oligomeric subunit C (AtpE) as being the primary target of bedaquiline.^{130,139,156,157} Binding of bedaquiline is located to a “cleft between two adjacent C subunits in the C-ring,” a region which contains the proton-binding acidic residue, Glu-61.^{139,156,157} The consequence is interference with proton movement and synthesis of ATP.¹⁵⁸

Although rapid depletion of ATP occurs following exposure of *Mtb* to bedaquiline, bactericidal action, as mentioned above, is delayed for several days, apparently as a result of induction of dormancy and utilisation of alternative energy sources and pathways such as glycolysis.^{43,142,159} It has even been proposed that the bactericidal mode of action of bedaquiline, at least in the case of *M. smegmatis*, results from the collapse of the transmembrane pH gradient and dissipation of the PMF which is lethal to mycobacteria.¹⁶⁰ This contention is consistent with observations that exposure of *M. smegmatis* to bedaquiline is associated with upregulated expression and utilisation of cytochrome *bd* oxidase, the non-proton-pumping terminal oxidase, which improves bacterial survival.^{74,160} In addition, and supportive of these findings, others have reported that selective knockout of cytochrome *bd* oxidase in *Mtb* results in increased susceptibility of the pathogen to bedaquiline,¹⁶¹ which, as mentioned earlier, is also evident following exposure of *M. smegmatis* to clofazimine.⁷⁴

Very recently Lamprecht and colleagues have reported that a combination of clofazimine, bedaquiline and the novel imidazopyridine amide antimycobacterial

agent, Q203, is extremely effective in accelerating the rate of extracellular and intramacrophage killing of *Mtb in vitro*, being superior to the individual agents and the various two drug combinations.¹⁶² Mechanistically, bedaquiline and Q203, via their inhibitory effects on the bacterial electron-transport chain were found to increase the intracellular reductive potential via elevated NADH/NAD⁺ ratios, which, in turn, augmented clofazimine-mediated generation of bactericidal ROS.¹⁶² Although interesting, the therapeutic potential of this drug combination remains to be addressed, as do concerns in relation to cross-resistance to these agents.^{162,163}

Alternative mechanisms of antimycobacterial action of bedaquiline

Like clofazimine, bedaquiline is a cationic amphiphilic drug, a property, which, albeit speculatively, may be related to its primary F₁/F₀-ATP synthase-directed mode of antimycobacterial action. Surprisingly, however, the existence of alternative, secondary mechanisms of bedaquiline -mediated antimycobacterial activity possibly related to the cationic amphiphilic properties, especially effects on membrane ion-transporting ATPases, appear to be largely unexplored.

Effects of bedaquiline on eukaryotic cells

This important aspect of bedaquiline research is, to our knowledge, also largely unexplored. The observed prolongation of the QT interval and potentially fatal cardiac arrhythmias, may, or may not, be related to interference with membrane-associated cation transporters, specifically K⁺ transport in cardiomyocytes secondary to cationic amphiphilic properties, an issue which requires investigation.²⁴ Very

recently, bedaquiline, like clofazimine,¹⁶⁴ has been reported to possess anti-tumour properties *in vitro*, which are related to inhibition of mitochondrial respiration and intracellular generation of ROS.^{165,166}

Clinical efficacy of bedaquiline

As mentioned above, bedaquiline may be added to a relevant BR for the treatment of DR-TB patients who do not respond to the current WHO-recommended treatment regimen. The dosing regimen consists of 400 mg orally once daily for two weeks, followed by 200 mg orally three times weekly with a total treatment duration of 24 weeks.^{121,155} Inclusion of bedaquiline in the treatment of DR-TB has led to improvements in treatment outcomes, resulting in shorter duration of treatment and low relapse rates. The shorter duration of therapy is associated with faster sputum and culture conversion.^{105,107,128,150} The improved sputum conversion demonstrated in two studies during the first two months of treatment was achieved in 48% and 84% of patients treated with bedaquiline/BR as opposed to 9% and 65% in the placebo/BR-treated groups, respectively.^{99,167,168} With respect to overall duration of chemotherapy, this ranged from 78 - 83 days in the bedaquiline/BR-treated group in comparison with 125 - 129 days in the placebo/BR-treated group.¹²¹

The rates of culture conversion were also significantly faster in patients treated with the bedaquiline/BR than those treated with placebo/BR. During the six months of therapy 79.5% and 81% of patients who received bedaquiline achieved culture conversion to negativity, while approximately 65% of those who received placebo/BR converted.^{99,124} Although not statistically significant, data reported by

Pym *et al.*⁹⁹ showed that the culture conversion rate is affected by the drug resistance profiles of *Mtb* isolates, decreasing as the degree of drug resistance of the isolates increases. In their study, the rates of culture conversion were 73.1%, 70.5% and 62% for MDR-, pre-XDR- and XDR-TB patients, respectively.⁹⁹ The use of bedaquiline has also been beneficial in preventing the emergence of resistance to the companion drugs in the regimens.^{2,99} Inclusion of bedaquiline in the drug regimen also resulted in lower relapse rates, with the majority of patients who achieved culture conversion to negativity maintaining this status for long periods, recording a median of 5.4 months of treatment-free follow-up.⁹⁹

Bedaquiline can also be used effectively for the treatment of patients infected with HIV.^{99,169} However, as with clofazimine, many ARV agents, such as efavirenz and lopinavir, have demonstrated DDIs with bedaquiline, necessitating replacement therapy with alternative agents such as nevirapine.^{170,171}

Mechanisms of resistance to bedaquiline

Since its introduction, an increase in the number of bedaquiline-resistant *Mtb* isolates has been reported.¹⁷² Based on this concern, the WHO has advised that bedaquiline resistance development be carefully monitored. To date, however, standardised assays for the detection of drug resistance in bedaquiline have not been developed. Implementation of surveillance to monitor the emergence of resistance via serial MIC determinations of isolates from patients on bedaquiline therapy, especially when there is a history of prior exposure to clofazimine, has been proposed. In this context, approximately 97% of clinical isolates at baseline drug susceptibility testing

(DST) have shown bedaquiline MIC values ranging from 0.0075 - 0.24 mg/L.^{99,125,173,174} Isolates with MIC values >0.24 mg/L, as well as those exhibiting a 4-fold increase in MIC from baseline occurring during treatment, are contenders for possible development of bedaquiline resistance and should be closely monitored. Used in conjunction with early detection of clinical signs of non-response to treatment, monitoring of drug resistance development involves sequential determination of the MIC values of *Mtb* present in sputum samples taken at baseline, and at weeks 8 and 24, following initiation of chemotherapy.^{99,175}

In the case of most DR-TB regimens, inclusion of bedaquiline together with a minimum of three other anti-TB agents has been recommended.^{2,176} These combination regimens, consisting of fewer anti-TB drugs, do, however, pose an increased risk of development of drug resistance.¹² In this respect, the WHO has recently issued an interim guideline¹⁴ for more discerning use of bedaquiline in DR-TB regimens. In this context, an effective treatment regimen containing four second-line drugs, including pyrazinamide, a fluoroquinolone and a second-line injectable agent, is recommended, with bedaquiline held in reserve for those clinical settings in which such regimens are deemed to be ineffective.⁴

Several molecular mechanisms underpinning bedaquiline resistance have been identified. Most prominent amongst these are mutations occurring at two separate genes. The first gene is the *atpE*, which encodes for F₁/F₀-ATP synthase. Currently, the occurrence of resistance due to mutations at this gene has been reported in approximately 30% of bedaquiline-resistant clinical isolates.^{107,177} To

date, this type of resistance has been exclusively associated with mutations at the C subunit of the enzyme.¹⁵⁸

