RESPONSE OF GROUND-DWELLING SPIDER ASSEMBLAGES

(ARACHNIDA, ARANEAE) TO MONTANE GRASSLAND MANAGEMENT

PRACTICES IN SOUTH AFRICA

RAYMOND JANSEN<sup>a</sup>, LUKHANYO MAKAKA<sup>1</sup>, IAN T. LITTLE<sup>2</sup> and

ANSIE DIPPENAAR-SCHOEMAN3

<sup>1</sup>Department of Environmental, Water and Earth Sciences, Tshwane University of Technology, Private

Bag X680, Pretoria, South Africa

<sup>2</sup>Percy FitzPatrick Institute of African Ornithology, DST/NRF Centre of Excellence, University of Cape

Town, Rondebosch, South Africa / Endangered Wildlife Trust, Private Bag X11, Modderfontein,

Johannesburg

1645, South Africa

<sup>3</sup>Agricultural Research Council (ARC), Plant Protection Research Institute (PPRI), Private Bag X134,

Queenswood 0121 / Department of Zoology and Entomology, University of Pretoria, Pretoria 0002, South

Africa

Corresponding author:

Raymond Jansen

Tel: +27 12 382 6347

Fax: +27 12 382 6345

e-mail: jansenr@tut.ac.za

1

**ABSTRACT.** 1. Frequent and extensive burning practices coupled with intensive grazing management are known to impact negatively on the vegetation diversity of grassland ecosystems. Few studies have investigated the impacts on spider diversity and community structure as a result of these management practices, and no studies have been conducted in high mountain grasslands on how these spider assemblages are influenced by this form of management.

- 2. Here, we present the results of a study conducted in the Mpumalanga grasslands on the eastern escarpment of South Africa. Ground-dwelling spiders were sampled in the summer season from 180 pit-fall traps in five study sites that varied from either being burnt annually and grazed heavily, burnt biennially and conservatively grazed, to communal land with no set management practice. Variations between sites were assessed and based on spider species composition and assemblage structure.
- 3. A total of 1145 individuals were collected representing 86 species from 60 genera and 43 families. Our results show that a majority of genera in these grasslands were represented by very few individuals, where a total of 37 species were represented by singletons and 17 species that were doubletons. The most abundant families were the Lycosidae (64.3%), Gnaphosidae (9.0%), Zodariidae (5.3%), Linyphiidae (4.7%) and Salticidae (3.1%).
- 4. Grazing intensity and fire frequency had no measurable effect on ground-dwelling spider abundance diversity or assemblage structure. Only when rare or single species occurrence was included, was there some form of association with sites. This study has provided for the first preliminary inventory of ground-dwelling spiders for this habitat.

Keywords. Spider assemblage; grassland management; burning; grazing

### Introduction

The Highveld grasslands of the Mpumalanga Province are endemic to the province and are highly threatened by human transformation primarily from agriculture, mining, afforestation and urbanization, which have resulted in the extensive fragmentation of these grasslands (Armstrong and van Hensbergen, 1999; Morris, 2001; Wessels et al., 2003). The North-eastern Mountain Grassland Biome, which encompasses a total area of 42 458 km<sup>2</sup> (Bredenkamp et al., 1996), for instance, has been transformed mainly for agricultural and forestry purposes (Bredenkamp et al., 1996; Morris, 2001). Mucina et al. (2006) mention that only 2.4% of Lydenberg Montane Grassland in Mpumalanga Province (North-eastern Sandy Highveld Grassland: Acocks 1988 or North-eastern Mountain Grassland: Bredenkamp et al., 1996) is formally conserved in protected areas. This 2.4% falls short of the IUCN's nominal recommendation of 10% protected area coverage for a biome (Rouget et al., 2006). In addition, this habitat is considered vulnerable in terms of its conservation status and has been identified as a priority habitat for conservation (SANBI, 2008). Within this habitat, there is also a high level of transformation, which is estimated at 45% by Bredenkamp et al. (1996), 55% by Morris (2001) and 23% by Mucina et al. (2006), with forestry alone contributing 20% of the transformation (Mucina et al., 2006). However, the extent of this degradation may be underestimated, as more subtle impacts that are often not as obvious, such as extensive and intensive grazing, in addition to frequent burning practices, may be overlooked. No studies to date have been undertaken on the changes in spider

communities within these grasslands and how varying forms of grassland management may impact upon their assemblage structure.

Although the Araneae constitute an abundant and highly successful group of invertebrates, little is still known of their diversity within most ecosystems in South Africa (Dippenaar-Schoeman and Wassenaar, 2002). Since many invertebrate taxa are undescribed, our knowledge of the geographical ranges of these animals is virtually non-existent (Foord, *et al.*, 2002). Indeed, research on spider communities within South African grasslands is lacking where the status of these communities and the presence of individual species is unknown and a large number of species are consistently still being described, e.g. Haddad & Wesolowska (2011) recently described a number of new jumping spiders from the grasslands of central South Africa.

Although spiders can be considered a ubiquitous component of invertebrate assemblages and important generalist predators in ecosystems (Wise, 1993), they are also known to be sensitive to fine-scale changes in environmental factors (Foord *et al.*, 2008). Even though spider populations may be high, even within disturbed habitats, spider communities are strongly influenced by most aspects of land management such as grazing and the frequency of burning within grassland ecosystems. As such, the structure of spider communities or the component of species that make up assemblages of spiders may be a useful tool for monitoring the effects of land use change within particular ecosystems, as spiders respond directly to prey abundance and the structure of vegetation (see Bell, *et al.* 2001 for a review). As such, they may prove to be a valuable group of organisms to indicate ecosystem health; particularly ecosystems such as South Africa's moist highland grasslands that are currently under severe threat.

Studies have indicated that the presence or absence of spider species may be related to the subtle changes in the vegetation structure as a result of grazing. Churchill and Ludwig (2004) mention that livestock grazing changes habitat structure and inhibits ecological succession in the Australian savanna. Dennis (2003) indicated that arthropod diversity can be altered by livestock grazing through changes in plant community composition and soil physical properties in Scotland. The effect of grazing management on spider communities is well documented in European pastures where grazing has been recognized as a form of disturbance which at low to moderate intensities results in high structural heterogeneity in the habitat (Ausden, et al., 2005; Dupre and Diekmann, 2001; Hodgson, 1986). Therefore, a light to moderate grazing intensity should effectively promote spider community conservation. Dennis et al. (2001) suggested that there was higher species richness and abundance of spiders in pastures grazed only by sheep than by both sheep and cattle. However, Gibson et al. (1992b) mentions that increased grazing in general reduces overall spider species richness. While studying the influence of grazing on the spider fauna in the American grasslands, Deltschev and Kajak (1974) found that intensive grazing had a highly detrimental affect on the spider assemblage of the former vegetation. In Africa, intensive grazing by domestic livestock has also been shown to alter savanna vegetation cover and significantly reduce the diversity of spider fauna (Warui et al., 2005). Furthermore, Abensperg-Traun et al. (1996) also recorded spider diversity as being strongly influenced by sheep grazing in Australia, stating that intense grazing had an adverse effect on the primitive arachnid sub-order Mygalomorphae, while moderate grazing favored wolf spiders (Family: Lycosidae) and those species belonging to the Idiopidae, known as brown trap-door

spiders. Churchill (1998) further recorded the Family Zodariidae as being strongly affected by grazing in the Mitchell Grasslands of the Northern Territory of Australia. However, no changes in ground-dwelling spider assemblage structure or species composition was reported in different grazing regimes within Hungarian pastures (Batary *et al.*, 2008; Samu *et al.*, 2010; Szinetár and Samu, 2012) or in Dutch grasslands (Kleijn, *et al.*, 2006). Some spider species may require specific features of the microclimate or structure which only exist in grazed grasslands (e.g. Thomas,1983), or even in grasslands with a mosaic structure, to provide conditions for different phases of their life cycles (Cherrill and Brown, 1990a and b; Weidemann *et al.*, 1990). It is clear that grazing with cattle or other large mammalian herbivores reduces relative vegetation cover and may thus influence spider diversity, abundance and community structure. However, this has not been investigated in the moist highland grassland of South Africa.

This study is the first attempt to document the spider community composition within South Africa's moist highland grasslands in Mpumalanga Province. Furthermore, this study will investigate whether spider assemblage structure differs between fine-scale grassland management practices and if particular assemblages can act as indicators of grassland ecosystem health, as spiders are known to be sensitive to fine-scale changes in environmental conditions (Foord *et al.*, 2008). In addition, spiders were selected because they are diverse, relatively easily collected, are functionally significant in ecosystems as predators and as food for higher predators, and interact with their abiotic and biotic environments in a manner that reflects ecological change (Churchill, 1997; Haddad *et al.*, 2010). In addition, the results of this study will contribute to the

South African National Survey on Arachnida (SANSA) database by inventorying spiders and by measuring species richness and abundance within these grasslands.

#### Materials and methods

This study was conducted in the moist highland grasslands in the Dullstroom district (25° 42' 846" S and 30° 09' 818" E), located between the towns of Belfast and Lydenburg (now called Mashishing) in Mpumalanga Province, Republic of South Africa (Fig. 1). The vegetation of the study area is classified as North-eastern Mountain Grassland by Bredenkamp et al. (1996) or Lydenburg Montane Grassland by Mucina et al. (2006) or North-eastern Mountain Sourveld by Acocks (1988). The region is characterized by a high rainfall in summer (Bredenkamp et al., 1996) and mist through most months of the year, with a mean annual rainfall ranging between 660-1180 mm per annum (Mucina et al., 2006). Temperature variation in the region is from below 0 °C in winter to 39 °C in summer, with an average of 15 °C per annum (Bredenkamp et al., 1996). The soils in this region are derived from shale and quartzite (Mucina et al., 2006) and are usually shallow (Bredenkamp et al., 1996). The Dullstroom district is situated within an altitudinal range of 1260-2160 m above sea level with undulating plains, mountain peaks and slopes, hills and valleys with the low lying slopes usually supporting grass growth (Mucina et al., 2006). The study was completed in areas over 1900 m above sea level.

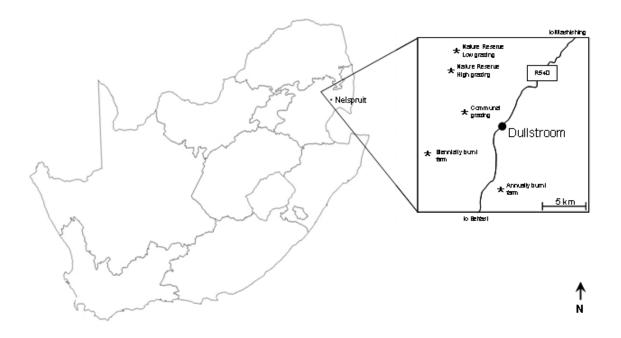


Fig. 1: Map of study area. The inset shows the locations of the study sites and their associated management regimes

# Study sites

The most dominant land use in the study area is livestock farming, primarily with cattle in the commercial farms and mixed herds of cattle, sheep and goats within the communal farms. In examining the impacts of burning and grazing on these highland grasslands, different grassland management practices were chosen that took into account landscape heterogeneity and farm management practices that included burning frequency, the proportional mix of herds and stocking rates. Five study sites were selected based on the farmer's willingness to allow the study to be undertaken on their property and that it reflected the common farming practices in the study area. Relative grazing intensity was defined as the amount of hectares of grazing land

available per large animal unit (ha/LAU). One large animal unit was estimated to be equivalent to one cow or five sheep or goats and represents the metabolic equivalent of a 454 kg cow (Owen-Smith and Danckwerts, 1997). Heavy, moderate and low grazing intensity were assumed to be a stocking rate of 1 LAU on 3 ha or less, 1 LAU on 4 - 10 ha and 1 LAU on 11 - 25 ha, respectively. The sites consisted of commercial farms that were stocked mainly with cattle at a heavy grazing intensity and burned annually (Site 1: Annual farm/AF), and biennially or every second year (Site 2: Biennial farm/BF), and a communal land (Sakhelwe Township) with no planned burning regime and holding a mixed proportion of cattle, goats and sheep at a high grazing intensity (Site 3: Communal Land/Com). Two other study sites were selected within an 8000 hectare (ha) conservation area managed by the provincial government conservation body (Mpumalanga Parks and Tourism Authority); Verloren Valei Nature Reserve. This area is grazed by indigenous herbivores stocked at a very low grazing intensity (Site 4: reserve Camp 3/V3) and another with free roaming mix proportion of indigenous ungulates at a moderate grazing intensity (Site 5: reserve Camp 7/V7). Verloren Valei Nature Reserve is an international RAMSAR site recognizing its importance for intact wetland conservation. Four of the five study sites were last burnt during the spring of 2008, except the communal land which was partially burnt (unplanned burning) in that year. The management practices of all the sites chosen for this study have been the same since 1983, with the exception of the communal lands that had no proper management records but is assumed to have varied little in the past 30 years. The study sites and their grazing intensity and ungulates species are presented in Appendix Α.

