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1 Progress in the Control of Bovine Tuberculosis in Spanish Wildlife

- 3 Christian Gortazar^{1*}, Joaquín Vicente¹, Mariana Boadella¹, Cristina Ballesteros¹, Ruth C.
- 4 Galindo¹, Joseba Garrido², Alicia Aranaz³, José de la Fuente^{1,4}
- 6 1. IREC (CSIC-UCLM-JCCM), Ronda de Toledo s/n, Ciudad Real 13071, Spain
- 7 2. NEIKER-TECNALIA, Inst Vasco Invest & Desarrollo Agrario, Dpt. Anim. Hlth., Bizkaia
- 8 48160, Spain
- 9 3. Departamento de Patología Animal (Sanidad Animal), Facultad de Veterinaria,
- 10 Universidad Complutense, 28040 Madrid, Spain
- 4. Department of Veterinary Pathobiology, Center for Veterinary Health Sciences, Oklahoma
- 12 State University, Stillwater, OK 74078, USA.
- * Corresponding author. Tel: 0034926295450, Fax: 0034926295451. E-mail address:
- 14 Christian.Gortazar@uclm.es (C. Gortazar).
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Abstract

17	Despite the compulsory test and slaughter campaigns in cattle, bovine tuberculosis (bTB) is
18	still present in Spain, and the role of wildlife reservoirs is increasingly recognized. We
19	provide an update on recent progress made in bTB control in Spanish wildlife, including
20	aspects of epidemiology, surveillance, host-pathogen interaction and wildlife vaccination.
21	At the high densities and in the particular circumstances of Mediterranean environments, wild
22	ungulates, mainly Eurasian wild boar and red deer, are able to maintain M. bovis circulation
23	even in absence of domestic livestock. Infection is widespread among wild ungulates in the
24	south of the country, local infection prevalence being as high as 52% in wild boar and 27% in
25	red deer. Risk factors identified include host genetic susceptibility, abundance, spatial
26	aggregation at feeders and waterholes, scavenging, and social behaviour. An increasing trend
27	of bTB compatible lesions was reported among wild boar and red deer inspected between
28	1992 and 2004 in Southwestern Spain. Sporadic cases of badger TB have been detected,
29	further complicating the picture.
30	Gene expression profiles were characterized in European wild boar and Iberian red deer
31	naturally infected with M. bovis. The comparative analysis of gene expression profiles in
32	wildlife hosts in response to infection advanced our understanding of the molecular
33	mechanisms of infection and pathogenesis, revealed common and distinctive host responses
34	to infection and identified candidate genes associated with resistance to bTB and for the
35	characterization of host response to infection and vaccination.
36	Ongoing research is producing valuable knowledge on vaccine delivery, safety and efficacy
37	issues. Baits for the oral delivery of BCG vaccine preparations to wild boar piglets were
38	developed and evaluated. The use of selective feeders during the summer was found to be a
39	potentially reliable bait-deployment strategy. Safety experiments yielded no isolation of M.

- 40 bovis BCG from faeces, internal organs at necropsy and the environment, even after oral
- 41 delivery of very high doses. Finally, preliminary vaccination and challenge experiments
- 42 suggested that a single oral BCG vaccination may protect wild boar from infection by a
- 43 virulent *M. bovis* field strain.
- 44 Keywords: Cervus elaphus; Disease control; Meles meles; Sus scrofa; Wildlife epidemiology

1. Introduction

46	Bovine tuberculosis (bTB), a chronic infectious disease shared between livestock and
47	wildlife, has a complex epidemiology, often with climate and habitat-mediated peculiarities.
48	Four potential wildlife bTB reservoirs (as reviewed in EFSA 2009) exist in mainland Spain:
49	Eurasian wild boar (Sus scrofa), red deer (Cervus elaphus), fallow deer (Dama dama) and
50	Eurasian badger (Meles meles). Eurasian badgers are scarce in Mediterranean habitats and
51	more abundant in Atlantic ones (Revilla and Palomares, 2002), and wild ungulates are
52	continuously expanding in range and in numbers throughout the whole Peninsula (e.g.
53	Gortazar et al. 2000, Delibes-Mateos et al. 2009), but are largely absent from the Islands.
54	Spain is a major livestock producer within the European Union. There are about 6,250,000
55	cattle in 143,000 herds. Since test and slaughter campaigns became compulsory, bTB in cattle
56	declined significantly from 12% herd prevalence in 1987 to 1.68% in 2008. However, cattle
57	bTB distribution is not uniform in the country. Island regions are almost bTB free, and most
58	of the northern Spanish mainland (Atlantic climate region) is also almost bTB free. In
59	contrast, several regions of central and southern Spain still have high bTB prevalence (Figure
60	1). In infected regions, bTB is consistently more prevalent among beef cattle and bullfighting
61	cattle, which often share wildlife habitats, than among the generally fenced dairy cattle
62	(http://rasve.mapa.es//Publica/Sanidad/documentos/INFORME%20FINAL%20TÉCNICO%2
63	<u>0TB%202008.pdf</u>).
64	Moreover, bTB is also present among goat and pig livestock, although information on
65	prevalence is lacking. About 2,500,000 pigs are raised in open air systems. Most belong to
66	the Iberian pig breed, which is raised in open evergreen oak woodland habitats of the south
67	western Iberian Peninsula. Contact with domestic ruminants and with wildlife occurs in this
68	ecosystem, and cases of <i>M. bovis</i> infection have been recorded (Gómez-Laguna et al. 2010).
69	Molecular typing suggests that M. bovis strains of pigs may be shared with livestock and

