

ANIMAL RESEARCH PAPER

Effects of xylanase supplementation on feed intake, digestibility and ruminal fermentation in Rambouillet sheep

L. H. VALLEJO¹, A. Z. M. SALEM^{1*}, L. M. CAMACHO², A. M. KHOLIF³, M. D. MARIEZCURRENA⁴, M. CIPRIANO², M. U. ALONSO¹, J. OLIVARES² AND S. LOPEZ⁵

¹ *Facultad de Medicina Veterinaria y Zootecnia, Universidad Autónoma del Estado de México, Toluca, México*

² *Unidad Académica de Medicina Veterinaria y Zootecnia, Universidad Autónoma de Guerrero, Cd. Altamirano-Iguala, Guerrero, México*

³ *Dairy Science Department, National Research Centre, 33 Bohouth St. Dokki, Giza, Egypt*

⁴ *Facultad de Ciencias Agrícola, Universidad Autónoma del Estado de México, Toluca, México*

⁵ *Instituto de Ganadería de Montaña (IGM) CSIC-Universidad de León, Departamento de Producción Animal, Universidad de León, E-24071 León, Spain*

(Received 16 May 2015; revised 13 January 2016; accepted 17 February 2016;
first published online 6 April 2016)

SUMMARY

The present study aimed to investigate the effects of adding xylanase enzyme (XY) to a basal diet containing 300 g maize stover and 700 g concentrate/kg dry matter (DM) on feed intake, ruminal fermentation, total tract and ruminal digestibility, as well as some blood parameters. Four male Rambouillet sheep (39 ± 1.8 kg body weight), with permanent rumen and duodenum cannulae were used in a 4×4 Latin square design. Sheep were fed a basal diet without xylanase addition (control, XY0), or with the addition of xylanase at 1 (XY1), 3 (XY3) or 6 (XY6) μg /g of diet DM for 84 days, with four 21-day experimental periods. Feed intake, digestibility and rumen fermentation parameters were determined on days 16–21 in each experimental period, and the apparent ruminal neutral detergent fibre (NDF) digestibility was determined on days 16 and 17. Treatments XY1 and XY3 increased feed intake, whereas digestibility was increased with XY6. Ruminal NDF digestibility increased when sheep were fed diets treated with xylanase. Ruminal pH, ammonia-N and acetic acid increased with xylanase treated diets. Propionic acid concentration increased with diet XY1 at 3 h post-feeding, but after 9 h post-feeding its concentration decreased in the rumen of sheep fed xylanase treated diets. Xylanase had no effect on blood urea, phosphorus and triglycerides. Addition of xylanase at 6 μg /g DM in a diet containing 300 g maize stover and 700 g concentrate/kg DM and fed to Rambouillet sheep improved feed digestibility and ruminal fermentation without affecting blood parameters.

INTRODUCTION

Improving feed utilization in ruminant nutrition is one of the most important features that determine farming profitability. Many strategies have been considered to improve feed utilization; including, for example, the use of live yeast (Elghandour *et al.* 2014), phytogenic extracts (Salem *et al.* 2014) or fibrolytic enzymes (Valdes *et al.* 2015).

A large number of commercial enzyme products, either from fungal or bacterial sources, in relatively concentrated and purified forms and containing

specific controlled enzyme activities, have been used in livestock feeding (Dean *et al.* 2013; Abdel-Aziz *et al.* 2015). Feeding fibre-degrading enzymes seems to improve feed utilization as well as animal performance (Khattab *et al.* 2011; Alsersy *et al.* 2015), but the mode of action remains unclear. Some possible modes of action have been postulated including hydrolysis of dietary fibre before ingestion, synergistic interaction with endogenous microbial enzymes within the rumen (Morgavi *et al.* 2004), favoured ruminal fermentation (Salem *et al.* 2013; Rojo *et al.* 2015), and enhanced ruminal microorganisms attachment and colonization to the plant cell wall (Wang *et al.* 2001).

* To whom all correspondence should be addressed. Email: asalem70@yahoo.com

Although some of the reported results on supplementing animal rations with fibrolytic enzymes are encouraging, they are also inconsistent. In some studies, exogenous enzymes improved feeding value and animal performance by enhancing fibre degradation, increasing intake and feed digestion *in vitro* (Salem *et al.* 2015a), *in situ* (Togtokhbayar *et al.* 2015) and *in vivo* (Salem *et al.* 2015b; Morsy *et al.* 2016). However, in other studies no effects of exogenous enzymes on feed intake and digestion were observed (Elwakeel *et al.* 2007; Dean *et al.* 2013). Although the reasons for this discrepancy are unknown, it could be due to differences in enzyme activity, application rate and composition, type of diet fed to the animals, physiological stage of the animal, time of enzyme delivery, ruminal activity and enzyme stability, enzyme-feed specificity and the portion of the diet to which enzymes are applied (Dean *et al.* 2013).

