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# Pb(II) binding to humic substances: an equilibrium and spectroscopic study

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1	Pb(II) binding to humic substances: an equilibrium and
2	spectroscopic study
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8	BRIEF: Pb(II) is proposed to bind to humic substances in two ways: a relatively strong multiligand
9	complex leading to humic aggregation, and a weaker 1:1 complex.
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### 16 ABSTRACT

17 The binding of Pb(II) to humic acids is studied through an approach combining equilibrium and 18 spectroscopic measurements. The methods employed are potentiometric and fluorometric titrations, 19 fluorescence excitation-emission matrices (EEM) and IR spectroscopy. Potentiometric titration curves 20 are analyzed using the NICA equations and an electrostatic model treating the humic particles as an 21 elastic polyelectrolyte network. EEMs are analyzed using parallel factor analysis, decomposing the 22 signal in its independent components and finding their dependence on Pb(II) activity. Potentiometric results are consistent with bimodal affinity distributions for Pb(II) binding, whereas fluorometric 23 24 titrations are explained by monomodal distributions. EEM analysis is consistent with three independent 25 components in the humic fluorescence response, which are assigned to moieties with different degree of 26 aromaticity. All three components show a similar quenching behavior upon Pb(II) binding, saturating at 27 relatively low Pb(II) concentrations. This is attributed to metal ion induced aggregation of humic 28 molecules, resulting in the interaction between the aromatic groups responsible for fluorescence; this is 29 also consistent with IR spectroscopy results. The observed behavior is interpreted considering that initial 30 metal binding (observed as strongly binding sites), correspond to bi- or multidentate complexation to 31 carboxylate groups, including binding between groups of different humic molecules, promoting 32 aggregation; further metal ions (observed as weakly binding sites) bind to single ligand groups.

33 KEYWORDS Potentiometric titration; fluorescence spectroscopy; IR spectroscopy; metal-humic
 34 binding; affinity spectra.

35

## 36 Introduction

Humic substances (HS) are important components of natural organic matter (NOM) in groundwater 37 and soils<sup>1,2</sup> HS interact with other chemical species in the natural environment, in particular binding 38 39 heavy metal cations.<sup>3–5</sup> Among different experimental studies, equilibrium binding data, usually titration curves, have been reported and modeled.<sup>3,6–8</sup> Recently, the electrostatic behavior of HS was treated 40 through the elastic polyelectrolyte network model (EPN)<sup>9</sup> which allows for expansion and shrinking as 41 the HS charge changes; it can be combined with the NICA<sup>10</sup> intrinsic isotherm, resulting in the NICA-42 EPN model which has been applied to proton binding to HS.<sup>9</sup> Fluorescence spectroscopy is an useful 43 tool in the study of metal binding to HS.<sup>4,11</sup> In the last years, excitation-emission matrix (EEM) 44 fluorescence spectroscopy with parallel factor analysis (PARAFAC) was successfully applied for the 45 identification and characterization of NOM.<sup>12-16</sup> For simple mixtures, PARAFAC components are 46 identified with individual fluorophores present; for HS, the identified components represent 47 48 fluorescence phenomena, which may be individual fluorophores or approximations to the effects of local 49 processes such as intramolecular charge transfer.

50 Even after many research efforts, the exact nature of HS and the way in which they interact with metal cations remains unclear. With the aim of gaining insight in this problem, the binding of Pb(II) to humic 51 52 acids (HA) is studied here by a combined analysis of titration and spectroscopic experiments. Equilibrium curves are obtained through potentiometric titrations and analyzed with the NICA-EPN 53 54 model for metal binding to obtain intrinsic binding parameters; also, fluorometric titrations are performed to compare the binding affinities. EEM measurements<sup>17</sup> with PARAFAC analysis<sup>13,18</sup> are 55 56 conducted to study the behavior of the different fluorophores with Pb(II) additions. Finally, IR-ATR 57 spectra are recorded to study the Pb-HA interactions.

58

# 59 Modeling of proton and metal binding data: the NICA-EPN model

60 Titration data was analyzed using the NICA isotherm for the intrinsic affinity<sup>7</sup> and the Elastic
61 Polyelectrolyte Network (EPN) submodel<sup>9</sup> for the electrostatic contribution. In the NICA-EPN model,

62 the amount bound of species *i* per unity of mass in the presence of species *j* is given by

63 
$$Q_{i} = g_{f} \sum_{k=1}^{M} \frac{n_{i,k}}{n_{i,k}} Q_{\max,k} \mathcal{G}_{i,k}(a_{1}^{\text{int}}, ..., a_{j}^{\text{int}}, ..., a_{n}^{\text{int}}) + (1 - g_{f}) \sum_{k=1}^{M} \frac{n_{i,k}}{n_{i,k}} Q_{\max,k} \mathcal{G}_{i,k}(a_{1}, ..., a_{j}, ..., a_{n}) + Q_{i,el}$$
(1)

64 where  $\mathcal{G}_{i,k}(a_i)$  is the fraction of type k sites occupied by species i, given in the NICA isotherm by