70	wildlife species (Parra et al. 2005). Regarding goats, current numbers in Spain are around
71	3,000,000 (http://www.mapa.es/estadistica/pags/anuario/2008/). Not only <i>M. caprae</i> , but also
72	M. bovis strains are identified in goats (Gutiérrez and García-Marín, 1999).
73	The role of wildlife reservoirs in bTB epidemiology is increasingly recognized, worldwide
74	(EFSA 2009). However, the definition of a wildlife reservoir is somewhat controversial. In a
75	broad sense, a reservoir is defined as one or more epidemiologically connected populations or
76	environments in which the pathogen can be permanently maintained and from which
77	infection is transmitted to the defined target population (Haydon et al. 2002). For being a
78	competent bTB reservoir, any host species must be susceptible, able to transmit the disease,
79	and abundant enough (Corner 2006). In Spain, the wildlife component of this reservoir is
80	composed of one or more sympatric host species, with marked regional differences.
81	Essentially, wild ungulates are responsible for bTB maintenance in Mediterranean regions of
82	continental Spain, and the badger could have some relevance in the more humid Atlantic
83	regions (see below).
84	Herein, we provide an update on recent progress made in bTB control in Spanish wildlife,
85	including aspects of epidemiology, surveillance, host-pathogen interaction and wildlife
86	vaccination.
87	
88	2. The wildlife factor in bTB epidemiology in Spain
89	The Iberian Peninsula in the southwestern end of Europe is largely dominated by
90	Mediterranean climate, with mild to cold, dry winters, hot and dry summers, and limited
91	rainfall (usually less than 600 mm per year), which is concentrated in spring and autumn. The
92	northern strip of the Peninsula, from Portugal to the Pyrenees, is characterized by an Atlantic

climate, with up to 2000 mm rainfall per year.

94	2.1. Island regions
95	Both Spanish island regions (the Balearic Islands in the Mediterranean and the Canary
96	Islands in the Atlantic) are almost bTB free. This is interesting from a wildlife perspective,
97	since these regions are lacking all four potential wildlife reservoirs. For comparison, bTB was
98	diagnosed among feral black pigs from the Italian Mediterranean island Sicily and the disease
99	still constitutes a problem for livestock in this region (Di Marco et al. 2008).
100	
101	2.2. Atlantic Spain
102	It is not known if wildlife represents a significant bTB reservoir in northern Spain. Eurasian
103	badgers, a well known bTB reservoir in Ireland and the UK, are more common and abundant
104	in Atlantic than in Mediterranean habitats in Spain (Revilla and Palomares, 2002), and
105	sporadic cases of M. bovis infection have been detected in different Spanish regions, even in
106	the north (Sobrino et al. 2008, A. Balseiro pers. comm.). However, prevalence figures based
107	on large enough sample sizes, which are difficult to obtain in this protected species, are
108	currently lacking. Moreover, recent monitoring data suggests that badger densities are
109	increasing (Sobrino et al. 2009).
110	Very few cases of <i>M. bovis</i> infection have been reported among wild ungulates from Atlantic
111	habitats in Spain, despite locally intense sampling. However, the bTB outbreak among red
112	deer and wild boar from the Brotonne forest in France shows that stable situations with a high
113	infection prevalence may well occur in Atlantic habitats (Zanella et al. 2008). One spillover
114	case has been reported in a roe deer (Capreolus capreolus, Balseiro et al. 2009).
115	Considering together both the continued reduction of cattle bTB prevalence up to levels close
116	to official eradication, and the nearly absence of wildlife bTB cases detected in Atlantic
117	Spain, the current role of wildlife in bTB epidemiology seems of little relevance in this

region, and eradication among cattle will hopefully be achieved soon. However, sporadic cases diagnosed in Eurasian badgers, along with increasing demographic trends in several potential bTB hosts (Gortazar et al. 2000, Acevedo et al. 2006, Sobrino et al. 2009), indicate that maintaining a targeted surveillance is advisable (see below).

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2.3. Mediterranean Spain

In semiarid Mediterranean habitats of central and southern Spain, several wild ungulates contribute to bTB maintenance in a multi-host system (Gortazar et al. 2008). The epidemiologically most relevant wildlife hosts include the Eurasian wild boar (Naranjo et al. 2008a), and two cervids of the subfamily cervinae: the red deer and the fallow deer. Spillover hosts include the red fox (Vulpes vulpes; Millán et al. 2008) and the endangered Iberian lynx (Lynx pardinus; Peña et al. 2006), among others. Habitat constraints determine that badgers exist only at very low densities in Mediterranean Spain (Revilla and Palomares, 2002). Bovine TB is widespread among wild ungulates in central and southern Spain (Vicente et al. 2006), local prevalence of culture confirmed infection being as high as 52% in wild boar and 27% in red deer (Gortazar et al. 2008). At the high densities and in the particular circumstances of Mediterranean environments, Eurasian wild boar and red deer are able to maintain Mycobacterium tuberculosis complex (MTBC) circulation in the absence of domestic livestock, for instance in fenced estates (Figure 2, panel a) and protected natural areas with no domestic ruminants (Gortazar et al. 2005). Red deer alone and wild boar alone maintain bTB circulation in the absence of other wildlife hosts (Vicente et al. 2006). The situation is worst among farm-like hunting enclosures called "cercones", where virtually all wild boar become infected (Acevedo et al. 2007). The reservoir role of fallow deer has been less studied, but is possibly relevant at a local scale (Aranaz et al. 2004, Gortazar et al. 2008,

142	Jaroso et al. unpublished data). Table 1 displays relevant facts about these three host species
143	in Mediterranean habitats.
144	Individual risk factors for M. bovis infection in wild ungulates include the host species, wild
145	boar consistently showing higher M. bovis infection prevalence than deer (Vicente et al.
146	2006, 2007b, Gortazar et al. 2008), and red deer showing higher prevalence than fallow deer
147	(Gortazar et al. 2008); sex in red and fallow deer (males more prevalent, Vicente et al. 2006,
148	Gortazar et al. 2008); age in deer and in wild boar (age-increasing trends; Vicente et al. 2006,
149	Gortazar et al. 2008); and host genetic susceptibility (e.g. Acevedo-Whitehouse et al. 2005
150	and Naranjo et al. 2008a in wild boar, Fernández-de-Mera et al. 2009b, in red deer).
151	In multi-host systems such as those occurring in Mediterranean Spain, the infection levels of
152	sympatric host species do also contribute to the risk factors for a given one (e.g. wild boar on
153	red deer and vice versa; Vicente et al. 2007b). Belonging to an M. bovis-infected social group
154	is a significant risk factor for infection in red deer and wild boar, but not for fallow deer
155	(Gortazar et al. submitted).
156	Environmental risk factors are often, but not always, linked to artificial management of
157	wildlife habitats and wildlife populations in high-wire fenced estates (Vicente et al. 2007a,
158	2007b). Fencing (Figure 2, panel a) can affect bTB epidemiology through a reduced host
159	genetic variability. Using microsatellite markers, Acevedo-Whitehouse et al. (2005) found
160	that genetic variability, fencing and wild boar abundance had significant effects on bTB
161	infection. The strongest effects were observed for genetic heterozygosity, with relatively less
162	heterozygous wild boar being more likely to be infected. Fencing may increase the chances of
163	mating amongst close relatives, and contribute to a reduced genetic variability and reduced
164	disease resistance. This was also suggested in red deer, where Fernández-de-Mera et al.
165	(2009a, 2009b) showed significant loss in variability of the drb2 MHC-II locus over only 16
166	years in a fenced red deer population.