Spiders were sampled over the summer months from the onset of November to the end of February. Within each site, four 25 hectares (ha) replicates were marked out with at least 500 m between replicates. Sampling sites were selected with sufficient distance between sites to avoid pseudo-replication (Hurlbert 1984), but were sufficiently close together to standardize as far as possible for extrinsic factors including soil type, rainfall, aspect, slope and temperature. In each of the replicates nine pittraps were laid in a 20 m diameter nested cross array with one central trap and with the two outside traps being only 5 m apart respectively (Fig. 2). Each trap was made up of a 10 cm diameter and 11 cm deep cup. This cup was fitted into a permanent sleeve that was sunk into the soil, with the sleeve lip flush with the soil surface. Trap covers were made up of zinc lids which were just 4 cm above the lip of the trap. These covers help with keeping out rain water and any falling debris (Mallis and Hurd, 2005). Sampling was done in three day intervals and in each sampling period 2 cm of a 70% ethylene glycol preservative was poured into each trap and the samples collected after three days. The timing of setting traps was weather dependant, based on rain avoidance.

Spiders are broadly grouped into web-builders and active hunters. The active hunters can further be divided into plant wanderers and ground wanderers (Haddad *et al.*, 2010). As pitfall sampling was used, only ground wandering spiders were sampled and identified in this study. Due to the large number of immature specimens collected and the unresolved taxonomy of many families (such as the Linyphiidae), some specimens could only be identified to genus level and are referred to as morphospecies

where necessary (Haddad *et al.*, 2010). All arachnids sampled were sorted and identified into morphospecies using the taxonomic keys in Dippenaar-Schoeman and Jocqué (1997). Data collected forms part of the South African National Survey of Arachnida (SANSA) and voucher specimens are housed in the National Collection of Arachnida (NCA) at the Plant Protection Research Institute of the Agricultural Research Council Pretoria.

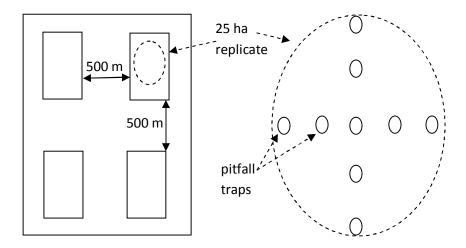


Fig. 2. Study site sample design indicating the four 25 ha replicates within each study site. Within each of these replicates is the nested cross array of nine pitfall traps

# Vegetation Analysis

Vegetation density: Vegetation density measurements were conducted on each site within each replicate at the beginning of each month. A disc pasture meter, described by Bransby and Tainton (1977), was used. The disc pasture meter consists of three main parts: a central aluminium rod, an aluminium sleeve that slides freely in the central rod and an aluminium disc that is attached to the base of the aluminium sleeve.

The central rod is marked at 0.5 cm interval starting from 0-60. A 500 m transect was set thrice on each replicate. In order to reduce the influence of edge effect, transects were laid 250 m from the edge of the replicate. The second transect was 150 m away from the first transect and the third was 200 m from the second transect. At 5 m intervals along each of the three 500 m long transects, the central rod of the disc pasture meter was held perpendicular to the ground surface and the sleeve with the attached disc released onto the sward from the upper end, thus the 60 cm mark. The settling height of the disc was then recorded yielding 100 readings per transect and a total of 300 readings per replicate. The numbers of rocky samples recorded at each 5 m along the 500 m transect were noted but eliminated from the analysis since it was considered not to be an indication of grazing pressure.

Vegetation percentage cover: A Modified-Whittaker design was established in all four replicates of each site to measure plant species cover (Stohlgren *et al.*, 1995). The Modified-Whittaker sampling design involves a 1,000 m<sup>2</sup> plot with nested subplots consisting of one 100 m<sup>2</sup>, two 10 m<sup>2</sup>, and ten 1 m<sup>2</sup> subplots. The 1 m<sup>2</sup> plots were surveyed with 1 m<sup>2</sup> frames that were nested at 0.5 m<sup>2</sup> to increase the accuracy of visual cover measurement. Plant species cover in each subplot was visually estimated using a cover class to represent the different percentage range for cover. The cover class and percentage range of coverage are presented in Table 1.

Table 1: Cover classes used when calculating plant vegetation cover

Cover class	Range of coverage (%) Mi	idpoint range (%)
1	<5	2.5
2	5-10	7.5
3	10-20	15
4	20-30	25
5	30-40	35
6	40-50	45
7	50-60	55
8	60-70	65
9	70-80	75
10	80-100	90

## Data Analysis

The number of spiders from each replicate within each site was pooled for that site to compare spider numbers and diversity between sites. ANOVA assumes that the data are sampled from populations with identical standard deviations. We tested this assumption using the method of Bartlett (Snedecor *et al.*, 1989). Bartlett statistic (corrected) = 85.012, the *P* value is < 0.0001. As such, Bartlett's test suggests that the differences among the standard deviations between sites is significant. We then Log<sub>10</sub> transformed the spider abundance data and performed a Kruskal-Wallis one-way analyses of variance (nonparametric ANOVA) to test if there were any change in spider abundance between sites. The Statistica statistical software package was used to perform these analyses (StatSoft version 6, 2009). Shannon-Wiener Index of diversity was calculated for each site and compared between habitats using ANOVA. Simple

linear (Pearson) correlation analysis was undertaken to investigate the relationship between the spider presence and abundance, vegetation density and vegetation cover.

Multivariate data analysis was conducted between all study sites using the statistical program PC-ORD (McCune and Mefford, 2006: PC-ORD 5.10) to note any significant difference between sites in terms of spider assemblage structure. Firstly, a presence/absence matrix of all species across all sites at all habitats was created. A Bray-Curtis similarity matrix (Bray and Curtis, 1957) was generated using the presence/absence transformation. Thereafter, three analyses were performed from the resulting matrix.

Firstly, a cluster analyses of the Bray-Curtis matrix using group average cluster option was performed. The percentage difference between sites calculated by the cluster program allows a two dimensional representation of how closely linked sites are to one another based on the presence of spider species at each study site.

Secondly, a non-metric multi-dimensional scaling (MDS) ordination (using Sørensen coefficient also known as the Czekanowski or Bray-Curtis coefficient distance measure) that constructs a map of the samples based on the underlying similarity matrix was undertaken; this was plotted as an MDS plot which provides a representation of the overall similarity among the sites across many dimensions.

An indicator species analyses was calculated that assesses species contributions/responses to management type analysis using a Monte Carlo test of significance with 5000 permutations (Dufrêne and Legendre, 1997). This method combines species abundance and occurrence. A species' uniqueness to a particular habitat and its frequency of being present in a particular habitat is expressed as a

percentage in comparison with other species in sampled habitats (Dufrêne and Legendre, 1997). A suitable percentage should be above 70% (Haddad *et al.*, 2010; McGeoch *et al.*, 2002; Van Rensberg *et al.*, 1999). All analyses were then repeated but with singletons removed.

## Results

A total of 1145 individuals were collected of which 558 were adults. The spiders caught represent 86 species from 60 genera and 43 families (Appendix B). The most abundant families were the Lycosidae (64.3%), Gnaphosidae (9.0%), Zodariidae (5.3%), Linyphiidae (4.7%), and Salticidae (3.1%) (Table 2; Appendix D). The most abundant species were an undetermined Lycosidae sp. 1 (Lycosidae, 15.9%), Lycosidae sp. 2 (Lycosidae, 15.2%) and a *Proevippa* species (Lycosidae, 12.7%) (Appendix C and D). These species constituted more than 40% of the total abundance.

Table 2: Family composition of ground dwelling spiders collected from all five study sites.

Species indicated with \* are plant dwellers that accidentally landed in pittraps

Family	Total_collected	% of Total	Total Species	% of Total
Amaurobiidae	1	0.09	1	1.16
Araneidae*	2	0.17	2	2.32
Clubionidae*	3	0.26	1	1.16
Corinnidae	27	2.34	8	9.3
Cyrtaucheniidae	5	0.44	2	2.32
Gnaphosidae	102	8.98	9	10.4
Hahniidae	1	0.09	1	1.16
Idiopidae	5	0.44	2	2.32

Linyphiidae	53	4.7	11	12.7
Liocranidae	29	2.53	1	1.16
Lycosidae	737	64.3	13	15.12
Nemesiidae	3	0.26	1	1.16
Orsolobiidae	1	0.09	1	1.16
Palpimanidae	19	1.66	1	1.16
Philodromidae	6	0.52	4	4.65
Phyxelididae	2	0.17	1	1.16
Prodidomidae	1	0.09	1	1.16
Salticidae	35	3.05	5	5.81
Scytodidae	2	0.17	1	1.16
Selenopidae	1	0.09	1	1.16
Sparassidae	3	0.26	1	1.16
Tetragnathidae*	2	0.17	2	2.32
Theraphosidae	2	0.17	2	2.32
Theridiidae	19	1.66	5	5.81
Thomisidae	23	2.01	4	4.65
Zodariidae	61	5.33	5	5.81
Total:	1145		86	

Most species were represented by only a few individuals (C). A total of 37 species were represented by singletons and 17 species by doubletons. The two reserve sites represented most of the species recorded and the greatest abundance per species in addition to having 14 spider species exclusively found on the reserve (Appendix C). However, in the camp where grazing pressure was increased (camp 7) spider number and diversity dropped (Table 3). Surprisingly, the study site burnt annually also held a high number of species. The farms burnt biennially, in addition to the communal lands, held the lowest abundance per species. An abundance curve for all study sites indicates that spider numbers were low following burning events in early summer, but recovered towards the end of summer (Fig. 3).

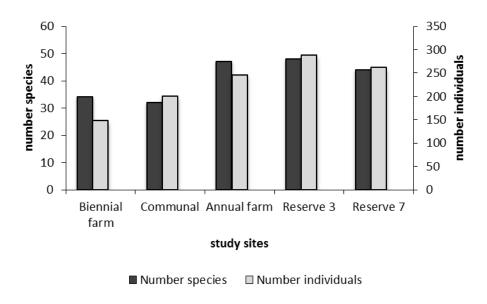


Fig. 3: Number of spider species and total number of individuals recorded within the five study sites

Table 3: Species diversity analyses of the spider communities between the five study sites

Study site	N	Sp	S	E	Н	D	SD
Annual farm	246	47	47	0.775	2.98	0.91	±7.3
Biennial farm	148	34	34	0.819	2.89	0.91	±4.5
Communal	201	32	32	0.769	2.66	0.88	±7.1
Reserve 3	288	48	48	0.675	2.62	0.83	±12.4
Reserve 7	262	44	44	0.752	2.85	0.89	±8.7

N = Total number of individual

Sp = Total number of species

S = Richness = number of non-zero elements in row

E = Evenness = H / In (Richness)

H = Diversity = - sum (Pi\*In(Pi)) = Shannon's diversity index

D = Simpson's diversity index for infinite population = 1 - sum (Pi\*Pi),

where Pi = importance probability in element i (element I relativized by row total)

SD = Standard Deviation on the mean number of individuals per species

No significant difference was recorded between sites based on individual species present or total number of individuals within each species located (Kruskal-Wallis oneway ANOVA: H = 5771, d.f. = 4, chi-square probability = 0.146, P > 0.05). Diversity

estimates indicated no clear differences between sites based on species richness, species evenness and species diversity measures (Table 3).

