

Drug Testing and Analysis



Metabolism of the tryptamine-derived new psychoactive substances 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT and their detectability in urine studied by GC-MS, LC-MSn, and LC-HR-MS/MS

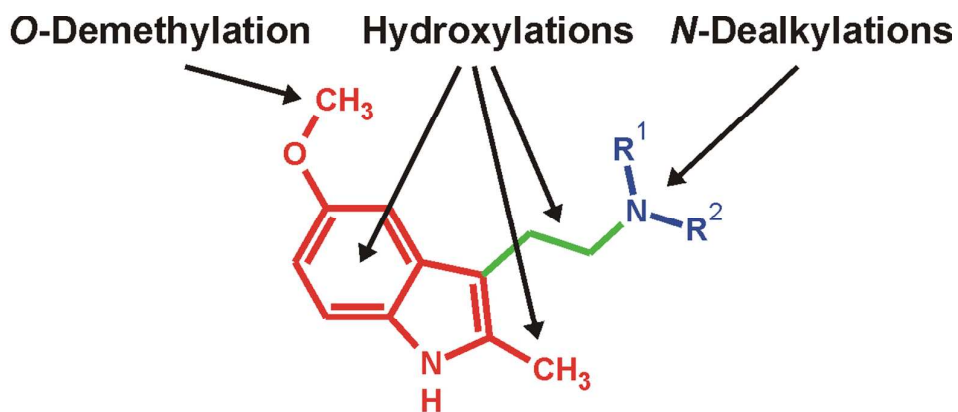
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Keywords:	new psychoactive substances, tryptamines, metabolism, cytochrome-P450, LC-MSn, LC-HR-MS/MS
Abstract:	Many N,N-dialkylated tryptamines show psychoactive properties and were encountered as new psychoactive substances. The aims of the presented work were to study the phase I and II metabolism and the detectability in standard urine screening approaches (SUSA) of 5-methoxy-2-methyl-N,N-diallyltryptamine (5-MeO-2-Me-DALT), 5-methoxy-2-methyl-N-allyl-N-cyclohexyltryptamine (5-MeO-2-Me-ALCHT), and 5-methoxy-2-methyl-N,N-diisopropyltryptamine (5-MeO-2-Me-DIPT) using GC-MS, LC-MSn, and LC-HR-MS/MS. For metabolism studies, urine was collected over a 24-h period after administration of the compounds to male Wistar rats at 20 mg/kg body weight (BW). Phase I and II metabolites were identified after urine precipitation with acetonitrile by LC-HR-MS/MS. 5-MeO-2-Me-DALT (24 phase I and 12 phase II metabolites), 5-MeO-2-Me-ALCHT (24 phase I and 14 phase II metabolites), and 5-MeO-2-Me-DIPT (20 phase I and 11 phase II metabolites) were mainly metabolized by O-demethylation, hydroxylation, N-dealkylation, and combinations of them as well as by glucuronidation and sulfation of phase I metabolites. Incubations with mixtures of pooled human liver microsomes and cytosols (pHLM and pHLC) confirmed that the main metabolic reactions in humans and rats might be identical. Furthermore, initial CYP activity screenings revealed that CYP1A2, CYP2C19, CYP2D6, and CYP3A4 were involved in hydroxylation, CYP2C19

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	and CYP2D6 in O-demethylation, and CYP2C19, CYP2D6, and CYP3A4 in N-dealkylation. For SUSAs, GC-MS, LC-MSn, and LC-HR-MS/MS were applied to rat urine samples after 1 or 0.1 mg/kg BW doses, respectively. In contrast to the GC-MS SUSAs, both LC-MS SUSAs were able to detect an intake of 5-MeO-2-Me-ALCHT and 5-MeO-2-Me-DIPT via their metabolites following 1 mg/kg BW administrations and 5-MeO-2-Me-DALT following 0.1 mg/kg BW dosage.

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2 **Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT and their**
3 **detectability in urine studied by GC-MS, LC-MSⁿ, and LC-HR-MS/MS**
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Abstract

Many *N,N*-dialkylated tryptamines show psychoactive properties and were encountered as new psychoactive substances. The aims of the presented work were to study the phase I and II metabolism and the detectability in standard urine screening approaches (SUSA) of 5-methoxy-2-methyl-*N,N*-diallyltryptamine (5-MeO-2-Me-DALT), 5-methoxy-2-methyl-*N*-allyl-*N*-cyclohexyltryptamine (5-MeO-2-Me-ALCHT), and 5-methoxy-2-methyl-*N,N*-diisopropyltryptamine (5-MeO-2-Me-DIPT) using GC-MS, LC-MSⁿ, and LC-HR-MS/MS. For metabolism studies, urine was collected over a 24-h period after administration of the compounds to male Wistar rats at 20 mg/kg body weight (BW). Phase I and II metabolites were identified after urine precipitation with acetonitrile by LC-HR-MS/MS. 5-MeO-2-Me-DALT (24 phase I and 12 phase II metabolites), 5-MeO-2-Me-ALCHT (24 phase I and 14 phase II metabolites), and 5-MeO-2-Me-DIPT (20 phase I and 11 phase II metabolites) were mainly metabolized by *O*-demethylation, hydroxylation, *N*-dealkylation, and combinations of them as well as by glucuronidation and sulfation of phase I metabolites. Incubations with mixtures of pooled human liver microsomes and cytosols (pHLM and pHLC) confirmed that the main metabolic reactions in humans and rats might be identical. Furthermore, initial CYP activity screenings revealed that CYP1A2, CYP2C19, CYP2D6, and CYP3A4 were involved in hydroxylation, CYP2C19 and CYP2D6 in *O*-demethylation, and CYP2C19, CYP2D6, and CYP3A4 in *N*-dealkylation. For SUSAs, GC-MS, LC-MSⁿ, and LC-HR-MS/MS were applied to rat urine samples after 1 or 0.1 mg/kg BW doses, respectively. In contrast to the GC-MS SUSA, both LC-MS SUSAs were able to detect an intake of 5-MeO-2-Me-ALCHT and 5-MeO-2-Me-DIPT via their metabolites following 1 mg/kg BW administrations and 5-MeO-2-Me-DALT following 0.1 mg/kg BW dosage.

Keywords: new psychoactive substances; tryptamines; metabolism; cytochrome-P450; LC-MSⁿ; LC-HR-MS/MS

Introduction

New psychoactive substances (NPS) have become increasingly popular among drug users around the world.^[1] As many of them are not regulated, these substances are marketed to avoid criminal prosecution for production, trading, drug possession, or consumption. NPS often mimic the effects of their traditional counterparts.^[1] Several hundred substances are classified as NPS where research on the potential harm for consumers is limited. Synthetic cannabinoids, phenylethylamines, synthetic cathinones, and tryptamines were the most prevalent representatives of NPS during the last few years.^[1] The biological effects associated with many psychoactive tryptamine derivatives are at least in part mediated by agonism at 5-HT_{1A} and 5-HT_{2A} receptors leading to hallucinogenic and stimulating effects.^[2-7] The synthesis and mode of action for many synthetic tryptamines were described by Shulgin and Shulgin.^[8] Examples of tryptamines described by them and subsequently appeared in case reports include 5-MeO-MIPT (5-methoxy-*N*-methyl-*N*-isopropyltryptamine)^[9] and 5-MeO-DIPT (5-methoxy-*N,N*-diisopropyltryptamine).^[10-14] Other tryptamines, such as 5-MeO-DALT (5-methoxy-*N,N*-diallyltryptamine),^[15,16] originally not described by Shulgin and Shulgin in 1997,^[8] have also appeared on the market and were linked to intoxication cases. Shulgin and Shulgin^[8] have also described the effects of tryptamines substituted on the 2-position of the indole ring such as 5-MeO-TMT (5-methoxy-2-*N,N*-trimethyltryptamine). From this perspective, it seems conceivable that other 2-methylated tryptamines might also appear on the drug market. In cases of intoxication with such substances, it is necessary to provide analytical detection methods, such as standard urine screening approaches (SUSAs)^[17] in order to facilitate adequate support of the therapeutic strategy used for clinical treatment. As shown by Michely *et al.*,^[18] tryptamines were mainly excreted as metabolites thus the targets for urine screening have to be elucidated by metabolism studies to include those in SUSAs. This work here is focused on the metabolism and the detectability of the three NPS 5-methoxy-2-methyl-*N,N*-diallyltryptamine (5-MeO-2-Me-DALT), 5-methoxy-2-methyl-*N*-allyl-*N*-cyclohexyltryptamine (5-MeO-2-Me-ALCHT), and 5-methoxy-2-methyl-*N,N*-diisopropyltryptamine (5-MeO-2-Me-DIPT), respectively.

