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Molecular epidemiology of rabies in KwaZulu Natal, South Africa.

ΒY

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Declaration

I hereby declare that this thesis, except where indicated, is my own research, and has not been submitted in part, or as a whole, for a degree at any other university.

Peter Coetzee

Signature:

Date: -----

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Dedications

Dedicated to my loving parents and

<u>brother</u>

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To my Creator, who gives me my strength, and who has directed my path.

Summary

MOLECULAR EPIDEMIOLOGY OF RABIES IN KWAZULU NATAL, SOUTH AFRICA

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In South Africa, two biotypes of type species 1 of the Lyssavirus genus are maintained independently among the members of the *Herpestidae* and *Canidae* families, respectively. Canid rabies is a relatively new addition to the African subcontinent, having been introduced from infectious cycles, which had existed among dogs in Angola, in the early 1940s. Two epidemics, believed to have originated from dog endemic regions which had existed in the southern Maputo district of Mozambique since 1952, have occurred among domestic dogs in the KwaZulu Natal province in recent years. The first of these epidemics started in 1964, and ended by 1968, while the second epidemic which started in 1976, has proven to be intractable, despite the concerted efforts which have been implemented to bring it under control. In order to contribute to the understanding of the molecular epidemiology of rabies in the KwaZulu Natal province, and to thereby assist in future surveillance and control efforts, we conducted a molecular sequence analysis of representative panel of viral isolates which were obtained from the province during the year 2003. A 591 nt. sequence encompassing the G-L intergenic region and glycoprotein cytoplasmic domain was sequenced for 128 viral isolates, which were obtained from the different magisterial districts and affected host species of the province, and was subsequently used to characterize these viruses phylogenetically.

Summary

Characterization of the KwaZulu Natal variants, and comparison of the obtained sequence data, to sequences data which was obtained from rabies endemic regions from elsewhere in South Africa and Zimbabwe, in general supported the pattern of spread which led to the introduction of rabies into the province, as was previously suggested from the literature. The phylogeny which was established from the analyses, indicated that the viral isolates from the province were highly related to each other, and could be divided into two groups, which although belonging to the canid biotype, were in general distinguishable from canid rabies virus isolates which were obtained from elsewhere in South Africa and Zimbabwe. The observation that these subfamilies showed a low genetic divergence, as well as that they shared a unique recent common ancestor, suggested that they were introduced recently into the northern reaches of the province, probably from the same geographical region (i.e. southern Mozambique).

Phylogenetic characterization of the KwaZulu Natal isolates further suggested that at least three enzootic fronts are currently responsible for the introduction of rabies into the northern and southern regions of the province. The first of these fronts was hypothesized to have spread directly across the southern Mozambique border (possibly via southeastern Swaziland), into the northeastern coastal regions of KwaZulu Natal, while the second front represented the south-eastwards spread of synergistic dog-jackal cycles from southeastern Mpumalanga, into the northern and northeastern regions of the province. The third front on the other hand, represented the possible spread of a remnant infectious cycle, left over from the 1964-1968 epidemic, from the northern region of the Eastern Cape, into southern KwaZulu Natal.

Phylogenetic characterization further proved useful for identifying the distribution of viral variants, and allowed us to propose a pathway by which the disease might have spread throughout the province. The proposed pathway of spread suggested that viral variants may have been translocated over long distances, and highlighted the role that major routes of human transportation may play in the dissemination of the disease. The regional characterization of viral variants from KwaZulu Natal, further demarked the location where the identified viral variants circulated in individual host populations, allowing us to place

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Summary

the current epidemic into an epidemiological framework which attempts to explain the long term persistence of the disease. This provided clues as to the intractability of the second epidemic, and allowed us to develop a proposal as to how current control strategies may be altered, in order to contain the current outbreak in the province.

The initial phylogeny which was established from the study provides an epidemiological framework, which will play an important role in determining the origin of future human spillover cases, and for tracking the spread of viral variants throughout the affected regions of the province. It is further envisaged that the data which was generated during the course of the project will be utilized in future surveillance efforts, targeted to the evaluation of the efficacy of potentially implemented control campaigns.