Moreover, fencing is commonly associated with other well known bTB risk factors such as
feeding and translocating (Figure 2, panel c), and in general with wildlife overabundance
(Gortazar et al. 2006). Artificial feeding and watering causes spatial aggregation and allows
maintaining ungulate densities above the carrying capacity of a given habitat. Disease
prevalence has been linked to spatial aggregation and high densities (Acevedo et al. 2007,
Vicente et al. 2007a, 2007b). However, high M. bovis prevalences are found even in
protected areas where no feeding and no translocations of wild ungulates take place, such as
the Doñana National Park (DNP, Gortazar et al. 2008), possibly because of the high densities
and habitat mediated spatial aggregation occurring in DNP. Aggregation and M. bovis
transmission may occur more often in certain habitats. For instance hardwood Quercus spp.
forest availability was associated with increased bTB risk in red deer and wild boar in central
Spain (Vicente et al. 2007b). Miller et al. (2003) suggested that woodland areas provide
shady, moist conditions under which M. bovis might survive longer in the environment.
Finally scavenging, including hunting gut-pile consumption (Figure 2, panel d), is most
probably a significant risk factor for wild boar and for carnivores. Infection through
consumption of contaminated materials may increase the probability of contacting M. bovis
(Gortazar et al. 2008). Ongoing research is evidencing that the amount of carrion scavenged
by wild boar in Mediterranean Spain compares to that by specialized scavengers (vultures),
and even locally this suid becomes the top consumer (unpublished data). Figure 2 suggests
that gut-piles may occasionally attract herbivores such as red deer.
The main purpose of controlling bTB in Spanish wildlife is to prevent the infection of the
domestic stock, and also humans. In this sense, ongoing research is addressing wildlife-
domestic livestock interaction to elucidate shared epidemiology and to identify control
measures such as those related to safe husbandry practices. Also, large scale studies on
spatiotemporal incidence patterns of bTB in livestock herds are including wildlife related

192	features, which will provide a first approximation to the relative contribution of wild animals,
193	and a fine biogeographical picture of the problem.
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195	3. Wildlife disease surveillance
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197	3.1. Defining the concepts
198	Two important concepts in wildlife disease epidemiology need to be defined: surveillance
199	and management. Surveillance is the ongoing recording of diseases in wildlife populations
200	with a view to disease management. Surveillance data are used to identify the areas to be
201	targeted for control, and to anticipate spatial and temporal trends so that pre-emptive
202	management interventions can be used to reduce disease risks (Artois et al. 2009). Wildlife
203	disease management begins with surveillance; knowing if diseases are present, their past and
204	current distribution, and the trends in their prevalence. Possibilities for disease management,
205	always in addition to surveillance, include disease control through different means, disease
206	eradication, which is usually not realistic, or taking no action, if the cost-benefit analysis
207	suggests that this is the most convenient option (Wobeser, 1994).
208	
209	3.2. Methods in bTB surveillance
210	The development of effective schemes for the surveillance of disease in wildlife populations
211	is becoming increasingly important (Artois et al. 2009). Tools available for bTB surveillance
212	include visual inspection for macroscopic bTB-compatible lesions or combinations of visual
213	inspection with culture (e.g. Vicente et al. 2006); systematic culture of selected tissues (e.g.
214	Gortazar et al. 2008); and even serology or combinations of serology and culture (e.g.

Aurtenetxe et al. 2008, Vicente et al. 2007b). In Spain, surveillance is based on large post-

216	mortem samples obtained from hunter-harvested wildlife. However, alternative techniques
217	need to be used in protected wildlife such as badgers, or in protected areas where hunting is
218	banned.
219	Bacteriological culture is the gold standard test for determining bTB infection prevalence. In
220	DNP red and fallow deer, taking culture samples from both the tonsil and retropharyngeal
221	lymph node increased the rate of isolation of M. bovis by 22% over culture of the
222	retropharyngeal lymph node alone (Martín-Hernando et al. 2010). Similarly, 19% of DNP
223	wild boar yielded M. bovis isolates from the tonsil but not from the mandibular LN samples
224	(unpublished data). Hence, pools of both tissues are now used for surveillance purposes.
225	Investigation of wild deer for bTB compatible lesions should ideally include examination of
226	the medial retropharyngeal, left tracheobronchial, mediastinal, mesenteric and ileocaecal
227	lymph nodes (LNs) (Martín-Hernando et al. 2010). Otherwise, the inspected organs should be
228	clearly stated to allow comparisons. In the wild boar visible lesions are most often (i.e. in 92-
229	100% of cases) located in the mandibular lymph nodes (Gortazar et al. 2003, Parra et al.
230	2006, Martín-Hernando et al. 2007). This makes targeted surveillance in this host easy.
231	However, for epidemiological purposes it is interesting to sample at least also the left
232	tracheobronchial LN of wild boar, to estimate the number of individuals with thoracic
233	extended lesions (Martín-Hernando et al. 2007). Ideally, lesion identification should be
234	carried out by trained staff in a systematic manner, and the presence of infection at the local
235	level should later be confirmed by culture. Such information is considered to be valuable for
236	exploring the magnitude and general distribution of bTB in wildlife, provided a large enough
237	sample size is obtained from an extensive area (Vicente et al. 2006). Further, molecular
238	characterization of the isolates from different species (including livestock) would provide
239	additional information to clarify both the local epidemiology of bTB and large spatiotemporal
240	patterns (Gortazar et al. 2005).

