

## Supplementary information

### Accessing the genomic information of unculturable oceanic picoeukaryotes by combining multiple single cells

Jean-François Mangot<sup>1\*</sup>, Ramiro Logares<sup>1</sup>, Pablo Sanchez<sup>1</sup>, Fran Latorre<sup>1</sup>, Yoann Seeleuthner<sup>2,3,4</sup>, Samuel Mondy<sup>2,3,4</sup>, Michael E. Sieracki<sup>5,6</sup>, Olivier Jaillon<sup>2,3,4</sup>, Patrick Wincker<sup>2,3,4</sup>, Colomban de Vargas<sup>7,8</sup>, Ramon Massana<sup>1\*</sup>

#### Author affiliations

<sup>1</sup> Department of Marine Biology and Oceanography, Institute of Marine Sciences (ICM)–CSIC, Pg. Marítim de la Barceloneta, 37-49, Barcelona E-08003, Spain.

<sup>2</sup> CEA, Institut de Génomique, Génoscope, 2 Rue Gaston Crémieux, Evry F-91000, France.

<sup>3</sup> CNRS, UMR 8030, CP5706, Evry, F-91000, France.

<sup>4</sup> Université d'Evry, UMR 8030, CP5706, Evry, F-91000, France.

<sup>5</sup> National Science Foundation, 4201 Wilson Boulevard, Arlington, VA 22230, USA.

<sup>6</sup> Bigelow Laboratory for Ocean Sciences, 60 Bigelow Drive, East Boothbay, ME 04544, USA.

<sup>7</sup> CNRS, UMR 7144, Station Biologique de Roscoff, Place Georges Teissier, Roscoff, F-29680, France.

<sup>8</sup> Sorbonne Universités, UPMC Université Paris 06, UMR 7144, Station Biologique de Roscoff, Place Georges Teissier, Roscoff, F-29680, France.

\* Correspondence and requests for materials should be addressed to J-F.M. ([jean-francois.mangot@wanadoo.fr](mailto:jean-francois.mangot@wanadoo.fr)) or R.M. ([ramonm@icm.csic.es](mailto:ramonm@icm.csic.es)).

Tel: (+34) 93 2309500; Fax: (+34) 93 2309555.

## Supplementary Tables

**Table S1. Main physical-chemical characteristics of the two sampled stations.**

Stations	Coordinates	Sampling date	Depth (m)	Temperature (°C)	Oxygen ( $\mu\text{mol kg}^{-1}$ )	Salinity (psu)	Chlorophyll ( $\text{mg Chl.m}^{-3}$ )
23	42° 10' 12" N, 17° 43' 12" E	18/11/2009	55	15.7	224.3	38.4	0.06
41	14° 33' 36" N, 70° 0' 36" E	30/03/2010	58	27.1	148.3	36.5	0.47

**Table S2. General sampling and sequencing characteristics of the different individual SAGs of MAST-4A and MAST-4E.**

Species	SAGs ID	Stations *	Sequencing depth (Gbp)	Sequencing platforms	CV**
MAST-4A	AA538-M19	23	6.9	Hiseq (Genoscope)	
	AA538-N22	23	8.5	Hiseq (Genoscope)	
	AA538-F10	23	6.0	Hiseq (Genoscope)	
	AA538-G04	23	4.7	Hiseq (Genoscope)	
	AA538-G20 <sup>†</sup>	23	4.6	Hiseq (Genoscope)	
	AA538-K07	23	4.0	Hiseq (Genoscope)	
	AA538-E21	23	5.7	Hiseq (Genoscope)	
	AA538-C11	23	2.7	Hiseq (Oregon)	
	AA538-E15	23	6.4	Hiseq (Genoscope)	
	AB537-A17	41	4.0	Hiseq (Oregon)	
	AA538-E19	23	2.4	Hiseq (Oregon)	
	AA538-G20_bis <sup>†</sup>	23	6.8	Hiseq (Oregon)	
	AA538-J18	23	4.8	Hiseq (Genoscope)	
	AB537-K04	41	3.5	Hiseq (Oregon)	
MAST-4E	AA538-A02	23	4.5	Hiseq (Genoscope)	
	AA538-A03	23	4.5	Hiseq (Genoscope)	
	AA538-C05	23	4.6	Hiseq (Genoscope)	
	AA538-F08	23	4.0	Hiseq (Genoscope)	
	AA538-J09	23	4.7	Hiseq (Genoscope)	
	AA538-A11	23	6.8	Hiseq (Genoscope)	
	AA538-L23	23	4.4	Hiseq (Genoscope)	
	AA538-M11	23	4.2	Hiseq (Genoscope)	
	AA538-N16	23	4.9	Hiseq (Genoscope)	
Mean (SE)	all MAST-4A SAGs		5.1 (1.7)		34.0%
	all MAST-4E SAGs		4.7 (0.8)		17.3%
	all SAGs		4.9 (1.4)		28.8%

