1	The inability of a bacteriophage to infect Staphylococcus aureus
2	does not prevent it from specifically delivering a photosensitiser to
3	the bacterium enabling its lethal photosensitisation
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5	Running Title: PDT Phage
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19	Keywords:
20	Tin (IV) chlorin e6 (SnCe6), MRSA, Photodynamic therapy (PDT)
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#### 24 Abstract

Objectives: It has been demonstrated that the efficiency of lethal photosensitisation can be improved by covalently binding photosensitising agents to bacteriophage. In this study we have investigated whether a bacteriophage requires the capacity to infect the bacterium to enhance lethal photosensitisation when linked to a photosensitizer.

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Methods: Tin (IV) chlorin e6 (SnCe6) was conjugated to bacteriophage  $\Phi$ 11, a transducing phage which can infect *Staphylococcus aureus* NCTC 8235-4, but not EMRSA 16. The conjugate and appropriate controls, were incubated with these bacteria and either exposed to laser light at 632.8 nm or kept in the dark.

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Results: The SnCe6 /  $\Phi$ 11 conjugate achieved a statistically significant reduction in the number of viable bacteria of both 8325-4 and EMRSA 16 strains by 2.31 log<sub>10</sub> and 2.63 log<sub>10</sub> respectively. The conjugate could not however instigate lethal photosensitisation in *E. coli*. None of the other combinations of controls; such as an equivalent concentration of SnCe6 only, an equivalent titre of bacteriophage only or experiments conducted without laser light; yielded significant reductions in the number of viable bacteria recovered.

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Conclusions: The inability of a bacteriophage to infect *S. aureus* does not prevent it
from specifically delivering a photosensitiser to a bacterium enabling its lethal
photosensitisation.

#### 47 Introduction

Light-activated antimicrobial agents (photosensitisers) are an appealing alternative to 48 conventional antibiotics for the treatment of localised bacterial infections. Lethal 49 photosensitisation (LP) has been demonstrated to be effective at killing a range of 50 bacteria including opportunistic pathogens, commensal cutaneous species,<sup>1</sup> 51 periodontal pathogens<sup>2</sup> and epidemic methicillin-resistant Staphylococcus aureus 52 (EMRSA).<sup>3</sup> LP has two main advantages over conventional antimicrobial 53 chemotherapy. Firstly, the bactericidal effect is limited to the area that is treated with 54 55 both the photosensitiser and light, preventing disturbance of the wider commensal microbial community.<sup>4</sup> Secondly, the non-specific mode of action of liberated singlet 56 57 oxygen  $({}^{1}O_{2})$  against bacteria is unlikely to induce the development of protective mechanisms and the subsequent proliferation of these genes through the wider 58 microbial community. 59

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The inherent reactivity of  ${}^{1}O_{2}$  limits its ability to diffuse through an aqueous 61 environment. The lifetime of  ${}^{1}O_{2}$  in pure water is ~4 µs, which results in a theoretical 62 diffusion distance of 125 nm, if one assumes that the moiety does not interact with a 63 biological molecule.<sup>5</sup> This short range action (on the scale of biological systems) 64 may possibly limit the effectiveness of LP. We have previously developed 65 66 methodologies to facilitate the close association of photosensitiser and bacteria using targeting systems based upon the covalent conjugation of the photosensitiser 67 tin (IV) chlorin e6 (SnCe6) onto immunoglobulin G (IgG).<sup>3, 6</sup> More recently, we have 68 found that covalently linking SnCe6 to S. aureus bacteriophage 75, commonly used 69 for typing, targets lethal photosensitisation to a range of strains of *S. aureus* 70 including MRSA.<sup>7</sup> In the study reported herein, we examined whether another 71

unrelated *S. aureus* bacteriophage, phage  $\Phi$ 11 a generalised transducing phage, could replace phage 75 in targeting lethal photosensitisation to *S. aureus* strains.

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## 75 Materials and Methods

## 76 Bacteria and Bacteriophage

77 The two strains of S. aureus used in these experiments were EMRSA-16 (NCTC 13143), one of the dominant nosocomial MRSA isolates in UK hospitals and 8325-4, 78 a prophage-free derivative of NCTC8325 which is methicillin-sensitive. These 79 80 strains were grown on Columbia agar (Oxoid Ltd., Basingstoke, United Kingdom) 81 supplemented with 5% (vol/vol) defibrinated horse blood (CBA). E. coli 10418 was 82 also incorporated as a Gram-negative control. In preparation for the lethal 83 photosensitisation experiments, a colony was inoculated into 20 mL of nutrient broth no. 2 (NB2) containing 10 mM CaCl<sub>2</sub> and grown aerobically for 16 hours at 37°C in a 84 shaking incubator. The cultures were then washed by centrifugation and re-85 86 suspension in PBS containing 10 mM CaCl<sub>2</sub> and adjusted to a final optical density of 0.05 at 600 nm (OD<sub>600</sub>), these cell suspensions contained approximately 1 x  $10^7$ 87 88 cfu/mL.