Cluster analyses was performed using Sørensen distance measure rather than Euclidean, as Sørensen gives less weight to outliers or singletons of which this study had numerous. The analyses was also performed after removing all singletons but the results remained the same. Cluster analysis based on spider species presence and absence (Fig. 4), grouped the communal land with the reserve's camp 7 and then to the reserve's camp 3. Both the reserve camp 3 and the communal lands have mixed stocking rates and are not burnt every year. There is a noticeable difference between farms that are burnt annually and heavily grazed than to grasslands that are burnt every two years and grazed moderately. Again, the study site burnt biennially grouped separately.

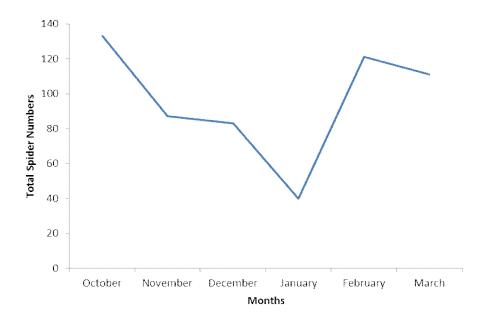


Fig. 4. Spider abundance curve for all study sites during the study period

According to the MDS ordination analysis based on spider species diversity on all the sites investigated (Fig. 5), the results were similar to the cluster analyses in that reserve camp 7 was more closely linked with farms burnt annually and with communal lands than to reserve camp 3 and the farm burnt biennially. Farms burnt every two years separate apart from any association with other study sites. Those habitats exposed to more intense grazing, such as the communal lands and the farm burnt annually, cluster closer to one another.

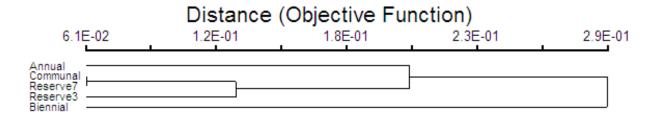


Fig. 5: Bray Curtis cluster analysis indicating similarity between sites based on spider species presence and absence (this includes the presence of the species alone)

A two-way cluster dendrogram analysis (also using Sørensen distance measure) was performed firstly including all species (86 species, Fig. 6) and then again, but removing single species occurrence (44 species included, Fig. 7). When all species including singletons were analysed, certain groups of assemblages were evident (Fig. 6). However, on removing these singletons, the evidence of assemblages relating to particular habitats fell away and only the biennial farm and annual farm had a few species associated with these habitats. The indicator species analyses revealed a number of species that can be regarded as potential indicators for specific grassland

habitats (Table 4), however, after performing a MONTE CARLO test of significance of observed maximum indicator value for species, these species were not found to be significant. The large indicator values are probably influenced by the abundance of certain dominant species.

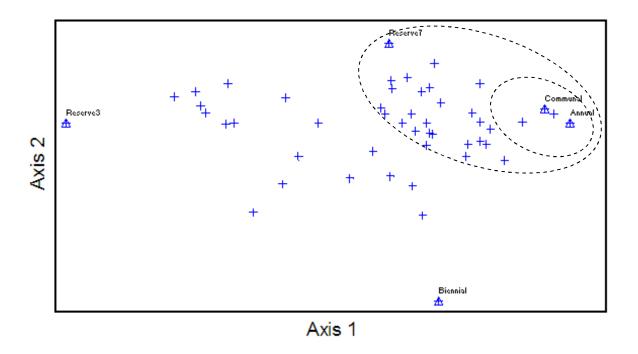


Fig. 6: Ordination analysis (Bray Curtis, distance measure is Sørensen) of spider diversity between sites. Crosses represent species and triangles represent sites. Samples with similar species composition are more closely clustered

Table 4: Percentage indicator values (>70%) of spider species for five different grassland managed sites relative to spider presence within each habitat and the number of individuals present in each habitat

Species	Annual Farm	Biennial Farm	Communal land	Reserve camp 3	Reserve camp 7	Total individuals	% indicator	P
Entypesa sp. 1	0	1	0	1	1	3	75	0.390
Olios sp. 1	0	1	0	1	1	3	75	0.390
<i>Drassodes</i> sp. 1	0	1	1	1	0	3	75	0.417
Camallina procurva	1	0	1	1	2	5	73	0.298
Linyphiidae sp. 2	1	0	1	1	3	6	77	0.298
Linyphiidae sp. 4	1	1	2	0	2	6	77	0.190
Xysticus mulleri Palpimanus	7	0	4	3	4	18	76	0.102
transvaalicus	4	1	0	12	2	19	77	0.477
Pardosa sp. 1	3	0	10	4	8	25	78	0.309
Cydrela sp. 1	19	8	5	4	6	42	82	0.207
Zelotes lightfooti	23	7	24	5	7	66	75	0.286
Trabea sp. 5	0	33	0	31	14	78	87	0.192
Proevippa sp. 2	32	13	33	26	41	145	87	0.192
Lycosidae sp.2	5	10	8	107	44	174	76	0.482
Lycosidae sp.1	50	18	51	9	54	182	79	0.102

*P* = Monte Carlo test for significance

Habitat structure has been described as an important aspect of spider assemblage structure (Gibson *et. al.*, 1992a). Because of this, a correlation analysis was undertaken to note any association between spider numbers and the density of vegetation, as determined by the disc pasture meter. A negative correlation was observed between the density of the vegetation and the number of spiders sampled (Table 5); however, in each case this correlation was not significant but it does imply that spider numbers are increased with reduced vegetation cover.

Table 5: Correlation analyses of mean values per study site of spider numbers against grass density (% grass), disk pasture meter (DPM) and vegetation cover (% veg)

	Mean	SD	r	Р
% Grass	83.3	5.46	-0.85	0.07
DPM	5.55	1.31	-0.80	0.10
% Veg	86.8	4.45	-0.83	0.08

### Discussion

Grazing is considered a disturbance effect at ecosystem scale (Díaz *et al.*, 2007; Dupre and Diekman, 2001) and has been recorded to influence species richness, assemblage structure and competition amongst species (Bilyeu *et al.*, 2007; Dupre and Diekman, 2001; Mucina *et al.*, 2006). Spiders are no exception, as pasture farming was found to negatively influence spider numbers and community composition within British (Gibson *et al.*, 1992a), Hungarian (Horváth, 2009), and African savanna ecosystems (Warui *et al.*, 2005). This is in contrast to our results where different levels of grazing and frequency of burning had no measurable effect on ground-dwelling spider abundance or assemblage structure. Only when rare or single species occurrence was included in the analyses, did we find an association of certain species with habitat types(see Fig. 7). The implications on spider assemblage and community structure based on various forms of grassland management through burning frequency and grazing intensity are discussed below.

Invertebrate communities often reflect the health or state of the ecosystem (Cameron *et al.*, 2003) and predatory invertebrates, such as spiders, can possibly be indicative of the prevalence or abundance of prey items. Certain species of spiders have feeding preferences for particular prey items and often the condition of the habitat may reflect the prevalence or abundance of the availability of prey items (Harwood *et al.*, 2001). As such, variations in the presence or absence of spiders and the structure of their communities can reveal underlying trends within the ecosystem. In this study, spider presence and absence did not differ significantly between the five study sites at both family and species levels. Moreover, specific families could not directly be associated with specific grassland management regimes, but certain trends were apparent at species level.

Species belonging to the family Lycosidae strongly dominated spider abundance in these grasslands (see Table 2). Jocqué & Alderweireldt (2005) hypothesize that the Lycosidae may have co-evolved with grasslands based on their relative rareness in dense forests and their appearance in the fossil record during the Miocene corresponds with the spread of grasslands. This family has also been found in large numbers in other habitat types such as semi-arid regions (e.g. Lotz *et al.*, 1991) and forest areas (Dippenaar-Schoeman and Wassenaar, 2002), but may be rare in habitats such as pine plantations (e.g. Van den Berg and Dippenaar-Schoeman, 1988) and Karoo regions (Haddad and Dippenaar-Schoeman, 2005). However, moderate grazing favoured wolf spiders (Lycosidae) that are more sun loving and their presence and numbers may be

influenced by the amount of sunshine (Dippenaar-Schoeman and Wassenaar, 2002). In more intensively grazed habitats that are also frequently burnt, conditions for wolf spiders are favored, and this may therefore be reflected in their numbers. Although not significant, there was a negative association with grassland vegetation density and cover observed in these grasslands where species of wolf spiders may be reduced where grass cover is increased. In addition, studies in agroecosystems have indicated that lycosids are apparently able to colonise disturbed habitats very quickly (Dippenaar-Schoeman, 1979; Van den Berg *et al.*, 1991), and are possibly very successful invaders of burt areas. Such findings may be useful in understanding how the spider communities in this study were structured, as some of the spiders were only found in particular sites. Indeed, the presence of overgrazed grasslands in the Steenkampsberg grasslands is high and more suited to this family of spiders. Vegetation structure, microclimatic factors and soil type may also have an influence on the local abundance of this particular group (Haddad and Dippenaar-Schoeman, 2005).

The Gnaphosidae were the second dominant group in this study and have been recorded to be the most abundant group amongst surveys in Southern Africa, especially in arid and semi-arid regions (Haddad and Dippenaar-Schoeman, 2002, 2005; Lots *et al.*, 1991). Indeed, it is one of the largest spider families recorded globally with regards to species and genera described (Platnick, 2012). It is a typical ground-dwelling family that can be expected to be collected in large numbers by pitfall traps left overnight, since the majority of species in this family are active nocturnal search and pursuit ground hunters. The Linyphiidae were the fourth most abundant group found in this study and they are the second most diverse family recorded globally. Eleven of the

twelve species recorded in this study were not identified down to genus level as the majority of species of this family, of generally very small spiders, have yet to be described. These small spiders are known to balloon regularly (Greenstone *et al.*, 1987) and have therefore very good colonisation properties and may be expected in recently burnt and heavily disturbed habitats. Pearce *et al.* (2005) found both the Linyphiidae and Lycosidae to dominate ballooning activity and ground densities in soya been fields. Gibson *et al.* (1992a) found the Linyphiidae to be the dominant group in heavily grazed pastures in England, which is characteristic of disturbed areas. Therefore, numbers could be expected to be high in the Steenkampsberg grasslands, as much of this habitat is heavily grazed and frequently burnt.

Morris (1990) suggested that managed grasslands often contain fewer individuals and species of invertebrates and highlighted the importance of architectural effects (vegetation and soil structure) of the habitat in influencing colonisation (Bristowe, 1941; Duffey, 1966). Our study found that habitats with increased vegetation cover and density represented spider communities with fewer individuals; however, this deduction can only be made for ground-dwelling species as we did not sample plant-dwelling or web-building species. It is further likely that those families of ground-dwelling species where high numbers of individuals were recorded have the ability to burrow and escape fire. The study site that was burnt annually has the second highest number of species recorded and it may be possible that fire does not play a large role in reducing ground-dwelling spiders within these grasslands. Indeed, Tainton and Mentis (1984) suggested that soil invertebrates are weakly affected by fire because soil temperatures are usually relatively low and they probably avoid fire by escaping underground or by using refugia

within or adjacent to the burn area. However, we recorded a large reduction in the prevalence of individuals during peak fire season (December and January) and a higher re-colonization trend following the fire season over all the habitats investigated (see Fig. 3). As such, fire may also directly influence individual survivorship and indirectly affect individuals through altering resource levels and patch conditions (Hobbs and Huenneke, 1992). This aspect needs to be investigated more extensively within these high altitude grasslands, but it remains that spider numbers did increase rapidly following burning events.