Experimental

Chemicals and reagents

5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT were available from previous studies and synthesized by Brandt *et al.*^[19-21] Isocitrate, isocitrate dehydrogenase, and 3'-phosphoadenosine-5'-phosphosulfate (PAPS) were received from Sigma-Aldrich (Taufkirchen, Germany), NADP⁺ from Biomol (Hamburg, Germany), and methanol (LC-MS grade), ammonium formate (analytical grade), formic acid (LC-MS grade), and acetonitrile (LC-MS grade) and all other chemicals or reagents (analytical grade) from VWR (Darmstadt, Germany). The baculovirus-infected insect cell microsomes (supersomes) containing human cDNA-expressed cytochrome P450 (CYP) enzymes CYP1A2, CYP2A6, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP3A4 (1 nmol/mL, each), or CYP2E1, CYP3A5 (2 nmol/mL, each), pooled human liver microsomes (pHLM, 20 mg microsomal protein/mL, 330 pmol total CYP/mg protein), pooled human liver cytosols (pHLC, 20 mg cytosolic protein/mL), UDP-glucuronyltransferase (UGT) reaction mix solution A (25 mM UDP-glucuronic acid), and UGT reaction mix solution B (250 mM Tris-HCl, 40 mM MgCl₂, and 0.125 mg/mL alamethicin) were obtained from Corning (Amsterdam, The Netherlands). After delivery, the enzyme preparations were thawed at 37°C, aliquoted, snap-frozen in liquid nitrogen, and stored at -80°C until use.

Urine samples

The compounds were orally administered in an aqueous suspension by gastric intubation to male Wistar rats (Charles Rivers, Sulzfeld) for toxicological diagnostic reasons.^[18] Doses were 20 mg/kg body weight (BW) for metabolism studies (identification of metabolites), 1 mg/kg BW for toxicological analyses, and 0.1 mg/kg BW for estimation of the detectability of very low dosed

1 applications, respectively. After administration, the rats were housed in metabolism cages for 24 h.
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3 Water was provided *ad libidum* and urine collected separately from feces. Before administration,
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5 blank urine samples were collected to evaluate the absence of interfering compounds. Urine
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7 samples were extracted, analyzed, and stored at -20°C immediately after collection.
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10 11 12 **Sample preparation for identification of phase I and II metabolites by LC-HR-MS/MS**

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15 As described elsewhere,^[22] 100 µL urine were mixed with 500 µL acetonitrile, vortexed for 2 min
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17 and centrifuged for further 2 min at 10,000 g. Five hundred µL of the supernatant were transferred
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19 into a glass vial and evaporated at 70°C under a stream of nitrogen. The residue was dissolved in 50
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21 µL of a mixture (1:1; v/v) of eluents A (2 mM aqueous ammonium formate plus 0.1% formic acid,
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23 pH 3) and B (2 mM aqueous ammonium formate with acetonitrile:methanol, 50:50, v/v; 1% water
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25 plus 0.1% formic acid) and a 1-µL aliquot was injected onto the LC-HR-MS/MS system.
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31 32 **Conditions for incubations for initial CYP activity screening studies**

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35 According to Richter *et al.*,^[23] final concentrations of 25 µM substrate were used. The microsomal
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37 incubations were performed for 30 min at 37°C, each containing one of the following enzymes at
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39 50 pmol/mL: CYP1A2, CYP2A6, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP2E1,
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41 CYP3A4, and CYP3A5. Further components in the final 50 µL incubation volume were phosphate
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43 buffer (90 mM, pH 7.4), 5 mM Mg²⁺, 5 mM isocitrate, 1.2 mM NADP⁺, 1.6 U/mL isocitrate
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45 dehydrogenase, and 200 U/mL superoxide dismutase. According to the Gentest manual, phosphate
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47 buffer was replaced with 45 and 90 mM Tris buffer for incubation of CYP2A6 and CYP2C19.
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49 Reactions were initiated by addition of the respective test substrate and stopped by addition of 50
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51 µL ice-cold acetonitrile. The solution was then centrifuged for 5 min at 10,000 g, 70 µL of the
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53 supernatant were transferred to an autosampler vial, and 1 µL was injected onto the LC-HR-MS/MS
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1 system. Blank and negative control samples were also prepared accordingly, whereby substrate or
2 enzymes were replaced with buffer, respectively.
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8 **Conditions for incubations with pHLM + pHLC**

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11 As described by Richter *et al.*,^[23] the *in vitro* drug metabolism studies using a mixture of pHLM
12 and pHLC were done with final protein concentrations of 1 mg/mL each. After 10 min of
13 preincubation at 37°C with 20 µL UGT reaction mix solution B, pHLM, pHLC, 90 mM phosphate
14 buffer (pH 7.4), 2.5 mM Mg²⁺, 2.5 mM isocitrate, 0.6 mM NADP⁺, 0.8 U/mL isocitrate
15 dehydrogenase, 100 U/mL superoxide dismutase, 10 µL UGT reaction mix A, and 40 µM aqueous
16 PAPS were added. All given concentrations are final concentrations. The reaction was initiated by
17 adding substrate at 25 µM (100 µL final volume) and stopped after 30 min incubation time with 50
18 µL ice-cold acetonitrile. The samples were centrifuged for 2 min at 10,000 g, 100 µL of the
19 supernatant transferred into an autosampler vial, and 1 µL injected onto the LC-HR-MS/MS
20 system. A blank and a negative control sample were also prepared, whereby substrate or enzyme
21 was replaced with buffer, respectively.
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39 **LC-HR-MS/MS apparatus for identification of phase I and II metabolites in urine and** 40 **incubations**