4. Host-pathogen interactions

The recent development of ELISA tests for the detection of antibodies against M. bovis will
facilitate studies on large areas and long time series. Two recent studies in wild boar coincide
in reporting sensitivity of 73-77% and specificity of 96-97%. A close association between
strong antibody response and the presence of gross lesions in individuals infected with M .
bovis has been observed in wild boar (Aurtenetxe et al. 2008; Lyashchenko et al. 2008). More
data on the utility of these techniques for other hosts such as red and fallow deer are needed.
3.3. Prevalence trends among Spanish wildlife
An increasing trend of bTB compatible lesions was reported among wild boar and red deer
inspected between 1992 and 2004 in Extremadura, south-western Spain (Parra et al. 2006). In
Doñana NP (Southern Spain), M. bovis infection prevalence increased from 1998-2003 to
2006-2007 by 100% in wild boar and by 50% in red deer (Romero et al. 2008, Gortazar et al.
2008). Among 14 wildlife populations included in a 10 year survey in south central Spain, 11
presented increasing levels of bTB compatible lesions and of incidence among juvenile wild
boar, in contrast to only 3 sites with decreasing bTB (unpublished data; Figure 3). However,
no nationwide figures on bTB trends are available for wildlife.
Field methods for estimating wild ungulate abundance and spatial aggregation (e.g. Acevedo
et al. 2007, 2008) have been implemented, allowing matching population monitoring with
disease surveillance. At a national scale, a Wildlife Disease Surveillance Program is being set
up. This scheme is expected to provide quality information on disease trends in Spanish
wildlife.

264	Substantial evidence suggests that genetic and environmental factors contribute to the
265	pathogenesis and differences in susceptibility of humans and mice to M. tuberculosis
266	(Fernando and Britton, 2006). In Eurasian wild boar, genetic factors have been associated
267	with resistance to bTB at the population level (Acevedo-Whitehouse et al. 2005). In this
268	study, wild boar microsatellite marker variability was correlated with bTB providing
269	evidence of both general and single-locus associative effects on bTB, and several loci
270	revealed high homology to regions of the genome with known immune function (Acevedo-
271	Whitehouse et al. 2005).
272	Recent studies demonstrated by suppression-subtractive hybridization and proteome analysis
273	differential gene expression in tonsils and mandibular lymph nodes of tuberculous and non-
274	tuberculous wild boar exposed to natural M. bovis infection (Naranjo et al. 2006a; 2007a).
275	Microarray gene expression profiling also showed differential gene expression in Iberian red
276	deer lymph nodes and wild boar peripheral blood mononuclear cells in response to natural M .
277	bovis infection (Fernández de Mera et al. 2008, Galindo et al. 2009). These studies showed
278	tissue-specific expression profiles that suggested differences in the role that tonsils and
279	mandibular lymph nodes play in response to M. bovis infection in wild boar (Naranjo et al.
280	2007b). Furthermore, these studies suggested candidate gene markers associated with bTB
281	resistance in wild boar and characterized genes that could be used to monitor host response to
282	pathogen infection and vaccination (Naranjo et al. 2006b; 2008b; Pérez de la Lastra et al.
283	2009). One of these genes, methylmalonyl-CoA mutase (MUT) was upregulated in M. bovis-
284	exposed uninfected animals and specific alleles were associated with resistance to bTB in
285	wild boar (Naranjo et al. 2006b; 2008b; Pérez de la Lastra et al. 2009). Other genes such as
286	complement component 3 (C3), interferon gamma (IFN-gamma), interleukin 4 (IL-4) and
287	Regulated on Activation, Normal T Expressed and Secreted cytokine, also known as CCL5
288	(RANTES) were downregulated in infected wild boar and upregulated in parentally and

orally BCG-immunized animals when compared to non-immunized controls (Pérez de la Lastra et al. 2009; Ballesteros et al. 2009c). These results also provided additional evidence that expression of selected genes correlates with protection to *M. bovis* infection after oral BCG vaccination in wild boar (Ballesteros et al. 2009c).

These studies identified new mechanisms by which wildlife hosts respond to *M. bovis* infection and how the pathogen circumvents host immune responses to establish infection. Furthermore, gene expression profile in vaccinated animals showed BCG-specific responses that are different from those observed in naturally *M. bovis*-infected wild boar which may be used to monitor BCG vaccination during experimental vaccine studies in this species. Gene expression studies in naturally-infected wildlife bTB reservoirs are important for functional genomics and vaccine studies to aid in disease control.

5. Wild boar oral vaccination with BCG

Whether wildlife vaccination will become a realistic option in the frame of bTB control in Spain will depend on factors such as the long term success of cattle bTB control in wildlife habitats, and the results of ongoing vaccination trials in other countries, such as Ireland and UK in badgers and New Zealand in possums (*Trichosurus vulpecula*). Current vaccination research in Spain is focused on wild boar. Delivery, safety and efficacy issues need to be addressed in order to be prepared for eventually licensing a vaccine. Recently, baits for the oral delivery of vaccine preparations to 2–4 month-old wild boar piglets, the preferred age for vaccination, were developed and evaluated. The use of selective feeders during the summer was found to be a potentially reliable bait-deployment strategy (Ballesteros et al. 2009a; 2009b). Safety experiments yielded no isolation of *M. bovis* BCG from faeces, internal organs at necropsy and the environment, even after oral delivery of very high doses. Finally,