Exogenous enzyme may affect some serum metabolites that reflect the nutritional and health status of animals (Morsy *et al.* 2016). Xylanase is an exogenous enzyme that may alter ruminal degradation of feeds and change concentrations of fermentation end-products (Lin *et al.* 1995). Moreover, it may also cause an indirect glucose sparing effect through the pentose-phosphate pathway (Jackson & Nicolson 2002).

The objective of the present study was to investigate effects of adding an exogenous xylanase enzyme at different application rates on feed intake, ruminal fermentation, total tract and ruminal digestibility, and blood urea, phosphorus and triglyceride concentrations in Rambouillet sheep fed a basal diet with 300 g maize stover and 700 g concentrate/kg dry matter (DM).

MATERIALS AND METHODS

All procedures involved in handling animals during the experimental period were conducted according to the official Mexican standard of animal care (NOM-051-ZOO-1995).

Study location

The experiment was conducted at the animal metabolic unit and the laboratory of animal nutrition of the Colegio de Postgraduados, Texcoco, Montecillo, Estado de México, México (2240 m a.s.l.). The climate is moderately humid with an average temperature of 15–18 °C and annual rainfall of 650 mm. The experiment was conducted during the autumn.

Table 1. *Ingredients and chemical composition of the basal diet fed to Rambouillet sheep (g/kg dry matter (DM), unless otherwise stated)*

	g/kg DM
Ingredients	
Ground sorghum grain	520
Maize stover	300
Molasses	80
Soybean meal	60
Urea	40
Chemical composition	
Dry matter (g/kg fresh matter)	870
Organic matter	950
Crude protein	154
Ether extract	57
Neutral detergent fibre	448
Acid detergent fibre	252
Phosphorus	4.3
Calcium	2.5

Enzyme activity

The enzyme product (Xylanase[®] plus, Dyadic[®] PLUS, Dyadic International, Inc., Jupiter, FL, USA) was assessed for endoglucanase and xylanase activities as described by Robyt & Whelan (1972) by catalytic hydrolysis of xylan from oat spelt and determining the released reducing groups using alkaline copper reagent. The product contained 34 000–41 000 units of xylanase/g, 12 000–15 000 units of β -glucanase/g and 45 000–55 000 units of cellulase/g.

Animals, housing and feeding

Four Rambouillet rams, weighing 39 ± 1.8 kg body weight (BW) and fitted with permanent cannulae in the rumen (2.5 cm internal diameter (i.d.)) and duodenum (T-type 0.8 cm i.d.) were used. The sheep were housed in individual cages equipped with high flow valve steel water bowls and fed a basal diet composed of 300 g maize stover and 700 g concentrate/kg DM (Table 1) *ad libitum* for 84 days. The basal diet was balanced for minerals and vitamins and formulated to match the nutrient requirements of sheep according to NRC (1985) recommendations plus a margin of 0.10. The ingredients and chemical composition of the basal diet are shown in Table 1. At the beginning of the experiment, sheep were treated with Ivermectin (Ivomec[®]-F-1 1 ml/50 kg BW, subcutaneous), Bacterin (Covexin[®] 10 ml/animal; intramuscular) and vitamins A, D and E (Vigantol[®] ADE 1 ml/animal, intramuscular).

The experiment was laid out according to a 4 × 4 Latin square design with four treatments, i.e. four application rates of xylanase (XY), namely 0 (control, XY0), 1 (XY1), 3 (XY3) and 6 (XY6) µl/g DM of the basal diet. In the first experimental period, treatments were assigned randomly to the experimental units (sheep). Experimental periods consisted of 21 days with days 1–15 considered as the adaptation period to the experimental diets and days 16–21 as the measurement and sample collection period. Sheep were fed twice daily in two equal meals at 07:00 and 19:00 h. The enzyme was added at the corresponding application rate, mixed with the diet individually and fed at 07:00 h. During the collection period, i.e. days 16–21, the amount of feed offered was recorded and orts collected and weighed for determination of daily feed intake. Additionally, feeds were sampled daily, composited weekly, dried at 60 °C to constant weight and stored for later chemical analysis.

Feed digestibility

Total tract digestibility was determined by total faecal collection during days 16–21 of each period. Faeces were collected daily before the morning feeding and stored at –10 °C for later analysis. A sub-sample of about 100 g/kg of the total faeces collected from each sheep was taken daily and composited for chemical analysis.

Apparent ruminal fibre digestibility was determined on days 16 and 17 following the procedure of Kozloski *et al.* (2014). Duodenal digesta samples (approximately 50 ml) were collected from each sheep 4 h after morning feeding and then at 4 h intervals over a period of 48 h. Samples were obtained from the duodenal cannula, collected in a 100 ml amber vial and immediately frozen until analysis. Samples were subsequently thawed and dried at 55 °C for 48 h, homogenized and analysed for neutral detergent fibre (NDF) and acid detergent fibre (ADF).

Dried feed, feed orts and faecal samples were ground through a Wiley mill (Arthur H. Thomas, Philadelphia, PA, USA) using a 1 mm screen and analysed according to AOAC (1997) for DM (#930-15), ash (#942-05), ether extract (EE; #920-39), nitrogen (N; #954-01) and ADF (#973-18), while NDF was analysed according to Van Soest *et al.* (1991). Organic matter (OM, g/kg DM) content was calculated by difference (1000-g ash/kg DM).