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44 The prepared samples were analyzed using a ThermoFisher Scientific (TF, Dreieich, Germany)
45 Dionex UltiMate 3000 RS LC system, consisting of a degasser, a quaternary pump, and an UltiMate
46 autosampler and coupled to a TF Q-Exactive Plus system equipped with heated electrospray
47 ionization (HESI)-II source. The instrument was used in positive ionization mode. According to the
48 manufacturer's manual, mass calibration was done before analysis using external mass calibration.
49 LC and MS settings were according to published procedures.^[23,24] The gradient elution was
50 performed on a TF Accucore PhenylHexyl column (100 x 2.1 mm, 2.6 µm). Chromatographic
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1 separation was performed at 40°C supplied by an analytical column heater (Dionex UltiMate 3000).
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3 The gradient was programmed as follows: 0-1 min 99% A, 1-10 min to 1% A, 10-11.5 min hold 1%
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5 A, and 11.5-13.5 min hold 99% A. The flow rate was set to 500 µL/min for 10 min and 800 µL/min
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7 from 10 to 13.5 min. The HESI-II source conditions were as follows: sheath gas, 60 arbitrary units
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9 (AU); auxiliary gas, 10 AU; spray voltage, 3.00 kV; heater temperature, 320°C; ion transfer
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11 capillary temperature, 320°C; and S-lens RF level, 60.0. Mass spectrometry analyses were
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13 performed using full-scan (FS) data and a subsequent data dependent acquisition (DDA) mode with
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15 an inclusion list of the *m/z* values of 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, 5-MeO-2-Me-
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17 DIPT and their potential metabolites. Additional DDA without inclusion list was performed to
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19 detect unexpected metabolites. The settings for FS acquisition were as follows: resolution, 35,000;
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21 microscans, 1; automatic gain control (AGC) target, 1e6; maximum injection time (IT), 120 ms;
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23 and scan range, *m/z* 130-1000; loop count, 5; dynamic exclusion, 1 s. The settings for DDA mode
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25 were as follows: resolution, 17,500; microscans, 1; AGC target, 2e5; maximum IT, 250 ms;
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27 isolation window, 1.0 *m/z*, HCD with stepped normalized collision energy (NCE), 17.5, 35, and
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29 52.5%; spectrum data type, profile; underfill ratio, 0.5% (corresponding to a signal with an
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31 abundance of 4e3); option exclude isotopes, enabled. Software for data evaluation was Thermo
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33 Fisher Xcalibur 2.2 SP1.48.
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41 **Standard urine screening approaches (SUSA)**

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45 The SUSAs were performed as described previously: GC-MS SUSA,^[17,25] LC-MSⁿ SUSA,^[26,27] and
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47 LC-HR-MS/MS SUSA.^[26,28]
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55 **Results and discussion**

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59 **Identification of phase I and II metabolites via LC-HR-MS/MS**

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3 Mass spectral interpretations were based on general fragmentation rules^[29] and by comparison with
4 the spectra of the parent compounds. All phase I and II metabolites tentatively identified based on
5 their MS² spectra are depicted in Figures S1-S6 in Supporting Information (SI). Additionally, the
6 spectra contain the supposed chemical structures, the accurate measured masses, the molecular
7 formulae, and the corresponding mass error values in parts per million (ppm).
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15 16 17 **Proposed fragmentation patterns for identification of phase I metabolites via LC-HR-MS/MS**

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20 In general, all metabolites were identified based on their m/z values of the protonated molecule
21 (PM), the calculated molecular formulae, and the fragmentation patterns compared to those of the
22 parent compounds. They were sorted by PM and with abbreviations for the corresponding
23 compound (5-MeO-2-Me-DALT, D; 5-MeO-2-Me-ALCHT, A; and 5-MeO-2-Me-DIPT, P). Due to
24 the large number of metabolites detected in this study, only representative examples have been
25 chosen to illustrate the fragmentation patterns. Following the suggestion described by Michely *et*
26 *al.*,^[18] the chemical structures were divided into three sections to allow a convenient description of
27 fragmentation patterns and positions of metabolic reactions. Section 1 represents the aromatic ring
28 system, section 2 the ethyl spacer, and section 3 the variable amine part of the molecule. As
29 expected, the fragment ions formed by section 1 and 2 were the same for 5-MeO-2-Me-DALT, 5-
30 MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT. All masses in the following sections are the
31 calculated (exact) masses.
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50 **5-MeO-2-Me-DALT**

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53 As shown in Figure S1 (see SI), 24 phase I metabolites of 5-MeO-2-Me-DALT could be identified.
54 5-MeO-2-Me-DALT (D1 in Figure S1; PM at m/z 285.1961, C₁₈H₂₅ON₂⁺) showed a fragmentation
55 comparable to DALT and 5-MeO-DALT^[18] and for most of the detected metabolites. The fragment
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ions of m/z 188.1070 ($C_{12}H_{14}ON^+$) and 110.0964 ($C_7H_{12}N^+$) represented typical α -cleavage of the tryptamine structure forming the characteristic fragment ions of sections 1 plus 2 (m/z 188.1070, $C_{12}H_{14}ON^+$) and section 3 (m/z 110.0964, $C_7H_{12}N^+$). The fragment ion of m/z 173.0835 ($C_{11}H_{11}ON^{\bullet+}$) resulted by a loss of a methyl radical of the methoxy group. For metabolite identification, the representative MS² fragment ions for section 3 were used in most cases. Unmodified section 3 led to a fragment ion of m/z 110.0964 ($C_7H_{12}N^+$), which suggested that the expected modification took place in sections 1 or 2. Modified section 3 led to fragment ions of m/z 70.0651 ($C_4H_8N^+$, *N*-deallylated) as shown e.g. in the MS² spectrum of *N*-deallyl-5-MeO-2-Me-DALT (D5, PM at m/z 245.1648, $C_{15}H_{21}ON_2^+$). As expected, the fragment ions of unchanged sections 1 and 2 (m/z 173.0835 and 188.1070) were also present. After *O*-demethylation, corresponding fragment ions of m/z 159.0679 ($C_{10}H_9ON^+$) and 174.0913 ($C_{11}H_{12}ON^+$), shown e.g. in the MS² spectrum of *O*-demethyl-5-MeO-2-Me-DALT (D11, PM at m/z 271.1805, $C_{17}H_{23}ON_2^+$), were detected, which was consistent with the detection of m/z 110.0964 ($C_7H_{12}N^+$). Furthermore, five isomers with the PM of m/z 301.1911, $C_{18}H_{25}O_2N_2^+$ could be observed and predicted to represent hydroxylated metabolites (D16-D20). Isomers 1-4 (D16-D19) showed fragment ions of m/z 110.0964 ($C_7H_{12}N^+$) indicating that the hydroxylation took place at sections 1 or 2. In the MS² spectrum of hydroxy-alkyl isomer 1 (D16), no specific shifts could be detected for the fragment ions (e.g. 188.1070 to 204.1019, +15.9949, +O) associated with sections 1 and 2 or any loss of water as in the MS² spectrum of hydroxy-alkyl isomer 2, which could indicate hydroxylation probably at the 2-methyl group of section 1. The fragment ions of m/z 176.0706 ($C_{10}H_{10}O_2N^+$) and 178.0863 ($C_{10}H_{12}O_2N^+$) represented a hydroxylated methyl group in section 1 with the positive charge at the carbon or at the nitrogen atom, respectively. In contrast, the MS² spectra of the hydroxy-aryl isomers 1 and 2 (D17 and D18) showed typical shifts of +15.9949 u (+O) of the fragment ions of sections 1 and 2, thus, resulting in the fragment ions of m/z 189.0784 ($C_{11}H_{11}O_2N^+$) and 204.1019 ($C_{12}H_{14}O_2N^+$). Furthermore, there were no fragment ions associated with a loss of water in these two spectra and thus the two isomers were predicted to be aryl hydroxylated metabolites (D17 and D18). In contrast, the MS² spectrum of hydroxy-alkyl isomer 2

(D19) showed such a loss of water (-18.0106 u, H₂O), typically observed with alkyl hydroxy groups, and therefore resulting in m/z 186.0913 (C₁₂H₁₂ON⁺), which suggested that hydroxylation occurred in the ethylamine side chain in section 2. The MS² spectrum of isomer 5 (D20) showed neither presence of the fragment ion of unchanged section 3 (m/z 110.0964, (C₇H₁₂N⁺), nor any typical shifts of this fragment ion. However, the fragment ions of unchanged sections 1 and 2 were detected and therefore the metabolite was inconsistent with a hydroxylated species. The presence of an *N*-oxide metabolite seemed likely as D20 had a longer retention time (5.0 min) than the parent compound (4.8 min) on the reversed phase column. This was observed for all *N*-oxides for which reference standards were available.^[30]

