

AN EPIDEMIOLOGICAL STUDY ON THE GENETIC RELATIONSHIPS OF FOOT AND MOUTH DISEASE VIRUSES IN EAST AFRICA

by Mesfin Sahle

A thesis submitted in partial fulfilment of the requirements for the degree of Doctor of Philosophy in the Department of Veterinary Tropical Diseases, Faculty of Veterinary Science, University of Pretoria, South Africa

February, 2004



Dedicated to my parents Sahle Fursa and Desta Sheferaw



ACKNOWLEDGMENTS

I would like to extent my sincere thanks to all lecturers and members of staff of Department of Veterinary Tropical Diseases, Faculty of Veterinary Science, University of Pretoria who contributed to the course work and assisted me during my studies. In particular I would like to thank the following people.

I acknowledge the most valuable assistance of my promoter Professor E.H. Venter and co-promoters Dr. W. Vosloo and Dr. R.M. Dwarka for constructive discussion and encouragement, every possible assistance and supervision all through my work.

The support given by Professor J.A.W. Coetzer, Head of Department of Veterinary Tropical Diseases, and his assistance in my personal problems during my staying here is gratefully acknowledged.

I would like to thank N. Ferris of the Animal Health Institute, Pirbright, UK for supplying us with the majority of the virus isolates used in this study.

I am grateful to Dr. A.D.S. Bastos for her assistance in constructive criticism of my project proposal and personal discussions. Particular thanks must be expressed to C.I. Boshoff and H.G. van Rensburg for their valuable discussion and all technical staff of the Exotic Diseases Division of the Onderstepoort Veterinary Institute for their technical assistance. Mention should also be made to my friends and colleagues in Ethiopia for their valuable discussions and assistance throughout my PhD project.

Finally, I would like to thank the National Animal Health Research Center and the Ethiopian Agricultural Research Institute for the award of a postgraduate scholarship under the Ethiopian Government and the South African Government for their assistance.

i



An epidemiological study on the genetic relationships of footand-mouth disease viruses in East Africa

by

Mesfin Sahle

Promoter

Prof. Estelle H. Venter

Department of Veterinary Tropical Diseases

University of Pretoria

Co-promoters

Dr. Wilna Vosloo

Dr. Rahana M. Dwarka

ARC- Onderstepoort Veterinary Institute, Exotic Diseases

Division

Degree

PhD

ABSTRACT

Within East African countries many of the known infectious diseases of animals occur commonly and are poorly controlled. Foot-and-mouth disease (FMD) is one of the contagious viral diseases that has great impact on economic development both in terms of direct and indirect losses. The epidemiology of the disease is complex due to the presence of six of the seven serotypes and the presence of large numbers of both wild and domestic susceptible animals in the region. Decision-making to determine the importance of FMD control relative to the economic consequences and what FMD control strategies should be applied based on the epidemiological information is required. In this regard the first step is to investigate the genetic relationships/variability of East African isolates and their phylogeographic distribution. These can provide base-line information for



designing control strategies by vaccination as well as the determination of the sources of infection.

Sufficient genetic information on the FMD serotypes O, SAT-1 and SAT-2 are lacking and therefore the number of viral lineages and genotypes or topotypes from East African countries could not be determined. Published studies on the relative occurrence and genotype distribution of FMD are largely confined to the southern and western part of the continent. In this study, the genetic profile of the 3 most prevalent serotypes (O, SAT-2 and SAT-1) recovered from outbreaks in East Africa between 1957 and 2003 was addressed. Phylogenetic analysis of partial and complete sequences of the 1D gene revealed the presence of distinct lineages and genotypes for East Africa as well as historical relationships of some of the genotypes with isolates from other regions. A great variation in the occurrence and distribution of these serotypes were found.

All the African and the Middle East/South East Asian isolates of serotype O included in this study clustered into one lineage having 8 distinct topotypes. These results indicated that between countries as well as inter-regional (east and west Africa) spread of viruses occurred in the past. Inter-regional spread of the virus between eastern Africa and western Africa was also confirmed for SAT-1 viruses. The fact that phylogenetic links are found with both serotypes implies that the spread of viruses was possibly associated with unrestricted animal movement due to nomadic movement in Africa. The phylogenetic relationships of SAT-1 viruses are more diversified in Africa. Eight lineages and 11 genotypes were identified when the optimal nucleotide sequence differences of ≥ 23% for lineages and ≥ 16% for genotypes were used as a cut-off values.

