



Database tool

Kin-Driver: a database of driver mutations in protein kinases

Franco L. Simonetti¹, Cristian Tornador², Nuria Nabau-Moretó³, Miguel A. Molina-Vila⁴ and Cristina Marino-Buslje^{1,*}

¹Fundación Instituto Leloir, Av. Patricias Argentinas 435. C1405BWE, Buenos Aires, Argentina, ²Pompeu Fabra University (UPF), Dept. de Tecnologies de la Informació i les Comunicacions. Tànger 122-140 08018, Barcelona, Spain, ³Computational Genomics Laboratory, Genetics Department, Institut de Biologia Universitat de Barcelona (IBUB), Facultat de Biologia, Av Diagonal 645 and ⁴Breakthrough Cancer Research Unit, Dexeus University Hospital, Sabino Arana 5-19, Barcelona, Spain.

*Corresponding author: Tel: +054 11 5238-7500; Fax: +054 11 5238-7501; Email: cmb@leloir.org.ar

Citation details: Simonetti,F.L., Tornador,C., Nabau-Moretó,N., *et al.* Kin-Driver: a database of driver mutations in protein kinases. *Database* (2014) Vol. 2014: article ID bau104; doi:10.1093/database/bau104

Received 26 June 2014; Revised 15 September 2014; Accepted 6 October 2014

Abstract

Somatic mutations in protein kinases (PKs) are frequent driver events in many human tumors, while germ-line mutations are associated with hereditary diseases. Here we present Kin-driver, the first database that compiles driver mutations in PKs with experimental evidence demonstrating their functional role. Kin-driver is a manual expert-curated database that pays special attention to activating mutations (AMs) and can serve as a validation set to develop new generation tools focused on the prediction of gain-of-function driver mutations. It also offers an easy and intuitive environment to facilitate the visualization and analysis of mutations in PKs. Because all mutations are mapped onto a multiple sequence alignment, analogue positions between kinases can be identified and tentative new mutations can be proposed for studying by transferring annotation. Finally, our database can also be of use to clinical and translational laboratories, helping them to identify uncommon AMs that can correlate with response to new antitumor drugs. The website was developed using PHP and JavaScript, which are supported by all major browsers; the database was built using MySQL server. Kin-driver is available at: <http://kin-driver.leloir.org.ar/>

Introduction

Cancer arises due to somatic mutations that result in a growth advantage for the tumor cells. These mutations are known as ‘drivers’ and can be divided into two groups: (i) ‘loss-of-function’ mutations, which inactivate tumor

suppressor genes (from here on ‘inactivating mutations’) and (ii) ‘activating’ or ‘gain-of-function’ mutations that transform proto-oncogenes into oncogenes. Somatic mutations in protein kinases (PKs) are frequent driver events in

many human tumor types and functionally relevant germline mutations are associated with hereditary disorders.

Clinical laboratories worldwide are analysing thousands of human tumor samples, looking for activating mutations (AMs) in certain PKs—such as EGFR, HER2 or BRAF—that correlate with good responses to new generations of antitumor drugs that are kinase inhibitors. Mutations either new or not functionally characterized are often found. In addition, whole-genomic sequencing of human malignancies and other diseases is identifying thousands of changes in PKs, but most of them are likely to be passenger mutations or even polymorphisms. Discriminating driver mutations in PKs is a significant challenge that is hampered by the fact that there are no curated sets of true driver and passenger alterations. The extent of this challenge was evidenced when three state-of-the-art methods, namely MutationAssessor (1), TransFITC (2) and FATHMM (3), were fed with well-established, tumor-associated AMs of PKs and failed to predict them as high impact or disease related (4). Therefore, it is uncertain that the current tools, which are generally based on conservation calculations, can be trusted to screen whole-genome sequencing data in search of driver mutations in PKs. New methods need to be developed and unambiguously assessed datasets of driver mutations are required to train and test them.