Using Turnbull's (1973) terms 'dominant', 'influents' and 'accessories' in describing the abundance of spiders in this study; the families Lycosidae, Gnaphosidae, Corinnidae and Linyphiidae can be regarded as 'dominant' (these families had the most number of species collected > 7); Salticidae, Theridiidae and Zodariidae as 'influents' (families with fewer species collected < 7) and the other 19 families as 'accessories' (families with the fewest species collected < 5). In this study, the presence or absence of a particular species may not be indicative of specific habitat requirements from a habitat specialist, as no particular species was significantly associated with any form of grassland management. It is indeed also likely that specimens were missed within other sites, thus we cannot conclude that a species that was not captured in our sampling was completely absent from a given site.

Within conserved grasslands, the impact of grazing on spider assemblages was reflected but not with an extensive drop in spider diversity (see Table 3). In Verloren Valei nature reserve camp 3, managed with infrequent burning and low grazing levels, reflected the most suitable management strategy for spiders, as it held the highest

diversity when compared to other study sites. Camp 7 in Verloren Valei is now managed with high number of indigenous ungulates and did differ from reserve camp 3 in reducing both diversity and abundance of ground-dwelling spiders, due to increased grazing pressure. However, the reduction was not dramatic, although the species found in this grazed camp were more similar to those found on commercially grazed and more frequently burnt habitats. Therefore, even within grasslands under formal conservation with a lower burning frequency, overstocking can reduce spider species richness and abundance; but probably more importantly, change spider species diversity, although this was not conclusively ascertained in this study. Horvath *et al.* (2009) suggested that a controlled management practice based on moderate grazing may result in a more species-rich spider assemblage as it increases habitat diversity which in turn, increases spider species numbers (Pozzi *et al.*, 1998). These studies, however, focused on webbuilders where vertical vegetation structure is of greater importance.

## Community structure

Multivariate analyses (ordination and cluster analyses) revealed that those habitats frequently burnt with moderate to high grazing levels reflected similar assemblages of ground-dwelling spider species (communal lands and the study site burnt annually) (see Fig. 5 and 5). This assemblage structure was also evident in performing a two-way cluster analyses based on species present between all the habitats sampled, with each habitat reflecting groups of dominant species. However, a number of singleton or rare species were observed that tended to skew the data and

analyses towards clustering groups or assemblages of spiders towards specific grassland management regimes (see Fig. 6). Upon removing these single occurrences from the analyses, no groups clustered around specific management practices (see Fig. 7). Furthermore, the indicator species analyses revealed the dominant species (most number of individuals) as the best indicators. However, they were highly prevalent in most sites sampled and these few species made up the majority of spiders sampled in this study. The indicator species analyses proposed by Dufrêne and Legendre (1997) may then not be suitable when comparing habitats where most species are only represented by very few individuals but rather make use of a two-way cluster analyses that gives all species equal weight.

This study did find that specific species commonly re-occur in habitats that follow similar management practices, such as those farms that were heavily grazed and more frequently burnt. A number of single species occurrences did occur in protected grasslands but this remains inconclusive to justify if these species were limited to these protected grasslands, and if a possibility exists whereby we may be losing specific species of ground-dwelling spiders that are more common in protected grasslands. We can also not say with confidence whether the various forms of grassland management practices were responsible for the occurrence of these rare individuals. However, it seems that with intensive grazing practices commonplace within the grasslands, this transformation is providing more suitable habitat to species that would not normally occur in these grasslands in such high numbers.

As was found in Hungarian pastures (Batary *et al.*, 2008; Samu, *et al.*, 2010; Szinetár and Samu, 2012) and Dutch grasslands (Kleijn *et al.*, 2006), grazing did not

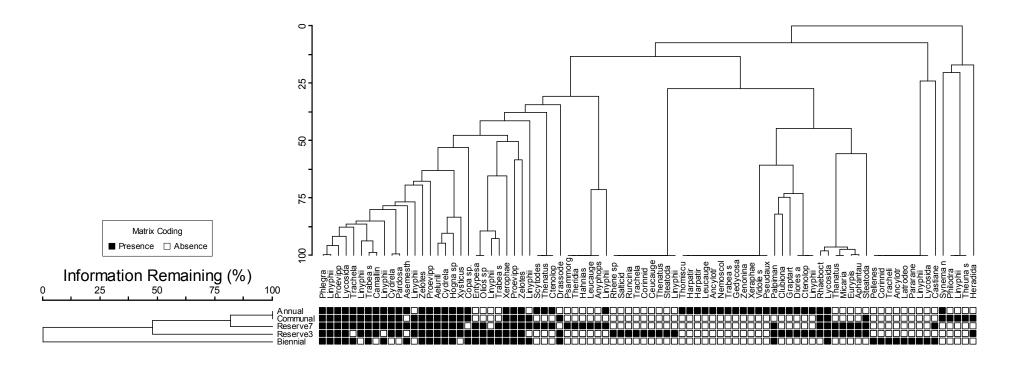


Fig. 7: Two-way cluster dendrogram depicting spider species diversity indicating the contribution each species has or groups of species have to community structure within each study site. This analyses includes single occurrences of a species on a site.

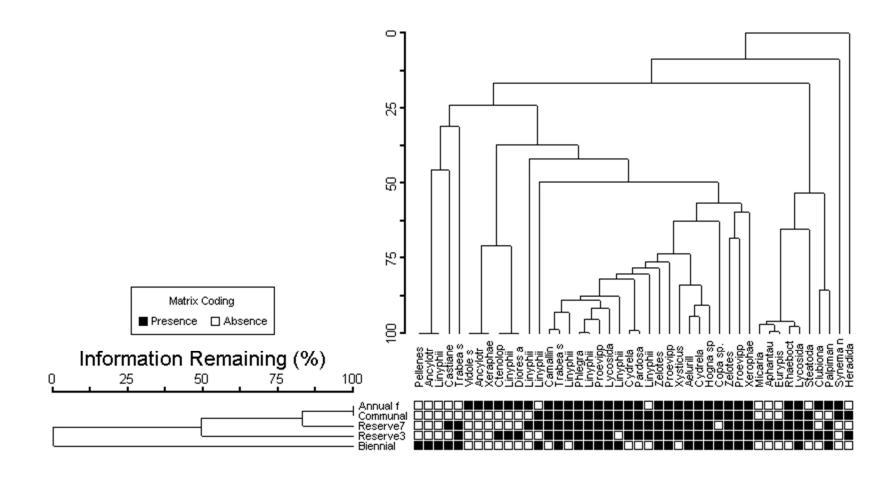


Fig. 8: Two-way cluster dendrogram excluding single occurrences of a species on a site

seem to have any measurable effect on either the presence of ground-dwelling spider species or their community composition in these moist highland grasslands. This study is a first in recording the ground-dwelling spider diversity in the moist highland grasslands of the Mpumalanga escarpment, a number of species ranges have now been extended where they were previously not recorded and a few species recorded in this study have yet to be described.

#### Conclusions and recommendations

This study has provided for a preliminary inventory of the ground-dwelling spider species within this montane escarpment grassland habitat (see Appendix B). It has not, however, conclusively illustrated the association or cluster of certain species to particular habitats associated with prevailing grassland management practices. There is preliminary evidence to suggest that some species may have a preference for certain habitats and that some spider families tend to increase in number when grass sward density and cover is reduced. It is likely that that both grazing and fire (burning) change the structural complexity of this shorter grassland type less than would have been in ecosystems such as savanna, where more intense grazing was observed to noticeably change spider species composition (e.g. Warui *et al.*, 2005). This larger change in habitat structure within other ecosystems, through a simplification of vegetation structure and possibly plant diversity and fire, has a direct impact on both the physical and chemical properties of soil litter and so may also reflect a more obvious change in

spider diversity within those ecosystems but not so within this highland grassland habitat type.

It is relatively simple to monitor spider assemblages in response to management practices, however, the biological requirements and the taxonomic status for many of the taxa recorded in this study remains unknown. As such, it remains problematic to suggest which groups or assemblages of the arachnids clearly reflect the effects of a management practice that can promote biodiversity conservation or individual spider species. However, this study has not conclusively demonstrated that spiders as a group do reflect management practices in moist highland grasslands, as stated by Gibson et al., (1992a) within other ecosystems. Batary et al. (2008), Samu et al. (2010) and Szinetár and Samu (2012) found similar results within Hungarian pastures where levels of grazing did not impact the diversity or species richness of ground-dwelling spiders. However, this was not the case with web-builders where vertical vegetation structure was influenced heavily by grazing intensity (Horváth, 2009). Ground-dwelling spider community attributes could not clearly be linked to vegetation structure or even species composition in this study but it is suggested that a management system designed to promote spider diversity should encompass grasslands that are grazed or cut at intervals, not too short to decrease plant species diversity but long enough to allow architectural diversity to develop that promote invertebrate conservation (Gibson et al.,1992b). This recommendation can be regarded as an applied version of the hypothesis that intermediate levels of disturbance tend to maximize diversity (Connell, 1979; Huston, 1979).

Singleton species occurrence dominated this study and the possibility exists that the survey undertaken in this study was insufficient and the habitat under sampled as suggested by Coddington et al. (2009). Indeed, high singleton frequencies characterize the majority of tropical arthropod surveys (Coddington, et al. 2009) and under-sampling is often clearly reflected when high singleton species are present in biological data sets (McGill, 2003). This has implications toward conservation management plans as one cannot make deductions from an incomplete inventory. It is therefore suggested that further studies should be designed to include seasons (Warui, 2004), years or even multiple years that would yield a more complete inventory so that conservation based decisions can be deduced from a more accurate biodiversity assessment. It is further suggested that studies expand to include a wider array of grassland management regimes to ascertain any associations with habitat variables. In addition, studies should incorporate web-builders that inhabit the vertical vegetation component to compliment a comprehensive spider survey for the region, which is currently lacking. Moreover, there is a clear need for further taxonomic description for many species and the need for a better autecological knowledge and understanding of their biology and their association with vegetation structure and plant species diversity.

## **Acknowledgements**

We wish to thank the farmers and landowners of the Dullstroom district in addition to the Mpumalanga Parks and Tourism Authority for granting us access to their land. We would also like to thank Tshwane University of Technology's Department of Research and Innovation for making the funds available to undertake the fieldwork for this

research. We are very grateful to Dr Stephan Foord, Dr Charles Haddad and an anonymous referee for their suggested comments on improving this manuscript. Finally, we would like to thank Thabo Mabuza for his tireless assistance in the field.

## References

- Abensperg-Traun, M., Smith, G.T., Arnold, G. W. & Steven, D.E. (1996) The effects of habitat fragmentation and livestock grazing on animal communities in remnants of gimlet, *Eucalyptus salubris* woodland in the Western Australian wheat-belt. I. Arthropods. *Journal of Applied Ecology*, **33**, 1281-1301.
- Acocks, J.P.H. (1988) Veld types of South Africa, 3<sup>rd</sup> edition. Botanical Research Institute, South Africa.
- Armstrong, A.J. & Van Hensbergen, H.J. (1999) Identification of priority regions for animal conservation in afforestable montane grasslands of the northern Eastern Cape Province, South Africa. *Biological Conservation*, **87**, 93-103.
- Ausden, M., Hall, M., Pearson, P. & Strudwick, T. (2005) The effects of cattle grazing on tall-herb fen vegetation and mollusks. *Biological Conservation*, **122**, 317-326.
- Batary, P., Baldi, A., Samu, F., Szuts, T. & Erdos, S. (2008) Are spiders reacting to local or landscape scale effects in Hungarian pastures? *Biological Conservation*, **141**, 2062-2070.
- Bell, J.R., Wheater, C.P. & Cullen, W.R. (2001) The implications of grassland and heathland management for the conservation of spider communities: a review. *J. Zool., Lond*,. **255**, 377-387.