5-MeO-2-Me-ALCHT and 5-MeO-2-Me-DIPT

According to Figures S3 and S5 (see SI), 24 phase I metabolites could be identified for 5-MeO-2-Me-ALCHT and 20 phase I metabolites for 5-MeO-2-Me-DIPT, respectively. As already mentioned above, the fragmentation patterns of the two parent compounds were comparable to those of 5-MeO-2-Me-DALT. The α -cleavages of 5-MeO-2-Me-ALCHT (A1 in Figure S3; PM at m/z 327.2431, C₂₁H₃₁ON₂⁺) and 5-MeO-2-Me-DIPT (P1 in Figure S5; PM at m/z 289.2274, C₁₈H₂₉ON₂⁺) formed the fragment ions of m/z 188.1070 (C₁₂H₁₄ON⁺) representing sections 1 and 2 and m/z 152.1434 (C₁₀H₁₈N⁺) and m/z 114.1277 (C₇H₁₆N⁺), respectively, representing the iminium ion of section 3. In contrast to 5-MeO-2-Me-DALT, for both compounds a further breakdown of section 3 could be detected, forming the fragment ions of m/z 70.0651 (C₄H₈N⁺) after loss of the cyclohexyl group and m/z 72.0807 (C₄H₁₀N⁺) after loss of one isopropyl group. The metabolite formed after *N,N*-bis-dealkylation (*N,N*-bis-deallyl, D2; *N*-deallyl-*N*-decyclohexyl, A2; and *N,N*-bis-deisopropyl metabolite, P2) and the further deaminated and oxidized carboxy metabolite (D3, A3, and P3) were identical for all three compounds, representing the conserved 2-methyltryptamine structure. Furthermore, all metabolites formed after *N*-deallylation of 5-MeO-2-Me-DALT (D4-D10) and *N*-decyclohexylation of 5-MeO-2-Me-ALCHT (A4-A9) were identical for both

1 compounds, except for D7 which was only found after *N*-deallylation and oxidation of 5-MeO-2-
2 Me-DALT.
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8 **Proposed fragmentation patterns for identification of phase II metabolites via LC-HR-**
9 **MS/MS**

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14 ***5-MeO-2-Me-DALT***

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19 As shown in Figure S2 (see SI), all 12 identified phase II metabolites were conjugated at the indole
20 part. The conjugates showed the same fragment ions as the underlying phase I metabolites after a
21 characteristic elimination of glucuronic acid (-176.0321 u) of the glucuronides and of sulfuric acid
22 (-79.9568 u) of the sulfates. The fragment ions of *m/z* 240.0325 (C₁₀H₁₀O₄NS⁺), 254.0482
23 (C₁₁H₁₂O₄NS⁺), and 284.0587 (C₁₂H₁₄O₅NS⁺) formed after sulfation and the fragment ions of *m/z*
24 336.1078 (C₁₆H₁₈O₇N⁺), 350.1234 (C₁₇H₂₀O₇N⁺), 366.1183 (C₁₇H₂₀O₈N⁺), and 380.1340
25 (C₁₈H₂₂O₈N⁺) after glucuronidation at the ethylindole part (section 1 and 2) after *O*-demethylation
26 or aryl-hydroxylation, respectively.
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39 ***5-MeO-2-Me-ALCHT and 5-MeO-2-Me-DIPT***

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43 As shown in Figures S4 and S6 (see SI), 14 phase II metabolites of 5-MeO-2-Me-ALCHT and 11 of
44 5-MeO-2-Me-DIPT could be identified. As already described for 5-MeO-2-Me-DALT, all
45 glucuronides and sulfates eliminated glucuronic acid (-176.0321 u) or sulfuric acid (-79.9568 u). In
46 addition, conjugation always took place at the ethylindole part after *O*-demethylation or aryl-
47 hydroxylation, forming the same fragment ions of the conjugated partial structure. As both
48 compounds have some common phase I metabolites following *N*-dealkylation, the resulting phase II
49 metabolites after further conjugation were consequently also identical (D6S, D8S, D4G, D6G, and
50 D8G₂ for 5-MeO-2-Me-DALT; A6S, A9S, A4G, A6G, and A9G for 5-MeO-2-Me-ALCHT).
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Proposed metabolic pathways

Based on the identified metabolites, the metabolic pathways for 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT were proposed and depicted in Figures 1-3. The numbering is given in Figures S1, S3, and S5 (see SI). As expected, the main metabolic reactions observed for all three test drugs were comparable.

5-MeO-2-Me-DALT

Four different hydroxy isomers could be detected. Two isomers of hydroxy-aryl (D17 and D18) were formed by hydroxylation at the indole part. Hydroxy-alkyl isomer 1 (D16) led to further oxidation and/or hydroxylation steps in section 1 (D15, D21, D22, and D23). Hydroxylation at section 2, forming hydroxy-alkyl isomer 2 (D19), was probably the preliminary step for the formation of two *N*-deallyl-oxo isomers 1 and 2 (D7 and D8). Moreover, *N*-oxidation (D20) alone and in combination with aryl hydroxylation (D24) was identified. The intermediate mono-hydroxy metabolite that led to the bis-hydroxylated product (both section 3; D25) was not detected. Initial *O*-demethylation (D11) and further oxidation led to *O*-demethyl-hydroxy-alkyl isomer 1 and 2 (D12 and D13) and *O*-demethyl-*N*-oxide (D14). Combination of *O*-demethylation and *N*-deallylation led to *O*-demethyl-*N*-deallyl metabolite (D4), following hydroxylation to *O*-demethyl-*N*-deallyl-hydroxy-alkyl (D6). After *N*-deallylation and further hydroxylation, two *N*-deallyl-hydroxy metabolites (D9, hydroxy-aryl and D10, hydroxy-alkyl) could be found. Finally, when D5 was further deallylated, a *N,N*-bis-deallyl metabolite (D2) was formed, that might have been deaminated and carboxylated to give *N,N*-bis-deallyl-deamino carboxylic acid (D3). Glucuronidation and sulfation could be observed for *O*-demethyl-*N*-deallyl (D4), *O*-demethyl-*N*-deallyl-hydroxy-alkyl (D6), *N*-deallyl-hydroxy-aryl (D9), *O*-demethyl (D11), and hydroxy-aryl isomer 2 (D18)

1 metabolites. For *O*-demethyl-hydroxy-alkyl isomer 2 (D13) and hydroxy-aryl isomer 1 (D17), only
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3 the glucuronidation species were detected.
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5-*MeO*-2-*Me*-ALCHT