Methods

Mutation recruitment

Recruitment procedure is described by Molina-Vila *et al.* (4). Briefly, in the case of proto-oncogenic kinases, abstracts and titles of PubMed manuscripts were mined with the kinase name, plus words ‘activating’, ‘gain of function’ or ‘constitutive activation’. For tumor suppressor kinases, the words ‘inactivating’ and ‘loss of function’ were used.

Furthermore, all UniProt entries for human kinases were mined for the same keywords to identify new variants. The references were manually checked to confirm its status.

For each annotated mutation, all samples with that mutation were retrieved from COSMIC using the Biomart perl API.

MSA construction

Human STK and TKs domains were obtained from Pfam families PF00069 and PF07714, respectively. To account for classification problems in Pfam families, some sequences incorrectly classified as TK were moved from this alignment to the corresponding one and realigned with

T-coffee (5). For each MSA, a sequence logo was calculated using seq2logo (6).

Mutation relative frequency calculation

A relative frequency was computationally calculated for all mutations of the 518 PKs of the COSMIC database release 70 (7) as the frequency of mutation in COSMIC for that gene times 1000 over the total number of tumor samples sequenced for that gene.

All mutations with a relative frequency above 2 (0.2%) were then checked in PubMed by introducing the name of the mutation (e.g. P267R) and added to the dataset if they were found to have functional effects. EGFR mutations conferring a response rate to erlotinib higher than 50%, according to the EGFR somatic mutations database (<http://www.somaticmutations-egfr.info/>), were also added.

Results

Kin-Driver database offers a comprehensive set of 560 primary AMs in the kinase and justamembrane (JM) domains of 39 PKs and 83 inactivating mutations in 5 kinases compiled by a two-step systematic search for each of the 518 PKs present in the ‘complete kinase’ study of the COSMIC database (7) (release 70). Only primary mutations with experimental evidence demonstrating their activating/inactivating role were included.

Kin-Driver is a MySQL relational database offering structural and sequence data cross-referenced with COSMIC and with our set of curated mutations. It also provides the frequencies of these mutations in actual tumor samples. The CosmicMart service is used to fetch the data, so frequencies for new mutations can easily be added and data are kept up to date with the periodic COSMIC releases.

Our database can be interrogated by protein name, gene name or keyword, amino acid position or specific mutation name (i.e. T790M). Range or specific mutations can also be used to look for driver mutations in other PKs in equivalent positions (see later). Finally, the database can be browsed by PK name, domain, tissue or type of histology, and these last two attributes obtained from the corresponding mutated samples are available in the COSMIC database.

Each individual mutation in Kin-Driver is displayed with its validation status (‘activating’, ‘inactivating’ or ‘unknown’), the mutation type (missense, insertion, deletion, nonsense, frameshift or indel), its absolute and relative frequencies in human tumors and the PubMed reference describing that particular mutation as activating/inactivating. Mutations can also be visualized in a multiple