- Bilyeu, D.M., Cooper, D.J. & Hobbs N.T. (2007) Assessing impacts of large herbivores on shrubs: tests of scaling factors for utilization rates from shoot-level measurements. *Journal of Applied Ecology*, **44**, 168-175.
- Bransby, D. J. & Tainton, N. M. (1977) The Disc Pasture Meter: Possible applications in grazing management. *Proceedings of Grassland Society of South Africa,* **12**, 115-118.
- Bray, J.R. & Curtis, J.T. (1957) An ordination of the upland forest communities of southern Wisconsin. *Ecological Monographs*, **27**, 325-449.
- Bredenkamp, G., Granger, J.E. & Van Rooyen, N. (1996) North-eastern Mountain

  Grassland. *Vegetation of South Africa, Lesotho and Swaziland* (ed. by A.B. Low & A.G. Rebelo), pp. 46-47. Department of Environmental Affairs and Tourism,

  Pretoria.
- Bristowe, W.S. (1941) *The comity of spiders*. Ray Society, London.
- Cameron, A, Johnston, R.J. & McAdam, J. (2003) Classification and evaluation of spider (Araneae) assemblages on environmentally sensitive areas in Northern Ireland. *Agriculture, Ecosystems and Environment*, **102**, 29-40.
- Cherrill, A.J. & Brown, V.K. (1990a) The life cycle and distribution of the wart-biter,

  Decticus verrucivorous (L.) (Tettigoniidae) within a chalk grassland in southern

  England. Biological Conservation, **53**, 125-143.
- Cherrill, A.J. & Brown, V.K. (1990b) The habitat requirements of the bush-cricket,

  \*Decticus verrucivorous\* (L.) (Orthoptera: Tettigoniidae). \*Biological Conservation\*,

  53, 145-157.

- Churchill, T.B. (1997) Spiders as indicator taxa: an overview for Australia. *Memoirs of the Museum of Victoria*, **56**, 331-337.
- Churchill, T.B. (1998) Spiders as ecological indicators in the Australian tropics: family distribution patterns along rainfall and grazing gradients. *Bulletin of the British Arachnological Society*, **11**, 325-330.
- Churchill, T.B. & Ludwig, J.A. (2004) Changes in spider assemblages along grassland and savanna grazing gradients in Northern Australia. *Rangeland Journal*, **26**, 3-16.
- Connell, J.H. (1979) Tropical rain forests and coral reefs as open non-equilibrium systems. *Population dynamics* (ed. by R.M. Anderson, L.R. Taylor & L.R. Turner), pp. 141-163. Blackwell Scientific, Oxford.
- Deltschev, K. & Kajak, A. (1974) Analysis of a sheep pasture ecosystem in the Pieniny Mountains (The Carpathians). XVI. Effect of pasture management on the number and biomass of spiders (Araneae) in two climatic regions (The Pieniny and the Sredna Gora Mountains). *Ekologia Polska*, **22**, 693-710.
- Dennis, P., Young, M.R. & Bentley, C. (2001) The effects of varied grazing management on epigeal spiders, harvestmen and pseudoscorpions of *Nardus* stricta grassland in upland Scotland. *Agriculture, Ecosystems and Environment*, **86**, 39-57.
- Dennis, P. (2003) Sensitivity of upland arthropod diversity to livestock grazing, vegetation structure and landform. *Journal of Food, Agriculture and Environment*, **1(2)**, 301-307.

- Díaz, S., Lavorel, S., Ntyre, S., Falczuk, V., Casanoves, F., Milchunas, D.G., Skarpe,
  C., Rusch, G., Sternberg, M., Noy-Meir, I., Landsberg, J., Zhang, W., Clark, H. &
  Campbell, B.D. (2007) Plant trait responses to grazing: A global synthesis.
  Global Change Biology, 13, 313–341.
- Dippenaar-Schoeman. (1979) Spider communities in strawberry beds: seasonal change in numbers and species composition. *Phytophylactica*, **11**, 1-4.
- Dippenaar-Schoeman, A.S. & Jocqué, R. (1997) *African spiders: an identification manual*. Agricultural Research Council, Durban, South Africa.
- Dippenaar-Schoeman, A.S. & Wassenaar, T.D. (2002) A checklist of the ground-dwelling spiders of coastal sand dune forests at Richards Bay, South Africa (Arachnidae: Araneae). *Bulletin of the British Arachnological Society*, **12**, 275-279.
- Coddington, J.A., Agnarsson, I., Miller, J.A., Kuntner, M. and Hormiga, G. (2009)

  Undersampling bias: the null hypothesis for singleton species in tropical arthropod surveys. *Journal of Animal Ecology*, **78**, 573-584.
- Duffey, E. (1966) Spider ecology and habitat structure. *Senckenbergiana Biologica*, **47**, 45-49.
- Dufrêne, M. & Legendre, P. (1997) Species assemblages and indicator species: the need for a flexible asymmetrical approach. *Ecological Monographs*, **67**, 345-366.
- Dupre, C. & Dlekmann, M. (2001) Differences in species richness and life-history traits between grazed and abandoned grasslands in southern Sweden. *Ecography*, **24**, 275–286.

- Foord, S.H., Ferguson, J.W.H. & Van Jaarsveld, A.S. (2002) Endemicity of Afromontane grasshopper assemblages: implications for grassland conservation. *African Journal of Ecology*, **40**, 318-327.
- Foord, S.H., Mafadza, M.M., Dippenaar-Schoeman, A.S. & Van rensburg, B.J. (2008)

  Micro-scale heterogeneity of spiders (Arachnida: Araneae) in the Soutpansberg,

  South Africa: a comparative survey and inventory in representative habitats.

  African Zoology, 43(2), 156-174.
- Gibson, C.W.D., Hambler, C. & Brown, V.K. (1992a) Changes in spider (Araneae) assemblages in relation to succession and grazing management. *Journal of Applied Ecology*, **29**, 132-142.
- Gibson, C.W.D., Brown, V.K., Losito, L. & McGavin, G.C. (1992b) The response of invertebrate assemblies to grazing. *Ecography*, **15**, 166-176.
- Greenstone, M.H., Morgan, C.E., Hultsch, A.L., Farrow, R.A.,
  & Dowse, J.E., 1987. Ballooning spiders in Missouri, USA, and
  new south Wales, Australia: family and mass distributions. *Journal of Arachnology*, 15, 163–170.
- Haddad, C.R. & Dippenaar-Schoeman, A.S. (2002) The influence of mound structure on the diversity of spiders (Araneae) inhabiting the abandoned mounds of the snouted harvest termite *Trinervitermes trinervoides* (Sjöstedt). *Journal of Arachnology*, **30**, 403-408.
- Haddad, C.R. & Dippenaar-Schoeman, A.S. (2005) Epigeic spiders in Nama Karoo grassland in the Northern Cape Province. *Navorsinge van die Nationale Museum, Bloemfontein, South Africa*, **1(21)**, 1-10.

- Haddad, C.R., Honiball, A.S., Dippenaar-Schoeman, A.S., Slotow, R. & Van Rensburg,
   B.J. (2010) Spiders as potential indicators of elephant-induced habitat changes in endemic sand forest, Maputaland, South Africa. *African Journal of Ecology*, 48, 446-460.
- Haddad, C.R. & Wesolowska, W. (2011) New species and new records of jumping spider (Araneae: Salticidae) from central South Africa. *African Invertebrates*, **52**(1): 51-134.
- Harwood, J.D., Sunderland, K.D., & Symondson, W.O.C., 2001. Living where the food is: web-location by linyphiid spiders in relation to prey availability in winter wheat. *Journal of Applied Ecology*, **38**, 88–99.
- Hobbs, R.J. & Huenneke, L.F. (1992) Disturbance, diversity and invasion: implications for conservation. *Conservation Biology*, **6(3)**, 324-337.
- Hodgson, J.G. (1986) Commonness and rarity in plants with special reference to the Sheffield flora. Part 1: The identity, distribution and habitat characteristics of the common and rare species. *Biological Conservation*, **36**, 199-252.
- Horváth, R., Magura, T., Szinetár C. & Tóthmérész, B. (2009) Spiders are not less diverse in small and isolated grasslands, but less diverse in overgrazed grasslands: A field study (East Hungary, Nyírség). *Agriculture, Ecosystems and Environment*, **130**, 16-22.
- Hurlbert, S.H. (1984). Pseudoreplication and the design of ecological field experiments. *Ecological Monographs*, **54**, 187-211.
- Huston, M. (1979) A general hypothesis of species diversity. *American Naturalist*, **113**, 81-101.

- Jocqué, R. & Alderweireldt, M. (2005) Lycosidae: the grassland spiders. *Acta zoological bugarica*, Suppl.1, 125-130.
- Kleijn, D., Baquero, R.A., Clough, Y., Diaz, M., De Esteban, J., Fernández, F., Gabriel, D., Herzog, F., Holzschuh, A., Jöhl, R., Knop, E., Kruess, A., Marshall, E.J.P., Steffan-Dewenter, I., Tscharntke, T., Verhulst, J., West, T.M. & Yela, J.L. (2006)
  Mixed biodiversity benefits of agri-environmental schemes in five European countries. *Ecology Letters*, 9, 243-254.
- Lotz, L.N., Seaman, M.T. & Kok, D.J. (1991) Surface active spiders (Araneae) of a site in semi-arid central South Africa. *Navorsinge van die Nasionale Museum, Bloemfontein*, **7**, 529-540.
- Mallis, R.E. & Hurd, L.E. (2005) Diversity among ground-dwelling spider assemblages: habitat generalists and specialists. *Journal of Arachnology*, **33**,101-109.
- McCune, B. & Mefford, M.J. (2006) PC-ORD Multivariate analysis of ecological data.

  Version 4. Oregon: MiM software design.
- McGill, B.J. (2003) A test of the unified theory of biodiversity. *Nature*, **422**, 881-885.
- McGoech, M.A., Van Rensburg, B.J. & Botes, A. (2002) The verification and application of bioindicators: a case study of dung beetles in a savanna ecosystem. *Journal of Applied Ecology*, **39**, 661-672.
- Morris, M.G. (1990) The effects of management on the invertebrate community of calcareous grassland. *Calcareous Grasslands Ecology and Management:*Proceedings of a Joint British Ecological Society / NCC symposium (ed. by S.H. Hillier, D.W.H. Walton & D.A. Wells), pp. 128-133. Bluntisham Books, UK.

- Morris, B. (2001) Mpumalanga Performance Report on Sustainable Development 2001.

  <a href="http://www.environment.gov.za/soer/reports/mpumalanga/">http://www.environment.gov.za/soer/reports/mpumalanga/</a>> 12 May 2012.

  website/documents%5 CFinal 2001 performance report.pdf> 12 April 2012.
- Mucina, L., Hoare, D.B., Lotter, M.C., Du Preez, P.J., Rutherford, M.C., Scott-Shaw,
  C.R., Bredenkamp, G.J., Powrie, L.W., Scott, L., Camp, K.G.T., Cilliers, S.S.,
  Bezuidenhout, H., Mostert, T.H., Siebert, S.J., Winter, P.J.D., Burrows, J.E.,
  Dobson, L., Ward, R.A., Stalmans, M., Oliver, E.G.H., Siebert, F., Schmidt, E.,
  Kobisi, K., Kose, L. (2006) Grassland Biome. *The Vegetation of South Africa,*Lesotho and Swaziland (ed. by L. Mucina & M.C. Rutherford), pp. 349-436.
  Strelitzia 19: South African National Biodiversity Institute, Pretoria.
- Owen-Smith, N. & Danckwerts, J.E. (1997) Herbivory. *Vegetation of Southern Africa* (ed. by R.M. Cowling & D.M. Richardson), pp. 397-420. Cambridge University Press, Cambridge.
- Pearce, S., Zalucki, M.P. & Hassan, E. (2005) Spider ballooning in soybean and non-crop areas of southeast Queensland. *Agriculture, Ecosystems and Environment*, **105**, 273-281.
- Platnick, N.I. (2012) The World Spider Catalog, Ver 12.5. The American Museum of Natural History. <a href="http://research.amnh.org/iz/spiders/catalog/">http://research.amnh.org/iz/spiders/catalog/</a> 16 May 2012
- Pozzi, S. Gonseth, Y. & Hanggi, A. (1998) Evaluation of dry grassland management on the Swiss occidental plateau using spider communities (Arachnida: Araneae).

