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Transcranial static magnetic field stimulation of M1 reduces corticospinal excitability without distorting sensorimotor integration in humans

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Dear Editor:

Transcranial static magnetic stimulation (tSMS) over the human motor cortex using powerful neodymium magnets induces a reduction in corticospinal excitability. It seems to act through intracortical inhibitory (ICI) mechanisms, which are GABA_A-dependent [1]; [2]. Similar effects have been documented in somatosensory cortex [3]. Given that tSMS modulates motor and sensory physiology [3], we wondered whether static magnetic fields could also affect sensorimotor integration. Here, we evaluated the effects of tSMS over M1 on short/long afferent-inhibition (SAI/LAI). SAI/LAI occur at cortical level when coupling (with the proper timing) an electrical pulse on a peripheral nerve (PNS) with a transcranial magnetic stimulation (TMS) pulse on M1 [4]. SAI is known to interact with ICI [5], therefore we hypothesized that tSMS-M1 would reduce corticospinal excitability with after-effects on SAI.

Seventeen un-medicated, healthy people participated (20–41yrs) (Ethic-Committee approval: CE-17/2015). In each session we determined (individually) the inter-stimulus-interval (ISI) which was used to evaluate SAI by recording 70 TMS motor-evoked potentials (MEP) in the relaxed first dorsal interosseous (FDI) muscle, at 0.2Hz. Ten MEP were unconditioned at an intensity high enough to obtain response amplitudes ≈ 1 mV, and 60 conditioned with PNS on the median nerve at the wrist, at the following intervals: 20-21-22-23-24-25 ms (Fig. 1a). Then, 36 *pre*-MEP were recorded. Of these, twelve were unconditioned (TEST) with a fixed intensity producing amplitudes ≈ 1 mV, and the other 24 conditioned by PNS (12 to evaluate SAI at the ISI determined above, and 12 to evaluate LAI with ISI = 200 ms [6]). MEP were acquired in 12 sequences (TEST-SAI-LAI). Next, tSMS was applied on the M1-FDI hot-spot as detailed previously [1], for 20min. One session was *real* tSMS (magnetic field of 0.5T), the other *sham*, with the order counterbalanced across subjects. Then, *Post*-MEP were recorded in two blocks: *i*) *Post-fixed intensity* (TMS intensity was the same as *pre*) and *ii*) *post-matched amplitude* (TMS intensity was adjusted to obtain TEST-amplitudes \approx *pre*-TEST); again the order was counterbalanced across subjects. Finally, we followed with the *post2* evaluation, starting 15min after the end of tSMS. The procedure was the same as in *post*.