65 
$$\mathcal{G}_{i,k}(a_1, \dots, a_j, \dots, a_n) = \frac{\left(\bar{K}_{i,k}a_i\right)^{n_{i,k}}}{\sum_{j=1}^n \left(\bar{K}_{j,k}a_j\right)^{n_{j,k}}} \times \frac{\left[\sum_{j=1}^N \left(\bar{K}_{j,k}a_j\right)^{n_{j,k}}\right]^{p_k}}{1 + \left[\sum_{j=1}^N \left(\bar{K}_{j,k}a_j\right)^{n_{j,k}}\right]^{p_k}}$$
(2)

In eqs 1 and 2, *N* is the number of species binding to the HA, *M* the number of sites,  $Q_{\text{max}}$  the total number of sites per unity of mass,  $Q_{\text{max},k}$  the same for type *k* sites (assuming 1:1 binding),  $q_k$  the fraction of type *k* sites ( $Q_{\text{max},k} = q_k Q_{\text{max}}$ ),  $n_{i,k}$  the nonideality parameter for species *i* on type *k* sites, ( $n_{1,k}$  being for H<sup>+</sup>),  $p_k$  is the heterogeneity parameter of type *k* sites and  $\overline{K}_{i,k}$  is the average binding constant of species *i* on type *k* sites. Two site types are considered: low affinity (k = 1, attributed to carboxylic groups) and high affinity (k = 2, usually attributed to phenolic groups),<sup>7,19</sup> so M = 2, and for binding of a metal cation in the presence of protons N = 2.

73 As it is widely accepted, HS are actually composed of relatively small molecules and molecular 74 fragments (as opposed to a macromolecular/polymeric view) which are entangled and/or bound by 75 diverse forces, like hydrogen bonding, metal ion bridging, van der Waals interactions, etc. forming particles of supramolecular nature which can shrink and swell.<sup>20,21</sup> In the EPN model,<sup>9</sup> these HS 76 77 particles are treated, from a purely mechano-statistical point of view, as an elastic network which can 78 exchange ions and solvent with the surrounding solution; further, the particles are considered as divided 79 in two fractions: an inner fraction  $g_{f_2}$  which behaves as a gel in Donnan equilibrium with the bulk solution, and an external fraction  $1 - g_f$ . The humic binding sites outside the gel are in equilibrium with 80 the bulk solution, whereas those inside the gel are in equilibrium at the internal ionic activity,  $a_i^{\text{int}}$ , 81 82 which is given by

83 
$$a_j^{\text{int}} = a_j \exp\left(-F\psi_D / RT\right)$$
(3)

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84 where  $\psi_D$  is the Donnan potential,  $a_j$  is the activity in the bulk and *F*, *R* and *T* have their usual meaning. 85 For a 1:1 electrolyte,  $\psi_D$  results to be, from the electroneutrality condition,<sup>9</sup>

86 
$$\psi_D = \frac{RT}{F} \operatorname{arcsinh}\left(\frac{g_f Q \delta \varphi_2}{2I(1-\varphi_2)}\right)$$
 (4)

where *Q* is the net charge of the humic particle,  $\delta$  the density of the dry humic substance (following Dinar <sup>22</sup>  $\delta$  = 1.5 g cm<sup>-3</sup>), *I* the ionic strength, and  $\varphi_2$  is the volume fraction of the HS in the gel, obtained from the swelling equilibrium condition:<sup>9</sup>

90 
$$\frac{1}{v_1} \left[ \chi \varphi_2^2 + \varphi_2 + \ln(1 - \varphi_2) \right] + \frac{1}{v_2} \left( \varphi_2^{1/3} - \frac{2\varphi_2}{f_P} \right) + 2I \left[ 1 - \sqrt{1 + \left( \frac{g_f Q \delta \varphi_2}{2I(1 - \varphi_2)} \right)^2} \right] = 0$$
(5)

91 where  $v_1$  is the water molar volume,  $v_2$  is the average molar volume of a humic equivalent chain,  $\chi$  is a 92 Van Laar heat of mixing parameter and  $f_P$  is the Flory functionality,<sup>23</sup> which was set equal to 3.  $Q_{i,el}$  is 93 the excess amount of species *i* electrostatically bound in the gel phase:

94 
$$Q_{i,el} = \left(c_i^{\text{int}} - c_i\right) \frac{g_f}{\delta \varphi_2}$$
(6)

where  $c_i^{\text{int}}$  and  $c_i$  are the concentrations of species *i* in the gel and the bulk solution, respectively. The Davies equation was employed to calculate activity coefficients throughout; it was verified that the ionic strength inside the gel was always within its validity limit (I < 0.3 M).

98 As it is well known,<sup>7</sup> eq 2 reduces to a Langmuir-Freundlich isotherm for non competitive (H<sup>+</sup>) binding:

99 
$$\mathcal{G}_{H,k}(a_{\rm H}) = \frac{\left(\bar{K}_{\rm H,k}a_{\rm H}\right)^{m_{\rm H,k}}}{1 + \left(\bar{K}_{\rm H,k}a_{\rm H}\right)^{m_{\rm H,k}}}$$
(7)

100 where  $m_{H,k} = n_{H,k} p_k$ .