313	preliminary vaccination and challenge experiments suggest that a single oral BCG
314	vaccination may protect wild boar from infection by a virulent M. bovis field strain
315	(Ballesteros et al. 2009c).
316	
317	6. Discussion
318	The ecology of M. bovis in Spain represents a multi-host system, with a relevant role for
319	wildlife, but also for domestic reservoirs and for the environment. There are huge differences
320	between regions, with almost no bTB and no wildlife reservoirs in the islands; almost no bTB
321	and few wildlife and habitat risk factors in the Atlantic mainland; and still high bTB
322	prevalence in the Mediterranean habitats of the mainland, partly explained by wildlife and
323	habitat risk factors. As bTB prevalence has dropped in livestock, the relative importance of a
324	potential wildlife reservoir may increase. Thus, wildlife aspects need to be considered in the
325	strategy to control bTB in Spain
326	(http://ec.europa.eu/food/animal/diseases/eradication/reportsanco-10584-
327	2007btbsubgroupsevillarev110-1-08.pdf). Moreover, wildlife numbers and wildlife
328	distribution are continuously changing, with potential impacts on epidemiology.
329	Regarding wildlife, steps have been taken towards disease control. Firstly, new regulations of
330	wild animal translocations (Royal Decree 1082/2009;
331	http://www.boe.es/boe/dias/2009/07/23/pdfs/BOE-A-2009-12206.pdf) and on disposal of
332	hunting carcass remains (both at the EU and the regional level), if properly enforced, will
333	contribute to limit bTB spread. Secondly, new diagnostic tools and setting up a Wildlife
334	Disease Surveillance scheme will allow documenting the spatial and temporal trends of bTB
335	in wildlife. In Spain, the variety of surveillance systems is broad, and a need exists to find
336	effective ways to share and exchange data among regions and coordinate on a global scale.

337	This will hopefully improve our ability to identify new health risks in wildlife populations
338	and enhance our capability to manage them when necessary (Artois et al. 2009).
339	Wildlife fencing, feeding, and keeping overabundant populations for hunting or other
340	recreational purposes, is still a problem of paramount importance in disease epidemiology,
341	not only regarding bTB (Gortazar et al. 2006). To contribute to solve this problem, census
342	methods have been developed (Acevedo et al. 2007) or adapted to Mediterranean habitats
343	(Acevedo et al. 2008). This allows getting accurate estimations of the wildlife densities
344	before and after a given management. Now it is the responsibility of the environment
345	authorities to achieve a reduction of the current wildlife densities to more sustainable levels,
346	and to contribute also to reduce the spatial aggregation of wildlife at feeders or waterholes
347	(Figure 2, panel b). However, the success of any such management action must be assessed
348	critically, including an analysis of the costs, of the ecological consequences and of the animal
349	and human health, welfare, and conservation benefits.
350	This, in turn, urgently requires applied research regarding waterhole ecology, wild boar and
351	deer space use and movements, carrion and gutpile consumption by birds and mammals, and
352	particularly on the effect of any attempt to control bTB in wildlife, for instance through
353	population control or testing the hypothesis that changes in food or water distribution may
354	affect transmission rates. Ongoing research is also addressing wildlife-domestic livestock
355	interaction in order to provide safer husbandry practices.
356	Despite the initial characterization of gene expression profiles in wildlife in response to M .
357	bovis infection and BCG vaccination, further experiments are required to identify specific
358	genes associated with protective response in these animals to monitor immune response after
359	vaccination and to establish gene markers for genetic studies and assisted breeding in farmed
360	game.

New vaccination and challenge trials are currently being conducted with larger numbers of
experimental subjects. In parallel, safety issues are being addressed and, in the field, bait
delivery experiments with biomarkers are being analyzed to evaluate the coverage of our
vaccination strategy and model the possible impact on bTB control in wildlife under the
conditions found in Spain. If these experiments progress as expected, work towards licensing
the use of oral BCG in free living wild boar will be the goal. However, vaccination will not
be a golden bullet but just one of several tools available for bTB control in wildlife in Spain.
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7. References
Acevedo, P., Escudero, M.A., Muñoz, R., Gortazar, C., 2006. Factors affecting wild boar
abundance across an environmental gradient in Spain. Acta Theriol. 51, 327-336.
Acevedo, P., Vicente, J., Hofle, U., Cassinello, J., Ruiz-Fons, F., Gortazar, C., 2007.
Estimation of European wild boar relative abundance and aggregation: a novel method in
epidemiological risk assessment. Epidemiol. Infect. 135, 519-527.

- Acevedo, P., Ruiz-Fons, F., Vicente, J., Reyes-Garcia, A.R., Alzaga, V., Gortazar, C., 2008.
- 384 Estimating red deer abundance in a wide range of management situations in Mediterranean
- 385 habitats. J. Zool. 276, 37-47.
- 386 Acevedo-Whitehouse, K., Vicente, J., Gortazar, C., Höfle, U., Fernández-de-Mera, I.G.,
- 387 Amos, W., 2005. Genetic resistance to bovine tuberculosis in the Iberian wild boar. Mol.
- 388 Ecol. 14, 3209-3217.
- Aranaz, A., de Juan, L., Montero, N., Sanchez, C., Galka, M., Delso, C., Alvarez, J., Romero,
- 390 B., Bezos, J., Vela, A.I., Briones, V., Mateos, A., Dominguez, L., 2004. Bovine tuberculosis
- 391 (Mycobacterium bovis) in wildlife in Spain. J. Clin. Microbiol. 42, 2602-2608.
- 392 Artois, M., Bengis, R., Delahay, R.J., Duchêne, M.J., Duff, P., Ferroglio, E., Gortazar, C.,
- Hutchings, M.R., Kock, R.A., Leighton, T., Mörner, T. & Smith G.C., 2009. Wildlife disease
- 394 surveillance and monitoring, In: Delahay, R., Smith, G.C., Hutchings, M.R. (Eds.)
- 395 Management of Disease in Wild Mammals. Springer, New York, pp. 187-214.
- 396 Aurtenetxe, O., Barral, M., Vicente, J., de la Fuente, J., Gortazar, C., Juste, R.A., 2008.
- 397 Development and validation of an enzyme-linked immunosorbent assay for antibodies against
- 398 *Mycobacterium bovis* in European Wild Boar. BMC Vet. Res. 4, 43-52.
- 399 Ballesteros, C., Gortazar, C., Canales, M., Vicente, J., Lasagna, A., Gamarra, J.A., Carrasco-
- 400 García, R., Fuente, J., 2009a. Evaluation of baits for oral vaccination of European wild boar
- 401 piglets. Res. Vet. Sci. 86, 388-393.
- 402 Ballesteros, C., Carrasco-García, R., Vicente J., Carrasco J., Lasagna, A., de la Fuente, J.,
- 403 Gortazar, C., 2009b. Selective piglet feeders improve age-related bait specificity and uptake
- rate in overabundant Eurasian wild boar populations. Wildl. Res. 36, 203-212.
- 405 Ballesteros, C., Garrido, J.M., Vicente, J., Romero, B., Galindo, R.C., Minguijón, E., Villar,
- 406 M., Martín-Hernando, M.P., Sevilla, I., Juste, R., Aranaz, A., de la Fuente, J., Gortazar, C.,