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11 The metabolic pathways were similar to those observed for 5-*MeO*-2-*Me*-DALT including *N*-
12 oxidation (A23), aliphatic (A20 and A21) and aromatic (A22) hydroxylation. However, aliphatic
13 hydroxylations did not affect section 2 but the 2-methyl substituent in section 1 and the cyclohexyl
14 ring in section 3 instead. Combination of both hydroxylation types (aromatic and aliphatic) showed
15 the bis-hydroxy metabolite (A25). An intermediate biotransformation step (aliphatic hydroxylation
16 at the 2-methyl substituent) was originally considered, but this could not be identified, possibly due
17 to fast oxidation to the carboxylic acid (A24). In contrast to 5-*MeO*-2-*Me*-DALT and 5-*MeO*-2-*Me*-
18 DIPT reflecting a symmetrical *N,N*-dialkylation pattern, 5-*MeO*-2-*Me*-ALCHT represents an
19 asymmetrically substituted amine. In consequence, besides the comparable *O*-demethylation (A17)
20 with further cyclohexyl hydroxylation (A18 and A19) and *N*-deallylation (A11), *N*-
21 decyclohexylation (A5) was found to be an important metabolic step as well. *N*-decyclohexylation
22 of 5-*MeO*-2-*Me*-ALCHT produced the same metabolite that resulted from *N*-deallylation of 5-
23 *MeO*-2-*Me*-DALT. Similarly, additional transformations included hydroxylations (A8 and A9),
24 oxidation to amide structure (A7), *O*-demethylation (A4), combinations of both (A6), and further *N*-
25 deallylation (A2), followed by oxidation to the corresponding carboxylic acid (A3). Retention times
26 and MS² spectra of these metabolites were the same for 5-*MeO*-2-*Me*-DALT and -ALCHT.
27 However, the alpha-position in metabolites A7 and D8 is not the probable position due to fact that
28 the intermediate should be an unstable hemiaminal. The *N*-deallylated metabolite (A11) could be
29 further metabolized by hydroxylation (A15 and A16), oxidation to an aldehyde (D14), *O*-
30 demethylation (A10), and combination of *O*-demethylation with hydroxylation (A12 and A13).
31 Glucuronidation and sulfation were found for *O*-demethyl-*N*-decyclohexyl-hydroxy-alkyl (A6), *N*-
32 decyclohexyl (A8), *O*-demethyl-*N*-deallyl (A10), *O*-demethyl-*N*-deallyl-hydroxy-cyclohexyl (A12),
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1 *O*-demethyl (A17), and *O*-demethyl-hydroxy-cyclohexyl (A18) metabolites, whereas *O*-demethyl-
2 *N*-decyclohexyl (A4) was only found as glucuronide. Two glucuronide isomers (A8G1 and A8G2)
3 were detected for the *N*-decyclohexyl-hydroxy-aryl metabolite (A8).
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10 **5-MeO-2-Me-DIPT**

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14 As expected, most 5-MeO-2-Me-DIPT metabolites were found to be equivalent to those described
15 above, such as aliphatic (P15 and P18) and aromatic (P16 and P17) hydroxylations, combination of
16 aliphatic and aromatic hydroxylation (P21), oxidation to an aldehyde (P14), and the corresponding
17 carboxylic acid (P19), *O*-demethylation (P11), and *N*-deisopropylation (P5). The aldehyde
18 metabolite (P14) could be further oxidized forming hydroxy-alkyl-oxo metabolite (P20).
19 Furthermore, *N,N*-bis-deisopropylation (P2) and deamination to the corresponding carboxylic acid
20 (P3) leading to the same metabolites as already seen for the DALT (D2 and D3) and ALCHT (A2
21 and A3) analogue. Combinations of *N*-deisopropylation with hydroxylations (P9 and P10) and *O*-
22 demethylation (P6) as well as combinations of *O*-demethylation with hydroxylations (P12 and P13),
23 and *O*-demethylation with *N*-deisopropylation (P4) were discovered. *N*-Deisopropylation in
24 combination with oxidation to two oxo isomers (P7 and P8) could also be found as for the DALT
25 analogue (D7 and D8). Glucuronidation and sulfation could be observed for *O*-demethyl-*N*-
26 deisopropyl (P4) and *O*-demethyl (P11) metabolites. For *O*-demethyl-*N*-deisopropyl-hydroxy-alkyl
27 (P6), *N*-deisopropyl-hydroxy-aryl (P9), *O*-demethyl-hydroxy-alkyl isomer 2 (P13), hydroxy-aryl
28 isomer 1 and 2 (P16 and P17), and aryl-alkyl-bis-hydroxy (P21) metabolites, only glucuronides
29 were found. Two glucuronide isomers (P9G1 and P9G2) were detected for the *N*-deisopropyl-
30 hydroxy-aryl metabolite (P9).
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61 **Comparison to published 5-MeO-DALT and 5-MeO-DIPT metabolism studies**

62 **5-MeO-2-Me-DALT**

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3 Michely *et al.*,^[18] described the metabolism of 5-MeO-DALT in rats. The main metabolic reactions
4 (*O*-demethylation, hydroxylations, and *N*-deallylation) were similar for both compounds. However,
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6 some metabolites, especially those modified at the 2-methyl group could only be found for 5-MeO-
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8 2-Me-DALT. Compared to 5-MeO-DALT, no bis- or tris-hydroxylations and no ring-rearranged
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10 metabolite could be found for the 2-methyl derivative probably due to the substitution of the 2
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12 position of the indole ring which could lead to a steric hindrance for the rearrangement. In addition,
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16 *N,N*-bis-deallylation followed by oxidative deamination and *N*-oxidation could only be detected for
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18 5-MeO-2-Me-DALT.
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20 21 22 23 **5-MeO-2-Me-DIPT**

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27 Narimatsu *et al.*,^[31-33] described, besides CYP kinetic studies, only the presence of a *O*-
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29 demethylated, a *N*-deisopropylated, and a mono-hydroxylated metabolite in rat and human liver
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31 microsomes, which all were also found in our study for 5-MeO-2-Me-DIPT.
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34 35 36 37 **Initial CYP activity screening studies**

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41 The ten most abundant human hepatic CYPs were incubated as described above to provide a
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43 qualitative description of metabolizing isoenzymes. As shown in Table 1, the involved isoenzymes
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45 were the same for the three NPS. *O*-Demethylation was catalyzed by CYP2C19 and CYP2D6, *N*-
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47 deallylation, or *N*-deisopropylation by CYP2C19, CYP2D6, and CYP3A4, and hydroxylations by
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49 CYP1A2, CYP2C19, CYP2D6, and CYP3A4. Additionally, the *N*-decyclohexylation of 5-MeO-2-
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51 Me-ALCHT was catalyzed by CYP2C9, CYP2C19, and CYP3A4.
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56 57 58 59 60 **Incubations with pHLM and pHLC**

1 Due to the lack of authentic human urine samples, *in vitro* testing was performed to evaluate the
2 transferability of rat metabolism results to humans. The combination of rat urine studies and the
3 incubation of pHLM and/or pHLC with the corresponding cofactors is a common technique that
4 produced reliable results and data for detection methods in human urine as shown in previous
5 studies.^[23,34-37] However, it should be kept in mind, that the comparison of *in vitro* and *in vivo*
6 models could be challenging due to sampling time of urine (24 h), time, amount, and route of
7 application, the used incubation conditions, and/or genetic variations.
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18 **5-MeO-2-Me-DALT**

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23 As summarized in Table 2, 22 out of 24 identified phase I metabolites of 5-MeO-2-Me-DALT were
24 detected in rat urine, 14 could be confirmed in the *in vitro* approach. In addition, two hydroxy-alkyl
25 metabolites (D16 and D19) were only detected in the *in vitro* model. This was probably based on
26 the 24 hours urine collection period and consequently further metabolism steps to the corresponding
27 oxo or carboxy metabolites occurred in rat. D4, D6, D9, D11, D13, D17, and D18 were found as
28 glucuronides, and D4, D6, D9, D11, and D18 as sulfates *in vivo*. D11, D17, and D18 were found as
29 glucuronides and D18 as sulfate *in vitro*. These differences could have been based on the incubation
30 time and lower formation of the corresponding phase I metabolites, especially in cases where
31 multiple metabolic reaction steps were needed.
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45 **5-MeO-2-Me-ALCHT**

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50 As shown in Table 3, 17 out of 24 identified *in vivo* phase I metabolites of 5-MeO-2-Me-ALCHT
51 could be recovered from the *in vitro* model. Eight glucuronides and six sulfates were detected in rat
52 urine and one glucuronide under *in vitro* conditions. The reasons for these differences were
53 considered comparable to those mentioned above. In comparison to 5-MeO-2-Me-DALT, no
54 additional metabolites could be found in the *in vitro* model when compared to the *in vivo* approach.
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5-MeO-2-Me-DIPT

According to Table 4, 15 out of 20 phase I metabolites identified *in vivo* could also be found in the *in vitro* model. Nine glucuronides and two sulfates were identified *in vivo*, whereas two glucuronides were detected *in vitro*.

