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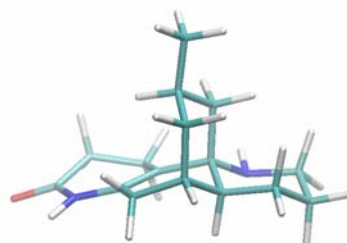
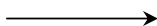
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## GRAPHICAL ABSTRACT



*N*-demethyl-sauroxine

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***N*-demethyl-sauroxine, a novel Lycopidine Group alkaloid from *Huperzia saururus***

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The present study describes the isolation and identification of *N*-demethyl-sauroxine, a novel Lycopodium alkaloid obtained from *Huperzia saururus* (Lam.) Trevis. (Lycopodiaceae). Its structure and relative stereochemistry were elucidated on the basis of its spectral data and chemical correlations. Additionally, acetylcholinesterase inhibitory activity was evaluated ( $IC_{50} = 209.6 \pm 1.1 \mu M$ ). The structure of the already identified alkaloid sauroxine was also re-validated through two dimensional NMR data.

*Keywords:* *Huperzia saururus*; *N*-demethyl-sauroxine; acetylcholinesterase inhibition; sauroxine NMR two dimensional data.

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*Huperzia saururus* (Lam.) Trevis. (“cola de quirquincho”, Lycopodiaceae) is a native Argentine species, used in the ethnomedicine as aphrodisiac<sup>1</sup> and memory improver.<sup>2</sup> As a consequence of decoctions consuming, cases of intoxication were detected, with manifestations such as nausea, vomits, abortion, even coma as it is explained by the Health Ministry in Argentina.<sup>1,3</sup>

Regarding chemical studies, we have identified *Lycopodium* alkaloids as the main compounds being sauroine and sauroxine (**1**) the majority alkaloids.<sup>4,5</sup> According to the hypothetic cholinergic stimulation implied by the adverse effects, biological assays developed with the alkaloid extract (AE) showed an important inhibitory action on the acetylcholinesterase enzyme (AChE).<sup>4</sup> Also it was demonstrated that AE exerts a facilitation of both, the long term potentiation (LTP) induction on hippocampal slices<sup>6</sup> and the memory retention according to Step-down Test results.<sup>7</sup> Besides, sauroine has similar properties in relation to the memory phenomenon, but it has not an inhibitory effect on AChE.<sup>5,8</sup>

In this paper, the isolation and structure elucidation of a novel *Lycopodium* alkaloid (**2**) is described. The obtaining of the new compound through a semi-synthesis strategy was also proposed, employing sauroxine, the second major alkaloid in *H. saururus*, as a substrate.

Additionally, due to the limited existing data of sauroxine,<sup>9,10</sup> its structure was re-validated.

#### PREFERRED POSITION FOR CHEMICAL STRUCTURES

Aerial parts of *H. saururus* dried and ground (2.45 kg) were alkalinized with 0.1 M NaOH (pH 12) and extracted by soxhlet with  $\text{ClCH}_3$  as solvent. 77 g of extract were obtained. This was solubilized with pH 2 HCl solution, then alkalinized until pH 12 with 0.1 N NaOH and partitioned with  $\text{ClCH}_3$  by using a liquid-liquid extractor. The organic phase, with a yield of 3.95 g, was purified by Sephadex LH-20 using  $\text{ClCH}_3/\text{EtOH}$  (1:1) as mobile phase. All the fractions that were positive to the Dragendorff's reagent were combined (2.21 g). This fraction was re-

submitted to Sephadex LH<sub>20</sub>, but acetone was used as mobile phase. Fraction 3 was purified by CC using Sephadex G-10 with H<sub>2</sub>O/EtOH (95:5). By posterior purification of the third fraction by TLC using ciclohexane/ClCH<sub>3</sub>/diethylamine (5:4:1) as mobile phase, **1** (9.7 mg, 0.00039%) was obtained; fraction 4 was purified in the same way but using ciclohexane/diethylamine (1:1) as mobile phase. Three bands were obtained, and the first one was purified by TLC with Cl<sub>2</sub>CH<sub>2</sub>/MeOH (4:1) leading to *N*-demethyl-sauroxine (**2**, 1.9 mg, 0.0000775%). It is noteworthy that this alkaloid was also obtained by using other extraction methods, and other extractant such as H<sub>2</sub>O, both at acid and neutral pH (data not shown), thus dismissing the possibility that **2** could be an artefact.