The second gene associated with bedaquiline resistance is the *rv0678*, which encodes the Rv0678 protein. The majority of bedaquiline-resistant mutants, reported in several studies, have mutations at the *rv0678* gene.^{105,107,177} In South Africa, all of the bedaquiline-resistant isolates, as well as some with the potential to develop bedaquiline resistance (>4-fold increase in MIC), have had mutations at the *rv0678* gene.⁹⁹ A case of bedaquiline resistance reported from Switzerland, also involved mutation of this gene.⁹⁷ Importantly, as mentioned above, *rv0678* gene mutations also lead to cross-resistance with clofazimine, potentially restricting treatment options.^{2,6,107}

The impact of *rv0678*-based resistance on the outcome of DR-TB chemotherapy appears variable. In the study by Pym *et al.*⁹⁹ 5 of 12 patients who harboured *Mtb* strains which demonstrated a >4-fold increase in their bedaquiline MICs during therapy, nevertheless had a successful treatment outcome.⁹⁹ On the contrary, Pule *et al.*⁹⁸ have emphasized the significance of efflux-mediated bedaquiline resistance as a contributor to treatment failure.

Rv0678-associated bedaquiline and clofazimine resistance can be attenuated *in vitro* via the use of efflux pump inhibitors, such as verapamil and the protonophores,^{106,178} timcodar, reserpine and valinomycin, all of which inhibit the pump by reducing the transmembrane potential.^{105,178} Verapamil is particularly effective, decreasing the bedaquiline and clofazimine MICs by at least 8-fold *in*

vitro.^{98,106,179} However, the clinical utility of this approach is dubious due to the high levels of verapamil required to achieve these effects, as well as metabolism of this agent by CYP3A4, indicative of a probable DDI when coadministered with bedaquiline and clofazimine.

The second factor contributing to the development of bedaquiline resistance in *Mtb* is the mutation rate, which is dependent on the dynamics of the bacterial population in the clinical lesion or culture. For bedaquiline, the rate of drug resistance development for *Mtb* in culture has been found to be $1/10^8$ cfu/mL. This rate of resistance is relatively low, being comparable to that of rifampicin.^{130,180} This bacterial density may be achievable in granuloma lesions of chronic TB patients.^{181,182} However, which of the two resistance genes has the highest mutation rate remains unknown, with *rv0678* seemingly the most likely contender.^{6,97,99}

An additional factor associated with bedaquiline resistance development is its half-life. Bedaquiline has a longer half-life (4 - 5.5 months) than those of other anti-TB agents in the current DR-TB regimens.^{2,142,170} After termination of therapy, its long half-life may favour selection of resistant populations.^{4,150}

Pharmacokinetic and pharmacodynamic properties of bedaquiline

Like clofazimine, bedaquiline has poor pharmacokinetic and pharmacodynamic properties. It is highly lipophilic, being distributed mainly in macrophage-rich tissues such as the lungs, while it is found in low concentrations in the blood, increasing with duration of chemotherapy.¹⁸³ In other bodily fluids, such as CSF, bedaquiline is

undetectable due to failure of this antibiotic to cross the blood-brain barrier.¹⁸⁴ During TB meningitis, when the blood-brain barrier is inflamed and disrupted, bedaquiline is detectable in the CSF especially in the early phase of treatment, albeit at very low concentrations, diminishing as treatment continues and the integrity of the blood-brain barrier is restored.^{184,185}

Like clofazimine, bedaquiline is metabolised by CYP3A4^{2,119,150} to a less active metabolite, M2, which is altered during coadministration with anti-TB and ARV agents.¹⁰¹ In the case of anti-TB drugs, two rifamycins, viz. rifampicin and rifapentine, which are potent CYP3A4 inducers, have been shown to increase bedaquiline clearance (4.78-fold and 3.96-fold for RIF and rifapentine, respectively), resulting in substantial reductions in bedaquiline tissue concentrations (79% and 75% for rifampicin and rifapentine, respectively), necessitating dosage adjustment to achieve the required chemotherapeutic levels.^{3,101,155,186} These adjustments may, in turn, contribute to the severity of AEs.¹⁰¹ In the case of the ARV drugs, lopinavir/ritonavir, these agents have been shown to increase bedaquiline retention time and serum concentrations.^{2,171} On the other hand, bedaquiline can be safely administered with nevirapine without dosage adjustment.²

Various AEs associated with the use of bedaquiline have been reported. The most common events are hepatic and cardiac in nature.¹⁸⁷ The former include increased liver enzyme levels. Despite the hepatic complications, patients with mild to moderate hepatic impairment such as those with hepatitis B and C and heavy alcohol use, can still be treated with bedaquiline.¹²⁴ In the case of the latter, corrected QT interval prolongation¹⁵⁵ and disturbances of the heart's electrical

rhythm are most prominent.^{8,119,172,176} The QT elongation is exacerbated when bedaquiline is used in combination with other antibiotics such as clofazimine, moxifloxacin and ketoconazole in DR-TB therapy.^{4,150,188} Accordingly, the use of bedaquiline in regimens containing any of these antimicrobial agents should be closely monitored.⁴ Because both clofazimine and bedaquiline carry risk of prolongation of the QT interval and cardiac arrhythmia, which is additive in patients treated with these agents, WHO guidelines recommend weekly electro-cardiograms (ECGs) during the first month, and thereafter monthly in patients treated with both agents.¹⁴ When bedaquiline is used without clofazimine, monthly ECGs are sufficient.

Other AEs, which are nonspecific, include nausea, dizziness, arthralgia, headache, hyperuricemia and vomiting.^{8,119,150,155} However, these events were reported to be mild and tolerable in most studies.^{99,186}

Worryingly, an increased mortality rate has been associated with bedaquiline therapy. In the pivotal phase II licensing study, a fatality rate of 12.7% was recorded in patients receiving a bedaquiline/BR, compared with 2.5% in the standard comparator group.^{121,124} In a subsequent study, a fatality rate of 7% was recorded in the group of patients receiving the bedaquiline/BR relative to placebo/BR.⁹⁹ The increased mortality rates reported in these studies were attributed mainly to respiratory disorders of infective and non-infective origin as opposed to bedaquiline toxicity.^{121,124}

Table 2. Similarities and differences between clofazimine and bedaquiline

Property	Similarities	Differences	
		Clofazimine	Bedaquiline
Structural properties	<ul style="list-style-type: none"> are strongly lipophilic. 	<ul style="list-style-type: none"> is a rimonophenazine. 	<ul style="list-style-type: none"> is a diaryquinoline.
Antimicrobial activity	<ul style="list-style-type: none"> have a slow onset of bactericidal action. active against both DS- and DR-<i>Mtb</i> strains. have limited activity against Gram-negative bacteria. 	<ul style="list-style-type: none"> has dose-independent antimicrobial activity. has broad spectrum antimicrobial activity against Gram-positive bacteria, parasites and fungi. 	<ul style="list-style-type: none"> has dose-dependent antimicrobial activity. has limited antimicrobial activity against mycobacterial species.
Microbial subpopulations	<ul style="list-style-type: none"> active against planktonic, slow/non-replicating bacteria. show high activity against slow/non-replicating bacteria. 	<ul style="list-style-type: none"> unknown. 	<ul style="list-style-type: none"> unknown.
Cellular target	<ul style="list-style-type: none"> target the mycobacterial membrane, collapsing the pH gradient and membrane potential. interfere with mycobacterial energy metabolism. are antagonised by cytochrome <i>bd</i> oxidase. 	<ul style="list-style-type: none"> probable multiplicity of microbial targets. 	<ul style="list-style-type: none"> primarily targets the mycobacterial F₁/F₀-ATPase.