Toxicological detection by standard urine screening approaches (SUSAs)

So far, no data about dosages or potency of the compounds are available. Therefore, the tested dosages for 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCMT, and 5-MeO-2-Me-DIPT were estimated based on those described for other tryptamine-derived NPS and were set to 1 mg/kg BW and 0.1 mg/kg BW. Michely *et al.*^[18] showed that such dosages were at or below common users doses of DALT and 5-MeO-DALT. The fact that the 0.1 mg/kg BW dosage was not detectable (except of 5-MeO-2-Me-DALT) could lead to a false negative result e.g. in context of forensic issues.

GC-MS SUSAs

Unfortunately, the three compounds and/or their metabolites could not be detected in rat urine after low dosages probably due to low concentration or instabilities caused by the sample preparation. However, after high dose administration, some metabolites were detected, indicating that the compounds might be detectable in a case of acute overdose.

LC-MSⁿ SUSAs

1 A list of all detected metabolites is given in Tables 5 and 6. 5-MeO-2-Me-DALT was detectable *via*
2 its metabolites after both low dose administrations. For 5-MeO-2-Me-ALCHT as well as 5-MeO-2-
3 Me-DIPT, only metabolites were detectable after 1 mg/kg BW dosages, but not after the lower 0.1
4 mg/kg BW dosages.
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10 **LC-HR-MS/MS SUSAs**

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17 As summarized in Table 7, metabolites of 5-MeO-2-Me-DALT and 5-MeO-2-Me-DIPT could be
18 identified with the accurate PM and a HR-MS/MS spectrum (identification, I) after 1 mg/kg BW
19 dose administrations. In addition, metabolites could be identified after the lower 0.1 mg/kg BW
20 dosage for 5-MeO-2-Me-DALT. For 5-MeO-2-Me-ALCHT, only detection (D) of the accurate PM
21 without the corresponding HR-MS/MS spectrum was possible. A second run with an inclusion list
22 on the masses of described metabolites would be necessary to confirm an intake of 5-MeO-2-Me-
23 ALCHT. The parent compounds were not identified, nor have they been detected in any of the urine
24 samples. Exemplified for 5-MeO-2-Me-DALT, Figure 4 shows a reconstituted ion chromatogram
25 of the 1 mg/kg BW dosage rat urine sample with various detected metabolites.
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39 **Conclusions**

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44 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT were extensively
45 metabolized in rats. *In vitro* metabolism studies with pHLM plus pHLC showed similar results,
46 which indicated that the described metabolites should also be detectable in human urine samples.
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49 CYP1A2, CYP2C9, CYP2C19, CYP2D6, and CYP3A4 were identified to be involved in the
50 biotransformation described in this study. An intake of an estimated users' dose of 1 mg/kg BW
51 was detectable *via* the metabolites by both LC-MS SUSAs for 5-MeO-2-Me-DALT and 5-MeO-2-
52 Me-DIPT. For 5-MeO-2-Me-ALCHT, a separate run using an inclusion list on the targets of interest
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1 would be necessary for detection. In addition, 5-MeO-2-Me-DALT metabolites were also
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3 detectable after the lower 0.1 mg/kg BW dosage by both LC-MS SUSAs.
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Table 1 CYP isoenzymes involved in metabolic pathways of 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT

Pathway	CYP 1A2	CYP 2A6	CYP 2B6	CYP 2C8	CYP 2C9	CYP 2C19	CYP 2D6	CYP 2E1	CYP 3A4	CYP 3A5
5-MeO-2-Me-DALT										
<i>O</i> -Demethylation							+	+		
<i>N</i> -Deallylation							+	+		+
Hydroxylations	+						+	+		+
5-MeO-2-Me-ALCHT										
<i>O</i> -Demethylation							+	+		
<i>N</i> -Deallylation							+	+		+
<i>N</i> -Decyclohexylation					+		+			+
Hydroxylations	+						+	+		+
5-MeO-2-Me-DIPT										
<i>O</i> -Demethylation							+	+		
<i>N</i> -Deisopropylation	+						+	+		+
Hydroxylations	+						+	+		+

Table 2 Appearance of 5-MeO-2-Me-DALT phase I and II metabolites in rat urine and in pHLM/HLC mix; G = glucuronide, S = sulfate; numbering according to Figure S1 (see SI).

Metabolite	Rat urine		pHLM/HLC	
	Phase I	Phase II	Phase I	Phase II
D2	+			
D3	+			
D4	+	+ G/S	+	
D5	+		+	
D6	+	+ G/S	+	
D7	+			
D8	+			
D9	+	+ G/S	+	
D10	+		+	
D11	+	+ G/S	+	+ G
D12	+		+	
D13	+	+ G	+	
D14	+			
D15	+		+	
D16			+	
D17	+	+ G		+ G
D18	+	+ G/S	+	+ G/S
D19			+	
D20	+		+	
D21	+		+	
D22	+		+	
D23	+			
D24	+			
D25	+		+	

Table 3 Appearance of 5-MeO-2-Me-ALCHT phase I and II metabolites in rat urine and in pHLM/HLC mix; G = glucuronide, G₁/G₂ = glucuronide isomer 1 and 2, S = sulfate; numbering according to Figure S3 (see SI).

Metabolite	Rat urine		pHLM/HLC	
	Phase I	Phase II	Phase I	Phase II
A2	+			
A3	+			
A4	+	+ G	+	
A5	+		+	
A6	+	+ G/S	+	
A7	+			
A8	+	+ G ₁ /G ₂ /S	+	
A9	+		+	
A10	+	+ G/S	+	
A11	+		+	
A12	+	+ G/S		
A13	+		+	
A14	+			
A15	+			
A16	+		+	
A17	+	+ G/S	+	+ G
A18	+	+ G/S	+	
A19	+		+	
A20	+		+	
A21	+		+	
A22	+		+	
A23	+		+	
A24	+			
A25	+		+	

Table 4 Appearance of 5-MeO-2-Me-DIPT phase I and II metabolites in rat urine and in pHLM/HLC mix; G = glucuronide, G₁/G₂ = glucuronide isomer 1 and 2, S = sulfate; numbering according to Figure S5 (see SI).

Metabolite	Rat urine		pHLM/HLC	
	Phase I	Phase II	Phase I	Phase II
P2	+			
P3	+			
P4	+	+ G/S	+	
P5	+		+	
P6	+	+ G		
P7	+			
P8	+			
P9	+	+ G ₁ /G ₂	+	
P10	+		+	
P11	+	+ G/S	+	+ G
P12	+		+	
P13	+	+ G	+	
P14	+		+	
P15	+		+	
P16	+	+ G	+	+ G
P17	+	+ G	+	
P18	+		+	
P19	+		+	
P20	+		+	
P21	+	+ G	+	

Table 5 Metabolites of 5-MeO-2-Me-DALT, m/z values of protonated molecule, characteristic MS² and MS³ fragment ions, and retention time (RT) detected by LC-MSⁿ SUSA in both low dose urines (1 mg/kg BW and 0.1 mg/kg BW). The numbers correspond to those shown in Figures S1 and S2 (see SI).