\* Stations 23 and 41 are located in the Mediterranean sea (Adriatic Sea) and Indian Ocean (Arabic Sea), respectively (<http://taraoceans.sb-roscoff.fr/EukDiv/#figureW1>).

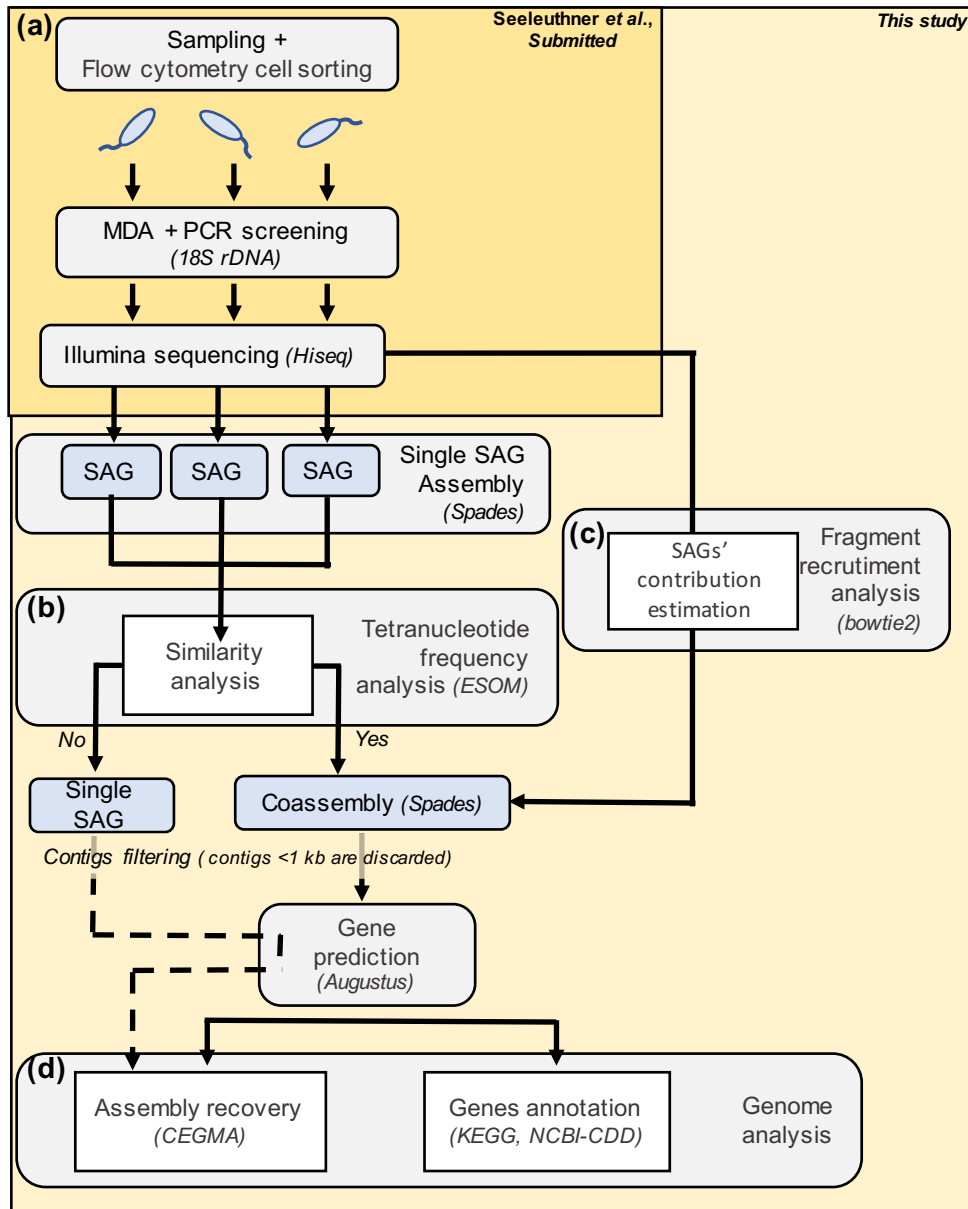
\*\* CV: Coefficient of variation = Standard error/mean.