It was observed that viruses from Uganda are evolving independently from viruses elsewhere on the continent and clustered into 3 discrete lineages. In contrast, viruses from countries neighbouring Uganda, Kenya and Tanzania, clustered into one lineage. Uganda also harboured 3 topotypes of SAT-2 virus isolates, one is distinct for Uganda and the other are shared with Kenya and Zaire (DRC). This study highlighted distinct lineages found in Uganda and needs further investigation.



Within SAT-2, 67 isolates from 22 African countries and Saudi Arabia clustered into 5 lineages which consisted of 15 genotypes. Clustering of viruses into distinct genotypes (topotypes) according to year of isolation and geographical origin was observed showing countries with common boundaries shared common epizootics in the past. These results also showed a link between eastern and southern African countries.

Attempts were also made to investigate the incidence of FMD in Ethiopia using sera collected from cattle, small ruminants and wildlife. The results obtained from the liquid phase blocking ELISA and the 3ABC ELISA indicated the presence of SAT-1 and SAT-2 in buffalo populations in the southern part of Ethiopia while results from small ruminants and other wildlife were not indicative of any significant role in the epidemiology of FMD. Serological results also indicated that SAT-1 is present in cattle, although this serotype has not been previously identified.

The cumulative molecular epidemiological results from this and previous studies indicated that genetic variability of FMD viruses can be independently maintained within country/countries or regions as well as inter-regions of Africa. The serological results from buffaloes in East Africa are also suggestive of a possible reservoir of the SAT types FMD in the region which has a great impact on the control of the disease. Furthermore, the numerous lineages and genotypes of FMD virus isolates in Africa having distinct or overlapping distributions as well as the genetic linkage between regions will complicate the epidemiology of the disease. Therefore, it is strategically important to consider a regional approach and the use of a vaccine which contains a cocktails of antigens of FMD virus strains.



TABLE OF CONTENTS

		Pages
Acknow	vledgments	1
Abstrac	xt	II
Table o	f contents	V
List of	Tables	IX
List of I	Figures	ΧI
Abbrev	iations	XIII
Chapte	r I: Literature review	
1.1	Introduction	1
1.2	Historic and economic significance of foot-and-mouth disease	1
1.3	Taxonomy of Picornaviruses	3
1.3.1	Types (serotypes) and subtypes	4
1.4	Foot-and-mouth disease virus	4
1.4.1	Physico-chemical properties of the FMD viruses	4
1.4.2	Virus morphology	5
1.4.3	Genome organization and protein processing	5
1.4.4	Genetic variation of FMD virus genome	7
1.4.5	Antigenic variation	8
1.5	Diagnosis of FMD	9
1.5.1	Clinical signs	10
1.5.2	Laboratory diagnosis	13
1.6	Epidemiology and control of FMD	14
1.6.1	Geographic distribution of FMD	15
1.6.2	Serotype distribution of FMD in Africa	15
1.6.3	Susceptible host range	19
1.6.4	The role of carriers in the epidemiology of the disease	20
1.6.5	Transmission of FMD	20
1.6.6	Control of FMD	21
1.6.6.1	Control by vaccination	22
1.6.6.2	Control by stamping out	22
1.6.6.3	Control of FMD in developing countries	22



1.7	Molecular Phylogeny	23
1.8	Summary and objective of the study	26
	II: Molecular epidemiology of serotype O foot-and-mouth viruses isolated from cattle in Ethiopia between 1977 - 2001	27
2.2	Materials and Methods	29
2.2.1	Viruses studied	29
2.2.2	Nucleic acid isolation	30
2.2.3	Complementary DNA synthesis (cDNA)	31
2.2.4	Polymerase chain reaction (PCR)	31
2.2.5	Agarose gel electrophoresis of PCR products	32
2.2.6	Nucleotide Sequencing	32
2.2.7	Data Analysis	33
2.3	Results	33
2.3.1	Phylogenetic analysis of all serotype O isolates included in this study	33
2.3.2	Pair-wise comparison of partial VP1 gene sequences	36
2.3.3	Amino acid variability	37
2.4	Discussion	37
	r III: Molecular epidemiology of serotype O foot-and-mouth	
the wor	e viruses from East African countries in relation to the rest of rid	
3.1	Introduction	42
3.2	Materials and Methods	42
3.2.1	Viruses studied	42
3.2.2	RT-PCR amplification of the 1D gene and nucleotide	
	sequencing	44
3.2.3	Sequence data analysis	45
3.3	Results	45
3.3.1	Genetic relationships of serotype O FMD viruses	45
3.4	Discussion	49
•	r IV: Genetic heterogeneity of SAT-2 foot-and-mouth disease in East Africa	
4.1	Introduction	52