Semi-synthesis of **2** was carried out by means of an adaptation of the Polonovsky reaction.<sup>11</sup> **1** (9.1 mg) was solubilized in MeOH (200 μL) and maintained at 0°C. Then, H<sub>2</sub>O<sub>2</sub> (20 μL) was added drop by drop. Reaction was left to develop at 25°C, through 6 h. Later, little amounts of MnO<sub>2</sub> were added to quenching the H<sub>2</sub>O<sub>2</sub> excess, and the necessary amount of MnO<sub>2</sub> was detected with KI-starch paper. This suspension was filtered by Celite® and taken to dryness, then solubilized in MeOH (200 μL) and, HCl 6 M was added until pH 2. Solution was taken to dryness again. Thus, sauroxine *N*-oxide hydrochloride was obtained (10.7 mg). This was dissolved in MeOH (200 μL) and FeSO<sub>4</sub>•7H<sub>2</sub>O (2 equiv) was added. This second step of reaction was developed as well at 25°C through 30 min. The solution was taken to dryness and solubilized with EDTA 0.1 M at pH 10 and then it was partitioned with Cl<sub>3</sub>CH ten times. Organic fractions were reunited and MgSO<sub>4</sub> (1.2 mg) was added. After filtration, it was taken to dryness. Thus, a mixture of **1** and **2** was obtained.

Both alkaloids were immediately separated by TLC, with ciclohexane/Cl<sub>3</sub>CH/diethylamine (5:4:1) as mobile phase. This way, 1.2 mg of **2** were obtained. Semi-synthetic and natural compounds were compared chromatographically by TLC.

Sauroxine (**1**). The structure of **1** was re-validated by the 2D NMR (<sup>1</sup>H-<sup>1</sup>H COSY, HMQC, HMBC, NOESY) data. The <sup>1</sup>H-<sup>1</sup>H COSY and TOCSY (Figure 1) revealed the connectivities of

C-2 to C-3, C-6 to C-7 and C-8, C-9 to C-11, and C-14 to C-16. The HMBC data (Figure 1) confirmed the  $\alpha$ -pyridone ring: H-2 showed correlations with C-1, C-4 and C-5, H-3 with C-2 and C-4, and N $\alpha$ -H with C-4. Connection to the B ring is showed through the following correlations: H-6 with C-4 and C-5, and H-7 with C-4 and C-5. Connectivities in B ring were H-6 to C-7, C-8 and C-12, H-7 to C-12 and C-13, and H-8 to C-6, C-7 and C-12. Piperidine C ring was constituted according to the correlations between H-9 with C-11 and C-13, H-10 with C-12, H-11 with C-9 and C-10, and H-12 with C-11. Finally, D ring correlations were H-7 with C-15, H-14 with C-15, H-16 with C-8, C-14 and C-15, and H-14 with C-4. NOESY correlations (Figure 1) allowed the statement of the stereochemistry of **1**. The most important were the existing between H-12 and H-6b, and H-3a.

Around thirty *Lycopodium* alkaloids belong to the Lycodine Class, where C/D mostly exhibit the *trans* configuration<sup>12</sup>,  $\alpha$ -obscurine being one example.<sup>13</sup> Differently, **1** is an stereoisomer of  $\alpha$ -obscurine and for this reason it is also called 12-epi- $\alpha$ -obscurine. In 1974, Nakashima et al., showed differences in <sup>13</sup>C NMR spectrum between the carbon signals at C-8 and C-14 for **1** and  $\alpha$ -obscurine<sup>10</sup>. In the present work, we improve the analysis showing these differences through 2D NMR, supporting the unique conformation of C/D ring in **1** and **2**.