101 Also, conditional affinity distributions can be obtained using constrained regularization methods.<sup>24</sup> If the 102 proton affinity spectrum  $g(\log K_H)$  is known, a conditional metal affinity distribution  $f(\log K_M)$  can be 103 estimated solving numerically the equation (assuming uncorrelated proton and metal affinity spectra):<sup>24</sup>

104 
$$Q_{M} = Q_{\max} \int_{-\infty}^{\infty} \left[ \int_{-\infty}^{\infty} \frac{K_{M} a_{M}}{1 + K_{H} a_{H} + K_{M} a_{M}} g(\log K_{H}) d\log K_{H} \right] f(\log K_{M}) d\log K_{M}$$
(8)

105 where  $Q_M$  is the amount of metal bound.

106

# 107 Materials and methods

108 The HAs employed were a purified Fluka humic acid (FHA) and IHSS standard Elliot soil humic acid 109 (EHA); these HS have been characterized in the literature, as presented in Table S1, Supporting Information. Acid-base potentiometric titrations were performed following usual procedures<sup>25</sup> at 1 g L<sup>-1</sup> 110 111 HA concentration and different ionic strengths in NaClO<sub>4</sub>. Pb(II)-humic potentiometric titrations also followed known procedures<sup>10</sup> at pHs of 4.0 and 5.5, and ionic strengths (NaClO<sub>4</sub>) of 0.1 M and 0.02 M. 112 113 Fluorometric titrations were performed, at a HA concentration of 20 mg L<sup>-1</sup>. In order to enable 114 comparisons, binding curves for this concentration were calculated using the fitted model parameters; 115 some potentiometric measurements were also done, but no processed due to the low reliability of ISE measurements at low HA concentrations;<sup>26</sup> however, they reflected the same general tendency. 116 Fluorescence EEMs of 20 mg L<sup>-1</sup> solutions of HA alone and with increasing amounts of Pb(II) were 117 118 recorded. UV-Vis spectra were also recorded to check the onset of humic flocculation. IR-ATR spectroscopy measurements of HA-Pb complexes were performed following usual techniques,<sup>27,28</sup> using 119 120 the electrolyte spectrum as reference to obtain the difference spectra. Experimental details are given in 121 Appendix A, Supporting Information.

The data treatment was, briefly, as follows. First from acid-base titration data the humic charge as a function of pH was computed and fitted to the NICA-EPN model, using a constrained Levenberg-Marquardt algorithm,<sup>29</sup> obtaining the corresponding NICA parameters as well as the EPN ones, namely  $\chi$ ,  $g_f$  and  $v_2$ . Then, curves of Pb(II) binding as a function of Pb(II) activity were obtained from titration data, and also fitted to the NICA-EPN model, taking the proton binding and EPN parameters from the proton experiments.

128 Fluorometric titration data was converted to relative fluorescence quenching,  $\theta_F$ , defined as:

129 
$$\theta_F = \frac{I_F^0 - I_F}{I_F^0 - I_F^\infty}$$
(9)

Here,  $I_F^0$  is the fluorescence emission intensity in the absence of Pb(II),  $I_F^\infty$  is the intensity observed at maximum quenching and  $I_F$  is the intensity at each point. This intensity values can be taken as the intensity at the wavelength of maximum emission or as the integrated emission spectra. Both ways led to the same results. Fluorescence EEMs were converted to Raman units following Stedmon et al.,<sup>18</sup> processed with PARAFAC,<sup>12,13</sup> yielding the number of independent fluorescent phenomena, their absorption and emission spectra, and their relative concentrations (scores) in each sample. The data treatment procedures are detailed in Appendix B, Supporting information.

137

138 **Results and discussion** 

139

Potentiometric titrations. Acid-base titration curve fitting to the NICA-EPN model shows very good agreement (Figure S1, Supporting Information). The intrinsic NICA parameters for proton binding are collected in Table 1, falling in the ranges reported in the literature;<sup>19,30</sup> also included are the total sites ( $Q_{max}$ ) and the electrostatic parameters  $g_{f}$ ,  $\chi$  and  $v_2$ . The proton binding constants found for low affinity sites are close to typical values for isolated carboxylic groups. For the high affinity type sites our results are close to the p $K_a$  for phenol (9.8); it has been reported that in HS most phenolic OH groups are isolated<sup>31,32</sup> which is consistent with these results.

The electrostatic parameters  $g_f$  and  $\chi$  are similar to those reported previously;<sup>9</sup> the first one represents the part of the humic particle where the electrostatic potential is on average equal to  $\psi_D$ , being about half of the binding sites, whereas a positive value for  $\chi$  corresponds to a repulsive heat of mixing humicwater, which is expected because HS have a significant fraction of non polar groups. There are significant differences in  $v_2$ , quite low for EHA and high for FHA; this parameter is the molar volume of the equivalent chain segments in a network representing the humic particle, thus the results found would indicate that FHA is relatively less entangled whereas EHA would be highly entangled.

