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1	The Diagnosis of Pancreatic Disease in Feline Platynosomosis
2	Liza S Köster ¹ *, Linda Shell ¹ , Jennifer Ketzis ² , Sreekumari Rajeev ² , and Oscar Illanes ²
3	¹ Department of Clinical Sciences, Ross University School of Veterinary Medicine, Basseterre,
4	West Indies
5	² Department of Biomedical Sciences, Ross University School of Veterinary Medicine,
6	Basseterre, West Indies
7	Running head: feline pancreatic disease and <i>Platynosomum</i> spp. infection.
8	Keywords: cat, cobalamin, fPL, pancreatitis, platynosomiasis, platynosomosis, Platynosomum
9	spp., fTLI
10	Abbreviations:
11	EPI: exocrine pancreatic insufficiency
12	fPL: feline pancreatic lipase
13	fTLI: feline trypsin-like immunoreactivity
14	IQR: interquartile range
15	RI: reference intervals
16	*Corresponding author: Liza S Köster, BVSc, DECVIM-CA, School of Veterinary Medicine,
17	College of Medical, Veterinary and Life Sciences, University of Glasgow, Bearsden Road,
18	Glasgow, G61 1QH, UK, email: <u>liza.koster@glasgow.ac.uk</u>

19 Abstract

20 Background *Platynosomum* spp. are cat-specific parasitic trematodes that parasitize the biliary 21 ducts and gallbladder. Due to the common connection to the major duodenal papilla of the 22 pancreas and common bile ducts in addition to the periductal proximity of the pancreas, it is 23 possible that platynosomosis could cause pancreatitis. 24 Hypothesis/Objectives To determine if platynosomosis, a commonly diagnosed parasitic disease 25 in cats on St. Kitts, has any association with pancreatic disease. To investigate this possibility, 26 the pancreas of free roaming cats with naturally acquired platynosomosis were evaluated via 27 ultrasound, serum concentrations of fPL, cobalamin, folate and fTLI, and histopathology. 28 Animals: Twenty free roaming, young adult feral cats, positive for feline immunodeficiency 29 virus, and diagnosed with *Platynosomum* spp. infection via fecal analysis. 30 Methods: The liver, biliary system, and pancreas were evaluated via ultrasonography during a 31 short duration anesthesia. Serum concentrations of fPL, fTLI, folate, and cobalamin were 32 measured. Sections of the right limb, left limb, and body of the pancreas were evaluated 33 histopathologically using H&E stain. 34 Results: None of the cats had sufficient criteria to fulfill the ultrasonographic diagnosis of 35 pancreatitis. One cat had an elevated fPL concentration in the range consistent with pancreatitis.

36 Four cats had cobalamin deficiencies and eleven had abnormal folate concentration. The fTLI

37 concentration was equivocal for the diagnosis of exocrine pancreatic insufficiency in one cat.

38 With a single exception, histopathology changes, when present (n=12), were mild, non-specific

39 and predominantly characterized by lymphocytic infiltrates and fibrosis. The exception was a cat

40 which presented a chronic interstitial and eosinophilic pancreatitis of slightly increased severity

41 likely the result of platynosomosis.

- 42 <u>Conclusion and clinical importance</u>: The results of this study suggest that platynosomosis rarely
- 43 induce pancreatic damage in cats. With only one exception, chronic pancreatitis diagnosed in
- 44 cats with fluke-induced cholangitis and cholangiohepatitis was subtle and interpreted as an
- 45 incidental background lesion unrelated to platynosomosis.