  \*Revue Suisse de Zoologie, 105, 465-485.
- Rouget, M., Jonas, Z., Cowling, R.M., Desmet, P.G., Driver, A., Mohamed, B., Mucina, L., Rutherford, M.C. & Powrie, L.W. (2006) Ecosystem Status and Protection

- Levels of Vegetation Types. *The Vegetation of South Africa, Lesotho and Swaziland* (ed. by L. Mucina & M.C. Rutherford), pp. 725-737. Strelitzia 19: South African National Biodiversity Institute.
- Samu, F., Kádár, F., Ónodi, G., Kertész, M., Szirányi, A., Szita, E., Fetykó, K., Neidert, D., Botos, E., Altbäcker, V. (2010). Different ecological responses of two generalist arthropod groups, spiders and carabid beetles (Araneae, Carabidae), to the effects of wildfire. *Community Ecology*, **11(2)**, 129-139.
- SANBI. 2008. South African National Biodiversity Institute: Grasslands Program. <a href="http://www.SANBI.co.za">http://www.SANBI.co.za</a> 11 November 2011.
- Statsoft Inc. (2009) Statistica (Data analysis software system), Version 9.
- Stohlgren, T.J., Falkner, M.B. & Schell, L.D. (1995) A Modified-Whittaker nested vegetation sampling method. *Vegetation*, **117**, 113-121.
- Szinetár, C. & Samu, F (2012) Intensive grazing opens spider assemblage to invasion by disturbance-tolerant species. *Journal of Arachnology*, **40**, 59-70
- Tainton, N.M. & Mentis, M.T. (1984) Fire in Grassland. Ecological Effects of Fire in South African Ecosystems (ed. by P. de V. Booyson & N.M. Tainton), pp. 115-147. Springer-Verlag, Berlin.
- Thomas, J.A. (1983) The ecology and conservation of *Lysandra bellagus* (Lepidoptera: Lycaenidae) in Britain. *Journal of Applied Ecology*, **20**, 59-83.
- Turnbull, A.L. 1973. Ecology of the true spiders (Araneomorphae). *Annual Reviews of Entomology*, **18**, 305-348.

- Van den Berg, A.M. & Dippenaar-Schoeman, A.S. (1988) Spider communities in a pine plantation at Sabie, Eastern Transvaal: a preliminary survey. *Phytophylactica*, **20**, 293-296.
- Van den Berg, A.M., Dippenaar-Schoeman, A.S. & Schoonbee, H.J. (1991) The effect of two pesticides in spiders in South African cotton fields. *Phytophylactica*, **22**, 435-441.
- Van Rensberg, B.J., McGeoch, M.A., Chow, S.L. & Van Jaarsveld, A.S. (1999)
  Conservation of heterogeneity among dung beetles in the Maputaland centre of endemism, South Africa. *Biological Conservation*, 88, 145-153.
- Warui, C.M., Villet, M.H., Young, T.P. & Jocque, R. (2005) Influence of grazing by large mammals on the spider community of a Kenyan savanna biome. *Journal of Arachnology*, **33**, 269-279
- Weidemann, S., Stiedl, O. & Kalmring, K. (1990) Distribution and population density of the bush cricket *Decticus verrucivorous* in a damp-meadow biotope. *Oecologia* (Berlin), 82, 369-373.
- Wessels, K.J., Reyers, B., Van Jaarsveld, A.S. & Rutherford, M. C. (2003) Identification of potential conflict areas between land transformation and biodiversity conservation in north-eastern South Africa. *Agriculture, Ecosystems and Environment*, **95**, 157-178.
- Wise, D.H. (1993) Spiders in ecological webs. Cambridge University Press, Cambridge.

## **APPENDICES**

**APPENDIX A:** Grazing equivalents of all ungulates and grazing intensity represented as hectares per large animal unit for all study sites

	COMMUNAL LANDS	BIENNIAL FARM	ANNUAL FARM	RESERVE camp 7	RESERVE camp 3	
Total ha	208.4	447.6	205.2	1740	4150	LAU equivalents
cattle	172	228	164	0	0	1
horses	2.73	0	0	0	0	1.1
goats	11.3	0	0	0	0	5.03
sheep	8.5	0	0	0	0	4.9
Blesbuck Black	0	0	0	26.7	46.7	4.4
Wildebeest	0	0	0	40.2	0	2.17
Zebra	0	0	0	375.3	0	1.51
TOTAL						
LAU	194.5	228	164	442.2	46.7	
ha/LAU	1.07	1.9	1.2	3.9	88.8	

Indigenous ungulates include: Blesbuck (*Damaliscus dorcas phillispi*), Black Wildebeest (*Connochaetes gnou*) and Zebra (*Equus burchelli*)

**APPENDIX B:** Spider species checklist according to family and life trait category. Those families with an \* are not regarded as ground dwelling species and probably accidentally landed in pittraps.

Family	Genus and species	Guild
^ · · · · · · · · · · · · · · · · ·		
Amaurobiidae*	Pseudauximus sp. 1	Retreat-web spider
Araneidae*		
	Nemoscolus sp. 1	orb-web
Clubionidae	Pararaneus cyrtoscapus Pocock, 1898	orb-web
Ciubioriidae	Clubiona sp. 1	sac spider
Corinnidae		out opido.
	Castianeira sp. 1	free-living ground dweller
	Cetonana sp. 1 (new)	free-living ground dweller
	Corinnidae sp. 1	free-living ground dweller
	Copa sp. 1 Graptartia mutillica Haddad, 2004	free-living ground dweller free-living ground dweller
	Trachelas pusillus Lessert, 1923	free-living ground dweller
	Trachelas sp. 1	free-living ground dweller
	Trachelinae sp. 1	free-living ground dweller
Cyrtaucheniidae		
Cyrtaucheniidae	Ancylotrypa sp. 1	ground burrow dweller (trapdoor spider)
	Ancylotrypa brevipalpis Hewitt, 1916	ground burrow dweller (trapdoor spider)
Gnaphosidae		
	Aphantaulax signicollis Tucker, 1923	free-living ground dweller
	Asemesthes sp. 1 maybe new	free-living ground dweller
	Camillina procurva Purcell, 1908 Drassodes sp. 1	free-living ground dweller free-living ground dweller
	Micaria sp. 1	free-living ground dweller
	Xerophaeus bicavus Tucker, 1923	free-living ground dweller
	Xerophaeus sp. 2	free-living ground dweller
	Zelotes lightfooti Purcell, 1907	free-living ground dweller
	Zelotes uquathus Fitzpatrick,2008	free-living ground dweller
Hahniidae		
	Hahnia tubicola Simon, 1898	sheet-web spider
Idiopidae		
	Ctenolopus oomi Hewitt, 1913	ground burrow dweller (trapdoor spider)
Linyphiidaa	Ctenolopus sp. 2 (new)	ground burrow dweller (trapdoor spider)
Linyphiidae	Linyphiidae sp. 1	Sheet-web spider
	Linyphiidae sp. 1	Sheet-web spider
	Linyphiidae sp. 3	Sheet-web spider
	••	•

	Linyphiidae sp. 4 Linyphiidae sp. 5 Linyphiidae sp. 6 Linyphiidae sp. 7 Linyphiidae sp. 8	Sheet-web spider Sheet-web spider Sheet-web spider Sheet-web spider Sheet-web spider
	Microlinyphia sterilis Pavesi, 1883	Sheet-web spider
	Linyphiidae sp. 10	Sheet-web spider
	Linyphiidae sp. 11	sheet-web spider
Liocranidae		
	Rhaeboctesis trinotatus Tucker, 1920	free-living ground dweller
Lycosidae	B /	
	Pardosa crassipalpis Purcell, 1903	free-living ground dweller
	Trabea purcelli Roewer, 1951	free-living ground dweller
	Trabea sp. 4	free-living ground dweller
	Trabea sp. 5	free-living ground dweller
	Proevippa fascicularis Purcell, 1903	free-living ground dweller
	Proevippa albiventris Simon, 1898	free-living ground dweller
	Proevippa sp. 2	free-living ground dweller
	Lycosidae sp. 1	free-living ground dweller
	Lycosidae sp. 2	free-living ground dweller
	Lycosidae sp. 5	free-living ground dweller
	Geolycosa sp. 3 Hogna sp. 5	free-living ground dweller free-living ground dweller
	Zenonina albocaudata Lawrence, 1952	free-living ground dweller
Nemesiidae	Zenonina albocaddata Lawrence, 1932	rree-living ground dweller
Nemesiidae	Entepesa schoutdeni Benoit, 1965	ground burrow dweller (trapdoor spider)
	Entepesa sp. 2	ground burrow dweller (trapdoor spider)
Orsolobiidae	Emopoda Sp. 2	ground burrow awener (trapacer spider)
Croolobilaac	Afrilobus australis Griswold and	
	Plantnick, 1987	free-living ground dweller
Palpimanidae		
•	Palpimanus transvaalicus Simon, 1893	free-living ground dweller
Philodromidae		
	Thanatus dorsolineatus Jézéquel, 1964	free-living plant dweller
	Thanatus vulgaris Simon, 1870	free-living plant dweller
	Thanatus sp. 2	free-living plant dweller
	Philodromus sp.1	free-living plant dweller
Phyxelididae		
	Vidole sothoana Griswold, 1992	ground retreat-web ground
Prodidomidae		
	Theuma sp. 1 (new)	free-living ground dweller
Salticidae		
	Aelurillus sp. 1	free-living ground dweller
	Pellenes sp. 1	free-living ground dweller
	Phlegra sp. 1	free-living ground dweller
	Rhene sp. 1	free-living ground dweller
	Salticidae sp. 3	free-living ground dweller
Scytodidae		
	Scytodes sp.	free-living ground dweller

Selenopidae

Anyphops sp. 1 free-living ground dweller

Sparassidae

Olios sp. 1 free-living plant dweller

Tetragnathidae

Leucauge festiva Blackwall, 1866 orb-web spider

Theraphosidae

Harpactira sp. ground burrow dweller (baboon spider)

ground burrow dweller (lesser baboon

spider)

Theridiidae

Euryopis sp. 1 free-living ground dweller

Latrodectus geometricus C.L. Koch,

1841

cob-web dweller Steatoda sp. 1 cob-web dweller Steatoda capensis Hann, 1990 Cob-web dweller

Theridion sp. 1 cob-web dweller

Thomisidae

Runcinia grammica L. Koch, 1937 free-living plant dweller Synema nigrotibiale Lessert, 1919 free-living plant dweller Thomisus australis Comellini, 1957 free-living plant dweller

Xysticus mulleri Lawrence, 1952 free-living ground dweller

Zodariidae

Cydrela schoemanae Jocqué, 1991

Cydrela sp. 2

Diores annettae Jocqué, 1990

Heradida sp. 1

Harpactirella sp.