*N*-demethyl-sauroxine<sup>14</sup> (**2**). The IR absorptions implied the presence of a ketone carbonyl group (1665.80 cm<sup>-1</sup>), an amide and amine group at 3223 cm<sup>-1</sup> and 3382 cm<sup>-1</sup>, respectively. **2** showed a molecular ion at m/z 260 (12.5 %) consistent with the formula C<sub>16</sub>H<sub>25</sub>N<sub>2</sub>O deduced from the HRESIMS (found: 261.1973; calcd; 261.1967). The base peak at m/z 203 (100 %) indicates the loss of the bridge ring (C-8, C-14, C-15, and C-16), plus a hydrogen atom from C-12 suggesting that **2** possess a lycodine-type skeleton<sup>15</sup>.

The <sup>13</sup>C analysis and HSQC-DEPT spectra indicated 16 carbon signals and showed the presence of only one methyl carbon, three methine carbons, eight methylene carbons, and four quaternary carbons. The carbon signals at  $\delta_{\text{C}} = 170.8$  and  $\delta_{\text{C}} = 59.5$  correspond to the carbonyl group (C-1) and the quaternary function (C-13). The signals at  $\delta_{\text{C-4}} = 110.6$  and  $\delta_{\text{C-5}} = 133.6$

correspond to the double bond C-C. HMBC experiment established that those signals have not correlations with any proton (Table 1). Comparing the  $^{13}\text{C}$  NMR spectrum of **1** and **2**, the most important differences were one less carbon signal and the absence of the signal belonging to the methyl group ( $\text{N}_\beta\text{-CH}_3$ ) of **1**. Furthermore, there is an increase of  $\delta_{\text{C-10}} = 18.6$  (22.8), and  $\delta_{\text{C-12}} = 32.6$  (40.1) signals, and a decrease of  $\delta_{\text{C-9}} = 47.6$  (39.9) and  $\delta_{\text{C-11}} = 24.9$  (22.4). All this is consistent with the effect observed in lycodine, in comparison with *N*-methyl-lycodine, where the *N*-methyl group is axially oriented as well.<sup>10</sup>

The  $^1\text{H}$ - $^1\text{H}$  COSY and TOCSY (Figure 2) revealed the connectivities of C-2 to C-3, C-6 to C-7 and C-8, C-9 to C-11, and C-14 to C-16. The HMBC data (Figure 2) led to confirm the  $\alpha$ -pyridone ring: H-2 showed correlations with C-1, C-3 and C-4, H-3 with C-4 and C-5, and  $\text{N}_\alpha\text{-H}$  with C-4. Connection to the B ring is showed through the following correlations: H-6 with C-4 and C-5. Connectivities in B ring were H-6 to C-7, C-8 and C-12. Connectivity in the D ring was H-8 to C-12. Finally, D ring correlations were H-14 with C-4, C-13 and C-15, H-16 with C-8, C-14 and C-15.

NOESY correlations (Figure 2) allowed the statement of the stereochemistry of **2**. The most important were those existing between H-12 and H-6b, and H3a. Together with the previous spectroscopic data, we can determine the C/D ring configuration as *cis*, and therefore, **1** and **2** are the only two Lycodine Group alkaloids having this conformation.

Semi-synthetic **2** was obtained as an amorphous solid, at a very low yield. Nevertheless, some important signals were detected in the  $^1\text{H}$  NMR spectrum such as the methyl group in C-16 and again, the absence of a methyl group in  $\text{N}_\beta$  compared to **1**. In addition, several carbon signals were identified: C-16 (22.1), C-2 (30.8), C-7 (33.8) and C-12 (42.6). As in the natural product, these chemical shifts are different from those observed in **1**.