<sup>†</sup> SAG sequenced by two different sequencing centers, the two sequencing replicates (AA538\_G20 and AA538\_G20\_bis) were kept for further analysis.

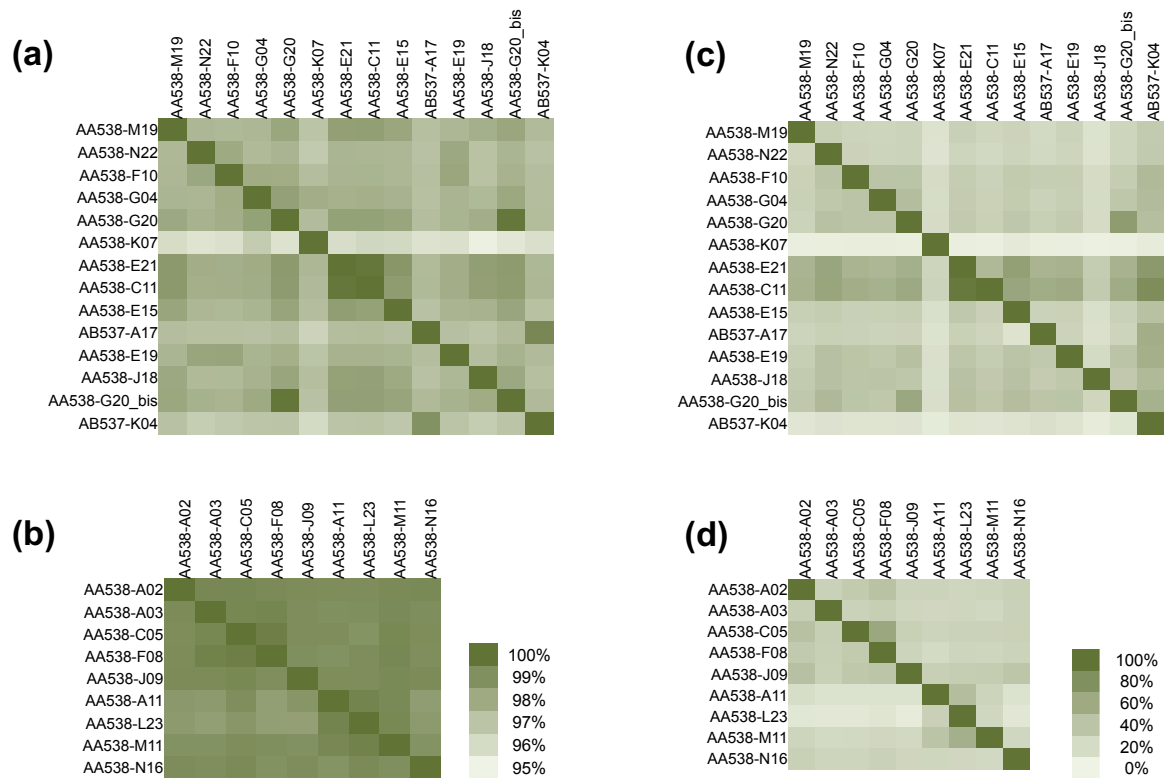
**Table S3. General functions present in MAST-4A and MAST-4E genomes based on protein classification according to KOGs of the 248 universal CEGMA eukaryotic genes.**

