1	Ecology: The Scientific Naturalist
2	Endless forms most hidden: katydids that masquerade as moss
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5 6	David W. Kikuchi ¹ , Gustavo H. Kattan ² , Carolina Murcia ³ , and Fernando Montealegre ⁴
7	¹ Department of Ecology, Evolution, and Behavior, University of Arizona, Tucson, AZ 85719, USA
8	² Departamento de Ciencias Naturales y Matemáticas, Pontificia Universidad Javeriana Seccional Cali,
9	Cali, Colombia
10	³ Department of Biology, University of Florida, Gainesville, FL, 32611, USA.
11	⁴ School of Life Sciences, University of Lincoln, UK
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15 16	In the cloud forests of the central range of the Colombian Andes, we discovered a species of katydid (Orthontera: Tettigoniidae) that imitates mosses to an uncanny degree and is exceedingly difficult to
17	(orthoptera. Tettigonnoac) that initiates mosses to an ancamy degree and is exceedingly anneal to detect (Fig. 1). The photographs were taken in the Santuario de Fauna y Flora Otún Ouimbaya, at an
18	elevation of 1900 m. With a mean annual precipitation of 2600 mm, the forest is humid and a variety of
19	mosses cover its rocks, tree trunks and palm leaves. The katydid has a dorsally green head and
20	pronotum, and bands on the abdomen that closely match the color of the moss where it hides. The legs,
21	pronotum and abdomen have green-colored triangular projections that imitate moss leaves, completing
22	the disguise. The insect remains motionless for prolonged periods of time.
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24	The insect in the picture belongs to the genus Adeclus (Orthoptera: Tettigoniidae: Pseudophyllinae:
25	Pleminiini). It is an adult, brachypterous male, and wing coloration pattern, pronotal and cerci shape
26	suggest this species is A. trispinosus (Cadena-Castañeda 2011). Resemblance to moss has evolved
27	independently across several taxa of orthopteroid insects (Mugleston et al 2013, Song et al 2015). It is
28	Very common in the Pleminini, a tribe with some 200 species described, and also in other groups of
29	Such campuflage bas also been observed in stick insects of the gonus Acapthoclonia (Phasmatedea) and
30	in the moss mantids genera Pogonogaster and Majangella (Mantodea) (Gutjérrez and Bacca 2014
32	Svenson et al. 2015)
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34	In the 1890s, British evolutionary biologist Edward B. Poulton recognized the mastery of orthopterans
35	over what he termed protective resemblance (Bidau 2014). Katvdids are masters of disguise, a strategy
36	that presumably helps them to avoid attack by visually-oriented predators. To a human observer, many
37	katydids exhibit an exceptional resemblance to a dead leaf, complete with midrib, holes and mildew
38	patches. Leaf-like wings have evolved independently in at least six lineages of Tettigoniidae (Mugleston
39	et al. 2013). Some species also resemble green leaves or lichens (Braun 2011, Bidau 2014). Katydids are
40	particularly diverse in humid tropical forests, where they include an astounding range of disguises. Of
41	the 378 species of katydids collected by Nickle and Castner (1995) at three sites in Loreto Province, Peru,

- 42 273 species had green or brown colors that helped conceal the animals in vegetation. Another 53
- 43 exhibited more refined forms of camouflage, resembling bark, twigs, leaves and lichens. Mapping these
- 44 diverse forms of camouflage (including moss resemblance) in Mugleston's et al. (2013) phylogeny
- 45 suggests multiple origins.
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47 How do animals disguise themselves so thoroughly as this katydid does? Currently, the concept of 48 camouflage recognizes a variety of strategies that make detection and recognition difficult (Ruxton et al. 49 2004, Stevens and Merilaita 2009, 2011). Crypsis refers to strategies that prevent detection. In a type of 50 crypsis known as background matching, an animal presents an appearance (color, pattern) that helps it 51 blend into the background. In contrast to background matching, disruptive coloration reduces the 52 probability of detection by breaking up an animal's outline so that its shape cannot be perceived (Cuthill 53 et al. 2005, Webster et al. 2013). Another form of camouflage known as masquerade works not to 54 prevent detection, but rather recognition (Skelhorn and Ruxton 2010). Masquerading animals resemble 55 irrelevant objects in the environment such as sticks or bird droppings so that as not to attract attention.

56 The katydid we found uses these different strategies to conceal itself: background matching, disruptive

- 57 coloration, and masquerade. Each of these strategies may come into play depending on the distance at
- which the insect is viewed; from afar, it may appear to simply be part of the mossy bark on which it
 rests, and escape detection through background matching. The greenish knobs and brown sections on
- 60 its limbs occur at a spatial frequency similar to the distribution of those elements on the bark (Fig. 1).
- 61 Closer inspection may fail to detect the katydid because the different parts of its disguise disrupt its
- 62 outline: the various chunks of color make it difficult to resolve its legs or body segments. Finally,
- 63 recognition may elude even the most assiduous assailants because its very body parts masquerade as
- 64 elements of a mossy tree: the legs resemble moss leaves; the long antennae pass for thin twigs. The fine
- 65 details of these components may increase the probability that they are perceived as distinct objects,
- 66 making the whole insect harder to notice through an effect that might operate similarly to disruptive
- 67 coloration, yet exploit object recognition rather than edge detection.
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69 Currently, the use of multiple camouflage strategies by the same individual is a burgeoning topic of