Proton binding	log	log	$Q_{\rm max}$	$q_1$	$q_2$	<i>m</i> <sub>H,1</sub>	<i>m</i> <sub>H,2</sub>	<b>g</b> <sub>f</sub>	χ	v <sub>2</sub> /
	$\overline{K}_{\mathrm{H,1}}$	$\overline{K}_{\mathrm{H,2}}$	mol kg <sup>-1</sup>							L mol <sup>-1</sup>
Elliot HA	4.42	9.43	5.0	0.52	0.48	0.51	0.20	0.48	0.47	1.7
Fluka HA	4.44	9.91	5.2	0.56	0.44	0.39	0.23	0.47	0.51	10.2
Pb(II) binding	$\log \overline{K}_{ m Pb,1}$	$\overline{K}_{ ext{Pb},2}$	<i>n</i> <sub>Pb,1</sub>	<i>n</i> <sub>Pb,2</sub>	<i>p</i> <sub>1</sub>	<i>p</i> <sub>2</sub>		$N_d^{a}$	$R^{2 b}$	RMSE <sup>c</sup>
Elliot HA	0.55	9.8	0.80	0.49	0.70	0.40		228	0.9990	0.058
Fluka HA	0.50	5.8	0.90	0.52	0.49	0.89		209	0.9993	0.039
<sup>a</sup> Total data points $\frac{b}{B^2} = 1$	$\Sigma(0)$	$(0)^{2}$	$\sqrt{\sum O^2} - ($	$(\nabla \alpha)^{\overline{2}}$	N ] c	PMSE -	$-\overline{\Sigma(0)}$	_0 )	$\frac{2}{N}$	

# 155 **Table 1.** NICA-EPN model fitting parameters

156 <sup>a</sup> Total data points. <sup>b</sup>  $R^2 = 1 - \sum (Q_i - Q_{i, calc})^2 / \left[ \sum Q_i^2 - \left( \sum Q_i \right)^2 / N_d \right]$ . <sup>c</sup> RMSE =  $\sqrt{\sum (Q_i - Q_{i, calc})^2 / N_d}$ 

157

Figure 1 shows the results for the HA-Pb titrations with good agreement; the generic NICA-Donnan 158 (ND)<sup>19</sup> predicted curves being also shown for comparison. The pH dependence is similar to that reported 159 by Milne et al.<sup>3</sup> and references therein. The influence of I has been much less studied; here it is observed 160 161 that the curves at different ionic strengths merge as Pb(II) activity increases; some literature studies show similar behavior: Cu binding to Suwannee River fulvic acid,<sup>33</sup> Am complexation with Gohy-573 162 humic acid,<sup>34</sup> and Cu complexation with Sable silt loam HA<sup>35</sup>. The NICA-EPN model predicts well this 163 dependence, whereas the generic ND curves are nearly parallel. Table 1 collects the Pb intrinsic binding 164 165 parameters and the fitting quality results for the full data set fitting (proton and metal curves.

Figure 2 shows the speciation of bound Pb(II) at pH = 5.5 and I = 0.1 M (other cases are similar), writing eq 1 in the form:

168 
$$Q_{\rm Pb} = Q_{\rm Pb,1}^{\rm int} \left( a_{\rm H}^{\rm int}, a_{\rm Pb}^{\rm int} \right) + Q_{\rm Pb,2}^{\rm int} \left( a_{\rm H}^{\rm int}, a_{\rm Pb}^{\rm int} \right) + Q_{\rm Pb,1}^{\rm ext} \left( a_{\rm H}, a_{\rm Pb} \right) + Q_{\rm Pb,2}^{\rm ext} \left( a_{\rm H}, a_{\rm Pb} \right) + Q_{\rm Pb,el}^{\rm ext}$$
(10)

169 where  $Q_{Pb,k}^{int} = g_f \left( n_{Pb,k} / n_{H,k} \right) Q_{max,k} \mathcal{G}_{Pb,k} \left( a_{Pb}^{int}, a_{H}^{int} \right)$  and so on. As it is observed, most Pb is bound to high 170 affinity sites in the activity range studied, specially for EHA. From  $a_{Pb} \approx 10^{-4}$  up the binding to low 171 affinity sites shows an increase. The electrostatic binding in the gel fraction (eq 6) is negligible in all 172 cases; note also that for FHA this contribution changes sign; this is due to inversion of the gel fraction 173 charge caused by the Pb loading.