Rumen and blood sampling and analysis

Rumen fluid was collected on day 18 of each experimental period, directly through the rumen cannula

from the ventral sac of each sheep at 3, 6 and 9 h after morning feeding. The rumen samples (approximately 50 ml/sheep) were filtered immediately through four layers of cheesecloth, strained and stored in 45 ml glass bottles. Ruminal fluid pH was then determined using a portable pH meter (Orion, model SA 210, USA). Subsequently, 4 ml of rumen fluid was mixed with 1 ml of a solution of metaphosphoric acid (250 g/l) in a test tube and stored at –18 °C for subsequent volatile fatty acid (VFA) analysis. A sub-sample of 5 ml of rumen fluid was acidified with 5 ml of 0.2 M hydrochloric acid (HCl) for ammonia-N analysis.

Concentrations of acetic, propionic and butyric acids in rumen fluid were measured by gas-liquid chromatography (Hewlett Packard, Little Falls, DE, USA) using a capillary column (30 m length, 0.32 mm i.d., 0.25 mm film thickness; Elite-FFAP, Perkin Elmer Instruments, Shelton, WA, USA) according to the method of Erwin *et al.* (1961). The injector temperature was set at 240 °C, flame ionization detector at 250 °C and oven at 140 °C with hydrogen gas (H₂) and air flows at 40 and 400 ml/min, respectively.

Concentrations of ammonia-N were determined photometrically in an ultraviolet light spectrophotometer (VARIAN CARY 1-E, CA, USA) set at a wavelength of 630 nm according to McCullough (1967).

On day 21 of each experimental period and 4 h after morning feeding, a sample of 10 ml of blood was collected via jugular vein of each sheep into clean dry test tubes, without anticoagulant. Blood samples were centrifuged at 5000 g at 4 °C for 20 min. Serum was separated into 2 ml Eppendorf tubes and frozen at –20 °C until analysis. Blood serum samples were analysed for concentrations of urea, phosphorus and triglycerides using specific kits (Stanbio Laboratory, Boerne, TX, USA) according to the manufacturer's specifications.

Statistical analysis

Data on feed intake, digestibility, ruminal fermentation parameters (at each time post-feeding) and blood parameters were examined by analysis of variance according to a 4 × 4 Latin square design with four periods and four experimental diets (XY0, XY1, XY3, XY6) using PROC MIXED of SAS (SAS Institute 2006). One ram was used within each period and treatment. The statistical model was:

$$Y_{ijkl} = \mu + A_i + P_j + T_k + \varepsilon_{ijkl}$$

where Y_{ijkl} is the observation for a given response variable, μ is the overall mean, A_i is the random effect of

Table 2. Feed intake, digestibility and blood metabolites in Rambouillet sheep fed a diet treated with increasing concentrations of xylanase (XY)

	Diets*					S.E.M. (n = 4)	P value				
	XY0	XY1	XY3	XY6	Treatment effect		Control v. enzyme	Linear	Quadratic	Cubic	
Feed intake (g DM/d)	1146	1211	1180	1004	56.2	0.043	0.034	0.066	0.508	0.684	
Total tract digestibility (g digested/g ingested)											
Dry matter	0.58	0.60	0.57	0.75	0.041	0.047	0.027	0.021	0.014	0.901	
Organic matter	0.59	0.62	0.59	0.77	0.039	0.032	0.019	0.018	0.014	1.000	
Crude protein	0.54	0.56	0.51	0.71	0.053	0.036	0.044	0.069	0.015	0.702	
Neutral detergent fibre	0.56	0.59	0.61	0.76	0.039	0.037	0.077	0.008	0.078	0.078	
Acid detergent fibre	0.46	0.50	0.53	0.70	0.051	0.006	0.020	0.031	0.243	0.305	
Apparent ruminal digestibility (g digested/g ingested)											
Neutral detergent fibre	0.36	0.39	0.42	0.46	0.024	0.045	0.036	0.122	0.078	0.078	
Acid detergent fibre	0.29	0.31	0.33	0.34	0.022	0.055	0.584	0.294	0.050	0.305	
Blood metabolites (mg/dl)											
Urea	44	49	50	49	4.4	0.358	0.643	0.346	0.147	0.355	
Phosphorus	0.23	0.29	0.27	0.22	0.098	0.574	0.431	0.233	0.699	0.808	
Triglycerides	8	10	6	9	2.0	0.511	0.945	0.917	0.339	0.369	

DM, dry matter. *Diet (Table 1) without addition of xylanase (XY0) or with addition of xylanase at 1 (XY1), 3 (XY3) and 6 (XY6) µl/g DM.

ram, P_j is the fixed effect of period, T_k is the fixed effect of rate of addition of enzyme (XY0, XY1, XY3, XY6) and ε_{ijkl} is the residual error. Tukey's test was used for multiple comparisons of means. Polynomial contrasts (linear, quadratic and cubic effects) were fitted to the four rates of addition of the enzyme. A treatment (average of all treatments receiving XY) v. control contrast was also performed. Significance was declared at a level of $P < 0.05$ and $P \leq 0.10$ was considered as a tendency approaching significance.