46 Introduction

47 Platynosomum spp. are cat-specific parasitic liver trematodes (flukes) that occupy the 48 hepatic ducts and gallbladder(1) of cats from tropical and subtropical regions of the world.(2) 49 Gross pathology findings in *Platynosomum* spp. infected domestic cats have included 50 hepatomegaly, a yellow and friable liver, biliary duct distension, increased bile consistency with 51 visible flukes, mesenteric lymphadenopathy, and ascites.(1) Both immature and mature flukes of 52 the *Platynosomum* spp. have been found in the pancreatic duct of cats.(3) Due to the common 53 connection to the major duodenal papilla of the pancreatic and common bile ducts, in addition to 54 the periductal proximity of the pancreas, it is possible that a liver fluke infection could cause 55 pancreatitis. Pancreatitis has been reported in a cat infected with *Eurytrema procyonis*, the 56 Raccoon pancreatic fluke.(4)

Hepatobiliary changes on ultrasonography in domestic cats infected with *Platynosomum*spp. are not specific to the fluke infection but 29% of cats have associated bile duct or
gallbladder alterations.(5) Abnormalities included distention of the gallbladder, with hyperechoic
walls and anechoic bile content and tortuous and distended bile ducts. Further, livers in infected
cats are described as enlarged, irregular in shape, and hyperechoic with heterogeneous texture.
Ultrasonographic descriptions of the pancreas in cats with platynosomosis have not been
described.

In the majority of feline pancreatitis cases, the etiology is unknown. Pancreatitis in cats has been classified as acute, chronic or chronic-active. Another classification scheme is based on histopathologic findings: necrotizing pancreatitis, pancreatic fibrosis with inflammation or without inflammation.(6) A histologically distinct suppurative pancreatitis, which is an acute form of the disease, has been documented in cats.(7) Acute pancreatitis in cats, which is usually 69 a clinically apparent condition, can occur in any age of cat, obese or underweight.(7). Although 70 the Siamese breed was overrepresented in one study (7) a review of other published cases could 71 not support that finding. Documented conditions or infectious diseases associated with 72 pancreatic pathology in cats are numerous and include: Toxoplasmosis(8, 9), flukes (Eurytrema 73 procyonis)(4), trauma(10), hypocobalaminemia(11), triaditis(12), hepatic lipidosis(13), diabetes 74 mellitus(14), and cavity effusions.(15) It is believed that acute and chronic pancreatitis occurs 75 commonly in cats. In one study of cats that were clinically normal at the time of death, the 76 prevalence of pancreatitis was 45% based on histopathology of pancreas, the prevalence of 77 pancreatitis increased to 67% when cats that were euthanized for a specific disease including 78 both gastrointestinal and non-gastrointestinal conditions.(16)Unfortunately ante-mortem 79 diagnosis of pancreatitis in domestic cats remains challenging due to inconsistent clinical 80 chemistries and the lack of specific clinical signs. Many cats suffer from subclinical pancreatitis 81 which can impair successful management of the comorbid diseases associated with pancreatitis. 82 One third of cats with diabetes mellitus are estimated to have subclinical pancreatitis based on 83 either elevated fPL or abdominal ultrasonography changes, which could have implications on 84 achieving remission.(14) In addition, the magnitude of fPL concentration has shown to be 85 associated with outcome in acute pancreatitis and is considered a prognostic variable in critically 86 ill cats.(17) The unfavorable reputation of abdominal ultrasonography in its ability to discern 87 pancreatic pathology has been recently highlighted in several studies where the correct diagnosis 88 of pancreatitis was made in a paltry 20% to 54% of cases.(18-20) While pancreatic 89 ultrasonography has the advantage of being a non-invasive method of assessing morphology, it is 90 limited by the lack of specificity and dependent on the skills of the sonographer. However, 91 ultrasonography is considered to be a reasonable clinically diagnostic tool when used in