There are several published studies of Pb binding to HS;<sup>3,36-42</sup> however, direct comparisons are 174 difficult because the different HS and conditions employed. The values of  $\overline{K}_{Pb,k}$  found here (with the 175 exception of high affinity sites for EHA) are within the ranges reported by Milne et al.,<sup>3</sup> Gustafsson,<sup>8,40</sup> 176 and Tipping.<sup>6</sup> Puy et al.<sup>37</sup> reported somewhat higher values of log  $\overline{K}_{Pb,1}$  (4.37) and log  $\overline{K}_{Pb,2}$  (7.57). The 177 high affinity constant for EHA found here is higher than the values reported by all these authors; in 178 179 some studies, Pb(II) binding to soil organic matter was found to be considerably higher than predicted 180 by current models, free Pb being about one order of magnitude lower than predicted (see Gustafsson et al.<sup>40</sup> and references therein). Thus, being EHA a soil HA, whereas FHA is originated from coal, the 181 difference in their  $\bar{K}_{Pb,2}$  values is consistent with the findings reported by Gustafsson et al.<sup>40</sup> 182

183

184 Fluorometric titrations. During titration of HA with Pb(II), fluorescent emission is quenched as 185 Pb(II) concentration increases, until a limiting emission is reached, which does not change as Pb(II) 186 concentration is further increased (Figure S2, Supporting Information). In all cases, the lowest emission intensity observed was taken as  $I_F^{\infty}$  in eq 9 to compute  $\theta_F$ . This quenching has been observed by a 187 number of researchers and for several cations.<sup>11,43</sup> Along the titrations, the absorbance spectra of the 188 189 HA-Pb solutions were checked. It was found to remain unchanged until the appearance of flocculation 190 and/or precipitation, were the absorbance started to decrease (Figure S3, Supporting Information); only 191 fluorescence measurements before this point were considered in the analysis, thus in all the results 192 presented here the UV spectra remained constant. Figure 3 compares the results for fluorometric 193 titrations with Pb binding curves calculated with the NICA-EPN model for EHA and FHA. The general

features are coincident with previous studies.<sup>3,44,45</sup> It is observed that the results are qualitatively similar 194 195 but not quantitatively coincident, showing the fluorometric response a tendency to saturation at pPb  $\sim$ 5, 196 which is not observed for the potentiometric curves. This behavior was also observed in preliminary 197 experiments conducted with other cations such as Cu(II) and Mn(II) (not shown). Similar comparisons were done previously by Saar and Weber<sup>43</sup> who measured binding of several cations to fulvic acids, 198 199 finding good correlation for Cu(II) but only poor agreement for Pb(II), and even poorer for Co(II) and Ni(II). Other studies<sup>11,46</sup> were instead focused on obtaining binding equilibrium constants directly from 200 201 quenching data.

202

**Excitation-Emission matrices.** The EEM plots for Pb(II) binding to HA show typical shapes observed for HS<sup>14,16,47</sup> (Figure S4, Supporting Information). The shape changes as Pb(II) concentration increases; the emission and excitation maxima shift towards lower wavelengths in both cases, but its intensity always decreases with increasing metal concentration. The changes in shape indicate that the EEMs are linearly independent, and consequently are analyzable using PARAFAC.

208 After PARAFAC processing changing the number of fluorophores,  $N_{f}$ , between 1 and 5 it was found, 209 in all cases, that  $N_f = 3$  gave an optimal fit of the model, revealed by a low value for residual standard 210 deviation,  $\sigma_R$ , and a high value for core consistency, CC, for that number (Figure S5, Supporting 211 Information). This result is consistent with that reported in literature for IHSS reference humic substances<sup>48</sup> and is also consistent with measurements of fluorescence lifetime of HS, showing three 212 distinct values.<sup>49</sup> Figure 4 shows contour maps of the different fluorophores for EHA and FHA, which 213 are similar to others HA.<sup>13,48,50</sup> The components have been named starting from the emission maximum 214 215 with the higher wavelength, the spectra showing only a small dependence with pH and ionic strength; 216 Table S2 (Supporting Information) collects the average peak emission and excitation wavelengths for FHA and EHA. The maxima location is related to the fluorophore structure:<sup>18,51</sup> the higher the degree of 217 218 conjugation, the higher the excitation wavelength; this suggests that the A component is the more

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219 aromatic, with aromaticity decreasing in the B and C components. The component C is also observed in fulvic acids.<sup>52</sup> The three have spectral characteristics consistent with the operationally defined humic 220 acids by Chen et al.<sup>53</sup> based on pyrolysis results.<sup>54</sup> The quenching efficiency for these components lies in 221 the same order: the more aromatic is more efficiently quenched. In similar studies with dissolved 222 organic matter and aquatic HS, 13 fluorophores were reported,<sup>55</sup> including aminoacid-like components 223 with emission  $\lambda_{em max} \leq 350$  nm; one of such fluorophores (tyrosine-like) was found<sup>56</sup> to give high 224 225 deviation from linearity in inner filter effects for  $A_{254}$  (the absorbance at 254 nm ) > 0.3, all the rest showed low deviations up to  $A_{254} \cong 1.0$ . In this study, no such fluorophore is found; moreover, the 226 227 fluorescence titrations are treated as relative values (eq 9) and the HA concentration and the full 228 absorbance spectra are constant in the measurements, thus the correction should be the same for all data, 229 even being nonlinear. Consequently, the PARAFAC resulting spectra were corrected for inner filter 230 effects. In view of the above considerations, the results shown in Figure 5 should be considered as 231 semiquantitative.