RESULTS

Feed intake, digestibility and blood parameters

Sheep fed XY1 and XY3 had greater ($P = 0.035$) feed DM intake than the control sheep (increases of 6 and 3% with XY1 and XY3, respectively). However, at the greatest application rate (XY6) feed intake was decreased slightly when compared with the control.

Sheep fed XY1 and XY6 had greater ($P < 0.05$) total tract DM, OM and crude protein (CP) digestibility than the control sheep. Dry matter digestibility increased by 30% with XY6 when compared with the control diet. Digestibility of ADF increased linearly ($P = 0.008$) with increasing enzyme application rates. The NDF digestibility of the enzyme treated diets tended ($P = 0.077$) to be greater than that of the control sheep (Table 2).

Sheep fed enzyme had greater ($P < 0.036$) ruminal NDF digestibility than control sheep. With XY6, ruminal NDF digestibility was increased by 28% when compared with the control diet. There was no difference in ruminal ADF digestibility between sheep fed the enzyme and the control diet (Table 2).

There were no differences in the blood concentrations of urea, phosphorus or triglycerides due to the different levels of xylanase addition (Table 2).

Ruminal fermentation

Ruminal pH of sheep fed with the enzyme was greater at all sampling times (P values were 0.050, 0.020 and 0.033 at 3, 6 and 9 h post-feeding, respectively) compared with control sheep. Within enzyme treatment, at the 3 h sampling, increasing enzyme application rate had no effect on pH. Sheep fed XY1 had maximum pH at the 6 h sampling and minimum pH at the 9 h sampling.

At the 6 h sampling, the ruminal ammonia-N concentration of sheep fed enzyme-treated diets was greater ($P = 0.048$) than the control sheep. Within enzyme treatment, XY6 showed the maximum ammonia concentrations (linear effect, $P = 0.028$), with an increase of 90% over the control value. At the 3 and 9 h sampling, there were no differences.

Table 3. Rumen fermentation at different times post-feeding of a diet treated with increasing concentrations of xylanase (XY) in Rambouillet sheep

	Diets*				S.E.M. (n = 4)	P value			
	XY0	XY1	XY3	XY6		Treatment effect	Control v. enzyme	Linear	Quadratic
pH									
3 h	6.0	6.3	6.2	5.8	0.17	0.031	0.050	0.198	0.385
6 h	5.8	6.1	6.0	6.1	0.13	0.057	0.020	0.041	0.051
9 h	5.8	5.9	6.0	6.0	0.06	0.116	0.033	0.039	0.236
Ammonia-N (mmol/l)									
3 h	17.3	15.9	15.5	16.6	3.11	0.976	0.730	0.933	0.690
6 h	7.7	9.0	12.7	14.6	1.86	0.012	0.048	0.028	0.929
9 h	8.6	8.3	8.2	8.2	0.43	0.906	0.517	0.604	0.650
Acetic acid (mmol/l)									
3 h	44.7	47.5	51.9	48.0	3.32	0.054	0.029	0.889	0.058
6 h	46.0	45.2	51.3	59.3	4.39	0.019	0.029	0.105	0.330
9 h	46.0	45.3	45.9	45.2	1.45	0.967	0.740	0.763	0.948
Propionic acid (mmol/l)									
3 h	43.2	47.3	45.0	42.3	1.91	0.035	0.047	0.379	0.026
6 h	42.7	42.9	39.1	45.7	2.75	0.047	0.046	0.530	0.213
9 h	47.5	45.7	44.2	45.9	2.19	0.047	0.041	0.672	0.037
Butyric acid (mmol/l)									
3 h	13.3	13.7	13.2	13.3	0.19	0.340	0.631	0.584	0.871
6 h	12.6	12.9	13.1	13.1	0.41	0.078	0.374	0.426	0.571
9 h	13.3	12.0	12.9	12.9	0.40	0.228	0.175	0.842	0.372

*Diet (Table 1) without addition of xylanase (XY0) or with addition of xylanase at 1 (XY1), 3 (XY3) and 6 (XY6) µl/g DM.

Acetic acid concentrations (mmol/l) were greater (quadratic effect, $P=0.029$) in enzyme treatments compared with the control at 3 and 6 h post-feeding. With XY6 these concentrations were increased by 7 and 28% compared with the control at 3 and 6 h post-feeding, respectively. At 9 h, there were no significant differences. Propionic acid concentrations (mmol/l) at 3 h post-feeding were greatest in XY1 (quadratic effect, $P=0.026$); however, at the 9 h sampling, all enzyme treatments had lower ($P=0.041$) propionic acid concentrations compared with the control. Among enzyme application rates, XY3 had the lowest propionic acid concentration (quadratic effect, $P=0.037$). No effects were observed on ruminal butyric acid concentrations between different treatments at all sampling times (Table 3).