- 407 2009c. First data on Eurasian wild boar response to oral immunization with BCG and
- 408 challenge with a *Mycobacterium bovis* field strain. Vaccine 27, 6662–6668.
- 409 Balseiro, A., Oleaga, A., Orusa, R., Robetto, S., Zoppi, S., Dondo, A., Goria, M., Gortazar,
- 410 C., Marin, J.F.G., Domenis, L., 2009. Tuberculosis in roe deer from Spain and Italy. Vet.
- 411 Rec. 164, 468-470.
- 412 Corner, L.A., 2006. The role of wild animal populations in the epidemiology of tuberculosis
- in domestic animals: how to assess the risk. Vet. Microbiol. 112, 303–312.
- Delahay, R.J., De Leeuw, A.N.S., Barlow, A.M., Clifton-Hadley, R.S., Cheeseman, C.L.,
- 415 2002. The status of *Mycobacterium bovis* infection in UK wild mammals: a review. Vet. J.
- 416 164, 90–105.
- Delibes-Mateos, M., Farfan, M.A., Olivero, J., Marquez, A.L., Vargas, J.M., 2009. Long-
- 418 term changes in game species over a long period of transformation in the Iberian
- 419 Mediterranean landscape. Environ. Manag. 43, 1256-1268.
- 420 Di Marco, V., Marianelli, C., Russo, M., Aronica, V., Bollo, E., Amedeo, S., Valenza, F.,
- 421 Capucchio, M.T., 2008. Black wild pigs of Nebrodi Park in Sicily: evidence of their possible
- 422 role as reservoir of *Mycobacterium tuberculosis* complex infection. European Society of
- Veterinary Pathology, 26th Meeting, 17-21 September, 2008, Dubrovnik, Croatia.
- 424 EFSA 2009. Draft report on wildlife reservoirs of bovine tuberculosis in Europe.
- 425 Fernández-de-Mera, I.G., Pérez de la Lastra, J.M., Ayoubi, P., Naranjo, V., Kocan, K.M.,
- 426 Gortazar, C., de la Fuente, J., 2008. Differential expression of inflammatory and immune
- response genes in mesenteric lymph nodes of Iberian red deer (*Cervus elaphus hispanicus*)
- 428 naturally infected with *Mycobacterium bovis*. Dev. Comp. Immunol. 32, 85-91.
- 429 Fernández-de-Mera, I.G., Vicente, J., Pérez De La Lastra, J.M., Mangold, A.J., Naranjo, V.,
- 430 Fierro, Y., De La Fuente, J., Gortazar, C., 2009a. Reduced major histocompatibility complex

- class II polymorphism in a hunter-managed isolated Iberian red deer population. J. Zool. 277,
- 432 157-170.
- Fernández de Mera, I.G., Vicente J., Naranjo V., Fierro Y., Garde J.J., de la Fuente J.,
- Gortazar C., 2009b. Impact of major histocompatibility complex class II polymorphisms on
- Iberian red deer parasitism and life history traits. Infec. Genet. Evol. 9, 1232-1239.
- 436 Fernando, S.L., Britton, W.J., 2006. Genetic susceptibility to mycobacterial disease in
- 437 humans. Immunol. Cell. Biol. 84, 125-137.
- 438 Galindo, R.C., Ayoubi, P., Naranjo, V., Gortazar, C., Kocan, K.M., de la Fuente, J., 2009.
- Gene expression profiles of European wild boar naturally infected with *Mycobacterium bovis*.
- 440 Vet. Immunol. Immunopathol. 129, 119-125.
- 441 Gómez-Laguna, J., Carrasco, L., Ramis, G., Quereda, J.J., Gómez, S., Pallarés, F.J., 2010.
- 442 Use of real-time and classic polymerase chain reaction assays for the diagnosis of porcine
- tuberculosis in formalin-fixed, paraffin-embedded tissues. J. Vet. Diagn. Invest. 22:123–127.
- Gortazar, C., Herrero, J., Villafuerte, R., Marco, J., 2000. Historical examination of the status
- of large mammals in Aragon, Spain. Mammalia 64, 411-422.
- Gortazar, C., Vicente, J., Gavier-Widen, D., 2003. Pathology of bovine tuberculosis in the
- European wild boar (Sus scrofa). Vet. Rec. 152, 779-780.
- 448 Gortazar, C., Vicente, J., Samper, S., Garrido, J.M., Fernandez-de-Mera, I.G., Gavin, P.,
- Juste, R.A., Martin, C., Acevedo, P., De La Puente, M. and Hofle, U., 2005. Molecular
- 450 characterization of *Mycobacterium tuberculosis* complex isolates from wild ungulates in
- 451 south-central Spain. Vet. Res. 36, 43-52.
- 452 Gortazar, C., Acevedo, P., Ruiz-Fons, F., Vicente, J., 2006. Disease risks and overabundance
- 453 of game species. Eur. J. Wildl. Res. 52, 81-87.