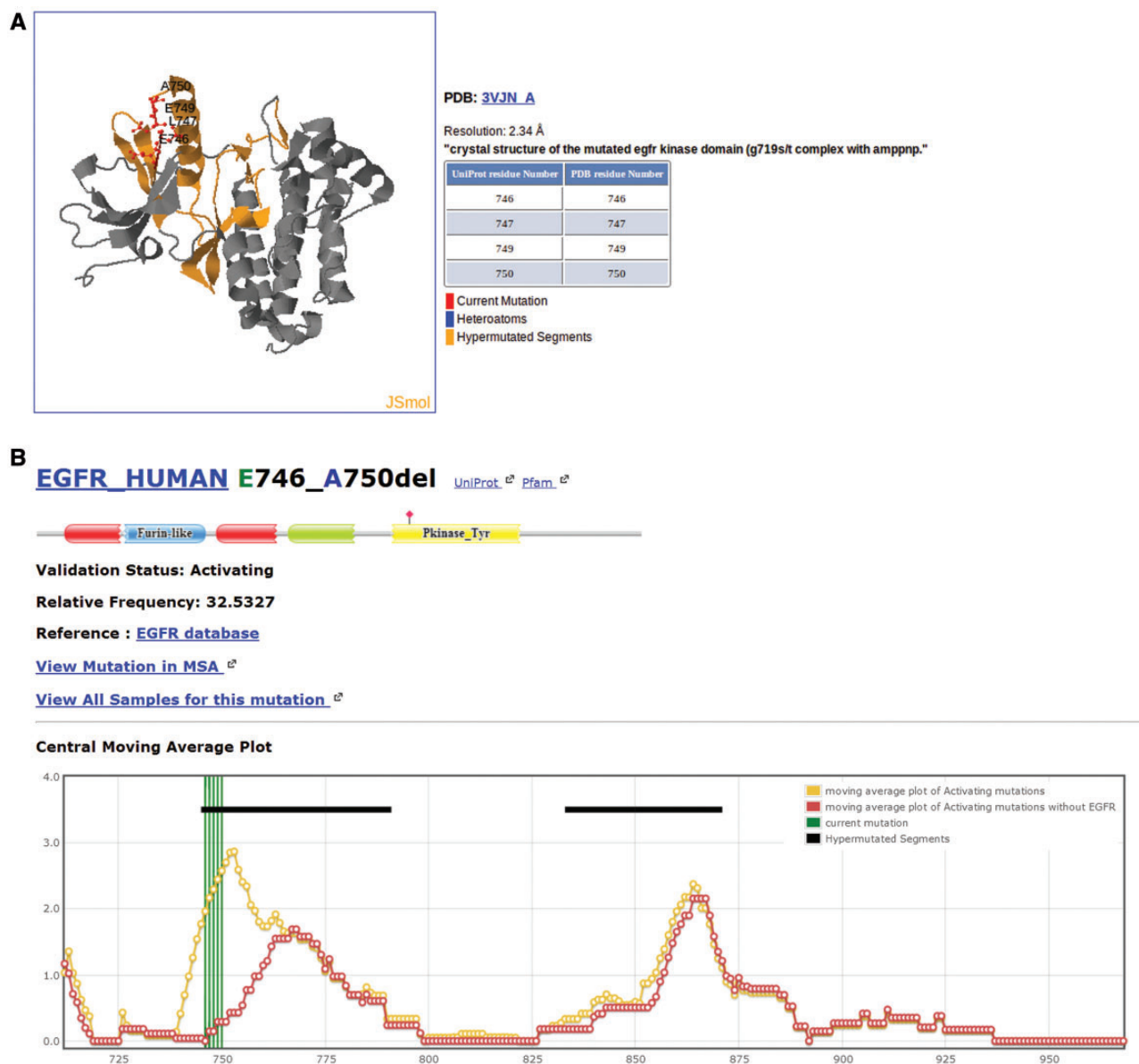


Figure 2. Snapshot of the Kin-Driver EGFR E746-A750del output. (A) Mutation EGFR E746-A750del mapped onto the structure of the selected human EGFR (pdb code 1M14). Other structures can be chosen for mapping. (B) Location of the mutation in a cmap plot ($n = 13$).

browsing can be useful to clinical laboratories; when confronted with a tumor sample, they can use Kin-Driver to find the genes affected by actionable mutations in this particular type of neoplasia, and they can subsequently direct their efforts to the analysis of those genes.

Although mutational data on PKs are currently cataloged in several databases (7, 12–14), they do not discriminate passengers from drivers. Kin-Driver is unique in offering a curated functional annotation, supported by experimental data. Also, the possibility of transferring the mutation to equivalent positions in homologous proteins through an MSA and the mapping into the desired protein structure with the Jsmol app (15) are unique features

of our database. In contrast, COSMIC (7) is the most comprehensive compilation of somatic mutations, but no functional status is given, no equivalences between PKs can be inferred, nor the position in the structure is shown. ProKinO (12) provides a kinase ontology and integrates basic mutational data (as recruited from COSMIC) with other types of data, but they do not offer functional or structural information. MoKCa and Canpredict (13, 14) offer a prediction of ‘cancer-associated’ mutations, but it is not based on experimental evidence and the pictures of the mutations in the protein structures are static. Finally, KinMutBase (16) has no information on activation status and is outdated.