DISCUSSION

Feed intake

Addition of xylanase to diets at low (i.e. XY1) and moderate (i.e. XY3) rates increased feed intake by

about 6 and 3%, respectively, compared with XY0; however, intake decreased by 9% when xylanase was applied at a greater concentration (i.e. XY6) compared with XY0. Therefore, addition of fibrolytic enzymes at certain concentrations may increase the intake of fibrous feeds. Beauchemin *et al.* (2003) concluded that high rates of enzyme application could be less effective than low rates of application in increasing feed intake, indicating the importance of determining the optimal rate of enzyme addition. The current results are in agreement with Gado *et al.* (2009), who observed about 13% greater DM intake in dairy cows due to enzyme supplementation at 40 g/day.

Digestibility

When xylanase was applied to the diet at the highest concentration (i.e. XY6), whole tract digestibility was increased (by 28–42%) compared with other xylanase concentrations. Improving digestibility, in particular, that of the fibre fractions is the main purpose of adding fibrolytic enzymes to ruminant feeds. Improved digestibility with xylanase at some

application rates supports the hypothesis that a suitable enzyme concentration could improve fermentation efficiency during the initial stages of digestion (Jalilvand *et al.* 2008).

The greater digestibility observed with the XY6 diet may be related, as previously mentioned, to improved rate of ruminal digestion of the potentially digestible NDF fraction (Yang *et al.* 1999) and to changes in gut viscosity (Hristov *et al.* 2000); although these features were not determined in the present study. Altered ruminal fermentation (Kholif & Aziz 2014; Rojo *et al.* 2015), enhanced microbial attachment and colonization to the plant cell wall (Wang *et al.* 2001) and complementary interactions with ruminal microbial enzymes (Morgavi *et al.* 2004) are different possible reasons for the improved rate of ruminal digestion.

In most reports, addition of fibrolytic enzymes to the ruminant feedstuffs increased the numbers of non-fibrolytic and fibrolytic bacteria in rumen fluid and provided more total polysaccharidase activity to digest feedstuffs (Giraldo *et al.* 2008). Mao *et al.* (2013) found that addition of cellulase and xylanase increased the numbers of total bacteria and *Fibrobacter succinogenes* in *in vitro* incubation medium resulting in enhanced fermentation. Results in the present study are consistent with Khattab *et al.* (2011) and Salem *et al.* (2013; 2015b), who observed greater feed digestibility in response to exogenous enzyme addition.

Ruminal NDF digestibility was increased by 7.5, 17 and 28% (compared with the control diet), respectively, with increasing xylanase application rates. Thus, the increased total tract fibre digestibility seems to be due, in part, to enhance fibre digestion in the rumen. Fibrolytic enzymes not only improve fibrolytic activity in the rumen, but also raise xylanase activity in the small intestine (Hristov *et al.* 1998, 2000). Hristov *et al.* (1998) reported that addition of enzymes elevated duodenal xylanase activity by 30% and cellulase activity by 2–5%. Hristov *et al.* (2000) showed that xylanase activity in the faeces was increased with enzyme supplementation, suggesting that xylanase and probably other exogenous fibrolytic enzymes, may work synergistically with the microbes within the large intestine.

Blood metabolites

None of the measured blood metabolites (urea, phosphorus and triglycerides) was affected by xylanase addition to feed and all were found within the

reference ranges (Boyd 1984). Serum urea concentration is an indicator of the nutritional status of sheep (Kumar *et al.* 1981), in particular regarding the provision of total and degradable protein in the feed. Normal serum urea values indicate that protein catabolism was not increased in the muscles and that kidney function was not adversely affected by diet.

Ruminal fermentation

Sheep fed xylanase had greater ruminal pH values compared with the control. One of the most important factors affecting fibre digestion is ruminal pH. For xylanase treatments, rumen pH ranged from 5.98 to 6.15, which was within the range considered acceptable for fibre digestion (Ørskov & Ryle 1990). Fibrolytic bacteria are very sensitive to ruminal pH changes (Sung *et al.* 2007). Greater ruminal pH values are more favourable for fibrolytic microbial activity than low ruminal pH (Sung *et al.* 2007).

Ruminal ammonia-N concentrations ranged from 7.7 to 17.3 mmol/l which were above the range that Satter & Slyter (1974) considered as sufficient for microbial protein synthesis. Greater ruminal ammonia-N concentrations in sheep fed the enzyme treated diets (XY6 and XY3) compared with the un-supplemented control support the possibility that xylanase enhances rumen protein degradation, probably in response to a shift in ruminal microbiota (Salem *et al.* 2013). Kholif & Aziz (2014) found that feeding goats on diets treated with a fibrolytic enzyme elevated ruminal ammonia-N concentration compared with un-supplemented control diets. The observed dose-effects reinforce the importance of defining the optimum application rate of enzyme for better feed utilization.