- 454 Gortazar, C., Torres J., Vicente J., Acevedo P., Reglero M., de la Fuente J., Negro J.J.,
- 455 Aznar-Martin J., 2008. Bovine Tuberculosis in Doñana Biosphere Reserve: The role of wild
- ungulates as disease reservoirs in the last Iberian lynx strongholds. PLoS ONE 3, e2776.
- 457 Gutiérrez, M., García-Marín, J.F., 1999. Brief communications and case reports:
- 458 Cryptococcus neoformans and Mycobacterium bovis causing granulomatous pneumonia in a
- 459 goat. Vet. Pathol. 36, 445-448.
- Haydon, D.T., Cleaveland, S., Taylor, L.H., Laurenson, M.K., 2002. Identifying reservoirs of
- infection: A conceptual and practical challenge. Emerg. Infect. Dis. 8, 1468-1473.
- Lyashchenko, K.P., Greenwald, R., Esfandiari, J., Chambers, M.A., Vicente, J., Gortazar, C.,
- Santos, N., Correia-Neves, M., Buddle, B.M., Jackson, R., O'Brien, D.J., Schmitt, S., Palmer,
- 464 M.V., Delahay, R.J., Waters, W.R., 2008. Animal-side serologic assay for rapid detection of
- 465 Mycobacterium bovis infection in multiple species of free-ranging wildlife. Vet. Microbiol.
- 466 132, 283-292.
- 467 Martin-Hernando, M.P., Hofle, U., Vicente, J., Ruiz-Fons, F., Vidal, D., Barral, M., Garrido,
- 468 J.M., de La Fuente, J., Gortazar, C., 2007. Lesions associated with Mycobacterium
- 469 *tuberculosis* complex infection in the European wild boar. Tuberculosis 87, 360-367.
- 470 Martin-Hernando, M.P., Torres, M.J., Aznar, J., Negro, J.J., Gandía, A., Gortazar, C. 2010.
- 471 Distribution of Lesions in Red and Fallow Deer Naturally Infected with Mycobacterium
- 472 *bovis.* J. Comp. Pathol. 142, 43-50.
- 473 Millan, J., Jimenez, M.A., Viota, M., Candela, M.G., Pena, L., Leon-Vizcaino, L., 2008.
- Disseminated bovine tuberculosis in a wild red fox (Vulpes vulpes) in Southern Spain. J.
- 475 Wildl. Dis. 44, 701-706.

- 476 Miller, R., Kaneene, J.B., Fitzgerald, S.D., Schmitt, S.M., 2003. Evaluation of the influence
- of supplemental feeding of white-tailed deer (Odocoileus virginianus) on the prevalence of
- bovine tuberculosis in the Michigan wild deer population. J. Wildl. Dis. 39, 84-95.
- Naranjo, V., Höfle, U., Vicente, J., Martín, M.P., Ruiz-Fons, F., Gortazar, C., Kocan, K.M.,
- 480 de la Fuente, J., 2006a. Genes differentially expressed in oropharyngeal tonsils and
- 481 mandibular lymph nodes of tuberculous and non-tuberculous European wild boars naturally
- 482 exposed to *Mycobacterium bovis*. FEMS Immunol. Med. Microbiol. 46, 298-312.
- Naranjo, V., Ayoubi, P., Vicente, J., Ruiz-Fons, F., Gortazar, C., Kocan, K.M., de la Fuente,
- 484 J., 2006b. Characterization of selected genes upregulated in non-tuberculous European wild
- boar as possible correlates of resistance to *Mycobacterium bovis* infection. Vet. Microbiol.
- 486 116, 224-231.
- Naranjo, V., Villar, M., Martín-Hernando, M.P., Vidal, D., Höfle, U., Gortazar, C., Kocan,
- 488 K.M., Vázquez, J., de la Fuente, J. 2007a. Proteomic and transcriptomic analyses of
- 489 differential stress/inflammatory responses in mandibular lymph nodes and oropharyngeal
- 490 tonsils of European wild boars naturally infected with *Mycobacterium bovis*. Proteomics 7,
- 491 220-231.
- Naranjo, V., Gortazar, C., Villar, M., de la Fuente, J. 2007b. Comparative genomics and
- 493 proteomics to study tissue-specific response and function in natural Mycobacterium bovis
- infections. Anim. Health Res. Rev. 8, 81-88.
- Naranjo, V., Gortazar, C., Vicente, J., de la Fuente, J., 2008a. Evidence of the role of
- European wild boar as a reservoir of *Mycobacterium tuberculosis* complex. Vet. Microbiol.
- 497 127, 1-9.

- Naranjo, V., Acevedo-Whitehouse, A., Vicente, J., Gortazar, C., de la Fuente, J., 2008b.
- 499 Influence of methylmalonyl-CoA mutase alleles on resistance to bTB in the European wild
- 500 boar (Sus scrofa). Anim. Genet. 39, 316-320.
- 501 Palomo, L.J., Gisbert, J., 2002. Atlas de los mamíferos terrestres de España. Dirección
- General de la Conservación de la Naturaleza-SECEM-SECEMU, Madrid, pp. 306-314.
- Parra, A., Larrasa, J., García, A., Alonso, J.M., Hermoso De Mendoza, J., 2005. Molecular
- 504 epidemiology of bovine tuberculosis in wild animals in Spain: A first approach to risk factor
- 505 analysis. Vet. Microbiol. 110, 293-300.
- Parra, A., Garcia, A., Inglis, N.F., Tato, A., Alonso, J.M., de Mendoza, M.H., de Mendoza,
- J.H., Larrasa, J., 2006. An epidemiological evaluation of *Mycobacterium bovis* infections in
- wild game animals of the Spanish Mediterranean ecosystem. Res. Vet. Sci. 80, 140-146.
- 509 Peña, L., Garcia, P., Jiménez, M.A., Benito, A., Alenza, M.D.P., Sánchez, B., 2006.
- 510 Histopathological and immunohistochemical findings in lymphoid tissues of the endangered
- 511 Iberian lynx (*Lynx pardinus*). Comp. Immunol. Microbiol. Infect. Dis. 29, 114-126.
- 512 Pérez de la Lastra, J.M., Galindo, R.C., Gortazar, C., Ruiz-Fons, F., de la Fuente, J. 2009.
- 513 Expression of immunoregulatory genes in peripheral blood mononuclear cells of European
- wild boar immunized with BCG. Vet. Microbiol. 134, 334-339.
- Revilla, E., Palomares, F., 2002. Spatial organization, group living and ecological correlates
- 516 in low-density populations of Eurasian badgers, *Meles meles*. J. Anim. Ecol. 71, 497-512.
- 817 Romero, B., Aranaz, A., Sandoval, A., Álvarez, J., de Juan, L., Bezos, J., Sánchez, C., Galka,
- 518 M., Fernández, P., Mateos, A., Domínguez, L., 2008. Persistence and molecular evolution of
- 519 Mycobacterium bovis population from cattle and wildlife in Doñana National Park revealed
- by genotype variation. Vet. Microbiol. 132, 87-95.