Clinical trials	<ul style="list-style-type: none"> • contribute to shortening of therapy. • result in low relapse rate after treatment cessation. • lead to increased QT prolongation. 	<ul style="list-style-type: none"> • is associated with lower mortality than bedaquiline therapy. 	<ul style="list-style-type: none"> • is associated with higher mortality than clofazimine
Drug resistance mechanisms	<ul style="list-style-type: none"> • have common mechanism of acquired resistance due to mutations in the mycobacterial transcriptional regulator Rv0678, resulting in upregulation of the MmpS5-MmpL5 drug efflux pump. 	<ul style="list-style-type: none"> • is also associated with mutations at <i>rv1979c</i> and <i>rv2535c</i> genes. • has a mutation rate of $1/10^{26}$ cfu/mL. 	<ul style="list-style-type: none"> • is also associated with mutations at <i>atpE</i> gene. • has a mutation rate of $1/10^8$ cfu/mL.
Pharmaco-properties	<ul style="list-style-type: none"> • accumulate in fatty tissues. • metabolised by CYP3A4 enzymes. • have DDI effect with several anti-TB and anti-retroviral agents. 	<ul style="list-style-type: none"> • discolours skin and tissues. 	<ul style="list-style-type: none"> • does not cause skin and tissue discolouration.

DDI, drug-drug interaction.

Summary

Similarities and differences between clofazimine and bedaquiline are summarised in Table 2.

Conclusion

Clofazimine and bedaquiline are lipophilic, prototype antimycobacterial agents of the riminophenazine and diarylquinoline classes, respectively. Both antibiotics target the outer membrane of *Mtb*, but differ with respect to target selectivity. The lipophilicity of both agents accounts for their unusual pharmacodynamic/pharmacokinetic properties, resulting in slow and prolonged tissue accumulation, which, in turn, appears to underpin their efficacy in countering the slow-growing, persistent pathogen. Given the differences in phospholipid composition between eukaryotic and prokaryotic cells, these two agents may act as templates for the design of novel membrane-active antimicrobial agents which interact selectively with the outer membrane of prokaryotic cells, enabling improved efficacy and reduced toxicity. With respect to therapeutic efficacy, the early promise of the STREAM trials in evaluating the utility and safety of short-course clofazimine- and/or bedaquiline-based DR-TB regimens is encouraging. However, compelling, definitive recommendations are awaited.

Transparency declarations

None to declare.

References

- 1 World Health Organization. *Global tuberculosis report 2015*. WHO, Geneva, 2015.
http://www.who.int/tb/publications/global_report/en/.
- 2 Chahine EB, Karaoui LR, Mansour H. Bedaquiline: A novel diarylquinoline for multidrug-resistant tuberculosis. *Ann Pharmacother* 2014; **48**: 107–15.
- 3 Zumla AI, Gillespie SH, Hoelscher M *et al*. New antituberculosis drugs, regimens, and adjunct therapies: needs, advances, and future prospects. *Lancet* 2014; **14**: 327–40.
- 4 Brigden G, Hewison C, Varaine F. New developments in the treatment of drug-resistant tuberculosis: clinical utility of bedaquiline and delamanid. *Infect Drug Resist* 2015; **8**: 367–78.
- 5 Upadhayaya RS, Vandavasi JK, Vasireddy NR *et al*. Design, synthesis, biological evaluation and molecular modelling studies of novel quinoline derivatives against *Mycobacterium tuberculosis*. *Bioorg Med Chem* 2009; **17**: 2830–41.
- 6 Somoskovi A, Bruderer V, Hömke R *et al*. A mutation associated with clofazimine and bedaquiline cross-resistance in MDR-TB following bedaquiline treatment. *Eur Respir J* 2015; **45**: 554–7.
- 7 Sloan DJ, Lewis JM. Management of multidrug-resistant TB: novel treatments and their expansion to low resource settings. *Trans R Soc Trop Med Hyg* 2016; **110**: 163–72.
- 8 Meintjes G. Management of drug-resistant TB in patients with HIV co-infection. *J Int AIDS Soc* 2014; **17**: 19508.
- 9 Palmero D, González Montaner P, Cufre M *et al*. First series of patients with XDR and pre-XDR TB treated with regimens that included meropenem-clavulanate in Argentina. *Arch Bronconeumol* 2015; **51**: e49–52.

- 10 Szumowski JD, Lynch JB. Profile of delamanid for the treatment of multidrug-resistant tuberculosis. *Drug Des Devel Ther* 2015; **9**: 677–82.
- 11 Wong EB, Cohen KA, Bishai WR. Rising to the challenge: new therapies for tuberculosis. *Trends Microbiol* 2013; **21**: 493–501.
- 12 Lessem E, Cox H, Daniels C *et al*. Access to new medications for the treatment of drug-resistant tuberculosis: patient, provider and community perspectives. *Int J Infect Dis* 2015; **32**: 56–60.
- 13 Padayatchi N, Gopal M, Naidoo R *et al*. Clofazimine in the treatment of extensively drug-resistant tuberculosis with HIV coinfection in South Africa: a retrospective cohort study. *J Antimicrob Chemother* 2014; **69**: 3103–7.
- 14 World Health Organization. *Companion handbook to the WHO guidelines for the programmatic management of drug-resistant tuberculosis*. Published online, November 2014.
www.who.int/tb/publications/pmdt_companionhandbook/en/.
- 15 Falzon D, Jaramillo E, Schünemann HJ *et al*. WHO guidelines for the programmatic management of drug-resistant tuberculosis: 2011 update. *Eur Respir J* 2011; **38**: 516–28.
- 16 Shim TS, Jo KW. Medical treatment of pulmonary multidrug-resistant tuberculosis. *Infect Chemother* 2013; **45**: 367–74.
- 17 van Altena R, de Vries G, Haar CH *et al*. Highly successful treatment outcome of multidrug-resistant tuberculosis in the Netherlands, 2000-2009. *Int J Tuberc Lung Dis* 2015; **19**: 406–12.
- 18 Dey T, Brigden G, Cox H *et al*. Outcomes of clofazimine for the treatment of drug-resistant tuberculosis: a systematic review and meta-analysis. *J Antimicrob Chemother* 2013; **68**: 284–93.

- 19 Dooley KE, Obuku EA, Durakovic N *et al.* World health Organization group 5 drugs for the treatment of drug-resistant tuberculosis: unclear efficacy or untapped potential? *J Infect Dis* 2013; **207**: 1352–8.
- 20 Guglielmetti L, Le Dû D, Jachym M *et al.* Compassionate use of bedaquiline for the treatment of multidrug-resistant and extensively drug-resistant tuberculosis: interim analysis of a French cohort. *Clin Infect Dis* 2015; **60**: 188–94.
- 21 Li K, Schurig-Briccio LA, Feng X *et al.* Multitarget drug discovery for tuberculosis and other infectious diseases. *J Med Chem* 2014; **57**: 3126–39.
- 22 Clofazimine. <http://www.drugbank.ca/drugd/DB00845>.
- 23 Bedaquiline. <http://www.drugbank.ca/drugs/DB08903>.
- 24 Feng X, Zhu W, Schurig-Briccio LA *et al.* Antiinfectives targeting enzymes and the proton motive force. *Proc Natl Acad Sci U S A* 2015; **112**: E7073–82.
- 25 Barry VC, Belton JG, Conalty ML *et al.* A new series of phenazines (rimino-compounds) with high antituberculosis activity. *Nature* 1957; **179**: 1013–5.
- 26 Reddy VM, O’Sullivan JF, Gangadharam PR. Antimycobacterial activities of riminophenazines. *J Antimicrob Chemother* 1999; **43**: 615–23.
- 27 Nunn AJ, Rusen ID, van Deun A *et al.* Evaluation of a standardised treatment regimen of anti-tuberculosis drugs for patients with multi-drug-resistant tuberculosis (STREAM): study protocol for a randomised controlled trial. *Trials* 2014; **15**: 353.
- 28 Moodley R, Godec TR, STREAM Trial Team. Short-course treatment for multidrug-resistant tuberculosis: the STREAM trials. *Eur Respir Rev* 2016; **25**: 29–35.