4.2	Materials and Methods	52
4.2.1	Viruses studied	52
4.2.2	Nucleic acid isolation and RT-PCR amplification	54
4.2.3	DNA purification and Cycle Sequencing	54
4.2.4	Phylogenetic Analysis	55
4.3	Results	55
4.3.1	Phylogenetic analysis	55
4.3.2	Sequence variations and distribution of mutations	59
4.4	Discussion	66
	r V: Molecular epidemiology of SAT-1 foot-and-mouth e viruses in East Africa isolated between 1971 and 2000 Introduction	69
5.2	Materials and Methods	70
5.2.1	Viruses used in this study	70
5.2.2	Nucleic acid isolation and RT-PCR amplification	71
5.2.3	DNA purification and Cycle Sequencing	71
5.2.4	Phylogenetic analysis	71
5.3	Results	72
5.3.1	Phylogenetic analysis	72
5.3.2	Sequence variations	76
5.4	Discussion	81
rumina	r VI: Serological survey to determine the role of small nts and wildlife in the epidemiology of foot and mouth e in Ethiopia and evaluation of serological tests Introduction	84
6.2	Materials and Methods	85
6.2.1	Study area	85
6.2.2	Serum samples	86
6.2.3	Antibody detection tests	89
6.2.3.1	Liquid phase blocking ELISA	89
6.2.3.2	The UBI FMDV non-structural enzyme-linked immunosorbent	90
6.2.4	Data analysis	91
6.3	Results	91
6.3.1	Analysis of sera from wildlife	92



6.3.2	Analysis of sera from small ruminants	93
6.3.3	Analysis of sera from cattle	94
6.4	Discussion	96
Chapte	er VII: General discussion and recommendations	
	General discussion and recommendation	99
Refere	nces	111
Appen	dix I	138
Appen	dix II	141



LIST OF TABLES

		Page
Table 1.1	Summary of the genus composition of the family	
	Picornaviridae	3
Table 1.2	Topotypes of FMD serotypes O, A, C, and South African	
	Territories types (SAT-1, -2 and -3) in Africa	17
Table 2.1	Summary of serotype O FMD viruses included in this study	29
Table 2.2	Average pair-wise distance comparison between genotypes	
	and/or clads of viruses studied	36
Table 3.1	Summary of serotype O FMD viruses included in this study	43
Table 3.2	Average pair-wise distance comparison between genotypes	
	1-11 of serotype O FMD viruses identified in this study	48
Table 3.3	Summary of genetically closely related viruses which	
	demonstrated < 6% nucleotide differences and possibly	
	originated from common sources	49
Table 4.1	Summary of SAT-2 FMD viruses included in this study	53
Table 4.2	Average pair-wise distance comparisons between lineages	
	(I-V) of SAT-2 FMD viruses included in this study	58
Table 4.3	Pair-wise sequence divergence using the P-distance model	60
Table 5.1	Summary of SAT-1 FMD viruses used in this study	70
Table 5.2	Average pair-wise distance comparisons between lineages of	
	SAT-1 FMD viruses included in this study	76
Table 6.1	Total number of serum samples collected per farming system	89
Table 6.2	Number of serum samples collected from randomly selected	
	district within the administrative regions	89
Table 6.3a	Summary of reactivity of serum samples tested by the Lpb	
	ELISA and 3ABC ELISA	92
Table 6.3b	Foot-and-mouth disease virus serotypes identified from	
	reactive sera of wildlife	93
Table 6.4	Summary of results of Lpb ELISA and 3ABC ELISA using	
	sera from small ruminants	94
Table 6.5	Summary of Lpb ELISA and 3ABC ELISA using sera from	
	cattle	95



Table 6.6	Cross tabulation comparison of reactive and non-reactive sera	
	from bovine, sheep and goats, wild ruminants and buffaloes	
	detected in Lpb ELISA and 3ABC-ELISA	96
Table 7.1	Summary of phylogenetic analysis of FMD virus isolates from	
	East Africa (EA) and elsewhere	100
Table 7.2	Summary of nucleotide sequence differences (%) used as a	
	cut-off values to define genotypes and lineages as well as	
	divergence (%) between genotypes within each serotype	101