Figure 5 present typical results for the scores (concentrations) of each fluorophore as a function of total Pb(II) concentration,  $c^{0}_{Pb}$ . Other results are presented in Figures S6 and S7 (Supporting Information). It is seen that component A has the higher scores and shows greater quenching, followed in both aspects by B and then C. Usually, the behavior is similar for all three components; in some cases, the component C shows no observable quenching. No essential differences are found between components; similar results were found by Yamashita and Jaffé.<sup>47</sup>

238

**IR-ATR spectra.** In Figure 6 the spectra of EHA in the absence and presence of Pb(II) are shown; as detailed in Appendix B (Supporting Information) the 4000-3000 cm<sup>-1</sup> was recorded with lower sensitivity due to water band saturation, thus presenting higher noise. The bare EHA spectrum is typical of HS,<sup>2</sup> having relatively broad bands resulting from the multiple chemical environments of the absorbing groups. Here, we will discuss mainly the carboxylic C=O stretching at 1720 cm<sup>-1</sup> and the carboxylate bands at 1585 ( $v_a$ , asymmetric stretching) and 1380 cm<sup>-1</sup> ( $v_s$ , symmetric);<sup>57,58</sup> these bands

are marked in the figure. Also, a sharp band is observed at *ca*. 1105 cm<sup>-1</sup>, attributed to C-O stretching of
polysaccharide-like substances<sup>59</sup> and a weak band at *ca*. 1260 cm<sup>-1</sup> attributable to phenol groups.<sup>60</sup>
Finally, the OH stretching region shows a negative band due to differences in OH absorption between
the HA solution and the electrolyte reference.

249 Two spectra with Pb in different concentrations are shown; using NICA-EPN parameters, the metal loadings are estimated as 0.36 and 2.4 mol kg<sup>-1</sup>, thus these can be considered as representative of low 250 251 and high loadings, respectively. In the low loading case, the carboxylic band decreases, whereas the carboxylate bands increase and shift to 1564 and 1386 cm<sup>-1</sup>, respectively, indicating that carboxylate 252 253 groups are involved in binding; the other bands mentioned do not change, indicating that OH groups are 254 not involved. Under high loading, those changes appear enhanced, except for  $v_s$ , where a sharp increase is found and new band arises at 1350 cm<sup>-1</sup>; the sharp increase has been observed in other cases,<sup>28,59</sup> 255 256 whereas the new band indicates a different carboxylate binding mode. According to Nakamoto,<sup>58</sup> the 257 difference  $\Delta = v_a - v_s$  is indicative of the binding mode, a decrease being attributable to a chelate (metal 258 bound to both O atoms) or a bridging complex (one metal bound to each O atom); here  $\Delta$  decreases from 205 to 178 cm<sup>-1</sup> at low loading, which is likely due to chelate formation, whereas the new band at high 259 loading shows  $\Delta = 208 \text{ cm}^{-1}$ , a small increase which could be indicative of monodentate binding. The C-260 261 O bands again do not show changes, whereas in the OH region a small increase is observed, which could 262 be attributable to OH<sup>-</sup> ions coordinating the Pb ions; involvement of HA OH groups in binding would 263 result in a decrease due to deprotonation. Thus, we conclude that Pb binding to the HA proceeds via the 264 carboxylate groups in different modes depending on metal loading, without appreciable involvement of 265 OH groups.

266

Affinity spectra. Figure 7 shows the affinity spectrum for Pb(II) binding to EHA corresponding to the
 NICA fitting results shown in Table 1, compared with the conditional spectra for fluorometric
 measurements obtained solving numerically eq 8; similar results are obtained for FHA (Figure S8,
 Supporting Information); Figure S9 (Supporting Information) shows the NICA affinity distributions for
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protons. These are bimodal distributions with peaks at log  $K_{\rm Pb}$  values ~0.5 for the low affinity sites, 271 being for high affinity 5.8 for FHA and 9.8 for EHA. On the other hand, fluorometric results show 272 273 monomodal distributions peaking at  $\sim 5$  for FHA and  $\sim 6-7$  for EHA, roughly coincident with the higher 274 affinity peak from the potentiometric experiments. This suggests (considering also IR results) that 275 fluorescent quenching is sensing only high affinity sites for Pb(II), whereas potentiometric analysis finds 276 two types. Comparison of potentiometric and fluorometric measurements has been conducted by several researchers,<sup>4,43,61,62</sup> usually finding that metal binding quenches the HS fluorescence; however, it was 277 noted that both techniques do not sense the same complexation phenomena:<sup>4</sup> for Cu binding to fulvic 278 acid, it was observed that the two measurements agreed only at low levels of copper loading,<sup>61</sup> which is 279 280 consistent with our findings. Also, Saar and Weber found poor agreement between potentiometric and fluorometric measurements for Pb.<sup>43</sup> These binding sites are expected to be, given the IR results. 281 282 carboxylates; this is also consistent with the fact that carboxylate groups are known to form stable complexes with Pb(II)<sup>63</sup> whereas phenol groups are not expected to bind Pb(II) at the present pH 283 values.<sup>64</sup> There are a number of studies which seek to obtain binding constants between metal cations 284 285 and humic substances from fluorescence measurements, based on the assumptions of 1:1 binding and that fluorescence was detecting all types of binding sites.<sup>46</sup> However, as already observed by Senesi,<sup>4</sup> the 286 287 two techniques are not equivalent and, as found here, fluorometry senses only the strongly binding sites. Therefore, the binding constants obtained exclusively through fluorometry should be considered valid 288 289 only in the context of this technique.