- 29** van Deun A, Maug AK, Salim MA *et al.* Short, highly effective, and inexpensive standardized treatment of multidrug-resistant tuberculosis. *Am Respir Crit Care Med* 2010; **182**: 684–92.
- 30** Trébucq A, Schwoebel V, Kuaban C *et al.* Expanding shortened MDR-TB treatment: the West African experience. *Int J Tuberc Lung Dis* 2014; **18**: Suppl. 1, S15.
- 31** Franzblau SG, O’Sullivan JF. Structure-activity relationships of selected phenazines against *Mycobacterium leprae* *in vitro*. *Antimicrob Agents Chemother* 1988; **32**: 1583–5.
- 32** Cholo MC, Steel HC, Fourie PB *et al.* Clofazimine: current status and future prospects. *J Antimicrob Chemother* 2012; **67**: 290–8.
- 33** Funk RS, Krise JP. Cationic amphiphilic drugs cause a marked expansion of apparent lysosomal volume: implications for an intracellular distribution-based drug interaction. *Mol Pharm* 2012; **9**: 1384-95.
- 34** Kazmi F, Hensley T, Pope C *et al.* Lysosomal sequestration (trapping) of lipophilic amine (cationic amphiphilic) drugs in immortalized human hepatocytes (Fa2N-4 cells). *Drug Metab Dispos* 2013; **41**: 897-905.
- 35** Lanoix JP, Lenaerts AJ, Neurmberger EL. Heterogeneous disease progression and treatment response in a C3HeB/FeJ mouse model of tuberculosis. *Dis Model Mech* 2015; **8**: 603-10.
- 36** Franzblau SG, White KE, O’Sullivan JF. Structure-activity relationships of tetramethylpiperidine-substituted phenazines against *Mycobacterium leprae* *in vitro*. *Antimicrob Agents Chemother* 1989; **33**: 2004–5.

- 37** Steel HC, Matlola NM, Anderson R. Inhibition of potassium transport and growth of mycobacteria exposed to clofazimine and B669 is associated with a calcium-independent increase in microbial phospholipase A₂ activity. *J Antimicrob Chemother* 1999; **44**: 209–16.
- 38** Matlola NM, Steel HC, Anderson R. Antimycobacterial action of B4128, a novel tetramethylpiperidyl-substituted phenazine. *J Antimicrob Chemother* 2001; **47**: 199–202.
- 39** Zhang D, Liu Y, Zhang C *et al.* Synthesis and biological evaluation of novel 2-methoxypyridylamino-substituted riminophenazine derivatives as antituberculosis agents. *Molecules* 2014; **19**: 4380–94.
- 40** Zhang D, Lu Y, Liu B *et al.* Identification of less lipophilic riminophenazine derivatives for the treatment of drug-resistant tuberculosis. *J Med Chem* 2012; **55**: 8409-17.
- 41** Grosset JH, Tyagi S, Almeida DV *et al.* Assessment of clofazimine activity in a second-line regimen for tuberculosis in mice. *Am J Respir Crit Care Med* 2013; **188**: 608–12.
- 42** Verma RK, Germishuizen WA, Motheo MP *et al.* Inhaled microparticles containing clofazimine are efficacious in treatment of experimental tuberculosis in mice. *Antimicrob Agents Chemother* 2013; **57**: 1050-2.
- 43** van Rensburg CE, Jooné GK, O'Sullivan JF *et al.* Antimicrobial activities of clofazimine and B669 are mediated by lysophospholipids. *Antimicrob Agents Chemother* 1992; **36**: 2729–35.
- 44** De Bruyn EE, Steel HC, van Rensburg EJ *et al.* The riminophenazines, clofazimine and B669, inhibit potassium transport in Gram-positive bacteria by a lysophospholipid-dependent mechanism. *J Antimicrob Chemother* 1996; **38**: 349–62.
- 45** van Rensburg CE, van Straten AM. An *in vitro* investigation of the susceptibility of *Enterococcus faecalis* to clofazimine and B669. *J Antimicrob Chemother* 1994; **33**: 356–8.

- 46** van Rensburg CE, Jooné GK, Sirgel FA *et al.* *In vitro* investigation of the antimicrobial activities of novel tetramethylpiperidine-substituted phenazines against *Mycobacterium tuberculosis*. *Chemotherapy* 2000; **46**: 43–8.
- 47** Makgatho ME, Anderson R, O’Sullivan JF *et al.* Tetramethylpiperidine-substituted phenazines as novel anti-plasmodial agents. *Drug Dev Res* 2000; **50**: 195–202.
- 48** Datta G, Bera T. The effects of clofazimine, niclosamide and amphotericin B, on electron transport of *Leishmania donovani* promastigotes. *Indian J Med Res* 2000; **112**: 15–20.
- 49** Barteselli A, Casagrande M, Basilico N *et al.* Clofazimine analogs with antileishmanial and antiplasmodial activity. *Bioorg Med Chem* 2015; **23**: 55–65.
- 50** Bellera CL, Balcazar DE, Vanrell MC *et al.* Computer-guided drug repurposing: identification of trypanocidal activity of clofazimine, benipine and saquinavir. *Eur J Med Chem* 2015; **93**: 338–48.
- 51** Tuvshintulga B, AbouLaila M, Davaasuren B *et al.* Clofazimine inhibits the growth of Babesia and Theileria parasites *in vitro* and *in vivo*. *Antimicrob Agents Chemother* 2016; doi:10.1128/AAC.01614-15.
- 52** Panic G, Vargas M, Scandale I *et al.* Activity profile of an FDA-approved compound library against *Schistosoma mansoni*. *PLoS Negl Trop Dis* 2015; **9**: e0003962.
- 53** Robbins N, Spitzer M, Yu T *et al.* An antifungal combination matrix identifies a rich pool of adjuvant molecules that enhance drug activity against diverse fungal pathogens. *Cell Rep* 2015; **13**: 1481–92.