- research. Background matching and disruptive coloration operate via distinct mechanisms, but are
- challenging to separate mechanistically (Webster et al. 2013). Sophisticated image analysis can tease
- them apart, however. Recent studies have shown that bird eggs rely on both background matching and

73 disruptive coloration for concealment (Stoddard et al. 2016), and that individuals can lay eggs on

- substrates where one strategy or the other is enhanced (Lovell et al. 2013). Moths can also enhance the
- 75 effects of background matching and disruptive coloration by repositioning their bodies after landing
- 76 (Kang et al. 2015). An interesting aspect of these studies is their emphasis on the importance of habitat
- 77 selection in determining which camouflage strategy predominates.
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- 79 There remain many interesting questions about the function of multiple camouflage strategies. They
- 80 may interact to protect animals when they are perceived both from afar and close up, to improve
- 81 camouflage on specific microhabitats, or increase the versatility of camouflage to work in different
- 82 microhabitats. To study these possibilities, we must take into account the perceptual systems of the

relevant predators: what is the spatial acuity of their vision, how do the retinas of their eyes detect 83 84 edges, and how do their brains recognize shapes and place them in categories (Stoddard 2012)? At the 85 same time, there is an urgent need to pair such organismal research with more detailed knowledge of 86 how camouflage is perceived in the wild. For example, to understand how prey will be perceived, we 87 need to know the distance at which predators search for their prey to know which perceptual processes 88 are most likely to function when camouflaged prey enter their field of vision. Birds such as 89 woodcreepers (Dendrocolaptinae) that move along branches searching for prey may have a very 90 different perceptual experience from birds that search for prey from perches, like some flycatchers 91 (Tyrannidae). Approaches used in studying the relationship between warning coloration (aposematism) 92 and camouflage may be helpful, as these two antipredator strategies can also co-exist within a single 93 organism to protect it from predators that see it at different distances (Tullberg et al. 2005, Barnett and 94 Cuthill 2014). Observing the behavior of either human or wild predators upon encountering 95 camouflaged prey would be helpful in better understanding the function of multiple camouflage 96 strategies in nature.

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98 The camouflage exhibited by our particular katydid seems quite specific. What are the evolutionary 99 consequences of this sort of specialization? Can camouflage specialization increase speciation rates? 100 Selection to maintain effective disguises can result in reproductive isolation between populations 101 specialized for different microhabitats (Nosil et al. 2002), which makes it reasonable to speculate that 102 camouflage may increasing diversification rates. Does camouflage also come at the price of elevated 103 extinction risk? This possibility must be considered because although antipredator defenses are often 104 thought of as leading to "escape-and-radiate" dynamics where diversification follows innovation that 105 allows expansion into new niches (Schluter 2000), recent work has shown unexpected extinction risk 106 associated with some antipredator adaptations (Arbuckle and Speed 2015). Highly specialized 107 camouflage would seem an ambiguous case because of its obvious benefits, but also potential costs 108 such as inhabiting habitats with low carrying capacities (Arbuckle and Speed 2015), vulnerability to 109 predators at high densities if predators form search images (Endler 1988), or metabolic trade-offs with 110 thermoregulation (Carrascal et al. 2001). Groups such as the Tettigoniidae provide a tantalizing opportunity for their exceptional diversity, wide geographic distribution, and striking array of disguises 111 112 suggest that many independent evolutionary experiments have already taken place. 113 114 Future evolutionary experimentation may be just around the corner as local microclimates shift in

115 response to anthropogenic activities, which can be challenging for invertebrate camouflage specialists 116 (Gröning et al. 2007). Although the Santuario de Fauna y Flora Otún Quimbaya will benefit from 117 government efforts to restore the local watershed and forest, the impact of global climate change on 118 habitat suitability for species that rely on camouflage remains an unknown. For better or worse, 119 different climate projections produce conflicting estimates for whether precipitation in the region will 120 increase or decrease. They agree, however, that relative humidity will decrease (Collins et al. 2013), 121 which may present a test of adaptability for taxa that hide amidst the moss. 122 123 References