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| μm | micrometer |
|---------|--------------------------------------|
| A | Adenosine |
| Ab | antibody |
| aa | amino acid |
| A.D. | Anno Domino |
| ABI | Applied Biotechnologies Incorporated |
| ABLV | Australian Bat Lyssavirus |
| AChR | acethycholine receptor |
| AIDS | Acquired Immune Deficiency Syndrome |
| approx. | approximately |
| Arg | arginine |
| ATP | adenosine triphosphate |
| B.C. | before Christ |
| BCG | bacille Calmette-Guerin |
| bp | base pair |
| С | cytosine |
| ca. | calculated |
| cDNA | complementary DNA |
| CNI | close neighbor interchange |
| CNS | central nervous system |
| CRU | cellular receptor unit |
| CVS | challenge virus standard |
| CW | COW |
| Da. | Dalton |
| ddNTP | dideoxyribonucleotide triphosphate |

| direct fluorescent antibody test |
|---------------------------------------|
| dog |
| defective interfering particle |
| deoxyribonucleic acid |
| deoxyribonucleotide triphosphate |
| dithiothreitol |
| Duvenhage Virus |
| for example |
| European Bat Lyssavirus 1 |
| European Bat Lyssavirus 2 |
| Eastern Cape |
| edition |
| electron microscopy |
| endoplasmic reticulum |
| Evelyn Rokitnicki Abelseth |
| and others |
| acceleration of gravity |
| guanosine |
| glycoprotein |
| glycine, aspartic acid and asparagine |
| glutamine |
| glycine |
| Global positioning system |
| genotype |
| hydrochloric acid |
| |

| HDCV | human diploid cell vaccine |
|------|---|
| HEP | Flury high egg passage |
| hm | human |
| HRIG | human rabies immunoglobin |
| i.e. | in other words |
| ICTV | International Committee for the Taxonomy of Viruses |
| IDT | Integrated DNA Technologies |
| IFA | indirect fluorescent antibody test |
| IL | interleukin |
| lle | isoleucine |
| Inc. | incorporated |
| IHNV | Infectious Hematopoietic Necrosis Virus |
| j | Canis mesomelas |
| kb | kilobase pairs |
| KCI | potassium chloride |
| km | kilometer |
| KZN | KwaZulu Natal |
| L | polymerase (L) protein |
| LBV | Lagos Bat Virus |
| Le | leader RNA |
| Lys | lysine |
| m | Cynictis penicillata |
| Μ | matrix protein |
| MAb | monoclonal antibody |

| methionine |
|---|
| magnesium chloride |
| millilitre |
| maximum likelihood |
| millimolar |
| Moloney Murine Leukemia Virus |
| Mokola Virus |
| maximum parsimony |
| messenger ribonucleic acid |
| nucleoprotein |
| non-virion protein |
| sodium hydroxide |
| nanogram |
| National Institute for Communicable Diseases |
| neighbourhood joining |
| nanometres |
| neurotrophin |
| nucleotide |
| Otocyon megalotis |
| open reading frame |
| Onderstepoort Veterinary Institute |
| degree Celsius |
| operational taxonomic unit |
| phosphoprotein |
| |

| PAGE | polyacrylamide gel electrophoresis |
|-----------|---|
| PAHO/AMRO | Panamerican Health Organization/Americas |
| PBS | phosphate buffered saline |
| PCEC | purified chick embryo cell vaccine |
| PCR | polymerase chain reaction |
| PEP | post exposure prophylaxis |
| РМ | Pitman Moore |
| pMol | pico molar |
| pp. | page |
| Pu | purine |
| PV | Pasteur virus |
| Ру | pyrimidine |
| RABV | rabies virus |
| RFLP | restriction fragment length polymorphism |
| RI | replicative intermediate |
| RIG | rabies immunoglobin |
| RNA | ribonucleic acid |
| RNP | ribonucleoprotein |
| RT | reverse transcriptase |
| RT-PCR | reverse transcriptase polymerase chain reaction |
| SAD | Street Alabama Dufferin |
| SEARG | South and East African Rabies Group |
| Ser | serine |
| SDS | sodium dodecyl sulphate |

| Shp | sheep |
|-------|---|
| SP | signal peptide |
| SPU | Special Pathogens Unit |
| т | thiamine |
| Taq | Thermus aquaticus |
| TBSV | Tomato Bushy Stunt Virus |
| TMV | Tobacco Mosaic Virus |
| TRAIL | Tumor Necrosis Factor Related Apoptosis Inducing Ligand |
| TTP | Transcirption termination and polyadenylation signal |
| U | unit |
| U | uracil |
| UPGMA | unweighted pair group method with arithmetic mean |
| USA | United States of America |
| UV | ultraviolet |
| V | volt |
| VNAb | virus neutralizing antibody |
| VRG | vaccinia recombinant glycoprotein |
| VSV | Vesicular Stomatitis Virus |
| WHO | World Health Organization |
| х | times |
| mg | milligram |
| μΙ | microliter |
| Ψ | G-L intergenic region, or pseudogene |

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