290

Fluorescence quenching. Quenching of fluorescence can be caused by several mechanisms, mainly dynamic (collisional) and static (complex formation).<sup>51</sup> For single fluorophores, the Stern-Volmer plot, a graphic of  $I_F^0/I_F$  as a function of the total quencher concentration, is useful to reveal the type of quenching taking place.<sup>51</sup> For HS, their complex nature introduces difficulties because the observed emission is the result of the combination of several fluorescence phenomena. Here, we treat the PARAFAC components as independent fluorophores to analyze the quenching behavior. In most cases,

297 the Stern-Volmer plots show an upward curvature (Figure S10, Supporting Information); for simple cases, this is indicative of mixed static-dynamic quenching;<sup>51</sup> however dynamic (collisional) quenching 298 is usually important at concentrations several orders of magnitude higher.<sup>65</sup> Furthermore, attempts to 299 deduce quenching parameters by usual methods<sup>51</sup> from these plots lead to unreasonably high values for 300 301 the dynamic quenching constant. Thus we conclude that static quenching is present in this case, as found before for other metal-HS systems.<sup>4</sup> The effect could be due to different causes;<sup>51</sup> for HS it has been 302 attributed to intersystem crossing;<sup>4</sup> that is, enhanced transference from the singlet excited state to the 303 triplet ground state. Aromatic carboxylic acids show usually this type of quenching.<sup>4,46,61,66</sup> However, in 304 the case of HS this is not necessarily true: some studies<sup>66</sup> show that complexes of model compounds 305 306 with heavy metal ions (such as Tb(III)) show quenching whereas binding of the same cation to HS does 307 not. Furthermore, a molecular quenching mechanism such as intersystem crossing would require the 308 carboxylic groups to be directly bound to the fluorescent aromatic groups, but this is not necessarily the 309 case: on one hand not all the fluorescent aromatic groups would have a carboxylic substituent, and on the other not every carboxylate would be attached to a fluorophore group.<sup>1,2,4</sup> Thus, we propose as an 310 311 alternative quenching mechanism the aggregation of humic molecules induced by Pb(II) binding. Quenching by dimerization of aromatic and highly conjugated substances is well known<sup>67,68</sup> and shows 312 313 increasing efficiency at higher wavelengths, as it is observed here; on the other hand, aggregation of HS by cation binding is well known.<sup>69</sup> Starting from this assumption, the quenching can be rationalized in 314 315 the following way, considering the process schematically depicted in Figure 8. When Pb(II) is added, it 316 first binds in bidentate form promoting aggregation by bridging two humic molecules; one or several 317 metal ions are expected to participate in the bridging. This process corresponds to the high affinity part 318 of the spectrum, which is thus assumed to correspond to bidentate complexation, but not necessarily all 319 Pb(II) bound in this way participate in bridging. Further increase in Pb(II) concentration results in singly 320 coordinated binding to carboxylate groups, corresponding to the low affinity part.

321 The process for the high affinity sites, which causes quenching, can be written as

$$322 \qquad 2HA + nPb^{2+} \Longrightarrow (HA)_2Pb_n \tag{11}$$

323 where *n* is the average number of  $Pb^{2+}$  ions bridging humic molecules, considering the HA 324 heterogeneity. The reaction will have a conditional equilibrium constant given by

325 
$$K = \frac{[(HA)_2 Pb_n]}{[HA]^2 c_{Pb}^n}$$
(12)

where  $c_{Pb}$  is the free Pb(II) concentration. Now, if it is assumed that free HA molecules show emission whereas those bound do not, it can be written for the total fluorophore concentration  $F_0$ :<sup>51</sup>

$$328 F_0 = F + 2C (13)$$

where *F* is the concentration of free fluorophores and *C* is the complexed fluorophores (not emitting) concentration. Identifying *F* with [HA] and *C* with  $[HA_2Pb_n]$ , it can be found from eqs 12 and 13, and taking into account that fluorescent intensity is proportional to fluorophore concentration, that

332 
$$\frac{I_F^0}{I_F} = \frac{F_0}{F} = \frac{1 + \sqrt{1 + 8KF_0 c_{Pb}^n}}{2}$$
(14)

333 Figure 9 shows the fit of FHA fluorophore scores (in Stern-Volmer form) to eq 14 using a common n 334 value for all curves, and different values for the product  $KF_0$ . The free Pb(II) concentration are obtained 335 from the potentiometric results with the Davies equation for activity coefficients. A good fitting is found 336 in general, the C component in some cases show a less satisfactory fitting, but it can be due to its lower 337 intensity, resulting in higher errors. EHA data shows similar fitting (Figure S11, Supporting Information). All FHA curves are fit with n = 1.6, whereas EHA fits with n = 1.3. Unfortunately, K and 338  $F_0$  cannot be separated, because the many unknown factors; the resulting values for their product are 339 340 presented in Table S3, Supporting Information.