Functioning Process	General Functions	Number of COGs expected	Number of COGs in MAST-4A	Number of COGs in MAST-4E
Information storage and processing	Translation, ribosomal structure and biogenesis	34	29	25
	RNA processing and modification	23	18	16
	Transcription	13	6	9
	Replication, recombination and repair	10	9	7
	Chromatin structure and dynamics	0	0	0
	Shared functions	6	5	4
Cellular processes and signalling	Cell cycle control, cell division, chromosome partitioning	2	2	2
	Nuclear structure	0	0	0
	Defence mechanisms	0	0	0
	Signal transduction mechanisms	4	3	3
	Cell wall/membrane/envelope biogenesis	1	0	0
	Cell motility	0	0	0
	Cytoskeleton	3	2	2
	Extracellular structures	0	0	0
	Intracellular trafficking, secretion, and vesicular transport	17	13	13
	Posttranslational modification, protein turnover, chaperones	42	30	31
Shared functions	6	3	2	
Metabolism	Energy production and conversion	22	13	12
	Carbohydrate transport and metabolism	11	8	9
	Amino acid transport and metabolism	2	2	2
	Nucleotide transport and metabolism	6	4	4
	Coenzyme transport and metabolism	2	2	1
	Lipid transport and metabolism	5	3	2
	Inorganic ion transport and metabolism	3	3	0
	Secondary metabolites biosynthesis, transport and catabolism	1	1	1
	Shared functions	4	4	4
Poorly characterized	General function prediction only	19	15	13
	Function unknown	6	4	3
Shared between different functioning process		6	5	4