LIST OF FIGURES

		Page
Fig. 1.1	Schematic representation of the FMD virus genome and empty	
	capsid construction and auto-assembly	7
Fig. 1.2	Clinical signs of FMD in indigenous Ethiopian cattle	11
Fig. 1.3	Map of Africa demonstrating the outbreaks of foot-and-mouth	
	disease between 1948 – 2002	18
Fig. 1.4	The probability of substitution of one nucleotide by another	
	between purines and/or pyrimidines	24
Fig. 2.1	UPGMA tree depicting genetic relationships of serotype O	
	FMD viruses from Ethiopia, Africa, the Middle East/Asia and	
	Europe	34
Fig. 2.2	Amino acid alignment of 165 amino acids of the C terminal end	
	of the VP1 protein of 36 type O FMD viruses	41
Fig. 3.1	Neighbour-joining tree depicting genetic relationships of	
	serotype O FMD viruses from Africa, the Middle East, Europe,	
	South America and Asia	47
Fig. 4.1	Neighbour-joining tree depicting genetic relationships of	
	serotype SAT-2 FMD viruses from East, West and southern	
	Africa and Saudi Arabia	57
Fig. 4.2	Amino acid sequence alignment of the 1D gene of 48 SAT-2	
	FMD viruses from East African countries	63
Fig. 4.3	Variability plot of sequences of 1D gene of SAT-2 viruses from	
	East Africa	65
Fig. 5.1	Neighbour-joining tree based on 396 nt of the 1D gene	
	depicting genetic relationships of 53 SAT-1 FMD viruses from	
	Africa	73
Fig. 5.2	Neighbour-joining tree based on 663 nt of 1D/2A gene	
	depicting phylogenetic relationships of 39 SAT-1 FMD virus	
	isolates from Africa	74
Fig. 5.3	Sequence alignment of 221 amino acids of the 1D gene of 36	
	SAT-1 FMD viruses	78
Fig. 5.4	Variability plot of complete sequences of the 1D gene of SAT-1	
	viruses from Africa	80



Fig. 6.1	Administrative map of the Federal Republic of Ethiopia,	
	depicting regions and zones	86
Fig. 6.2	Map of Ethiopia showing the different farming systems	87
Fig 7.1	Distribution of various Topotypes of FMD virus isolates	
	identified in Africa	
	A: Serotype O	108
	B: SAT-1	109
	C: SAT-2	110

LIST OF ABBREVIATIONS

BHK Baby Hamster Kidney cells

bp base pair

CBPP Contagious bovine pleuropneumonia

CCPP Contagious caprine pleuropneumonia

CFT Complement fixation test

cDNA Complementary deoxyribonucleic acid

dNTPs deoxynucleotide triphosphates

ed. editor

edt. edition

e.g for example

ELISA Enzyme-linked immunosorbent assay

EtOH Ethyl alcohol

Fig. Figure

FMD Foot and mouth disease

FMDV Foot and mouth disease virus

g Gram

x g Unit of acceleration

GDP Gross domestic production

H₂O₂ Hydrogen peroxide

H₂SO₄ Sulphuric acid

IBRS-2 Instituto Biologico Rim Sunio

Lpb ELISA Liquid phase blocking ELISA

kDa Kilo dalton

masl meter above sea level

mbsl meter below see level

M Molar

ML Maximum likelihood

MM Master mix

MOD Maximum optical density

MOE Ministry of Agriculture of Ethiopia

MP Maximum parsimony

m/v mass by volume

NaAc Sodium acetate



NJ Neighbour-joining

nt nucleotide

OD Optical density

OIE Office International des Epizooties

OPD o- phenelene diamine

PBS Phosphate buffered saline

PBS-C Phosphate buffer saline and casein

PBS-T Phosphate buffer saline and twee-20

PK Pig kidney

RT-PCR Reverse transcriptase polymerase chain reaction

RGD Arginine-Glycine-Asparic acid

Rnase Human placental ribonuclease

S Sedimentation coefficient

ssRNA Single stranded ribonucleic acid

SAT South African Territories

TCID₅₀ Tissue culture infective dose

UK United Kingdom

UPGMA Unweighted Pair-Group method using arithmetic average

VNT Virus neutralization test

WRL World Reference Laboratory