92 combination with serological markers of pancreatitis. The current test of choice for non-invasive 93 diagnosis of pancreatitis is the assessment of feline pancreatic specific lipase (fPL) 94 concentration.(21) Recently a study demonstrated that the sensitivity and specificity of 95 ultrasonography in detecting acute pancreatitis diagnosed by abnormal fPL, was 84% (95% 96 confidence interval = 60-97%), and 75% (95% confidence interval = 48-93%) respectively.(13) 97 This same study found that ultrasonographic changes of pancreatic limb thickening, loss of 98 margination and peripancreatic fat hyperechogenicity in combination with an abnormal fPL 99 assay were highly indicative of acute pancreatitis in cats. Computed tomography offers little 100 additional benefit in the ability to diagnose pancreatitis in cats, with a sensitivity of 20% reported 101 in one study.(18) Endosonography is reported to have superior visualization of the normal 102 pancreas with better resolution in pancreatic pathology, however the diagnosis of pancreatitis 103 would not have been altered in the study that compared its usefulness to that of abdominal 104 ultrasonography, making the clinical application of this invasive diagnostic test limited.(22) 105 Exocrine pancreatic insufficiency (EPI) is assumed to develop as a consequence of end-106 stage chronic pancreatitis in the feline species; however cats less than six months of age also can 107 develop EPI.(23) Reported prevalence ranging from 0.013 - 0.103% has been increasing, 108 possibly due to increased awareness and availability of commercial assays. (23-26) Typically cats 109 diagnosed with EPI have hypocobalaminemia due to malabsorption, while folate concentrations 110 are normal or increased. (25, 26) The gold standard for diagnosis of EPI in domestic cats has not 111 been established, but feline trypsin-like immunoreactivity (fTLI) has been validated for use in 112 serum from starved cats, with concentrations $\leq 8 \text{ ug/L}$ consistent with the diagnosis of EPI.(27) 113 While very few of the cats have had biopsy confirmation of the disease the diagnosis is 114 supported by clinical response to pancreatic enzyme supplementation, reported in 87% of

115 cases.(25) While one study diagnosed EPI using fTLI with a cut-off of $12 \mu g/L(11)$, the

recommendation of Gastrointestinal Laboratory, Texas A&M, refers to a cut-off of 8 ug/L for the
diagnosis and between 8-12 µg/L as equivocal.

The objective of this study was to determine if platynosomosis, a commonly diagnosed parasitic disease in cats on St. Kitts, could be associated with pancreatitis and its sequela EPI. To investigate this, the morphology, functional reserve, and architecture of the pancreas of free roaming cats naturally infected with *Platynosomum* spp. were evaluated using abdominal ultrasound, serum concentrations of fPL, fTLI and cobalamin, and histopathology.

123 Materials and Methods

124 This prospective study was conducted in conjunction with a larger investigation on the 125 diagnosis and treatment of platynosomosis in domestic cats on St. Kitts, West Indies. Cats were 126 recruited from the Ross University School of Veterinary Medicine (RUSVM) Feral Cat Program 127 (FCP), a trap, spay/neuter and release (TNR) program, during the period August 2014 to July 128 2015. All procedures in this study as well as those within the FCP and the larger platynosomosis 129 study were conducted under the following RUSVM Institutional Animal Care and Use 130 Committee (IACUC), approved protocols: FCP (13-9-017), FCP retrovirus testing (15-2-006), 131 necropsy and organ harvesting (14-3-009), and Platynosomum *spp*. investigation (15-1-004). The 132 cat housing facility also was inspected and approved by the IACUC. As per the Feline 133 Immunodeficiency Virus (FIV) "test and remove policy" of the TNR program, all cats more than 134 six months old were tested using a patient side commercial FIV antibody testa and euthanized if 135 tested positive. Only FIV positive cats were included in the study. A fresh stool sample was 136 collected for diagnosis of *Platynosomum* spp. infection by standard parasitological methods 137 described elsewhere.(28)

Prior to euthanasia, eight cats were allocated to another study which investigated the efficacy of praziquantel as treatment of platynosomosis. These eight cats were examined and euthanized at either 12 days (n=4) or 24 days (n=4) post FIV testing and after treatment with praziquantal. Their appetite, appearance, urination and defecation habits were monitored twice daily.

Food was withheld for 12 hours prior to anesthesia for each procedure, but water was
available *ad libitum*. Abdominal ultrasonography was performed under a short duration
anesthesia, using a combination of ketamine hydrochloride (3-5 mg/kg)b, buprenorphine
hydrochloride (0.01-0.016 mg/kg)c, and dexmedetomidine hydrochloride (11-20 μg/kg)d
administered intramuscularly. All cats were euthanized using pentobarbital (1ml/4.5kg)e
administered intravenously while they were anesthetized.