No.	Target for SUSA	Protonated molecule, m/z	MS ² fragment ions, m/z , and relative intensity, %	MS ³ fragment ions, m/z , and relative intensity, %, on the ion given in bold	RT, min
D4	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl- <i>N</i> -deallyl-)	231	174 (100)	174 : 146 (100), 133 (23), 131 (8)	4.3
D11	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-)	271	174 (100), 110 (4)	174 : 146 (100), 133 (10), 131 (10)	6.2
D11 G	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-) glucuronide	447	350 (100), 271 (74), 174 (79)	350 : 174 (100), 253 (26), 146 (2)	4.5
D13 G	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-HO-allyl-aryl-) glucuronide	463	366 (100), 190 (6)	366 : 190 (100), 172 (4)	4.6
D17 G	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-HO-allyl-aryl-) glucuronide	477	380 (100), 204 (18), 301 (7)	380 : 204 (100), 172 (7)	5.0

Table 6 Metabolites of 5-MeO-2-Me-ALCHT and 5-MeO-2-Me-DIPT, m/z values of protonated molecule, characteristic MS² and MS³ fragment ions, and retention time (RT) detected by LC-MSⁿ SUSA in low dose urine (1 mg/kg BW). The numbers correspond to those shown in Figures S3-S6 (see SI).

No.	Target for SUSA	Protonated molecule, m/z	MS ² fragment ions, m/z , and relative intensity, %	MS ³ fragment ions, m/z , and relative intensity, %, on the ion given in bold	RT, min
A10 G	5-MeO-2-Me-ALCHT-M (<i>O</i> -demethyl- <i>N</i> -deallyl-) glucuronide	449	174 (100), 273 (99), 350 (20), 159 (13)	174 : 146 (100) 273 : 174 (100)	6.1
A17 G	5-MeO-2-Me-ALCHT-M (<i>O</i> -demethyl-) glucuronide	489	350 (100), 313 (85), 174 (84), 152 (3)	350 : 174 (100)	7.6
P11	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-)	275	174 (100), 114 (4)	114 : 72 (100) 174 : 146 (100), 131 (13), 133 (9)	6.0
P20	5-MeO-2-Me-DIPT-M (HO-alkyl-oxo-)	319	218 (100), 114 (6)	218 : 190 (100), 186 (98), 172 (75), 158 (55)	6.6
P21	5-MeO-2-Me-DIPT-M (HO-alkyl-HO-aryl-)	321	220 (100), 205 (3), 114 (1)	220 : 172 (100), 186 (98), 190 (85), 160 (68), 174 (27)	9.2
P11 G	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-) glucuronide	451	350 (100), 174 (68), 275 (38)	174 : 146 (100), 131 (11), 133 (5)	4.3
P13 G	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-HO-alkyl-) glucuronide	467	366 (100), 190 (76)	190 : 134 (100) 366 : 190 (100), 172 (4)	4.2
P16 G	5-MeO-2-Me-DIPT-M (HO-aryl-) glucuronide isomer 1	481	380 (100), 204 (11)	204 : 172 (100), 204 (5)	4.6
P17 G	5-MeO-2-Me-DIPT-M (HO-aryl-) glucuronide isomer 2	481	380 (100), 204 (30)	204 : 173 (100), 172 (82), 163 (54), 204 (16)	5.4

Table 7 Metabolites of 5-MeO-2-Me-DALT, 5-MeO-2-Me-ALCHT, and 5-MeO-2-Me-DIPT, calculated exact mass of the protonated molecule, and retention time (RT) detected by LC-HR-MS/MS SUSA in rat urine (1 mg/kg body weight (BW) or 0.1 mg/kg BW). D = detection of protonated molecule in MS¹, I = identification via MS¹ and MS²; the numbers correspond to those shown in Figures S1-S6 (see SI).

No.	Target for SUSA	Calculated exact masses of protonated molecule, <i>m/z</i>	1 mg/kg BW dose	0.1 mg/kg BW dose	RT, min
D4	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl- <i>N</i> -deallyl-)	231.1492	I	I	3.0
D6	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl- <i>N</i> -deallyl-HO-alkyl-)	247.1441	D	D	2.6
D11	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-)	271.1805	I	I	3.6
D13	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-HO-alkyl-) isomer 2	287.1754	I		3.3
D17	5-MeO-2-Me-DALT-M (HO-aryl-) isomer 1	301.1911	D	D	3.8
D18	5-MeO-2-Me-DALT-M (HO-aryl-) isomer 2	301.1911	I		4.0
D21	5-MeO-2-Me-DALT-M (HOOC-)	315.1703	D		3.7
D11 S	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-) sulfate	351.1373	D		3.8
D4 G	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl- <i>N</i> -deallyl-) glucuronide	407.1813	D	D	2.7
D11 G	5-MeO-2-Me-DALT-M (<i>O</i> -demethyl-) glucuronide	447.2123	I	I	3.3
D17 G	5-MeO-2-Me-DALT-M (HO-aryl-) glucuronide	477.2231	D	D	3.4
A17	5-MeO-2-Me-ALCHT-M (<i>O</i> -demethyl-)	313.2274	D		4.9
A8 G ₁	5-MeO-2-Me-ALCHT-M (<i>N</i> -decyclohexyl-HO-aryl-) glucuronide isomer 1	437.2282	D		3.1
A10 G	5-MeO-2-Me-ALCHT-M (<i>O</i> -demethyl- <i>N</i> -deallyl-) glucuronide	449.2282	D		3.8
A17 G	5-MeO-2-Me-ALCHT-M (<i>O</i> -demethyl-) glucuronide	489.2595	D		4.3
P4	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl- <i>N</i> -deisopropyl-)	233.1648	I		3.1
P11	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-)	275.2118	I		3.7
P12	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-HO-alkyl-) isomer 1	291.2067	I		2.8
P20	5-MeO-2-Me-DIPT-M (HOOC-)	319.2016	D		3.9
P4	5-MeO-2-Me-DIPT-M	409.1969	D		2.8

G	(<i>O</i> -demethyl- <i>N</i> -deisopropyl-) glucuronide				
P11 G	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-) glucuronide	451.2439	D		3.2
P13 G	5-MeO-2-Me-DIPT-M (<i>O</i> -demethyl-HO-alkyl-) glucuronide	467.2389	D		3.2

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Legends to Figures

Figure 1 Proposed metabolic pathways of 5-MeO-2-Me-DALT. Phase II metabolites: glucuronides (G), and sulfates (S). Structures in brackets represent proposed (but not detected) intermediates; numbering according to Figure S1 (see SI).

Figure 2 Proposed metabolic pathways of 5-MeO-2-Me-ALCHT. Phase II metabolites: glucuronides (G), and sulfates (S). Structures in brackets represent proposed (but not detected) intermediates; numbering according to Figure S3 (see SI).

Figure 3 Proposed metabolic pathways of 5-MeO-2-Me-DIPT. Phase II metabolites: glucuronides (G), and sulfates (S). Structures in brackets represent proposed (but not detected) intermediates; numbering according to Figure S5 (see SI).

Figure 4 Reconstructed LC-HR-MS ion chromatogram with the corresponding protonated exact masses indicating the given 5-MeO-2-Me-DALT metabolites in rat urine after 1 mg/kg BW dose administration; numbering according to Figures S1 and S2 (see SI).