- 54 Singh S, Bouzinbi N, Chaturvedi V *et al.* *In vitro* evaluation of a new drug combination against clinical isolates belonging to the *Mycobacterium abscessus* complex. *Clin Microbiol Infect* 2014; **20**: O1124–7.
- 55 Cariello PF, Kwak EJ, Abdel-Massih RC *et al.* Safety and tolerability of clofazimine as salvage therapy for atypical mycobacterial infection in solid organ transplant recipients. *Transpl Infect Dis* 2015; **17**: 111–8.
- 56 Bennie CJ, To JL, Martin PA *et al.* *In vitro* interaction of some drug combinations to inhibit rapidly growing mycobacteria isolates from cats and dogs and these isolates' susceptibility to cefovecin and clofazimine. *Aust Vet J* 2015; **93**: 40–5.
- 57 Ferro BE, Meletiadiis J, Wattenberg M *et al.* Clofazimine prevents the regrowth of *Mycobacterium abscessus* and *Mycobacterium avium* type strains exposed to amikacin and clarithromycin. *Antimicrob Agents Chemother* 2015; **60**: 1097–105.
- 58 Obregón-Henao A, Arnett KA, Henao-Tamayo M *et al.* Susceptibility of *Mycobacterium abscessus* to antimycobacterial drugs in preclinical models. *Antimicrob Agents Chemother* 2015; **59**: 6904–12.
- 59 Roy K, Sil A, Das NK *et al.* Effectiveness and safety of clofazimine and pentoxifylline in type 2 lepra reaction: a double-blind, randomized, controlled study. *Int J Dermatol* 2015; **54**: 1325–32.
- 60 Xu J, Lu Y, Fu L *et al.* *In vitro* and *in vivo* activity of clofazimine against *Mycobacterium tuberculosis* persisters. *Int J Tuberc Lung Dis* 2012; **16**: 1119–25.
- 61 Cholo MC, Boshoff HI, Steel HC *et al.* Effects of clofazimine on potassium uptake by a Trk-deletion mutant of *Mycobacterium tuberculosis*. *J Antimicrob Chemother* 2006; **57**: 79–84.

- 62 Mothiba MT, Anderson R, Fourie B *et al.* Effects of clofazimine on planktonic and biofilm growth of *Mycobacterium tuberculosis* and *Mycobacterium smegmatis*. *J Glob Antimicrob Resist* 2015; **3**: 13–8.
- 63 Williams K, Minkowski A, Amoabeng O *et al.* Sterilizing activities of novel combinations lacking first- and second-line drugs in a murine model of tuberculosis. *Antimicrob Agents Chemother* 2012; **56**: 3114–20.
- 64 Zhang Z, Li T, Qu G *et al.* *In vitro* synergistic activity of clofazimine and other antituberculous drugs against multidrug-resistant *Mycobacterium tuberculosis* isolates. *Int J Antimicrob Agents* 2015; **45**: 71–5.
- 65 Zou L, Liu M, Wang Y *et al.* Determination of *in vitro* synergy between linezolid and other antimicrobial agents against *Mycobacterium tuberculosis* isolates. *Tuberculosis (Edinb)* 2015; **95**: 839–42.
- 66 López-Gavín A, Tudó G, Vergara A *et al.* *In vitro* activity against *Mycobacterium tuberculosis* of levofloxacin, moxifloxacin and UB-8902 in combination with clofazimine and pretomanid. *Int J Antimicrob Agents* 2015; **46**: 582–5.
- 67 Irwin SM, Gruppo V, Brooks E *et al.* Limited activity of clofazimine as a single drug in a mouse model of tuberculosis exhibiting caseous necrotic granulomas. *Antimicrob Agents Chemother* 2014; **58**: 4026–34.
- 68 Tyagi S, Ammerman NC, Li SY *et al.* Clofazimine shortens the duration of the first-line treatment regimen for experimental chemotherapy of tuberculosis. *Proc Natl Acad Sci U S A* 2015; **112**: 869–74.
- 69 Zhang M, Sala C, Hartkoorn RC *et al.* Streptomycin-starved *Mycobacterium tuberculosis* 18b, a drug discovery tool for latent tuberculosis. *Antimicrob Agents Chemother* 2012; **56**: 5782–9.

- 70** Yano T, Kassovska-Bratinova S, Teh JS *et al.* Reduction of clofazimine by mycobacterial type 2 NADH:quinone oxidoreductase: a pathway for the generation of bactericidal levels of reactive oxygen species. *J Biol Chem* 2011; **286**: 10276–87.
- 71** Piccaro G, Poce G, Biava M *et al.* Activity of lipophilic and hydrophilic drugs against dormant and replicating *Mycobacterium tuberculosis*. *J Antibiot* 2015; **68**: 711-4.
- 72** Grant SS, Kaufmann BB, Chand NS *et al.* Eradication of bacterial persisters with antibiotic-generated hydroxyl radicals. *Proc Natl Acad Sci U S A* 2010; **109**: 12147–52.
- 73** Lechartier B, Cole ST. Mode of action of clofazimine and combination therapy with benzothiazinones against *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2015; **59**: 4457–63.
- 74** Lu P, Heineke MH, Koul A *et al.* The cytochrome *bd*-type quinol oxidase is important for survival of *Mycobacterium smegmatis* under peroxide and antibiotic-induced stress. *Sci Rep* 2015; **5**: 10333.
- 75** Nazarov PV, Lider VA. Mechanism of the membrane stabilizing action of vitamins K and E under conditions of chronic phenol poisoning in albino rats. *Vopr Pitan* 1996; **2**: 11–4.
- 76** Lu Y, Zheng M, Wang B *et al.* Clofazimine analogs with efficacy against experimental tuberculosis and reduced potential for accumulation. *Antimicrob Agents Chemother* 2011; **55**: 5185–93.
- 77** Steel HC, Cockeran R, Anderson R. Platelet-activating factor and lyso-PAF possess direct antimicrobial properties *in vitro*. *APMIS* 2002; **110**: 158–64.
- 78** LaDow JE, Warnock DC, Hamill KM *et al.* Bicephalic amphiphile architecture affects antibacterial activity. *Eur J Med Chem* 2011; **46**: 4219–26.

- 79** Kapoor R, Eimerman PR, Hardy JW *et al.* Efficacy of antimicrobial peptoids against *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2011; **55**: 3058–62.
- 80** Bhattacharyya D, Sen PC. The effect of binding of chlorpromazine and chloroquine to ion transporting ATPases. *Mol Cell Biochem* 1999; **198**: 179–85.
- 81** Datiles MJ, Johnson EA, McCarty RE. Inhibition of the ATPase activity of the catalytic portion of ATP synthases by cationic amphiphiles. *Biochim Biophys Acta* 2008; **1777**: 362–8.
- 82** Hong S, Pedersen PL. ATP synthase and the actions of inhibitors utilized to study its roles in human health, disease, and other scientific areas. *Microbiol Mol Biol Rev* 2008; **72**: 590–641.
- 83** Te Brake LH, Russel FG, van den Heuvel JJ *et al.* Inhibitory potential of tuberculosis drugs on ATP-binding cassette drug transporters. *Tuberculosis (Edinb)* 2016; **96**: 150–7.
- 84** Anderson R, Smit MJ. Clofazimine and B669 inhibit the proliferative responses and Na⁺, K⁺-adenosine triphosphatase activity of human lymphocytes by a lysophospholipid-dependent mechanism. *Biochem Pharmacol* 1993; **46**: 2029–38.
- 85** Ren YR, Pan F, Parvez S *et al.* Clofazimine inhibits human Kv1.3 potassium channel by perturbing calcium oscillation in T lymphocytes. *PLoS One* 2008; **3**: e4009.
- 86** Faouzi M, Starkus J, Penner R. State-dependent blocking mechanism of Kv1.3 channels by the antimycobacterial drug clofazimine. *Br J Pharmacol* 2015; **172**: 5161–73.
- 87** van Rensburg CE, van Staden AM, Anderson R. The riminophenazine agents, clofazimine and B669 inhibit the proliferation of cancer cell lines *in vitro* by phospholipase A₂-mediated oxidative and nonoxidative mechanisms. *Cancer Res* 1993; **53**: 318–23.