149 Ultrasonography was performed by an internal medicine specialist, using an 8.5 MHz to 150 14 MHz sector scannerf. All images were saved onto the server (DICOM). The cat was 151 positioned in dorsal recumbency and the abdomen was clipped using electronic clippersg. After a 152 complete abdominal ultrasound, the pancreas was evaluated and measured using electronic 153 calipers in either longitudinal or transverse plane, whichever was most appropriate for the area 154 being examined.

155 The procedure for pancreatic measurements and grading followed the criteria
156 recommended by Williams *et al.* (2013).(13) Thickness was the maximum ventro-dorsal width
157 measured using electronic calipers, with greater than 1 cm considered abnormal.(29, 30) As per
158 the classification scheme developed by Zimmerman et al (2013)(10), four ultrasonographic
159 criteria of pancreatitis (change in echogenicity, hyperechoic peripancreatic fat, pancreatic
160 enlargement, and peritoneal fluid) were considered. Finding three or more of these criteria was

161 considered evidence for ultrasonographic changes consistent with pancreatitis.

162	Serum, collected at the time of euthanasia, was stored at -80°C until it was shipped to
163	IDEXX laboratoryh for determinations of fPL, folate, cobalamin and fTLI. Reference intervals
164	(RI) for fPL was reported as $0 - 3.5 \mu$ g/L. In addition, as part of the gastroenterology panel
165	offered by IDEXX laboratories, folate (RI: 8.9 – 19.9 µg/L), cobalamin (RI: 276 – 1425 ng/L),
166	and feline trypsin-like immunoreactivity (fTLI) (RI: 12 - 82 ug/L) were determined. As per the
167	reference laboratory, a diagnosis of pancreatitis is considered with fPL values > 3.5 ug/L or fTLI
168	values > 100 ug/L. EPI was considered with fTLI values $\leq 8 \ \mu g/L$.
169	Immediately after euthanasia a gross post-mortem examination was conducted. Samples
170	from the body and both limbs of the pancreas were removed with at least 6 sections taken, 2
171	from each lobe or from the body submitted in separate containers, with a few cases where the
172	pieces of pancreas were submitted in a single container. These samples were labeled, fixed in 10
173	% neutral buffered formalin for at least 48-hours, embedded in paraffin, cut and 5 micron
174	sections were stained with hematoxylin and eosin for routine light microscopy evaluation.
175	Two board certified pathologists, blinded to all other results, reviewed, described, and
176	graded the histological sections using the classification suggested by De Cock et al. (2007).(16)
177	Excel software was used to calculate the median and interquartile range (IQR) of the
178	continuous data.
179	Results
180	Animals

181 Twenty young adult cats (17 intact males, 3 intact females) were included in this study.
182 None of the eight cats that were recruited for the praziquantel study were removed due to
183 complications of treatment and good appetite and normal activity were recorded throughout the

184 duration of this study. All cats tested positive for *Platynosomum* spp. egg on standard
185 parasitological fecal testing, in addition histopathology confirmed biliary fluke infestation.

186 *Histopathology*

187 Table 1 depicts the histopathology of the twenty cats that were necropsied. Case 12 had 188 evidence of mild, acute pancreatitis based on the presence of small intralobular lesions 189 containing a few neutrophils within the inflammatory cell infiltrate. Eleven cats, including case 190 12, had mild chronic pancreatitis characterized by interlobular lymphocytic inflammation and 13 191 cats had mild periductular fibrosis. Mild peri-pancreatic fat inflammation was noted in case 3, 192 nodular hyperplasia in cases 8 and 13 with minimal amyloid deposition within pancreatic islets 193 also seen in case 13. Chronic mild to moderate interstitial lymphocytic and eosinophilic 194 pancreatitis was detected in the right pancreatic lobe of case 14, which also exhibited a moderate 195 hyperplastic and eosinophilic lymphadenitis within regional pancreaticoduodenal lymph nodes. 196 Pancreatic lesions in this animal were interpreted as significant, even though they involved less 197 than 20% of the affected lobe (Figure 1 and 2), and the result of platynosomosis. Cholangitis, 198 mild to severe, was confirmed in all 20 cats and cholecystitis, mild and moderate severity, in 18 199 of the cats.