As it was previously mentioned, the purified alkaloid extract of *H. saururus* was previously evaluated in our lab in relation to its effect on the acetylcholinesterase enzyme. As the results obtained exhibited a strong inhibitory effect ( $\text{IC}_{50} = 0.58 \mu\text{g/mL}$ )<sup>4</sup>, we evaluated some of the

purified alkaloids as well, searching for the one with the best results. Thus, sauroine showed no inhibitory effect<sup>5</sup>, sauroxine had an  $IC_{50} = 8.9 \pm 0.4 \mu\text{g/mL}$  ( $32.3 \mu\text{M}$ )<sup>17</sup>, 6-hydroxyglycopidine showed an  $IC_{50} = 78.1 \pm 3,5 \mu\text{g/mL}$  ( $298.8 \pm 13.3 \mu\text{M}$ )<sup>17</sup>, and in the present paper we evaluated the same activity for **2**. An  $IC_{50} = 54.5 \mu\text{g/mL}$  ( $209.6 \pm 1.1 \mu\text{M}$ ) was obtained. As it can be seen, hitherto no compound has similar inhibitory activity to the extract, so we postulate a possible synergistic effect among the alkaloids present.

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### Supplementary data

Supplementary data associated to this article can be found in the online version, at...

### References and notes

1. Amorín, J. L. *Farmacobotánica* **1974**, *116*, 3-6.
2. Martínez Crovetto, R. *Miscelánea* **1981**, *69*, 15.
3. Manual de intoxicaciones para agentes de atención primaria. Resolución N° 652. Ministerios de Salud y Acción Social. Buenos Aires, September 30<sup>th</sup>, **2002**.  
[http://www.msal.gov.ar/hm/site/pngcam/normas/r%2002\\_652%20.pdf](http://www.msal.gov.ar/hm/site/pngcam/normas/r%2002_652%20.pdf). Accessed June 21<sup>st</sup>, 2008.
4. Ortega, M. G.; Agnese, A. M.; Cabrera, J. L. *Phytomedicine* **2004a**, *11*, (6) 539-543.



5. Ortega, M. G.; Agnese, A. M.; Cabrera, J. L. *Tetrahedron Lett.* **2004b**, *45*, (38) 7003-7005.
6. Ortega, M. G.; Vallejo, M. G.; Cabrera, J. L.; Pérez, M. F.; Almirón, R. S.; Ramírez, O. A.; Agnese, A. M. *J. Ethnopharmacol.* **2006**, *104*, 374-378.
7. Vallejo, M. G.; Ortega, M. G.; Cabrera, J. L.; Carlini, V. P.; Rubiales de Barioglio, S. E., Agnese, A. M. *J. Ethnopharmacol.* **2007**, *111*, 685-687.
8. Vallejo, M. G.; Ortega, M. G.; Cabrera, J. L.; Carlini, V. P.; Rubiales de Barioglio, S. E.; Almirón, R. S.; Ramírez, O. A.; Agnese, A. M. *J. Nat. Prod.* **2009**, *72*, 156-158.
9. Ayer, W.; Habgood, T.; Deulofeu, V.; Juliani, H. *Tetrahedron* **1965**, *21*, 2169-2172.
10. Nakashima, T. T.; Singer, P. P.; Browne, L. M.; Ayer, W. A. *Can. J. Chem.* **1975**, *53*, 1936-1942.
11. McCamley, K.; Ripper, J.; Singer, R. D.; Scammells, P. J. *J. Org. Chem.* **2003**, *68*, 9847-9850.
12. Ma, X.; Gang, D. R. *Nat. Prod. Rep.* **2004**, *21*, 752-772.
13. Ayer, W. A.; Berezowsky, J. A.; Iverach, G. G. *Tetrahedron* **1962**, *18*, 567-573.
14. *N*-demethyl-sauroxine (**2**, very pale yellow oil;  $[\alpha]_D^{26} +17.46$  (c 0.32, MeOH)), IR (BrK)  $\nu_{\max}$  3383, 3223, 2929, 1741 and 1666  $\text{cm}^{-1}$ .
15. Anet, F. A. L.; Rao, M. V. *Tetrahedron Lett.* **1960**, *20*, 9-12.
16. Ellman, G. L.; Courtney, K. D.; Andres jr, V.; Featherstone, R. M. *Biochem. Pharmacol.* **1961**, *7* (2), 88-95.
17. Puiatti, M.; Borioni, J. L.; Vallejo, M. G.; Cabrera, J. L.; Agnese, A. M.; Ortega, M. G.; Pierini, A. B. *J. Mol. Graphics Modell.* **2013**, *44*, 136-144.