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Bacteriophage  $\Phi 11$  is a generalised transducing phage present in *S. aureus* NCTC 8235 as a prophage.<sup>8</sup> This bacteriophage can infect the NCTC8325 derivative 8325-4, but not EMRSA16. Phage  $\Phi 11$  was propagated in *S. aureus* 8325-4 using the phage overlay method and SnCe6 was covalently conjugated to the bacteriophage using methods described previously.<sup>7</sup> The concentration of SnCe6 bound to the phage was determined by spectral analysis against a calibration curve generated from known concentrations of SnCe6. In different experiments, between 7.5 x 10<sup>6</sup> and  $4.7 \times 10^7$  cfu of bacteriophage were used in conjugation reactions and the amount of SnCe6 bound varied between 3.5 and 7 µg/mL.

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## 100 Lethal Photosensitisation Experiments

Fifty microlitres of the  $\Phi$ 11-SnCe6 conjugate was added to 50 µL of bacterial 101 102 suspension in a sterile 96-well plate. The controls consisted of: SnCe6 alone (at the 103 same concentration as the conjugate),  $\Phi$ 11 alone (at the same titre as the conjugate) 104 and PBS control. All of the mixtures were incubated in the dark, with stirring, for 30 105 minutes prior to exposure to laser light. The relevant samples were then sequentially 106 exposed to laser light (632.8 nm) from a helium / neon (HeNe) gas laser with a 107 measured power output of 29.2 mW (Spectra-Physics, Darmstadt-Kranichstein, 108 Germany) for a period of 5 minutes; the mixtures were magnetically stirred 109 throughout the course of an experiment. Additional 'dark controls' were also 110 conducted for these four variables without laser light. The number of viable bacteria 111 remaining in the samples was determined immediately following exposure to the 112 laser light by serial dilution and enumeration of colony forming units on CBA. Each 113 experimental variable was repeated as a duplicate.

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The duplicate experiments with both strains of *S. aureus* were conducted a total of four times (n=8), whilst those for *E. coli* were repeated twice (n=4). The null hypothesis was that there was no difference between the  $log_{10}$  counts of the number of colony forming units using various different experimental parameters, this was analysed by student's t-test to yield p-values.

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#### 122 **Results**

123 When compared to the control, which was not exposed to laser light nor to

124 photosensitiser, the  $\Phi$ 11-SnCe6 conjugate in the presence of laser light yielded a

125 2.31 log<sub>10</sub> reduction (p=<0.05) in the number of viable bacteria recovered from the

126 culture of *S. aureus* 8325-4 and a 2.63  $log_{10}$  reduction (p=<0.05) for the culture of

127 EMRSA16. In the presence of laser light, the  $\Phi$ 11-SnCe6 conjugate did not result in

significant killing of *E. coli* 10418. None of the other combinations of controls (i.e.

129 SnCe6 only, phage only and 'dark controls') produced significant bacterial kills

130 (figure 1).

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## 132 **Discussion**

133 We have previously shown that bacteriophage 75, a serotype F staphylococcal 134 phage, could be used to target lethal photosensitisation to a range of S. aureus strains including strains it could not infect. The capacity of bacteriophage 75 to 135 136 target LP to a range of S. aureus strains was surprising since this phage has a 137 restricted host range. The question we asked in the current study was whether other 138 staphylophage could target LP to a range of S. aureus strains, once conjugated to a photosensitiser, or if this was a specific trait of phage 75. We did this by 139 investigating the capacity of bacteriophage  $\Phi$ 11, a prototypic group B-transducing 140 phage,<sup>9</sup> to target lethal photosensitisation to *S. aureus*. 141 142

143 When SnCe6 was conjugated to *S. aureus* bacteriophage  $\Phi$ 11, *S. aureus* strains 144 8325-4 and EMRSA16 in the presence of laser light there was an increase in the 145 killing of these bacteria by 2.39 log<sub>10</sub> and 2.35 log<sub>10</sub> respectively, when compared to 146 the equivalent concentration of SnCe6 alone (i.e. free SnCe6 that was not 147 conjugated to the bacteriophage). Since it is known that staphylophage have the capacity to bind to all strains of *S. aureus*<sup>10</sup> and bacteriophage  $\Phi$ 11 is not capable of 148 infecting strain EMRSA16, the kill achieved by the  $\Phi$ 11-SnCe6 conjugate suggests 149 150 that the photosensitiser-bacteriophage conjugate only needs to bind to the bacterial cell to induce killing in the presence of laser light. The selectivity of the 151 152 photosensitier-bacteriophage conjugate in targeting lethal photosensitisation to S. aureus was demonstrated by the inability to cause significant killing of E. coli in the 153 154 presence of laser light.

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156 Our results demonstrate that it is possible to use different serotypes of

157 staphylophage as vehicles to deliver photosensitiser payloads to the surface of *S*.

aureus thus enabling selective lethal photosensitization of this bacterium in the

159 presence of laser light. Such designer composites would not only possess all of the

advantages that photodynamic therapy has over conventional antibiotic therapy, as

161 described in the introduction, but they would also ensure there was minimal collateral

162 damage to the host and its indigenous microflora.

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- 164 **Transparency Declarations**
- 165 None

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- 167 Funding
- 168 This project was funded internally

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# 172 Figure Legend



174 Figure 1. The number of viable bacteria recovered following exposure of SnCe6-

175 bacteriophage 11 conjugate to laser light (leftmost columns) compared to controls.

176 The designations L+ / L- and S+ / S- refer to the presence or absence of light and /

- 177 or photosensitiser respectively.
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