200 Ultrasono

Ultrasonographic evaluation

The left limb, right limb, and the body of the pancreas were visualized in 14, 14, and 13 cats respectively of the 20 cats examined. The median thickness and IQR of the left limb, right limb, and body of the pancreas was 8.1 mm (IQR: 6.2 mm; 8.5 mm; n = 14), 4.95 mm (4.1 mm; 6.7 mm; n = 14), and 6.6 mm (4.3 mm; 7.4 mm; n = 13) respectively. One of the twenty cats (case 14) had a thickened (> 1 cm) pancreas, with the left pancreatic limb measuring 10.8 mm. The margination of the right or left limb was described as irregular in two and five cats 207 respectively, two of which also had irregular margination of the pancreatic body. An additional208 four cats had irregular margination of the body without abnormal limb margination.

209 Echogenicity of the left limb, right limb, and pancreatic body was described as 210 normoechoic in eight, nine, and five cats respectively; hyperechoic in two, three, and two cats 211 respectively; hypoechoic in four, two, and six cats respectively. Peripancreatic fat was 212 considered normoechoic in all cats without evidence of free fluid. The median pancreatic duct 213 was 1.3 mm (0.9 mm; 1.65 mm; n = 11) with a diameter of greater than 10 mm in seven cats. 214 Based on the ultrasonographic criteria of pancreatitis, 13 cats had a score of 1 and 7 cats had a 215 score of 0; thus none of the cats were considered to have pancreatitis based on ultrasound results 216 (Table 1).

217 *fPL*, cobalamin, folate, and fTLI results

218 With few exceptions, the concentrations of fPL, cobalamin, folate, and fTLI for the 20 219 cats in this study were within the normal reference range (Table 1). One cat (case 20) had 220 increased fPL (22.7 µg/L) and high normal fTLI (79.1 µg/L) concentrations consistent with acute 221 pancreatitis as well as low folate concentrations. Four cats (cases 5, 6, 8 and 17) had cobalamin 222 concentrations below reference range. Five cats (cases 1, 8, 17, 18, and 20) had low folate, and 223 six cats (cases 4, 9, 13, 14, 15, and 16) had increased folate concentration. Case 6 had a fTLI 224 concentration below published reference range (10.2 μ g/L) but not within the diagnostic 225 reference range ($<8 \mu g/L$) for EPI, in addition to a low cobalamin concentration (163 ng/L). 226 Discussion 227 In this population of twenty clinically healthy, young adult, feral cats diagnosed with 228 platynosomosis and confirmed cholangitis and cholecystitis, pancreatic histopathology supported

the diagnosis of mild acute pancreatitis in one cat and mild chronic pancreatitis in 11 (55%). Five

of these cats also had abnormal clinical chemistry results that may have been related topancreatic disease yet none of the cats had significant ultrasound changes.

While abdominal ultrasonographic parameters of pancreatitis are not well defined,

- 233 particularly in the case of chronic forms, abdominal ultrasonography remains a practical non-234 invasive imaging modality used for antemortem diagnosis of pancreatitis. Fortunately age-235 related pancreatic changes in echogenicity and width are not significant in cats as they are in 236 humans; (30) thus there was no need to account for age in our study. In this study, while 237 pancreatic changes were visible, none of the cats had sufficient criteria (pancreatic limb 238 thickening, loss of margination and peripancreatic fat hyperechogenicity) to fulfill the 239 ultrasonographic diagnosis of pancreatitis. Hyperechoic peripancreatic fat, considered a sensitive 240 index of pancreatitis in cats (68%) (13), was not found in any of the cats in this study. 241 Unfortunately the often multifocal histological distribution of pancreatitis does not aid the ability 242 to diagnose pancreatitis by ultrasonography.(13). Ferreri et al (2003) showed that 54% and 46% 243 of cats with acute and chronic pancreatitis respectively had unremarkable ultrasonographic 244 changes of the pancreas.(20) In addition ultrasonographic changes were unable to distinguish 245 acute from chronic pancreatitis. More recently, Oppliger et al (2014), found similar results in 246 that 39% of cats diagnosed with pancreatitis based on fPL had unremarkable ultrasonographic 247 changes and an agreement between fPL concentration and histopathology was not found.(31) 248 The current test of choice for non-invasive diagnosis of pancreatitis is the fPL 249 concentration (21). In our study, only one cat had an fPL concentration in the range consistent 250 with pancreatitis. In this cat the pancreas was not detectable with ultrasonography and the only
- 251 noticeable pathology was mild fibrosis.
- 252