- 521 Sobrino, R., Martin-Hernando, M.P., Vicente, J., Aurtenetxe, O., Garrido, J.M., Gortazar, C.,
- 522 2008. Bovine tuberculosis in a badger (*Meles meles*) in Spain. Vet. Rec. 163, 159-160.
- 523 Sobrino, R., Acevedo, P., Escudero, M.A., Marco, J., Gortazar, C., 2009. Carnivore
- 524 population trends in Spanish agrosystems after the reduction in food availability due to rabbit
- decline by rabbit haemorrhagic disease and improved waste management. Eur. J. Wildl. Res.
- 526 55, 161-165.
- 527 Vicente, J., Hofle, U., Garrido, J.M., Fernandez-de-Mera, I.G., Juste, R., Barral, M.,
- 528 Gortazar, C., 2006. Wild boar and red deer display high prevalences of tuberculosis-like
- 529 lesions in Spain. Vet. Res. 37, 107-119.
- Vicente, J., Delahay, R.J., Walker, N.J., Cheeseman, C.L., 2007a. Social organization and
- 531 movement influence the incidence of bovine tuberculosis in an undisturbed high-density
- badger *Meles meles* population. J. Anim. Ecol. 76, 348-360.
- Vicente, J., Höfle, U., Garrido, J.M., Fernández-De-Mera, I.G., Acevedo, P., Juste, R., Barral,
- 534 M., Gortazar, C., 2007b. Risk factors associated with prevalence of tuberculosis-like lesions
- in wild boar and red deer in South Central Spain. Vet. Res. 38, 451-464.
- Vicente, J., Hofle, U., Fernandez-De-Mera, I.G., Gortazar, C., 2007c. The importance of
- parasite life history and host density in predicting the impact of infections in red deer.
- 538 Oecologia 152, 655-664.
- Wobeser, G.A., 1994, Disease in Wild Animals, Investigation and management, 1st edition
- Edition. Plenum, New York, pp. 131-234.
- Zanella, G., Durand, B., Hars, J., Moutou, F., Garin-Bastuji, B., Duvauchelle, A., Ferme, M.,
- 542 Karoui, C., Boschiroli, M.L., 2008. *Mycobacterium bovis* in wildlife in France. J. Wildl. Dis.
- 543 44, 99-108.

Table 1.- Current situation in Spain, status regarding bovine tuberculosis (bTB), and selected

545 bTB risk factors among three wild ungulates.

546

	Host species			
Fact	Eurasian wild boar	Red deer	Fallow deer	References
Situation in Spain				
- Current distribution (1)	Widespread	Patchy	Local	Palomo and Gisbert, 2002
- Population trend	Increasing	Increasing	Stable to increasing	Gortazar et al. 2000, Acevedo et al. 2006
- Abundance	Range 1 to 90/km ² , even higher in farm-like enclosures	Mean 21, locally up to 69/km ²	Usually lower than red deer, but locally up to $50/\text{km}^2$	Acevedo et al. 2007, 2008
Status regarding bTB				
- Prevalence	42.5% (max 100%)	13.7% (max 50%)	? (max 18.5%)	Vicente et al. 2006, Gortazar et al. 2008
- % of generalized bTB	57.8-61.2%	57.1-70%	73.3%	Vicente et al. 2006, Martín- Hernando et al. 2007, 2010
- % of lung lesions	38.1%	30%	80%	Martín-Hernando et al. 2007, 2010
Known bTB risk factors	s			
- Individual	Age, Genetics,	Age, Sex, Genetics	Age, Sex	Acevedo-Whitehouse et al. 2005, Vicente et al. 2006, Gortazar et al. 2008, Fernández-de-Mera et al. 2009
- Intra and inter- specific	Social group, Red deer bTB prevalence	Social group, Wild boar bTB prevalence	Not known	Vicente et al. 2007, Gortazar et al. (Submitted)
- Environmental	Density, Fencing, Hardwood <i>Quercus</i> spp. forest availability, Spatial aggregation of wild boar at artificial watering sites	Density, Hardwood Quercus spp. forest availability, Spatial aggregation of wild boar at artificial watering and feeding sites	Not known	Acevedo-Whitehouse et al. 2005, Vicente et al. 2007a, 2007b
- Ability to cross fences	High	Low	Low	Unpublished data from GPS-tagged wild boar
- Use of carrion	Frequent	Occasional	No	Unpublished data from camera trap surveys

547

548 (1) All three are absent from the island regions.

549	Figure legends
550	
551	Figure 1 Recent trends in cattle herd bovine tuberculosis (bTB) prevalence (in %) in three
552	ecologically contrasting Spanish regions, the Balearic Islands (Mediterranean climate, no
553	potential wildlife reservoirs), Galicia (Atlantic climate, potential wildlife reservoirs
554	abundant), and Castilla - La Mancha (Mediterranean climate, potential wildlife reservoirs
555	abundant and frequent high wire fencing & feeding). Prevalence data from MARM
556	(http://www.mapa.es).
557	
558	Figure 2 Risk factors for bovine tuberculosis in Spanish wildlife. (a) High wire fencing
559	nowadays occupies over 50% of suitable wildlife habitat in some provinces. While most
560	fences are not wild boar proof, they reduce the genetic variability and disease resistance of
561	red deer and wild boar. (b) Feeding and artificial watering causes aggregation of wild
562	ungulates, in the picture wild boar. (c) Wildlife translocations imply the risk of spreading
563	diseases. (d) Carrion consumption. Red deer stag consuming material from a hunting gut pile.
564	The picture was taken by a movement triggered camera set up after a driven hunt to record
565	carrion consumption.
566	
567	Figure 3 Ten year trends in the prevalence of bovine tuberculosis compatible lesions in
568	wild boar from south central Spain. In panel (a), total lesion prevalence for each of the 14
569	populations in 2001-2004 and 2007-2009 is shown. Panel (b) shows the prevalence by age
570	class for a population with declining trend (NV) and a population with an increasing trend
571	(CB). These differences are most evident among yearlings.