- 88** Officioso A, Alzoubi K, Manna C *et al.* Clofazimine induced suicidal death of human erythrocytes. *Cell Physiol Biochem* 2015; **37**: 331–41.
- 89** Yoon GS, Keswani RK, Sud S *et al.* Clofazimine biocrystal accumulation in macrophages upregulates IL-1RA production to induce a systemic anti-inflammatory state. *Antimicrob Agents Chemother* 2016; pii: AAC.00265-16.
- 90** Hwang TJ, Dotsenko S, Jafarov A *et al.* Safety and availability of clofazimine in the treatment of multidrug and extensively drug-resistant tuberculosis: analysis of published guidance and meta-analysis of cohort studies. *BMJ Open* 2014; **4**: e004143.
- 91** Aung KJ, Van Deun A, Declercq E *et al.* Successful '9-month Bangladesh regimen' for multidrug-resistant tuberculosis among over 500 consecutive patients. *Int J Tuberc Lung Dis* 2014; **18**: 1180–7.
- 92** Kuaban C, Noeske J, Rieder HL *et al.* High effectiveness of a 12-month regimen for MDR-TB patients in Cameroon. *Int J Tuberc Lung Dis* 2015; **19**: 517–24.
- 93** Piubello A, Harouna SH, Souleymane MB *et al.* High cure rate with standardised short-course multidrug-resistant tuberculosis treatment in Niger: no relapses. *Int J Tuberc Lung Dis* 2014; **18**: 1188–94.
- 94** Prideaux B, Via LE, Zimmerman MD, Eum S *et al.* The association between sterilizing activity and drug distribution into tuberculosis lesions. *Nat Med* 2015; **21**: 1223-7.
- 95** Swanson R, Ammerman N, Ngcobo B *et al.* Clofazimine contributes sustained antimicrobial activity after treatment cessation in the mouse model of tuberculosis chemotherapy. *Antimicrob Agents Chemother* 2016; **60**: 2864-9.

- 96** Kurbatova EV, Dalton T, Ershova J *et al.* Additional drug resistance of multidrug-resistant tuberculosis in patients in 9 countries. *Emerg Infect Dis* 2015; **21**: 977–83.
- 97** Bloemberg GV, Keller PM, Stucki D *et al.* Acquired resistance to bedaquiline and delamanid in therapy for tuberculosis. *N Eng J Med* 2015; **20**: 1986–8.
- 98** Pule CM, Sampson SL, Warren RM *et al.* Efflux pump inhibitors: targeting mycobacterial efflux systems to enhance TB therapy. *J Antimicrob Chemother* 2016; **71**: 17–26.
- 99** Pym AS, Diacon AH, Tang SJ *et al.* Bedaquiline in the treatment of multidrug- and extensively drug-resistant tuberculosis. *Eur Respir J* 2016; **47**: 564–74.
- 100** Horita Y, Doi N. Comparative study of the effects of antituberculosis drugs and antiretroviral drugs on cytochrome P450 3A4 and P-glycoprotein. *Antimicrob Agents Chemother* 2014; **58**: 3168–76.
- 101** Svensson EM, Murray S, Karlsson MO *et al.* Rifampicin and rifapentine significantly reduce concentrations of bedaquiline, a new anti-TB drug. *Antimicrob Chemother* 2015; **70**: 1106–14.
- 102** Xu HB, Jiang RH, Xiao HP. Clofazimine in the treatment of multidrug-resistant tuberculosis. *Clin Microbiol Infect* 2012; **18**: 1104–10.
- 103** Tang S, Yao L, Hao X *et al.* Clofazimine for the treatment of multidrug-resistant tuberculosis: prospective, multicenter, randomized controlled study in China. *Clin Infect Dis* 2015; **60**: 1361–7.
- 104** Gopal M, Padayatchi N, Metcalfe JZ *et al.* Systematic review of clofazimine for the treatment of drug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2013; **17**: 1001–7.
- 105** Andries K, Villellas C, Coeck N *et al.* Acquired resistance of *Mycobacterium tuberculosis* to bedaquiline. *PLoS One* 2014; **9**: e102135.

- 106** Gupta S, Cohen KA, Winglee K *et al.* Efflux inhibition with verapamil potentiates bedaquiline in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2014; **58**: 574–6.
- 107** Hartkoorn RC, Uplekar S, Cole ST. Cross-resistance between clofazimine and bedaquiline through upregulation of MmpL5 in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2014; **58**: 2979–81.
- 108** Koser CU, Javid B, Liddell K *et al.* Drug resistance mechanisms and tuberculosis drugs. *Lancet* 2015; **385**: 305–7.
- 109** Zhang S, Chen J, Cui P *et al.* Identification of novel mutations associated with clofazimine resistance in *Mycobacterium tuberculosis*. *J Antimicrob Chemother* 2015; **70**: 2507–10.
- 110** Baijnath S, Naiker S, Shobo A *et al.* Evidence for the presence of clofazimine and its distribution in the healthy mouse brain. *J Mol Histol* 2015; **46**: 439–42.
- 111** Baik J, Stringer KA, Mane G *et al.* Multiscale distribution and bioaccumulation analysis of clofazimine reveals a massive immune system-mediated xenobiotic sequestration response. *Antimicrob Agents Chemother* 2013; **57**: 1218–30.
- 112** Srikanth CH, Joshi P, Bikkasani AK *et al.* Bone distribution study of anti leprotic drug clofazimine in rat bone marrow cells by a sensitive reverse phase liquid chromatography method. *J Chromatogr B Analyt Technol Biomed Life Sci* 2014; **960**: 82–6.
- 113** Swanson RV, Adamson J, Moodley C *et al.* Pharmacokinetics and pharmacodynamics of clofazimine in a mouse model of tuberculosis. *Antimicrob Agents Chemother* 2015; **59**: 3042–51.
- 114** Baik J, Rosania GR. Molecular imaging of intracellular drug-membrane aggregate formation. *Mol Pharm* 2011; **8**: 1742–9.

- 115** Baik J, Rosania GR. Macrophage sequester clofazimine in an intracellular liquid crystal-like supramolecular organization. *PLoS One* 2012; **7**: e47494.
- 116** Keswani RK, Yoon GS, Sud S *et al.* A far-red fluorescent probe for flow cytometry and image-based functional studies of xenobiotic sequestering macrophages. *Cytometry* 2015; **87**: 855–67.
- 117** Yoon GS, Sud S, Keswani RK *et al.* Phagocytosed clofazimine biocrystals can modulate innate immune signalling by inhibiting TNF α and boosting IL-1RA secretion. *Mol Pharm* 2015; **12**: 2517–27.
- 118** Keswani RK, Baik J, Yeomans L *et al.* Chemical analysis of drug biocrystals: a role for counter ion transport pathways in intracellular drug disposition. *Mol Pharm* 2015; **12**: 2528–36.
- 119** Worley MV, Estrada SJ. Bedaquiline: a novel antitubercular agent for the treatment of multidrug-resistant tuberculosis. *Pharmacotherapy* 2014; **34**: 1187–97.
- 120** Phillely JV, Wallace RJ Jr, Benwill JL *et al.* Preliminary results of bedaquiline as salvage therapy for patients with nontuberculous mycobacterial lung disease. *Chest* 2015; **148**: 499–506.
- 121** Diacon AH, Pym A, Grobusch MP *et al.* Multidrug-resistant tuberculosis and culture conversion with bedaquiline. *N Engl J Med* 2014; **371**: 723–32.
- 122** Geffen N. Anything to stay alive: the challenges of a campaign for an experimental drug. *Dev World Bioeth* 2016; **16**: 45–54.
- 123** Lessem E, Bernardo J, Reed C *et al.* Informed use of bedaquiline for tuberculosis. *Lancet* 2015; **385**: 1724.