Psammorygma aculeatum Karsch,

1878

free-living ground dweller free-living ground dweller free-living ground dweller free-living ground dweller

free-living ground dweller

**APPENDIX C:** Individual number of ground dwelling spider species located within each of the five study sites

Species	Annual farm	Biennial farm	Communal land	Reserve 3	Reserve 7
Rhene sp. 1	0	0	0	1	0
Salticidae sp. 3	0	0	0	1	0
Thomiscus australis	1	0	0	0	0
Runcinia grammica	0	0	0	1	0
Psammorgna					
aculeatum	0	0	0	0	1
Trachelas sp. 1	0	0	0	1	0
Corinnidae sp. 2	0	1	0	0	0
Trachelinae sp.	0	1	0	0	0
Corinnidae sp. 1	0	0	0	1	0
Harpactira sp. 1	1	0	0	0	0
Harpactirella sp. 1	1	0	0	0	0
Leucauge festiva	1	0	0	1	1
Philodromus sp	0	0	1	0	0
Thanatus vulgaris	0	0	0	1	0
Theridion sp. 1	0	0	0	0	1
Latrodecetus					
geometricus	0	1	0	0	0
Steatoda capensis Pararaneus	0	0	0	1	0
cyrtoscapus	0	1	0	0	0
Linyphiidae sp. 9	0	0	1	0	0
Linyphiidae sp. 11	0	0	0	1	0
Trabea purcelli	1	0	0	0	0
Geolycosa sp. 3	1	0	0	0	0
Lycosidae sp. 5	0	1	0	0	0
Zenonina albocauda	1	0	0	0	0
Hahnia tubicola	0	0	0	0	1
Pseudauximus sp.	1	0	0	0	0
Anyphops sp	0	0	0	0	1
Theuma sp. 1	0	0	1	0	0
Nemoscolus sp. 1	1	0	0	0	
Pellenes sp. 1	0	2	0	0	0
Graptartia mutillica	1	0	0	1	0
Scytodes sp.1 Thanatus	1	0	0	0	1
dorsolineatus	0	0	0	1	1
Thanatus sp. 2	1	0	0	0	1
Ancylotrypa brevipalpis	0	2	0	0	0
Ctenolophus sp. 2	1	0	0	0	1
Linyphiidae sp. 10	0	1	0	1	0
Linyphiidae sp. 7	0	2	0	0	0
Micaria sp.	0	0	0	1	1
Vidole sothoana	2	0	0	0	0