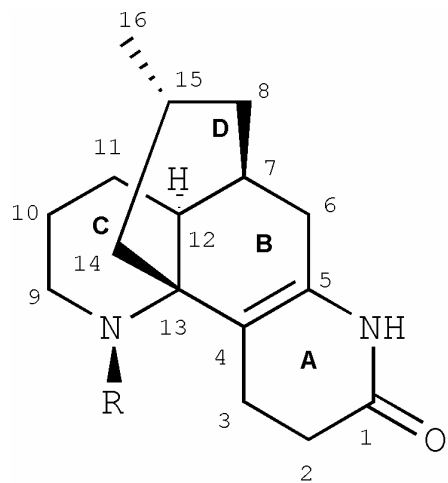
**Table 1.** NMR data of *N*-demethyl-sauroxine

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position	$\delta_C^a$	$\delta_H^b$ (multi, <i>J</i> in MHz)
1	170.8 (C)	---
2a	30.5 (CH <sub>2</sub> )	2.42 (1H, br t, 7.7)
2b		2.68 (1H, br t, 7.7)
3a	19.7 (CH <sub>2</sub> )	2.52-2.59 (1H, m)
3b		2.75 (1H, br t, 10.1)
4	110.6 (C)	---
5	133.5 (C)	---
6a	34.4 (CH <sub>2</sub> )	1.84 (1H, d, 18,8)
6b		2.60 (1H, dt, 18.8; 4,3)
7	32.7 (CH)	2.06-2.11 (1H, m)
8a	35.5 (CH <sub>2</sub> )	1.31-1.36 (1H, m)
8b		1.42 (1H, br d, 13,3)
9a	39.9 (CH <sub>2</sub> )	3.02 (1H, t, 12.3, 3.0)
9b		3.19 (1H, d, 12.3)
10a	22.4 (CH <sub>2</sub> )	1.50-1.57 (1H, m)
10b		1.82 (1H, br d, 12.2)
11	22.8 (CH <sub>2</sub> )	1.88-2.02 (1H, m)
12	40.1 (CH)	2.03-2.10 (1H, m)
13	59.5 (C)	---
14	32.2 (CH <sub>2</sub> )	1.65-1.73 (1H, m)
15	26.2 (CH)	1.72 (1H, m)
16	21.7 (CH <sub>3</sub> )	0.95 (3H, d, 5.7)

<sup>a</sup> 100 MHz, CDCl<sub>3</sub>

<sup>b</sup> 400 MHz, CDCl<sub>3</sub>

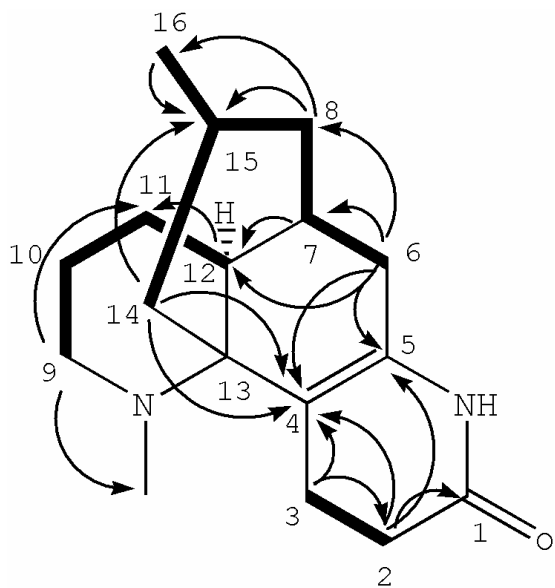


- 1: R = Me  
2: R = H

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**Figure 1.** Selected 2D NMR correlations for **1**.