- 124** Mingote LR, Namutamba D, Apina F *et al.* The use of bedaquiline in regimens to treat drug-resistant and drug-susceptible tuberculosis: a perspective from tuberculosis-affected communities. *Lancet* 2015; **385**: 477–9.
- 125** Bélard S, Heuvelings CC, Janssen S *et al.* Bedaquiline for the treatment of drug-resistant tuberculosis. *Expert Rev Anti Infect Ther* 2015; **13**: 535–53.
- 126** Conradie F, Meintjes G, Hughes J *et al.* Clinical access to bedaquiline programme for the treatment of drug-resistant tuberculosis. *S Afr Med J* 2014; **104**: 164–6.
- 127** Patel RV, Riyaz SD, Park SW. Bedaquiline: a new hope to treat multidrug-resistant tuberculosis. *Curr Top Med Chem* 2014; **14**: 1866–74.
- 128** Udwardia ZF, Amale RA, Mullerpattan JB. Initial experience of bedaquiline use in a series of drug-resistant tuberculosis patients from India. *Int J Tuberc Lung Dis* 2014; **18**: 1315–8.
- 129** Furin J, Brigden G, Lessem E *et al.* Global progress and challenges in implementing new medications for treating multidrug-resistant tuberculosis. *Emerg Infect Dis* 2016; **22**: e151430.
- 130** Andries K, Verhasselt P, Guillemont J *et al.* A diarylquinoline drug active on the ATP synthase of *Mycobacterium tuberculosis*. *Science* 2005; **307**: 223–7
- 131** Cole ST, Alzari PM. Microbiology. TB – a new target, a new drug. *Science* 2005; **307**: 214–5.
- 132** Jain PP, Degani MS, Raju A *et al.* Identification of a novel class of quinolone-oxadiazole hybrids as anti-tuberculosis agents. *Bioorg Med Chem Lett* 2016; **26**: 645–9.
- 133** Qiao CJ, Wang XK, Xie F *et al.* Asymmetric synthesis and absolute configuration assignment of a new type of bedaquiline analogue. *Molecules* 2015; **20**: 22272–85.

- 134** Balemans W, Vranckx L, Lounis N *et al.* Novel antibiotics targeting respiratory ATP synthesis in Gram-positive pathogenic bacteria. *Antimicrob Agents Chemother* 2012; **56**: 4131–9.
- 135** Jain PP, Degani MS, Raju A *et al.* Rational drug design based synthesis of novel arylquinolines as anti-tuberculosis agents. *Bioorg Med Chem Lett* 2013; **23**: 6097–105.
- 136** Gelber R, Andries K, Paredes RM *et al.* The diarylquinoline R207910 is bactericidal against *Mycobacterium leprae* in mice at low dose and administered intermittently. *Antimicrob Agents Chemother* 2009; **53**: 3989–91.
- 137** Raju RM, Raju SM, Zhao Y *et al.* Leveraging advances in tuberculosis diagnosis and treatment to address nontuberculous mycobacterial disease. *Emerg Infect Dis* 2016; **22**: 365–9.
- 138** Wang H, Zhang X, Bai Y *et al.* Comparative efficacy and acceptability of five anti-tubercular drugs in treatment of multidrug resistant tuberculosis: a network meta-analysis. *J Clin Bioinforma* 2015; **5**: 5. doi: 10.1186/s13336-015-0020.
- 139** Segala E, Sougakoff W, Nevejans-Chauffour A *et al.* New mutations in the mycobacterial ATP synthase: new insights into the binding of the diarylquinoline TMC207 to the ATP synthase C-ring structure. *Antimicrob Agents Chemother* 2012; **56**: 2326–34.
- 140** Gold B, Roberts J, Ling Y *et al.* Rapid, semiquantitative assay to discriminate among compounds with activity against replicating or nonreplicating *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2015; **59**: 6521–38.
- 141** Reddy VM, Einck L, Andries K *et al.* *In vitro* interactions between new antitubercular drug candidates SQ109 and TMC207. *Antimicrob Agents Chemother* 2010; **54**: 2840–6.
- 142** Field SK. Bedaquiline for the treatment of multidrug-resistant tuberculosis: great promise or disappointment? *Ther Adv Chronic Dis* 2015; **6**: 170–84.

- 143** Tasneen R, Betoudji F, Tyagi S *et al.* Contribution of oxazolidinones to the efficacy of novel regimens containing bedaquiline and pretomanid in a mouse model of tuberculosis. *Antimicrob Agents Chemother* 2015; **60**: 270–7.
- 144** Makarov V, Lechartier B, Zhang M *et al.* Towards a new combination therapy for tuberculosis with next generation benzothiazinones. *EMBO Mol Med* 2014; **6**: 372–83.
- 145** Tasneen R, Li SY, Peloquin CA *et al.* Sterilizing activity of novel TMC207- and PA-824-containing regimens in a murine model of tuberculosis. *Antimicrob Agents Chemother* 2011; **55**: 5485–92.
- 146** Wallis RS, Jakubiec W, Mitton-Fry M *et al.* Rapid evaluation in whole blood culture of regimens for XDR-TB containing PNU-100480 (sutezolid), TMC207, PA-824, SQ109, and pyrazinamide. *PLoS One* 2012; **7**: e30479.
- 147** Ibrahim M, Truffot-Pernot C, Andries K *et al.* Sterilizing activity of R2079-10 (TMC207)-containing regimens in the murine model of tuberculosis. *Am J Respir Crit Care Med* 2009; **180**: 553–7.
- 148** Dhillon J, Andries K, Phillips PJ *et al.* Bactericidal activity of the diarylquinoline TMC207 against *Mycobacterium tuberculosis* outside and within cells. *Tuberculosis* 2010; **90**: 301–5.
- 149** Diacon AH, Dawson R, von Groote-Bidlingmaier F *et al.* Bactericidal activity of pyrazinamide and clofazimine alone and in combination with pretomanid and bedaquiline. *Am J Respir Crit Care Med* 2015; **191**: 943–53.
- 150** Leibert E, Danckers M, Rom WN. New drugs to treat multidrug-resistant tuberculosis: the case for bedaquiline. *Ther Clin Risk Manag* 2014; **10**: 597–602.
- 151** Vocat A, Hartkoorn RC, Lechartier B *et al.* Bioluminescence for assessing drug potency against nonreplicating *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2015; **59**: 4012–9.

- 152** Irwin SM, Prideaux B, Lyon ER *et al.* Bedaquiline and pyrazinamide treatment responses are affected by pulmonary lesion heterogeneity in *Mycobacterium tuberculosis* infected C3HeB/FeJ mice. *ACS Infect Dis* 2016; **2**: 251-67.
- 153** Maloney PC, Kashket ER, Wilson TH. A proton motive force drives ATP synthesis in bacteria. *Proc Natl Acad Sci U S A* 1974; **71**: 3896–900.
- 154** Hicks DB, Liu J, Fujisawa M *et al.* F₁F₀-ATP synthases of alkaliphilic bacteria: lessons from their adaptations. *Biochim Biophys Acta* 2010; **1797**: 1362–77.
- 155** Goulooze SC, Cohen AF, Rissmann R. Bedaquiline. *Br J Clin Pharmacol* 2015; **80**: 182–4.
- 156** de Jonge MR, Koymans LH, Guillemont JE *et al.* A computational model of the inhibition of *Mycobacterium tuberculosis* ATPase by a new drug candidate R207910. *Proteins* 2007; **67**: 971–80.
- 157** Koul A, Dendouga N, Vergauwen K *et al.* Diarylquinolines target subunit c of mycobacterial ATP synthase. *Nat Chem Biol* 2007; **3**: 323–4.
- 158** Preiss L, Langer JD, Yildiz Ö *et al.* Structure of the mycobacterial ATP synthase Fo rotor ring in complex with the anti-TB drug bedaquiline. *Sci Adv* 2015; **1**: e1500106.
- 159** Koul A, Vranckx L, Dhar N *et al.* Delayed bactericidal response of *Mycobacterium tuberculosis* to bedaquiline involves remodelling of bacterial metabolism. *Nat Commun* 2014; **5**: 3369.
- 160** Hards K, Robson JR, Berney M *et al.* Bactericidal mode of action of bedaquiline. *J Antimicrob Chemother* 2015; **70**: 2028–37.