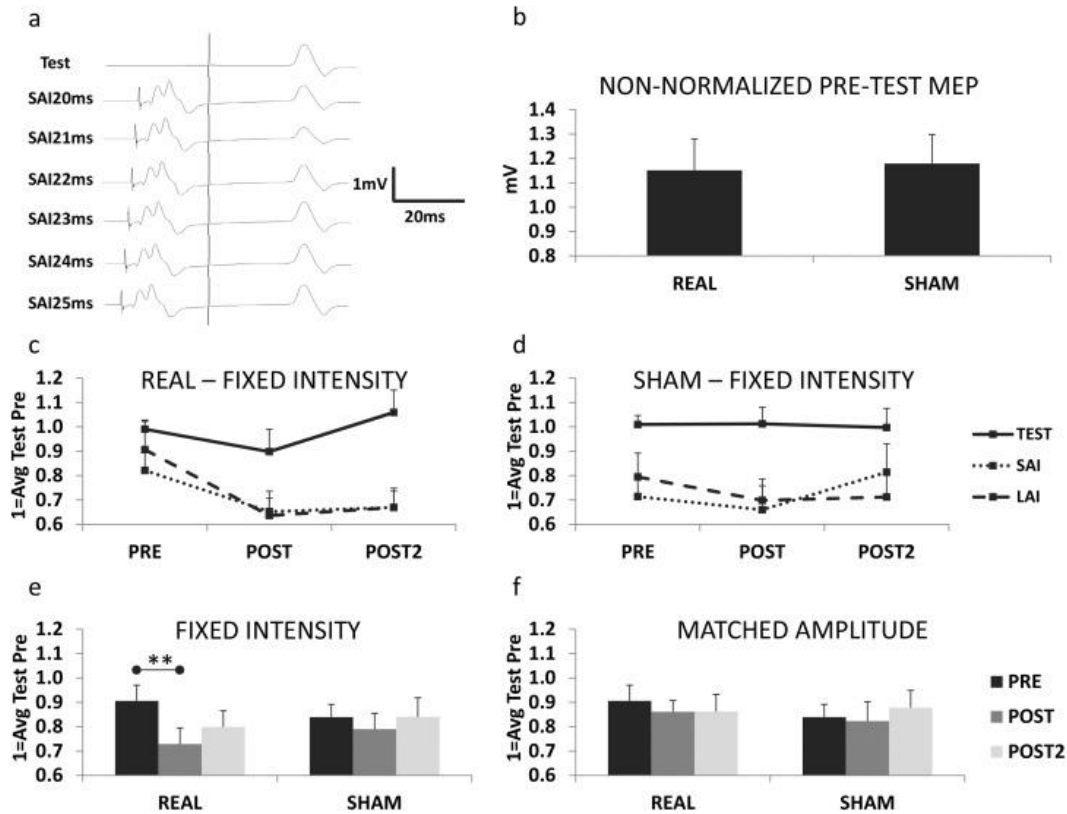


Fig. 1. a.) SAI inter-stimulus-interval (ISI) determination in one subject (an ISI of 21 ms was chosen because it generated the greatest inhibition). b.) Non-normalized TEST amplitudes obtained during *real* and *sham* sessions did not differ at *pre* (Student t-test). The average of *pre* values in *real* and *sham* sessions was 1.16 mV (SE 0.12); this value is used as the unitary value in the ordinate of the graphs shown in the rest of the figure. c,d.) TEST, SAI and LAI changes over testing times (for *fixed-intensity blocks*) were different for *real* and *sham* (ANOVA $p = 0.035_{\text{STIM} \times \text{CONDITION} \times \text{TIME}}$). Subsequent ANOVA for each stimulation mode (*real* and *sham*) showed a reduction of MEP over time for *real*-tSMS (ANOVA $p = 0.016_{\text{TIME}}$) observed for TEST, SAI and LAI (ANOVA $p > 0.05_{\text{TIME} \times \text{CONDITION}}$); at *post* MEP were reduced $\approx 20\%$ (*post-hoc*; $p = 0.005$, $p = 0.015_{\text{Bonferroni}}$). For *sham*-tSMS MEP did not change significantly over time. e) Time effects for c.) and d.) with all Conditions pooled. f) MEP responses pooling Conditions for *matched-amplitude blocks* on a time basis. In all analyses shown in figures c-f, the amplitude of the TEST was always larger than SAI and LAI (ANOVA $p < 0.01_{\text{CONDITION}}$). ** $p < 0.01$.

For TMS application and MEP recording we followed standard methods described elsewhere [1]. PNS was applied with a Digitimer-DS7A (500 μ sec pulse-duration; cathode proximal). MEP amplitudes were normalized (intra-subject normalization). For this, for each subject we calculated the average of the values of the TEST at *pre* across the *real* and *sham* sessions. This value was used as denominator for all values of the given subject at all testing time-points, either for the TEST, SAI and LAI. Graphs show means and standard error of the mean (SE). Results were considered significant if $p < 0.05$.

The essential message from our results is straightforward: tSMS reduces corticospinal excitability but has no effect on SAI or LAI. Fig. 1b shows similar non-normalized TEST amplitudes for *real* and *sham* sessions before magnet application (similar results were obtained for SAI and LAI). For *fixed-intensity blocks*, *real*-tSMS reduced the normalized amplitudes of the TEST, SAI and LAI at *post* (by $\approx 20\%$), but all recovered at *post2*; *sham*-tSMS produced no effects (Fig. 1c-e). To check whether SAI and LAI responses to tSMS were influenced by the size of the TEST, responses to tSMS were acquired while keeping the amplitude of the TEST constant at *pre*, *post* and *post2* (*matched-amplitude blocks*). In this case the amplitude of the TEST, SAI and LAI did not change significantly over time (Fig. 1f, TIME effect; all conditions pooled). In *fixed-intensity* and *matched-amplitude blocks* TEST were larger than SAI and LAI at all testing times.

These results corroborate the finding that 20 min of tSMS reversibly reduces corticospinal excitability [1]. tSMS affected SAI and LAI similarly, and both followed the same pattern of inhibition shown affecting TEST. However, the evaluation of SAI and LAI while maintaining the same amplitude of the TEST along testing-times (*matched-amplitude blocks*) reveals that SAI/LAI were not modified by tSMS. Therefore, tSMS reduces corticospinal excitability without affecting SAI and LAI.

The mechanisms underlying SAI are mainly cortical and are related to the cholinergic and GABAergic systems [5] ; [7]. The fact that in our hands tSMS seems not to directly affect SAI circuits but has been shown to modulate ICI [2] leads us to conclude that SAI and ICI might be representing the operational mechanism of distinct subtypes of GABAergic inhibitory interneurons [8], which in one case is affected by tSMS, but not in the other. We cannot discount an effect of tSMS on intrinsic excitability of the corticospinal motoneurons; in the case of a putative inhibition of M1 pyramidal cell bodies the response to TMS would wane independently of the input to the cells from cortico-cortical connections, something compatible with our results. The mechanisms responsible for LAI are less understood but it has been proposed that LAI implicates interneurons acting via GABA_B receptors. Nevertheless, whatever the exact mechanism, it seems to be unaffected by tSMS on M1.

In conclusion, tSMS-M1 reduces corticospinal excitability without affecting SAI and LAI. Further research should explore the effect of tSMS on some other expressions of sensorimotor integration.

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