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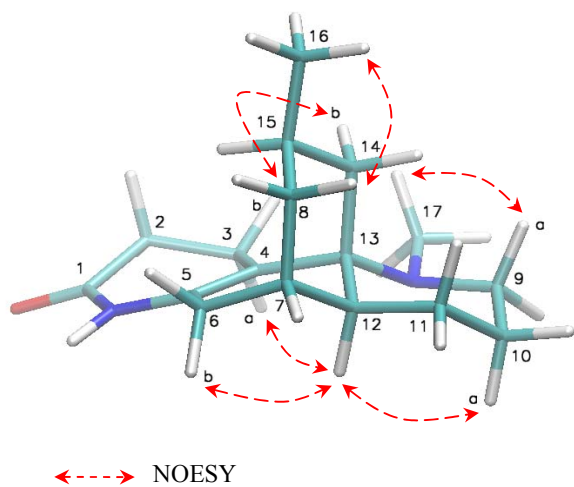


—  $^1\text{H}$ - $^1\text{H}$  COSY and TOCSY  
→ HMBC

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**Figure 2.** Selected NOESY correlations for **1**.

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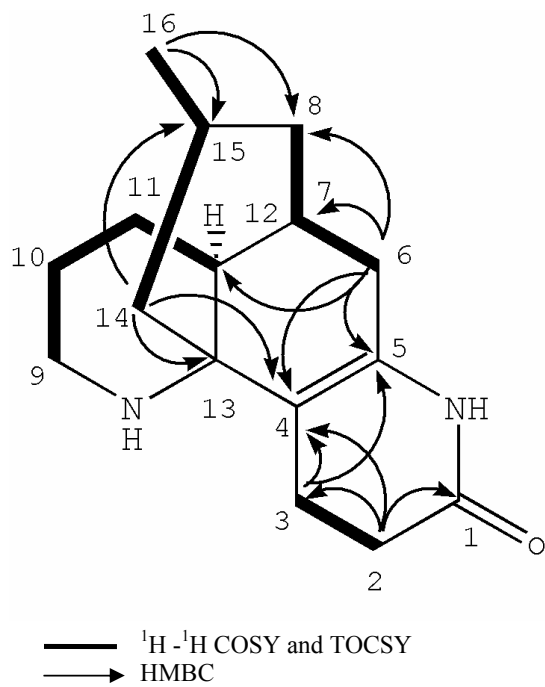


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**Figure 3.** Selected 2D NMR correlations for **2**.

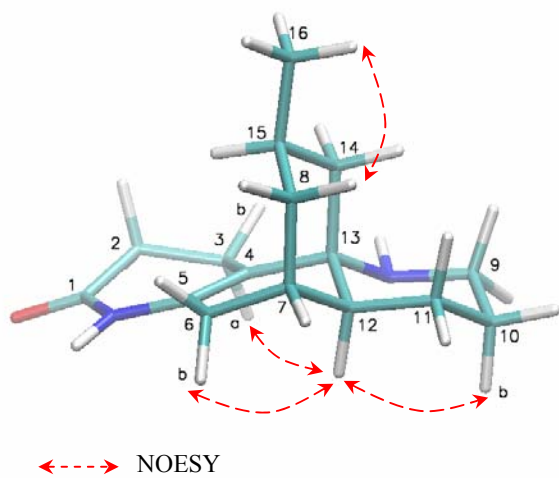
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**Figure 4.** Selected NOESY correlations for **2**.

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