## Supplementary Figures

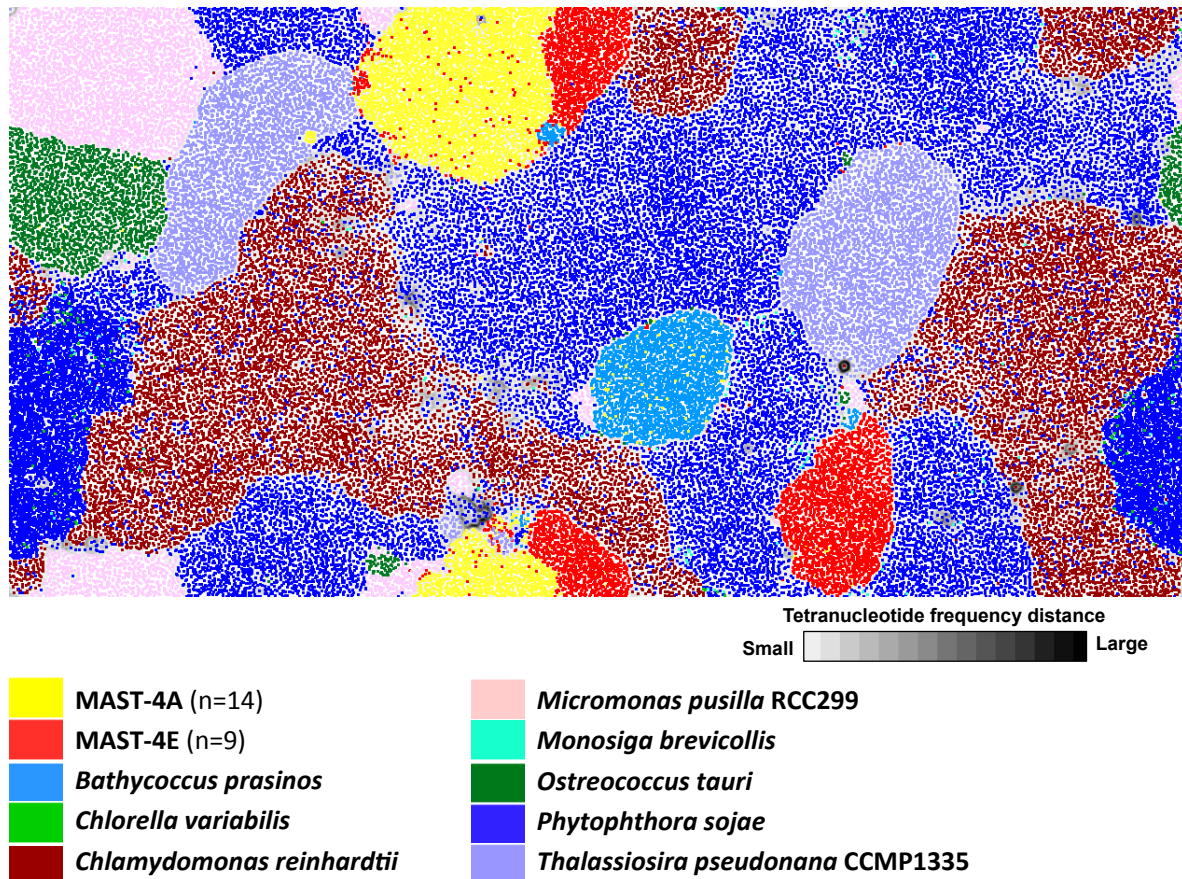


**Supplementary Fig. S1. Schematic pipeline of a single-cells co-assembly performed in this study.** Details on the sampling, single-cell sorting and SAG sequencing will be available in a concomitant study (Seeleuthner *et al.*, submitted). The rest of the main steps of our co-assembly strategy are described in this paper.



**Supplementary Fig. S2. Cross-SAG Blast analysis between MAST-4A and MAST-4E**

**SAGs.** Mean pairwise genomic similarity of MAST-4A **(a)** and MAST-4E **(b)** SAGs are represented, together with the percentage of shared nucleotidic regions for MAST-4A **(c)** and MAST-4E **(d)**. Values derive from blasting full-length contigs of each SAG against full-length contigs of each sister SAG. Query and subject SAGs are listed in the left and top of each heatmap, respectively.