Thus, we conclude that fluorescence quenching is consistent with metal-induced aggregation of HS; this aggregation proceeds at a molecular scale, to be distinguished of precipitation, however, more experimental information is needed to confirm this mechanism. Other spectroscopic studies are consistent with the present findings. Xia et al.<sup>70</sup> studied by X-ray absorption the interaction of Pb(II) with a humic extract from a silt loam soil suggesting the presence of two C atoms in the second ACS Paragon Plus Environment

coordination shell, which would indicate bidentate binding. A study by EPR measurements combined
 with DFT analysis is consistent with a structure of Pb (II) bound to two carboxylates.<sup>71</sup>

Concluding, the NICA-EPN model here introduced describes adequately the binding of proton and Pb(II) to humic acids; fluorescence quenching has a different dependence with Pb concentration compared with potentiometric titrations: it is proposed that the addition of Pb(II) causes dimerization (or aggregation) of humic molecules. Thus, fluorescent quenching senses multiligand Pb(II) binding (which, from IR results, should be a chelate type complex), and the second site type (with lower affinity) observed in potentiometric measurements is proposed be due to 1:1 binding.

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**Supporting Information Available.** Additional figures and tables. Appendix A details the experimental procedures, and Appendix B does the same for data analysis. This material is available free of charge via the Internet at <u>http://pubs.acs.org</u>.

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### **363 FIGURE CAPTIONS**

Figure 1. Pb(II) binding curves at different pH and ionic strengths for Elliot soil humic acid (a) and purified Fluka humic acid (b). The dotted lines are the NICA-Donnan predicted curves with the generic recommended parameters of Milne et al.<sup>3</sup> 1 g  $L^{-1}$  HA in NaClO<sub>4</sub> at the ionic strengths shown.

367

Figure 2. Speciation of Pb(II) binding at pH = 5.5 and I = 0.1 M in NaClO<sub>4</sub> (1 g L<sup>-1</sup> HA) for Elliot soil humic acid (a) and purified Fluka humic acid (b). The closed symbols (total and high affinity binding) correspond to the left side scale, and open symbols (low affinity and electrostatic binding) to the right side scale.

372

Figure 3. Amount of metal bound (closed symbols) and relative fluorescence quenching (open symbols) for simultaneous potentiometric and fluorescence titration of EHA (a) and FHA (b) with Pb(II). 20 mg  $L^{-1}$  HA in NaClO<sub>4</sub> at the ionic strengths shown.

376

Figure 4. Fluorescence components as obtained from PARAFAC. (Top) Elliot soil humic acid, pH = 4.0; I = 0.02 M; (bottom) Purified Fluka humic acid, pH = 5.5, I = 0.1 M. Blue is zero intensity, red is maximum (arbitrary scale). 20 mg L<sup>-1</sup> HA in NaClO<sub>4</sub>.

380

Figure 5. Scores of the fluorescence components (shown in Figure 3) deduced form PARAFAC as a function of total Pb(II) concentration, for 20 mg L<sup>-1</sup> FHA at pH = 5.5, I = 0.02 M (NaClO<sub>4</sub>).

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Figure 6. ATR-IR spectra of Elliot soil HA (0.28 g L<sup>-1</sup>) in the absence and in the presence of Pb(II) at different concentrations. pH = 5.5, I = 0.1 M (NaClO<sub>4</sub>). See text for details.

- 387 Figure 7. Affinity spectrum found for EHA by NICA fitting (solid lines) and conditional spectra found
- 388 from analysis based on eq 8 of fluorometric quenching experiments (dashed lines).

390	Figure 8. Simplified scheme of the humic-Pb(II) interaction. In the first stage (high affinity sites) the
391	Pb <sup>2+</sup> ions promote aggregation of humic molecules through bridging; the Pb ions bind to carboxylate
392	groups in chelate form as indicated by IR results. In the second stage, 1:1 binding takes place yielding
393	the low affinity sites; mainly single bond Pb binding is assumed, as suggested by IR results.
394	
395	Figure 9. Stern-Volmer plots as a function of free Pb(II) concentrations for FHA fluorophores (symbols).
396	Lines show the fitting to eq 14.
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Figure 1 71x63mm (300 x 300 DPI)



Figure 2 70x61mm (300 x 300 DPI)



Figure 3 72x57mm (300 x 300 DPI)



Figure 4 50x30mm (300 x 300 DPI)



Figure 5 38x19mm (300 x 300 DPI)



Figure 6 58x39mm (300 x 300 DPI)



Figure 7 46x25mm (300 x 300 DPI)



Figure 8 101x113mm (300 x 300 DPI)



Figure 9 75x51mm (300 x 300 DPI)



TOC Art 29x12mm (300 x 300 DPI)