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- Arbuckle, K., and M. P. Speed. 2015. Antipredator defenses predict diversification rates. Proceedings of
 the National Academy of Sciences 2015:1–6.
- Barnett, J. B., and I. C. Cuthill. 2014. Distance-dependent defensive coloration. Current Biology
 24:R1157–R1158.
- Bidau, C. J. 2014. Patterns in Orthoptera biodiversity. I. Adaptations in ecological and evolutionary
 contexts. Journal of Insect Biodiversity 2:1–39.
- Braun, H. 2011. The little lichen dragon An extraordinary katydid from the Ecuadorian Andes
 (Orthoptera, Tettigoniidae, Phaneropterinae, Dysoniini). Zootaxa 39:33–39.
- Carrascal, L. M., J. A. Díaz, D. L. Huertas, and I. Mozetich. 2001. Behavioral Thermoregulation by
 Treecreepers : Trade-Off between Saving Energy and Reducing Crypsis. Ecology 82:1642–1654.
- Collins, M., R. Knutt, J. Arblaster, J.-L. Dufresne, T. Fichefet, P. Friedlingstein, X. Gao, W. J. Gutowski, T.
 Johns, G. Krinner, M. Shongwe, C. Tebaldi, A. J. Weaver, and M. Wehner. 2013. Long-term climate
 change: projections, commitments, and irreversibility. Page *in* T. F. Stocker, D. Qin, G.-K. Plattner,
 M. Tignor, S. K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bev, and P. M. Midgley, editors. Climate
 change 2013: the physical science basis. Contribution of Working Group I to the Fifth Assessment
 Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge,
 UK.
- Cuthill, I. C., M. Stevens, J. Sheppard, T. Maddocks, C. A. Párraga, and T. S. Troscianko. 2005. Disruptive
 coloration and background pattern matching. Nature 434:72–74.
- Endler, J. A. 1988. Frequency-dependent predation, crypsis, and aposematic coloration. Philosophical
 Transactions of the Royal Society of London 319:505–523.
- Gröning, J., S. Krause, and A. Hochkirch. 2007. Habitat preferences of an endangered insect species,
 Cepero's ground-hopper (Tetrix ceperoi). Ecological Research 22:767–773.
- Gutiérrez, Y., and T. Bacca. 2014. Phasmatodea (insecta) de la Reserva Natural Río Ñambí, Nariño,
 Colombia 18:210–221.
- Kang, C., M. Stevens, J. Y. Moon, S. I. Lee, and P. G. Jablonski. 2015. Camouflage through behavior in
 moths: The role of background matching and disruptive coloration. Behavioral Ecology 26:45–54.
- Lovell, P. G., G. D. Ruxton, K. V. Langridge, and K. A. Spencer. 2013. Egg-laying substrate selection for
 optimal camouflage by quail. Current Biology 23:260–264.
- Mugleston, J. D., H. Song, and M. F. Whiting. 2013. A century of paraphyly: A molecular phylogeny of
 katydids (Orthoptera: Tettigoniidae) supports multiple origins of leaf-like wings. Molecular
 Phylogenetics and Evolution 69:1120–1134.
- Nickle, D. A., and J. L. Castner. 1995. Strategies Utilized by Katydids (Orthoptera: Tettigoniidae) against
 Diurnal Predators in Rainforests of Northeastern Peru. Journal of Orthoptera Research:75–88.
- Nosil, P., B. J. Crespi, and C. P. Sandoval. 2002. Host-plant adaptation drives the parallel evolution of
 reproductive isolation. Nature 417:440–443.
- 161 Ruxton, G. D., T. N. Sherratt, and M. P. Speed. 2004. Avoiding attack. Oxford University Press, New York.
- 162 Schluter, D. 2000. The Ecology of Adaptive Radiation. Oxford University Press, Oxford.
- Skelhorn, J., and G. D. Ruxton. 2010. Predators are less likely to misclassify masquerading prey when
 their models are present. Biology letters 6:597–599.
- Stevens, M., and S. Merilaita. 2009. Animal camouflage: current issues and new perspectives.
 Philosophical Transactions of the Royal Society. Series B, Biological sciences 364:423–427.
- Stevens, M., and S. Merilaita. 2011. Animal camouflage: Function and mechanisms. Animal camouflage:
 mechanisms and function:1–16.
- Stoddard, M. C. 2012. Mimicry and masquerade from the avian visual perspective. Current Zoology
 58:630–648.
- 171 Stoddard, M. C., K. Kupán, H. N. Eyster, W. Rojas-Abreu, M. Cruz-López, M. A. Serrano-Meneses, and C.
- 172 Küpper. 2016. Camouflage and Clutch Survival in Plovers and Terns. Scientific Reports 6:32059.

173 174	Svenson, G. J., N. B. Hardy, H. M. Cahill Wightman, and F. Wieland. 2015. Of flowers and twigs: Phylogenetic revision of the plant-mimicking praying mantises (Mantodea: Empusidae and
175	Tullborg B S S Marilaita and C Wiklund 2005 Anosomatism and crupsis combined as a result of
177	distance dependence: functional versatility of the colour nattern in the swallowtail butterfly larva
178	Proceedings. Biological sciences / The Royal Society 272:1315–1321.
179	Webster, R. J., C. Hassall, C. M. Herdman, JG. J. Godin, and T. N. Sherratt. 2013. Disruptive camouflage
180	impairs object recognition. Biology Letters 9:20130501.
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186	Captions: Now you see it, now you don't. (a) Picture of a moss katydid in its overall context. Tip: the
187	animal is located in the center bottom of the picture, facing left. (b) Close up of the same individual that
188	shows coloring and anatomical features that resemble the moss on which it is sitting. The long antennae
189	are reddish brown with specs of green and are almost twice as long as the animal (see Fig 1a again).
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