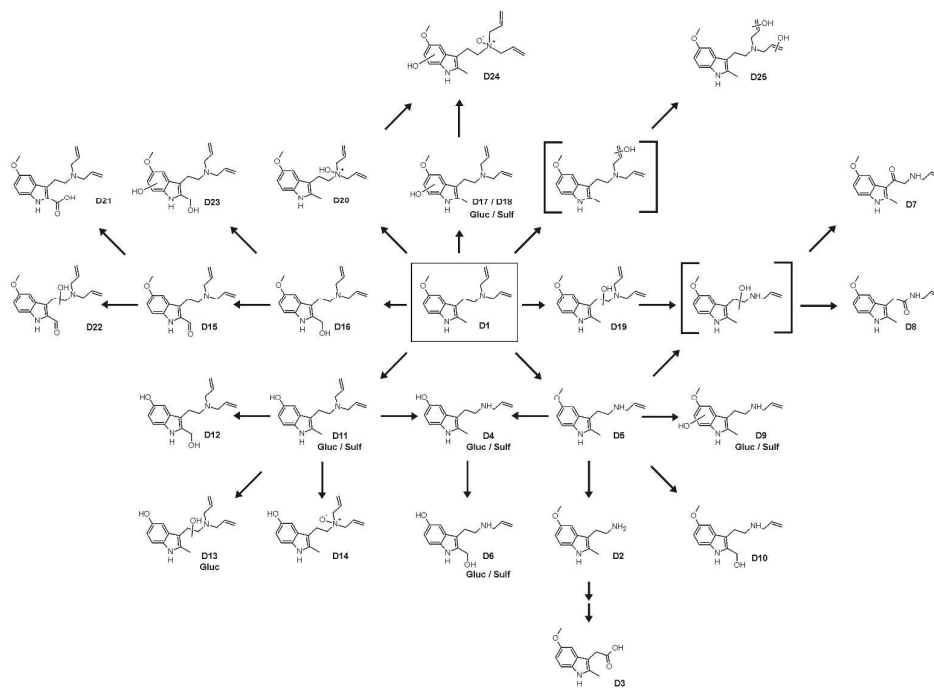


Figure 1 Proposed metabolic pathways of 5-MeO-2-Me-DALT. Phase II metabolites: glucuronides (G), and sulfates (S). Structures in brackets represent proposed (but not detected) intermediates; numbering according to Figure S1 (see SI).

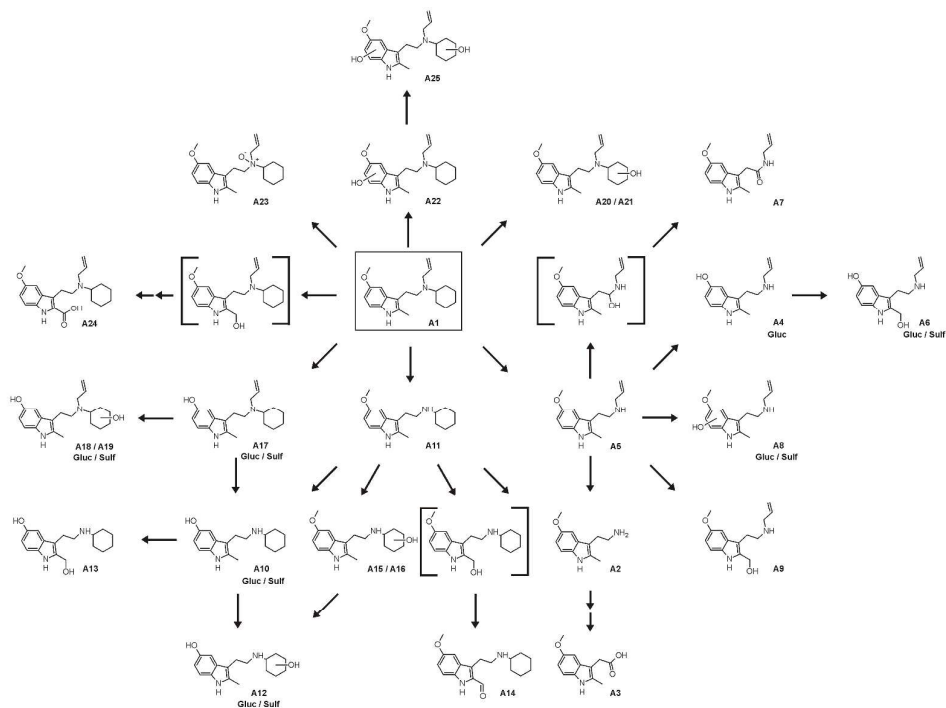


Figure 2 Proposed metabolic pathways of 5-MeO-2-Me-ALCHT. Phase II metabolites: glucuronides (G), and sulfates (S). Structures in brackets represent proposed (but not detected) intermediates; numbering according to Figure S3 (see SI).

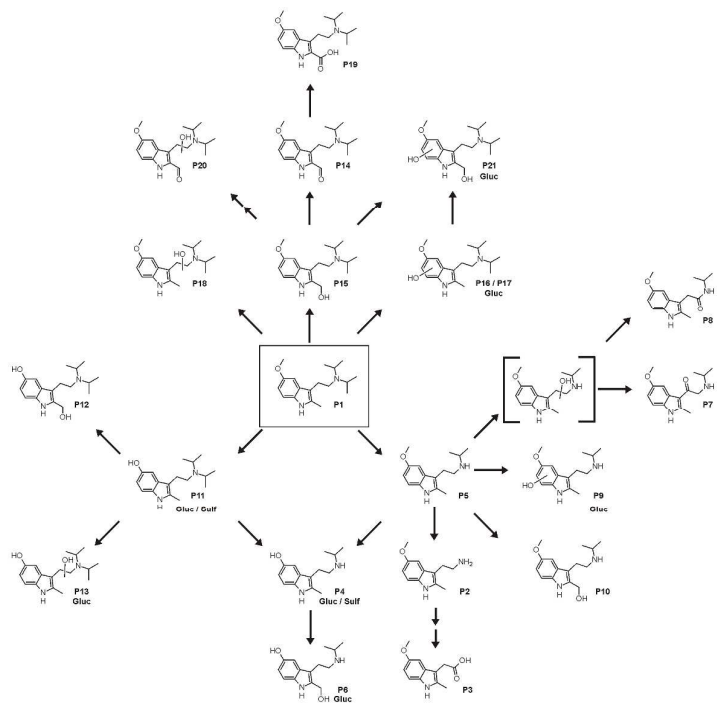


Figure 3 Proposed metabolic pathways of 5-MeO-2-Me-DIPT. Phase II metabolites: glucuronides (G), and sulfates (S). Structures in brackets represent proposed (but not detected) intermediates; numbering according to Figure S5 (see SI).

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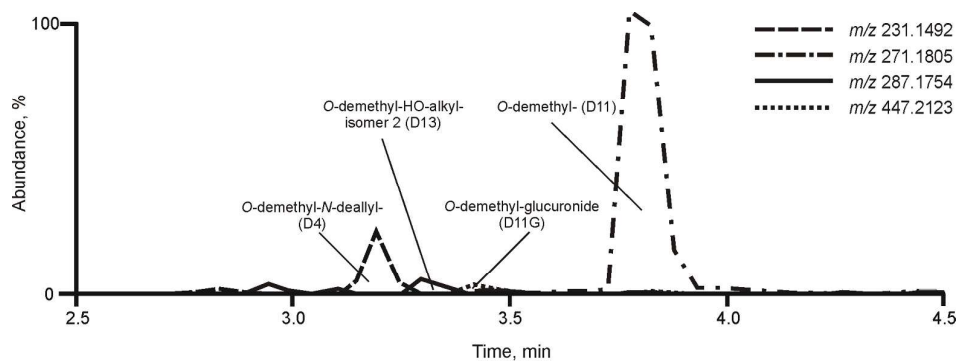


Figure 4 Reconstructed LC-HR-MS ion chromatogram with the corresponding protonated exact masses indicating the given 5-MeO-2-Me-DALT metabolites in rat urine after 1 mg/kg BW dose administration; numbering according to Figures S1 and S2 (see SI).