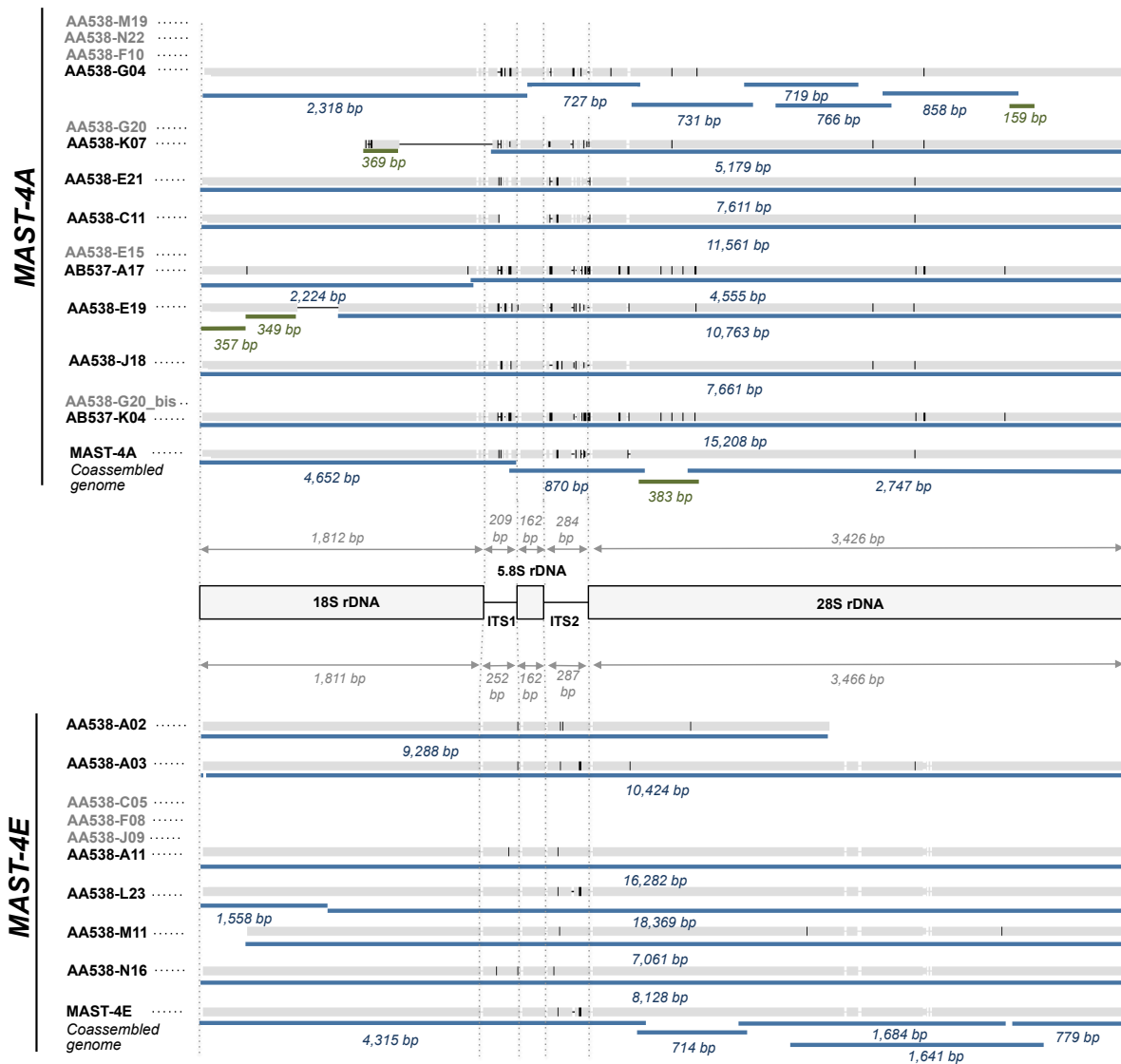
**Supplementary Fig. S3. Comparing tetranucleotide frequencies among selected genomes in an ESOM map.** Published protist genomes belonging to separate supergroups are combined together with MAST-4A and MAST-4E SAGs. Bestmatches of contigs of 2.5-5 kbp in size are represented by individual points, coloured according to their provenance as MAST-4A (yellow), MAST-4E (red), *Ostreococcus tauri* (dark green), *Micromonas pusilla* (light blue), *Bathycoccus prasinos* (blue), *Chlorella variabilis* (light green), *Chlamydomonas reinhardtii* (dark red), *Thalassiosira pseudonana* (purple), *Phytophthora sojae* (dark blue) and *Monosiga brevicollis* (pink). Large differences in tetranucleotide frequencies represent natural divisions between taxonomic groups.







identities of the retrieved CEGs were calculated among SAGs (“SAGs vs SAGs”) and between SAGs and co-assembly (“SAGs vs Coass.”).



**Supplementary Fig. S5. Retrieval of the rDNA operon in SAGs of the two MAST-4 lineages.** Sequences of MAST-4A (top) and MAST-4E (bottom) individual SAGs containing the rDNA operon were aligned with their corresponding co-assembled genomes. SAGs without rDNA operon contigs are shown in grey. The position and length of contigs from each SAG and co-assemblies necessary to reconstruct the rDNA operon are shown (contigs <500 bp in green and >500 bp in blue). Differences in individual SAGs against the consensus sequence are marked.