Greater acetic acid concentrations were obtained with xylanase-treated diets (especially with XY6) compared with the control. Improving fibre digestion usually alters rumen fermentation and affects the production of individual VFA. The greater acetic acid concentrations with xylanase addition could be associated with improved digestion of structural carbohydrates (Soltan *et al.* 2013). Changes in individual VFA concentrations observed when fibrolytic enzymes were added to feed suggest that these exogenous enzymes could affect microbial growth and activity, causing a shift in the metabolic pathways by which specific microbes utilize substrates (Almaraz *et al.* 2010). Shifts in ruminal fermentation may be the result of altered fibre structure, which could stimulate microbial colonization (Giraldo *et al.* 2008), or a

shift in the species profile of fibre-colonizing bacteria in response to enzyme addition (Wang *et al.* 2001). Gado *et al.* (2009) and Salem *et al.* (2013) also observed greater acetic acid concentrations in the rumen when animals were fed diets supplemented with exogenous enzymes.

Among the tested xylanase application rates, concentrations of 3 and 6 µl xylanase/g DM of the basal diet resulted in enhanced digestibility and ruminal fermentation in Rambouillet sheep. However, with 6 µl xylanase/g DM of the basal diet feed intake decreased, whereas ruminal ammonia-N and individual VFA increased compared with the other rates of enzyme addition. Generally, addition of xylanase had no effects on blood serum concentrations of urea, phosphorus and triglycerides.

The authors acknowledge the financial support from the IAEA (Vienna, Austria) Research Contract number MEX16307 within the D3-10-27 Coordinated Research Project. First author would like to thank the Mexican National Council for Science and Technology (Consejo Nacional de Ciencia y Tecnología-CONACYT) for the PhD scholarship received.

REFERENCES

- ABDEL-AZIZ, N. A., SALEM, A. Z. M., EL-ADAWY, M. M., CAMACHO, L. M., KHOLIF, A. E., ELGHANDOUR, M. M. Y. & BORHAMI, B. E. (2015). Biological treatments as a mean to improve feed utilization in agriculture animals – an overview. *Journal of Integrative Agriculture* **14**, 534–543.
- ALMARAZ, I., GONZÁLEZ, S. S., PINOS-RODRÍGUEZ, J. M. & MIRANDA, L. A. (2010). Effects of exogenous fibrolytic enzymes on *in sacco* and *in vitro* degradation of diets and on growth performance. *Italian Journal of Animal Science* **9**, 6–10.
- ALSERSY, H., SALEM, A. Z. M., BORHAMI, B. E., OLIVARES, J., GADO, H. M., MARIEZCURRENA, M. D., YACUOT, M. H., KHOLIF, A. E., EL-ADAWY, M. & HERNANDEZ, S. R. (2015). Effect of Mediterranean saltbush (*Atriplex halimus*) ensiling with two developed enzyme cocktails on feed intake, nutrient digestibility and ruminal fermentation in sheep. *Animal Science Journal* **86**, 51–58.
- AOAC (1997). *Official Methods of Analysis of the Association of Official Analytical Chemist*, Vol. **1**, 16th edn, Washington, DC: Association of Official Analytical Chemists.
- BEAUCHEMIN, K. A., COLOMBATTO, D., MORGAVI, D. P. & YANG, Y. Z. (2003). Use of exogenous fibrolytic enzymes to improve feed utilization by ruminants. *Journal of Animal Science* **81**, (E Suppl.) **2**, E37–E47.
- BOYD, J. W. (1984). The interpretation of serum biochemistry test results in domestic animals. *Veterinary Clinical Pathology* **13**, 7–14.
- DEAN, D. B., STAPLES, C. R., LITTELL, R. C., KIM, S. C. & ADESOGAN, A. T. (2013). Effect of method of adding a fibrolytic enzyme to dairy cow diets on feed intake digestibility, milk production, ruminal fermentation, and blood metabolites. *Animal Nutrition and Feed Technology* **13**, 337–353.
- ELGHANDOUR, M. M. Y., VÁZQUEZ CHAGOYÁN, J. C., SALEM, A. Z. M., KHOLIF, A. E., MARTÍNEZ CASTAÑEDA, J. S., CAMACHO, L. M. & CERRILLO-SOTO, M. A. (2014). Effects of *Saccharomyces cerevisiae* at direct addition or pre-incubation on *in vitro* gas production kinetics and degradability of four fibrous feeds. *Italian Journal of Animal Science* **13**, 295–301.
- ELWAKEEL, E. A., TITGEMEYER, E. C., JOHNSON, B. J., ARMENDARIZ, C. K. & SHIRLEY, J. E. (2007). Fibrolytic enzymes to increase the nutritive value of dairy feedstuffs. *Journal of Dairy Science* **90**, 5226–5236.
- ERWIN, E. S., MARCO, G. J. & EMERY, E. M. (1961). Volatile fatty acid analysis of blood and rumen fluid by gas chromatography. *Journal of Dairy Science* **44**, 1768–1771.
- GADO, H. M., SALEM, A. Z. M., ROBINSON, P. H. & HASSAN, M. (2009). Influence of exogenous enzymes on nutrient digestibility, extent of ruminal fermentation as well as milk production and composition in dairy cows. *Animal Feed Science and Technology* **154**, 36–46.
- GIRALDO, L. A., TEJIDO, M. L., RANILLA, M. J., RAMOS, S. & CARRO, M. D. (2008). Influence of direct-fed fibrolytic enzymes on diet digestibility and ruminal activity in sheep fed a grass hay-based diet. *Journal of Animal Science* **86**, 1617–1623.
- HRISTOV, A. N., MCALLISTER, T. A. & CHENG, K.-J. (1998). Stability of exogenous polysaccharide-degrading enzymes in the rumen. *Animal Feed Science and Technology* **76**, 161–168.
- HRISTOV, A. N., MCALLISTER, T. A. & CHENG, K.-J. (2000). Intraruminal supplementation with increasing levels of exogenous polysaccharide-degrading enzymes: effects on nutrient digestion in cattle fed a barley grain diet. *Journal of Animal Science* **78**, 477–487.
- JACKSON, S. & NICOLSON, S. W. (2002). Xylose as a nectar sugar: from biochemistry to ecology. *Comparative Biochemistry and Physiology Part B. Biochemistry and Molecular Biology* **131**, 613–620.
- JALILVAND, G., ODONGO, N. E., LOPEZ, S., NASERIAN, A., VALIZADEH, R., SHAHRODI, E., KEBREAB, E., FRANCE, J. (2008). Effects of different levels of an enzyme mixture on *in vitro* gas production parameters of contrasting forages. *Animal Feed Science and Technology* **146**, 289–301.
- KHATTAB, H. M., GADO, H. M., KHOLIF, A. E., MANSOUR, A. M. & KHOLIF, A. M. (2011). The potential of feeding goats sun dried rumen contents with or without bacterial inoculums as replacement for berseem clover and the effects on milk production and animal health. *International Journal of Dairy Science* **6**, 267–277.
- KHOLIF, A. E., GOUDA, G. A., MORSY, T. A., SALEM, A. Z. M., LOPEZ, S. & KHOLIF, A. M. (2015). *Moringa oleifera* leaf meal as a protein source in lactating goat's diets: feed intake, digestibility, ruminal fermentation, milk yield and