232

Eleven cats had histopathological changes of chronic pancreatitis. One of these cats, case

253 6, had abnormal concentrations of both fTLI (10.2 μ g/L) and cobalamin (163 ng/L) suggesting a 254 diagnosis of EPI which is assumed to be a consequence of end-stage chronic pancreatitis in the 255 feline species. Mild lymphocytic inflammation and periductular fibrosis was found on pancreatic 256 histopathology. The history of this cat is unknown, but the cat's body condition score was 257 graded as 2.5/5 (4/9)(32) with a body mass of 2.75 kg, which is considered underweight for an 258 adult male domestic shorthair. However, without the clinical history of chronic enteropathy, the 259 diagnosis of EPI remains equivocal in this individual.

260 The potential multifocal nature of pancreatitis and small sample sizes are possible reasons 261 that histopathological lesions could have been missed. The lack of a control group is another 262 limitation to the study. Since ultrasonography was not performed by a boarded radiologist, the 263 sensitivity for this procedure could have been influenced by operator experience. Lastly, all cats 264 included in this study were FIV positive which could have potentially affected pancreatic 265 pathology, therefore the results of this study do not necessarily apply to FIV negative cats. 266 Except for the one case, case 14, with chronic periductal eosinophilic inflammatory cell 267 infiltration and fibrosis, likely the result of platynosomosis, microscopic findings in the pancreas 268 of these *Platynosomum spp*-infected cats were subtle, non-specific background lesions 269 interpreted as clinically insignificant and not likely related to platynosomosis. Thus, our findings 270 suggest that *Platynosomum* spp.-induced pancreatic lesions in cats with platynosomosis are rare. 271 While one cat, case 20, had serological indices consistent with pancreatitis based on fPL 272 concentration, and another cat, case 6, had suspected EPI based on a combination of low fTL and 273 cobalamin concentrations, histopathology did not correlate with the diagnoses in either cat. The 274 significance of hypocobalaminemia in four cats with playnosomosis is not yet known but 275 warrants screening of newly diagnosed cases of platynosomosis.

276 Funding and conflict of interest

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382

384 Tables

385	Table 1. The severity of cholangitis and cholecystitis determined by histopathology, pancreatic
386	score based on ultrasound, histopathology, and serological testing in 20 cats diagnosed with
387	<i>Platynosomum</i> sp. infections. A cumulative score \geq 3 was consistent with an ultrasonographic
388	diagnosis of pancreatitis. Feline pancreatic lipase immunoreactivity, cobalamin, and fTLI
389	concentrations were reported as abnormal if they were outside the laboratory reference ranges.
390	

392 Figures

- **393** Fig 1. Chronic interstitial pancreatitis in case number 14. Numerous eosinophils are present
- 394 within the inflammatory cell infiltrate. Hematoxylin and eosin. Bar: 100 microns.

395

- **396** Fig 2. Chronic pancreatitis in case number 14. Eosinophils are a predominant feature within the
- 397 periductal and interstitial inflammatory cell infiltrate. Hematoxylin and eosin. Bar E, 40X.

398

401 Manufacturer details:

aSNAP FIV/FeLV Combo Test, IDEXX Laboratories, Westbrook, Maine, USA

bPfizer Inc. New York, NY

Buprenex, Reckett Benckiser Healthcare, Hull, England

dDexdomitor manufactured by Orion Pharma, Finland and distributed by Zoetis Inc, Kalamazoo

MI

eEuthasol, pentobarbitone sodium, Virbac, Fort Worth, Texas, USA

fEsaote, MyLabTM, Genoa, Italy

gOster® clippers, USA

hGI panel, IDEXX Laboratories, Westbrook, Maine, USA