Conclusions

To the best of our knowledge, Kin-Driver is the first comprehensive curated dataset of driver mutations in human cancer. It compiles data otherwise disseminated in several databases that offer poor functional information and in hundreds of articles describing the effects of one or a few mutations each. Our database can be used as a gold standard to develop and validate new bioinformatics methods to predict driver mutations. Second, it sets an easy and intuitive environment for the visualization and analysis of mutations in PKs. And third, it can help researchers and clinicians to recognize relevant mutations in human malignancies.

Conflict of interest. None declared.

References

1. Reva, B., Antipin, Y. and Sander, C. (2011) Predicting the functional impact of protein mutations: application to cancer genomics. *Nucleic Acids Res.*, 39, e118.
2. Gonzalez-Perez, A., Deu-Pons, J. and Lopez-Bigas, N. (2012) Improving the prediction of the functional impact of cancer mutations by baseline tolerance transformation. *Genome Med.*, 4, 89.
3. Shihab, H.A., Gough, J., Cooper, D.N. et al. (2013) Predicting the functional, molecular, and phenotypic consequences of amino acid substitutions using hidden Markov models. *Hum. Mutat.*, 34, 57–65.
4. Molina-Vila, M.A., Nabau-Moretó, N., Tornador, C. et al. (2014) Activating mutations cluster in the “molecular brake” regions of protein kinases and do not associate with conserved or catalytic residues. *Hum. Mutat.*, 35, 318–328.
5. Notredame, C., Higgins, D. and Heringa, J. et al. T-Coffee: a novel method for fast and accurate multiple sequence alignment. *J. Mol. Biol.*, 302, 205–217.
6. Thomsen, M.C.F. and Nielsen, M. (2012) Seq2Logo: a method for construction and visualization of amino acid binding motifs and sequence profiles including sequence weighting, pseudo counts and two-sided representation of amino acid enrichment and depletion. *Nucleic Acids Res.* first published online May 25, 2012 doi:10.1093/nar/gks469
7. Forbes, S.A., Bindal, N., Bamford, S. et al. (2011) COSMIC: mining complete cancer genomes in the catalogue of somatic mutations in cancer. *Nucleic Acids Res.*, 39, D945–D950.
8. Punta, M., Coghill, P.C., Eberhardt, R.Y. et al. (2012) The Pfam protein families database. *Nucleic Acids Res.*, 40, D290–D301.
9. Consortium, T.U. (2012) Reorganizing the protein space at the Universal Protein Resource (UniProt). *Nucleic Acids Res.*, 40, D71–D75.
10. Hamburg, M.A. and Collins, F.S. (2010) The path to personalized medicine. *N. Engl. J. Med.*, 363, 301–304.
11. Kelloff, G.J. and Sigman, C.C. (2012) Cancer biomarkers: selecting the right drug for the right patient. *Nat. Rev. Drug Discov.*, 11, 201–214.
12. Gosal, G., Kochut, K.J. and Kannan, N. (2011) ProKinO: an ontology for integrative analysis of protein kinases in cancer. *PLoS One*, 6, e28782.
13. Richardson, C.J., Gao, Q., Mitsopoulous, C. et al. (2009) MoKCa database—mutations of kinases in cancer. *Nucleic Acids Res.* 37(suppl. 1); D824–D831.
14. Kaminker, J.S., Zhang, Y., Watanabe, C. et al. (2007) CanPredict: a computational tool for predicting cancer-associated missense mutations. *Nucleic Acids Res.*, 35(suppl. 2); W595–W598.
15. Hanson, R.M., Prilusky, J., Renjian, Z. et al. (2013) JSmol and the Next-Generation Web-Based Representation of 3D Molecular Structure as Applied to Proteopedia. *Isr. J. Chem.*, 53, 207–216.
16. Ortutay, C., Väliaho, J., Stenberg, K. et al. KinMutBase: a registry of disease-causing mutations in protein kinase domains. *Hum. Mutat.* 25, 435–442.