- composition, and its fatty acids profile. *Small Ruminant Research* **129**, 129–137.
- KHOLIF, A. M. & AZIZ, H. A. (2014). Influence of feeding cellulolytic enzymes on performance, digestibility and ruminal fermentation in goats. *Animal Nutrition and Feed Technology* **14**, 121–136.
- KOZLOSKI, G. V., STEFANELLO, C. M., MESQUITA, F. R., ALVES, T. P., RIBEIRO FILHO, H. M. N., ALMEIDA, J. G. R. & MORAES GENRO, T. C. (2014). Technical note: evaluation of markers for estimating duodenal digesta flow and ruminal digestibility: acid detergent fiber, sulfuric acid detergent lignin, and n-alkanes. *Journal of Dairy Science* **97**, 1730–1735.
- KUMAR, N., SINGH, U. B. & VERMA, D. N. (1981). Effect of different levels of dietary protein and energy on growth of male buffalo calves. *Indian Journal of Animal Science* **51**, 513–517.
- LIN, Y., VONK, R. J., SLOOFF, M. J. H., KUIPERS, F. & SMIT, M. J. (1995). Differences in propionate-induced inhibition of cholesterol and triacylglycerol synthesis between human and rat hepatocytes in primary culture. *British Journal of Nutrition* **74**, 197–207.
- MAO, H. L., WU, C. H., WANG, J. K. & LIU, J. X. (2013). Synergistic effect of cellulase and xylanase on *in vitro* rumen fermentation and microbial population with rice straw as substrate. *Animal Nutrition and Feed Technology* **13**, 477–487.
- MCCULLOUGH, H. (1967). The determination of ammonia in whole blood by direct colorimetric method. *Clinica Chimica Acta* **17**, 297–304.
- MORGAVI, D. P., BEAUCHEMIN, K. A., NSEREKO, V. L., RODE, L. M., McALLISTER, T. A. & WANG, Y. (2004). *Trichoderma* enzymes promote *Fibrobacter succinogenes* S85 adhesion to, and degradation of, complex substrates but not pure cellulose. *Journal of the Science of Food and Agriculture* **84**, 1083–1090.
- MORSY, T. A., KHOLIF, A. E., KHOLIF, S. M., KHOLIF, A. M., SUN, X. & SALEM, A. Z. M. (2016). Effects of two enzyme feed additives on digestion and milk production in lactating Egyptian buffaloes. *Annals of Animal Science* **16**, 209–222.
- NRC (1985). *Nutrient Requirements of Sheep*, 6th edn, Washington, DC: National Academy Press.
- ØRSKOV, E. R. & RYLE, R. (1990). *Energy Nutrition in Ruminants*. New York: Elsevier Science Publishers.
- ROBYT, J. F. & WHELAN, W. J. (1972). Reducing value methods for maltodextrins. 1. Chain-length dependence of alkaline 3,5-dinitrosalicylate and chain length independence of alkaline copper. *Analytical Biochemistry* **45**, 510–516.
- ROJO, R., KHOLIF, A. E., SALEM, A. Z. M., ELGHANDOUR, M. M. Y., ODONGO, N. E., MONTES DE OCA, R., RIVERO, N. & ALONSO, M. U. (2015). Influence of cellulase addition to dairy goat diets on digestion and fermentation, milk production and fatty acid content. *Journal of Agricultural Science, Cambridge* **153**, 1514–1523.
- SALEM, A. Z. M., GADO, H. M., COLOMBATTO, D. & ELGHANDOUR, M. M. Y. (2013). Effects of exogenous enzymes on nutrient digestibility, ruminal fermentation and growth performance in beef steers. *Livestock Science* **154**, 69–73.