Symema nigrotibiale			_	_	_	_	
Castianeira 0 2 0 0 1 Entypesa schoutedeni 0 1 0 1 0 1 1 Ancylotrypa sp. 1 3 0 0 0 0 0 Ctenolophus oomi 2 0 0 0 1 0 Ctenolophus oomi 2 0 0 0 1 0 Ctenolophus os 1 1 0 0 0 2 0 Clubiona sp. 1 1 0 0 0 2 0 Linyphiidae sp. 1 2 0 0 0 1 0 0 Asemesthes sp. 1 1 1 0 0 0 1 1 Aphantaulax signicollis 0 0 0 0 2 1 Aphantaulax signicollis 0 0 0 0 2 1 Trachelas pusillus 1 1 1 1 0 1 1 0 Trachelas pusillus 1 1 1 1 0 0 1 Linyphiidae sp. 6 1 0 0 0 0 3 Linyphiidae sp. 6 1 0 0 0 0 3 Linyphiidae sp. 5 0 1 2 0 1 Euryopis sp. 1 0 0 1 1 1 3 Linyphiidae sp. 4 1 1 2 0 0 2 Linyphiidae sp. 4 1 1 1 2 0 2 Linyphiidae sp. 4 1 1 1 2 0 2 Linyphiidae sp. 2 1 1 0 1 1 1 3 Linyphiidae sp. 2 1 1 0 1 1 1 3 Linyphiidae sp. 3 0 0 3 1 1 1 Cydrela sp. 2 1 0 3 1 3 1 3 Linyphiidae sp. 3 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1							
Entypesa schoutedeni		0	· ·		·	0	
Olios sp. 1         0         1         0         1         1           Ancylotrypa sp. 1         3         0         0         0         0           Ctenolophus comi         2         0         0         1         0           Clubiona sp. 1         1         0         0         2         0           Linyphiidae sp. 1         2         0         0         1         0           Asemesthes sp. 1         1         1         0         0         0         0           Asemesthes sp. 1         1         1         0         0         0         1         Aphantaulax signicollis         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0         0		0	2	0	0	1	
Ancylotrypa sp. 1 3 0 0 0 0 0 Ctenolophus oomi 2 0 0 0 1 0 0 Ctenolophus oomi 2 0 0 0 1 0 0 Ctenolophus oomi 2 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	Entypesa schoutedeni	0	1	0	1	1	
Clubiona sp. 1	Olios sp. 1	0	1	0	1	1	
Clubiona sp. 1	Ancylotrypa sp. 1	3	0	0	0	0	
Clubiona sp. 1	Ctenolophus oomi	2	0	0	1	0	
Xerophaeus.sp. 2         3         0         0         0         0           Asemesthes sp. 1         1         1         0         0         1           Aphantaulax signicollis         0         0         0         2         1           Drassodes sp. 1         0         1         1         1         0         0           Linyphiidae sp. 6         1         0         0         0         3         Linyphiidae sp. 6         1         0         0         3         Linyphiidae sp. 5         0         1         2         0         1         Linyphiidae sp. 5         0         1         2         0         1         1         2         1         1         1         2         1         1         1         2         1         1         1         2         1         2         1         1         1         2         2         2         1         1         1         2         2         2         2         0         2         2         0         2         2         0         2         2         0         2         2         0         2         2         0         2         0         2	Clubiona sp. 1	1	0	0	2	0	
Xerophaeus.sp. 2         3         0         0         0         0           Asemesthes sp. 1         1         1         0         0         1           Aphantaulax signicollis         0         0         0         2         1           Drassodes sp. 1         0         1         1         1         0         0           Linyphiidae sp. 6         1         0         0         0         3         Linyphiidae sp. 6         1         0         0         3         Linyphiidae sp. 5         0         1         2         0         1         Linyphiidae sp. 5         0         1         2         0         1         1         2         1         1         1         2         1         1         1         2         1         1         1         2         1         2         1         1         1         2         2         2         1         1         1         2         2         2         2         0         2         2         0         2         2         0         2         2         0         2         2         0         2         2         0         2         0         2	Linyphiidae sp. 1	2	0	0	1	0	
Asemesthes sp. 1 1 1 0 0 0 1 Aphantaulax signicollis 0 0 0 0 2 1 Drassodes sp. 1 0 1 1 1 1 0 0 1 Drassodes sp. 1 0 1 1 1 1 0 0 1 Drassodes sp. 1 0 1 1 1 1 0 0 1 Drassodes sp. 1 1 1 1 1 0 0 1 Drassodes sp. 1 1 1 1 1 0 0 1 Drassodes sp. 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1		3	0	0	0	0	
Aphantaulax signicollis 0 0 0 0 2 1 Drassodes sp. 1 0 1 1 1 1 0 Trachelas pusillus 1 1 1 1 0 1 Linyphiidae sp. 6 1 0 0 0 3 Linyphiidae sp. 5 0 1 2 0 1 Euryopis sp. 1 0 0 0 0 3 2 Camillina procurva 1 0 1 1 2 Linyphiidae sp. 2 1 0 1 1 2 Linyphiidae sp. 4 1 1 2 0 2 Linyphiidae sp. 4 1 1 2 0 2 Diores annettae 5 0 0 0 2 Diores annettae 5 0 0 0 2 0 Zelotes uquathus 1 3 1 1 1 Cydrela sp. 2 1 0 3 1 3 Linyphiidae sp. 3 1 1 1 1 Cydrela sp. 2 1 0 3 1 3 Linyphiidae sp. 3 0 0 3 1 4 Xerophaeus bicavus 1 3 1 2 3 Steatoda sp. 1 0 0 3 5 3 Steatoda sp. 1 0 0 3 5 3 Steatoda sp. 1 0 0 3 5 3 Phlegra sp. 1 2 3 2 2 2 3 Copa sp. 1 9 3 1 1 0 Linyphiidae sp. 8 4 3 3 2 4 Xysticus mulleri 7 0 4 3 4 Aelurillus sp. 1 7 3 2 3 4 Aelurillus sp. 1 7 3 2 3 4 Palpimanus transvaalicus 4 1 0 12 2 Proevippa fascicularis 1 7 5 4 2 Proevippa fascicularis 1 7 5 4 2 Proevippa albiventris 16 2 12 1 3 Cydrela schoemanae 19 8 5 4 6 Hogna sp. 5 15 8 7 8 5 Trabea sp. 4 5 7 Trabea sp. 5 0 33 0 31 14 Proevippa sp. 2 32 13 33 26 41 Lycosidae sp. 2 5 10 8 51 9 54  Total individuals per site 246 148 201 288 262			1	0	0	1	
Drassodes sp. 1         0         1         1         1         0           Trachelas pusillus         1         1         1         0         1           Linyphiidae sp. 6         1         0         0         0         3           Linyphiidae sp. 5         0         1         2         0         1           Euryopis sp. 1         0         0         0         3         2           Camillina procurva         1         0         1         1         2           Linyphiidae sp. 2         1         0         1         1         3           Linyphiidae sp. 4         1         1         2         0         2           Diores annettae         5         0         0         2         0         2           Zelotes uquathus         1         3         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1	•	0	0	0	2	1	
Trachelas pusillus         1         1         1         0         1           Linyphiidae sp. 6         1         0         0         0         3           Linyphiidae sp. 5         0         1         2         0         1           Euryopis sp. 1         0         0         0         3         2           Camillina procurva         1         0         1         1         2           Linyphiidae sp. 2         1         0         1         1         3         1         1         3           Linyphiidae sp. 4         1         1         2         0         2         0         2         0         2         0         2         0         2         0         2         0         2         0         2         0         2         0         2         0         0         3         1         1         1         1         1         1         1         1         1         1         1         1         1         1         2         0         2         0         0         3         1         1         1         1         1         1         1         1         1	-	-					
Linyphidae sp. 6	•	-					
Linyphiidae sp. 5         0         1         2         0         1           Euryopis sp. 1         0         0         0         3         2           Camillina procurva         1         0         1         1         2           Linyphiidae sp. 2         1         0         1         1         3           Linyphiidae sp. 4         1         1         2         0         2           Diores annettae         5         0         0         2         0         2           Zelotes uquathus         1         3         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         1         <		•	-				
Euryopis sp. 1 0 0 0 1 3 2 Camillina procurva 1 0 1 1 1 2 Linyphilidae sp. 2 1 1 0 1 1 1 3 Linyphilidae sp. 4 1 1 1 2 0 2 Diores annettae 5 0 0 0 2 0 Zelotes uquathus 1 3 1 1 1 1 Cydrela sp. 2 1 0 3 1 3 Linyphilidae sp. 3 1 1 1 1 1 Cydrela sp. 2 1 0 3 1 3 Linyphilidae sp. 3 0 0 3 1 4 Xerophaeus bicavus 1 3 1 2 3 Steatoda sp. 1 0 0 3 5 3 Steatoda sp. 1 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 0 3 5 5 3 Steatoda sp. 1 0 0 0 0 3 5 5 3 Steatoda sp. 2 15 8 7 8 5 5 Steatoda sp. 2 15 8 7 8 5 5 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 107 44 Steatoda sp. 2 10 8 10 8 10 9 54				_			
Camillina procurva       1       0       1       1       2         Linyphiidae sp. 2       1       0       1       1       3         Linyphiidae sp. 4       1       1       2       0       2         Diores annettae       5       0       0       2       0         Zelotes uquathus       1       3       1       1       1         Cydrela sp. 2       1       0       3       1       3         Linyphiidae sp. 3       0       0       3       1       4         Xerophaeus bicavus       1       3       1       2       3         Steatoda sp. 1       0       0       3       5       3         Phlegra sp. 1       3       2       2       2       3         Copa sp. 1       9       3       1       1       0         Linyphiidae sp. 8       4       3       3       2       2       2       3         Copa sp. 1       9       3       1       1       0       1       1       0       1       1       0       1       1       0       1       1       1       0       1       2		_	_		_		
Linyphiidae sp. 2			_				
Linyphiidae sp. 4 1 1 1 2 0 2 0 2 Diores annettae 5 0 0 0 2 0 Zelotes uquathus 1 3 1 1 1 1 Cydrela sp. 2 1 0 3 1 3 Linyphiidae sp. 3 0 0 3 1 3 Linyphiidae sp. 3 0 0 3 1 4 Xerophaeus bicavus 1 3 1 2 3 Steatoda sp. 1 0 0 3 5 3 Phlegra sp. 1 3 2 2 2 2 3 Copa sp. 1 3 2 2 2 2 3 Copa sp. 1 9 3 1 1 0 Linyphiidae sp. 8 4 3 3 3 2 4 Xysticus mulleri 7 0 4 3 4 Aelurillus sp. 1 7 3 2 3 4 Aelurillus sp. 1 7 3 2 3 4 Aelurillus sp. 1 7 5 4 2 Pardosa crassipalpis 3 0 10 4 8 Rhaeboctesis trinotatus 2 0 2 19 6 Trabea sp. 4 5 3 6 7 12 Proevippa albiventris 16 2 12 1 3 Cydrela schoemanae 19 8 5 4 6 Hogna sp. 5 15 8 7 8 5 Zelotes lightfooti 23 7 24 5 7 Trabea sp. 5 0 33 0 31 14 Proevippa sp. 2 32 13 33 26 41 Lycosidae sp. 1 50 18 51 9 54  Total individuals per site 246 148 201 288 262		•	-				
Diores annettae         5         0         0         2         0           Zelotes uquathus         1         3         1         1         1           Cydrela sp. 2         1         0         3         1         3           Linyphiidae sp. 3         0         0         3         1         4           Xerophaeus bicavus         1         3         1         2         3           Steatoda sp. 1         0         0         3         5         3           Phlegra sp. 1         3         2         2         2         3           Copa sp. 1         9         3         1         1         0           Linyphiidae sp. 8         4         3         3         2         4         4         3         4         4         4         4         4         4         4         4         4         4         4         4         4         3         4         4         4         4         4         4         4         4         4         4         4         4         4         4         4         4         4         4         2         2         2         2 <td< td=""><td></td><td>=</td><td>-</td><td>-</td><td></td><td></td><td></td></td<>		=	-	-			
Zelotes uquathus 1 3 1 1 1 1 Cydrela sp. 2 1 0 3 1 3 1 3 Linyphiidae sp. 3 0 0 3 1 4 4 Xerophaeus bicavus 1 3 1 2 3 Steatoda sp. 1 0 0 3 5 3 7 Nelegra sp. 1 3 2 2 2 2 3 Copa sp. 1 9 3 1 1 0 0 Linyphiidae sp. 8 4 3 3 2 2 4 Xysticus mulleri 7 0 4 3 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4	•	=	·				
Cydrela sp. 2       1       0       3       1       3         Linyphiidae sp. 3       0       0       3       1       4         Xerophaeus bicavus       1       3       1       2       3         Steatoda sp. 1       0       0       3       5       3         Phlegra sp. 1       3       2       2       2       3         Copa sp. 1       9       3       1       1       0         Linyphiidae sp. 8       4       3       3       2       4         Xysticus mulleri       7       0       4       3       4         Aelurillus sp. 1       7       3       2       3       4         Palpimanus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       4       5       3       6       7       12         Proevippa asp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1			_				
Linyphiidae sp. 3 0 0 0 3 1 4 4  Xerophaeus bicavus 1 3 1 2 3  Steatoda sp. 1 0 0 0 3 5 3  Phlegra sp. 1 3 2 2 2 2 3  Copa sp. 1 9 3 1 1 0  Linyphiidae sp. 8 4 3 3 2 2 4  Xysticus mulleri 7 0 4 3 4  Aelurillus sp. 1 7 3 2 3 4  Palpimanus  transvaalicus 4 1 0 12 2  Proevippa fascicularis 1 7 5 4 2  Pardosa crassipalpis 3 0 10 4 8  Rhaeboctesis  trinotatus 2 0 2 19 6  Trabea sp. 4 5 3 6 7 12  Proevippa albiventris 16 2 12 1 3  Cydrela schoemanae 19 8 5 4 6  Hogna sp. 5 15 8 7 8 5  Zelotes lightfooti 23 7 24 5 7  Trabea sp. 2 32 13 33 26 41  Lycosidae sp. 2 5 10 8 107 44  Lycosidae sp. 1 50 18 51 9 54	•	=		-			
Xerophaeus bicavus       1       3       1       2       3         Steatoda sp. 1       0       0       3       5       3         Phlegra sp. 1       3       2       2       2       3         Copa sp. 1       9       3       1       1       0         Linyphiidae sp. 8       4       3       3       2       4         Xysticus mulleri       7       0       4       3       4         Aelurillus sp. 1       7       3       2       3       4         Palpimanus       transvaalicus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2       2         Pardosa crassipalpis       3       0       10       4       8       8         Rhaeboctesis       trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12       7         Proevippa albiventris       16       2       12       1       3       3       2       1       3       4       6       6       7       1			_				
Steatoda sp. 1       0       0       3       5       3         Phlegra sp. 1       3       2       2       2       3         Copa sp. 1       9       3       1       1       0         Linyphiidae sp. 8       4       3       3       2       4         Xysticus mulleri       7       0       4       3       4         Aelurillus sp. 1       7       3       2       3       4         Palpimanus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31		-	_				
Phlegra sp. 1       3       2       2       2       3         Copa sp. 1       9       3       1       1       0         Linyphiidae sp. 8       4       3       3       2       4         Xysticus mulleri       7       0       4       3       4         Aelurillus sp. 1       7       3       2       3       4         Palpimanus       transvaalicus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0<							
Copa sp. 1       9       3       1       1       0         Linyphiidae sp. 8       4       3       3       2       4         Xysticus mulleri       7       0       4       3       4         Aelurillus sp. 1       7       3       2       3       4         Palpimanus       7       3       2       3       4         Palpimanus       1       7       5       4       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       1       1       7       12       1       1       1       1       1       1       1       1       1       1       1       1       1       1       1       1       1       1		_	_				
Linyphiidae sp. 8				2			
Xysticus mulleri       7       0       4       3       4         Aelurillus sp. 1       7       3       2       3       4         Palpimanus       1       7       3       2       3       4         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       10       4       8       8         Rhaeboctesis       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107							
Aelurillus sp. 1       7       3       2       3       4         Palpimanus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       10       4       8         trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54     <			3	3		4	
Palpimanus         transvaalicus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54	<del>-</del>	7	0			4	
transvaalicus       4       1       0       12       2         Proevippa fascicularis       1       7       5       4       2         Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54     Total individuals per site 246       148       201       288       262		7	3	2	3	4	
Proevippa fascicularis         1         7         5         4         2           Pardosa crassipalpis         3         0         10         4         8           Rhaeboctesis         trinotatus         2         0         2         19         6           Trabea sp. 4         5         3         6         7         12           Proevippa albiventris         16         2         12         1         3           Cydrela schoemanae         19         8         5         4         6           Hogna sp. 5         15         8         7         8         5           Zelotes lightfooti         23         7         24         5         7           Trabea sp. 5         0         33         0         31         14           Proevippa sp. 2         32         13         33         26         41           Lycosidae sp. 2         5         10         8         107         44           Lycosidae sp. 1         50         18         51         9         54    Total individuals per site  246  148  201  288  262						_	
Pardosa crassipalpis       3       0       10       4       8         Rhaeboctesis       trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54     Total individuals per site  246     148     201     288     262         Total species unique to       246       148       201       288       262			-				
Rhaeboctesis         trinotatus       2       0       2       19       6         Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54     Total individuals per site  246  148  201  288  262  Total species unique to							
trinotatus     2     0     2     19     6       Trabea sp. 4     5     3     6     7     12       Proevippa albiventris     16     2     12     1     3       Cydrela schoemanae     19     8     5     4     6       Hogna sp. 5     15     8     7     8     5       Zelotes lightfooti     23     7     24     5     7       Trabea sp. 5     0     33     0     31     14       Proevippa sp. 2     32     13     33     26     41       Lycosidae sp. 2     5     10     8     107     44       Lycosidae sp. 1     50     18     51     9     54    Total individuals per site  246  148  201  288  262  Total species unique to		3	0	10	4	8	
Trabea sp. 4       5       3       6       7       12         Proevippa albiventris       16       2       12       1       3         Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54     Total individuals per site  246  148  201  288  262  Total species unique to		•	•	•	40		
Proevippa albiventris         16         2         12         1         3           Cydrela schoemanae         19         8         5         4         6           Hogna sp. 5         15         8         7         8         5           Zelotes lightfooti         23         7         24         5         7           Trabea sp. 5         0         33         0         31         14           Proevippa sp. 2         32         13         33         26         41           Lycosidae sp. 2         5         10         8         107         44           Lycosidae sp. 1         50         18         51         9         54    Total individuals per site  Total species unique to							
Cydrela schoemanae       19       8       5       4       6         Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54    Total individuals per site 246 148 201 288 262 Total species unique to	•				-		
Hogna sp. 5       15       8       7       8       5         Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54    Total individuals per site 246 148 201 288 262 Total species unique to							
Zelotes lightfooti       23       7       24       5       7         Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54     Total individuals per site  246  148  201  288  262  Total species unique to	-						
Trabea sp. 5       0       33       0       31       14         Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54    Total individuals per site Total species unique to							
Proevippa sp. 2       32       13       33       26       41         Lycosidae sp. 2       5       10       8       107       44         Lycosidae sp. 1       50       18       51       9       54    Total individuals per site 246 148 201 288 262 Total species unique to	_						
Lycosidae sp.2       5       10       8       107       44         Lycosidae sp.1       50       18       51       9       54    Total individuals per site          246       148       201       288       262         Total species unique to       246       148       201       288       262							
Lycosidae sp.1       50       18       51       9       54         Total individuals per site       246       148       201       288       262         Total species unique to	Proevippa sp. 2	32	13	33	26	41	
Total individuals per site 246 148 201 288 262 Total species unique to	Lycosidae sp.2	5	10	8	107	44	
site 246 148 201 288 262 Total species unique to	Lycosidae sp.1	50	18	51	9	54	
site 246 148 201 288 262 Total species unique to							
site 246 148 201 288 262 Total species unique to	Total individuals per						
	site	246	148	201	288	262	
this site 12 8 3 9 5			_	_	_	_	
	tnis site	12	8	3	9	5	

**APPENDIX D:** Spider family prevalence per site

Family	Annual Farm	Biannual Farm	Communal	Reserve camp 3	Reserve camp 7	Total
Amaurobiidae	1	0	0	0	0	1
Araneidae	1	1	0	0	0	2
Clubionidae	1	0	0	2	0	3
Corinnidae	11	8	2	4	2	27
Cyrtaucheniidae	3	2	0	0	0	5
Gnaphosidae	30	15	28	13	16	102
Hahniidae	2	0	0	1	1	4
Idiopidae	1	0	0	0	1	2
Linyphiidae	9	8	12	7	17	53
Liocranidae	2	0	2	19	6	29
Lycosidae	130	95	132	197	183	737
Nemesiidae	0	1	0	1	1	3
Palpimanidae	4	1	0	12	2	19
Philodromidae	1	0	1	2	2	6
Phyxelididae	2	0	0	0	0	2
Prodidomidae	0	0	1	0	0	1
Salticidae	10	7	4	7	7	35
Scytodidae	1	0	0	0	1	2
Selenopidae	0	0	0	0	1	1
Sparassidae	0	1	0	1	1	3
Tetragnathidae	1	0	0	1	1	3
Theraphosidae	2	0	0	0	0	2
Theridiidae	0	1	3	9	6	19
Thomisidae	9	0	6	4	4	23
Zodariidae	25	8	10	8	10	61
Total	246	148	201	288	262	1145