- 161** Berney M, Hartman TE, Jacobs WR Jr. A *Mycobacterium tuberculosis* cytochrome *bd* oxidase mutant is hypersensitive to bedaquiline. *MBio* 2014; **5**: e01275-14.
- 162** Lamprecht DA, Finin PM, Rahman MA *et al.* Turning the respiratory flexibility of *Mycobacterium tuberculosis* against itself. *Nat Commun* 2016; **7**:12393: doi: 10.1038.
- 163** Pethe K, Bifani P, Jang J *et al.* Discovery of Q203, a potent clinical candidate for the treatment of tuberculosis. *Nat Med* 2013; **19**: 1157–60.
- 164** van Rensburg CEJ, Anderson R, O'Sullivan JF. Riminophenazine compounds: pharmacology and antineoplastic potential. *Crit Rev Oncol Hematol* 1997; **25**:55-67.
- 165** Fiorillo M, Lamb R, Tanawitz HB *et al.* Bedaquiline, an FDA-approved antibiotic inhibits mitochondrial function and potently blocks the proliferative expansion of stem-like cancer cells (CSCs). *Aging*; **8**: 1-15.
- 166** Zaccagnino A, Managò A, Leanza L *et al.* Tumor-reducing effect of the clinically used drug clofazimine in a SCID mouse model of pancreatic ductal adenocarcinoma. *Oncotarget* 2016; doi: 10.18632/oncotarget.11299. 1-18.
- 167** Diacon AH, Pym A, Grobusch M *et al.* The diarylquinoline TMC207 for multidrug-resistant tuberculosis. *N Engl J Med* 2009; **360**: 2397–405.
- 168** Diacon AH, Dawson R, Van Groote-Bidlingmaier F *et al.* 14-day bactericidal activity of PA-824, bedaquiline, pyrazinamide and moxifloxacin combinations: a randomised trial. *Lancet* 2012; **380**: 986–93.
- 169** Ndjeka N, Conradie F, Schnippel K *et al.* Treatment of drug-resistant tuberculosis with bedaquiline in a high HIV prevalence setting: an interim cohort analysis. *Int J Tuberc Lung Dis* 2015; **19**: 979–85.

- 170** Svensson EM, Dooley KE, Karlsson MO. Impact of lopinavir-ritonavir or nevirapine on bedaquiline exposures and potential implications for patients with tuberculosis-HIV coinfection. *Antimicrob Agents Chemother* 2014; **58**: 6406–12.
- 171** Pandie M, Wiesner L, McIlleron H *et al.* Drug-drug interactions between bedaquiline and the antiretrovirals lopinavir/ritonavir and nevirapine in HIV-infected patients with drug-resistant TB. *J Antimicrob Chemother* 2016; **71**: 1037–40.
- 172** Hervé C, Bergot E, Veziris N *et al.* Tuberculosis in 2015: from diagnosis to the detection of multi-resistant cases. *Rev Mal Respir* 2015; **32**: 784–90.
- 173** South African Department of Health. Introduction of new drugs and drug regimens for the management of drug resistant tuberculosis in South Africa: Policy Framework Version 1.1: June 2015. <http://www.nicd.ac.za/assets/files/document.pdf>
- 174** Torrea G, Coeck N, Desmaretz C *et al.* Bedaquiline susceptibility testing of *Mycobacterium tuberculosis* in an automated liquid culture system. *J Antimicrob Chemother* 2015; **70**: 2300–5.
- 175** Keller PM, Hönke R, Ritter C *et al.* Determination of MIC distribution and epidemiological cut off values for bedaquiline and delamanid in *Mycobacterium tuberculosis* using the MGIT 960 system equipped with TB eXiST. *Antimicrobial Agents Chemother* 2015; **59**: 4352–5.
- 176** Kakkar AK, Dahiya N. Bedaquiline for the treatment of resistant tuberculosis: promises and pitfalls. *Tuberculosis* 2014; **94**: 357–62.
- 177** Huitric E, Verhasselt P, Koul A *et al.* Rates and mechanisms of resistance development in *Mycobacterium tuberculosis* to a novel diarylquinoline ATP synthase inhibitor. *Antimicrob Agents Chemother* 2010; **54**: 1022–8.

- 178** Grossman TH, Shoen CM, Jones SM *et al.* The efflux pump inhibitor timcodar improves the potency of antimycobacterial agents. *Antimicrob Agents Chemother* 2015; **59**: 1534–41.
- 179** Srikrishna G, Gupta S, Dooley KE *et al.* Can the addition of verapamil to bedaquiline-containing regimens improve tuberculosis treatment outcomes? A novel approach to optimizing TB treatment. *Future Microbiol* 2015; **10**: 1257–60.
- 180** Petrella S, Cambau E, Chauffour A *et al.* Genetic basis for natural and acquired resistance to the diarylquinoline R207910 in mycobacteria. *Antimicrob Agents Chemother* 2006; **50**: 2853–6.
- 181** Mitchison D, Davies G. The chemotherapy of tuberculosis: past, present and future. *Int J Tuberc Lung Dis* 2012; **16**: 724–32.
- 182** Gideon HP, Phuah JY, Myers AJ *et al.* Variability in tuberculosis granuloma T cell responses exists, but a balance of pro- and anti-inflammatory cytokines is associated with sterilization. *PLoS Pathog* 2015; **11**: e1004603.
- 183** Alffenaar JW, Bolhuis M, van Hateren K *et al.* Determination of bedaquiline in human serum using liquid chromatography-tandem mass spectrometry. *Antimicrob Agents Chemother* 2015; **59**: 5675–80.
- 184** Akkerman O, Odish OF, Bolhuis MS *et al.* Pharmacokinetics of bedaquiline in cerebrospinal fluid and serum in multidrug-resistant tuberculous meningitis. *Clin Infect Dis* 2016; **62**: 523–4.
- 185** Nau R, Sörgel F, Eiffert H. Penetration of drugs through the blood-cerebrospinal fluid/blood-brain barrier for treatment of central nervous system infections. *Clin Microbiol Rev* 2010; **23**: 858–83.
- 186** Winter H, Egizi E, Murray S *et al.* Evaluation of the pharmacokinetic interaction between repeated doses of rifapentine or rifampin and a single dose of bedaquiline in healthy adult subjects. *Antimicrob Agents Chemother* 2015; **59**: 1219–24.

187 Kwon YS, Koh WJ. Synthetic investigational new drugs for the treatment of tuberculosis. *Expert Opin Investig Drugs* 2016; **25**: 183–93.

188 Pontali E, Sotgiu G, D'Ambrosio L *et al.* Bedaquiline and multidrug-resistant tuberculosis: a systematic and critical analysis of the evidence. *Eur Respir J* 2016; **47**: 394–402.