- SALEM, A. Z. M., KHOLIF, A. E., ELGHANDOUR, M. M. Y., BUENDÍA, G., MARIEZCURRENA, M. D., HERNANDEZ, S. R. & CAMACHO, L. M. (2014). Influence of oral administration of *Salix babylonica* extract on milk production and composition in dairy cows. *Italian Journal of Animal Science* **13**, 10–14.
- SALEM, A. Z. M., AMMAR, H., KHOLIF, A. E., ELGHANDOUR, M. M. Y. & ORTIZ, L. B. (2015a). Effect of glucoamylase enzyme extract on *in vitro* gas production and degradability of two diets with 25% of corn or sorghum grains. *Indian Journal of Animal Science* **85**, 183–188.
- SALEM, A. Z. M., ALSERSY, H., CAMACHO, L. M., EL-ADAWY, M. M., ELGHANDOUR, M. M. Y., KHOLIF, A. E., RIVERO, N., ALONSO, M. U. & ZARAGOZA, A. (2015b). Feed intake, nutrient digestibility, nitrogen utilization, and ruminal fermentation activities in sheep fed *Atriplex halimus* ensiled with three developed enzyme cocktails. *Czech Journal of Animal Science* **60**, 185–194.
- SAS Institute (2006). *SAS 9.0 User's Guide: Statistics, version 9.0*. Cary, NC: SAS Institute.
- SATTER, L. D. & SLYTER, L. L. (1974). Effect of ammonia concentration on rumen microbial protein production *in vitro*. *British Journal of Nutrition* **32**, 199–208.
- SOLTAN, Y. A., ABDALLA, A. L., SILVA, L. R. F., NATEL, A. S., MORSY, A. S. & LOUVANDINI, H. (2013). Response of different tropical pasture grass species to treatments with fibrolytic enzymes in terms of *in vitro* ruminal nutrient degradation and methanogenesis. *Animal Nutrition and Feed Technology* **13**, 551–568.
- SUNG, H. G., KOBAYASHI, Y., CHANG, J., HA, A., HWANG, I. H. & HA, J. K. (2007). Low ruminal pH reduces dietary fiber digestion via reduced microbial attachment. *Asian-Australasian Journal of Animal Sciences* **20**, 200–207.
- TOGTOKHBAYAR, N., CERRILLO, M. A., RODRÍGUEZ, G. B., ELGHANDOUR, M. M. M. Y., SALEM, A. Z. M., URANKHAICH, C., JIGIDPUREV, S., ODONGO, N. E. & KHOLIF, A. E. (2015). Effect of exogenous xylanase on rumen *in vitro* gas production and degradability of wheat straw. *Animal Science Journal* **86**, 765–771.
- VALDES, K. I., SALEM, A. Z. M., LOPEZ, S., ALONSO, M. U., RIVERO, N., ELGHANDOUR, M. M. Y., DOMÍNGUEZ, I. A., RONQUILLO, M. G. & KHOLIF, A. E. (2015). Influence of exogenous enzymes in presence of *Salix babylonica* extract on digestibility, microbial protein synthesis and performance of lambs fed maize silage. *Journal of Agricultural Science, Cambridge* **153**, 732–742.
- VAN SOEST, P. J., ROBERTSON, J. B. & LEWIS, B. A. (1991). Methods for dietary fibre, neutral detergent fibre, and non-starch carbohydrates in relation to animal nutrition. *Journal of Dairy Science* **74**, 3583–3597.
- WANG, Y., McALLISTER, T. A., RODE, L. M., BEAUCHEMIN, K. A., MORGAVI, D. P., NSEREKO, V. L., IWAASA, A. D. & YANG, W. (2001). Effects of an exogenous enzyme preparation on microbial protein synthesis, enzyme activity and attachment to feed in the rumen simulation technique (Rusitec). *British Journal of Nutrition* **85**, 325–332.
- YANG, W. Z., BEAUCHEMIN, K. A. & RODE, L. M. (1999). Effects of an enzyme feed additive on extent of digestion and milk production of lactating dairy cows. *Journal of Dairy Science* **82**, 391–403.