

14q13 Distal Microdeletion Encompassing NKX2-1 and PAX9: Patient Report and Refinement of the Associated Phenotype

Mattia Gentile,¹* Delia De Mattia,² Angela Pansini,¹ Federico Schettini,² Antonia Lucia Buonadonna,¹ Manuela Capozza,² Romina Ficarella,¹ and Nicola Laforgia²

¹Department of Medical Genetics, Hospital Di Venere - ASL BARI, Bari, Italy

²Neonatology and NICU, Department of Biochemical Sciences and Human Oncology, University of Bari, Bari, Italy

Manuscript Received: 12 January 2016; Manuscript Accepted: 15 April 2016

Chromosome 14q11-q22 deletion syndrome (OMIM 613457) is a rare genomic disorder whose associated phenotype is heterogeneous, depending on the size, and, mostly, on the deleted region. We report the clinical and molecular characterization of a female newborn, whose phenotype was characterized by poor growth, dysmorphic facial features, subclinical hypothyroidism, and mild reduction of CD3CD8 Lymphocytes with increased CD4/CD8 ratio. By array-CGH, we identified a 4.08 de novo interstitial deletion of the 14q13.2q21.1 region, which includes 16 OMIM genes. Our patient phenotype is compared with other published cases, for a better classification of the 14q11-q22 deletion syndrome. We demonstrated that the 14q13.2q21.1 deletion, which encompasses NKX2-1, but not FOXG1 gene and HPE8 region, identifies a well defined, more benign, microdeletion syndrome. This report confirms that an early identification with accurate characterization of the genomic disorders is of great relevance, enabling proper genetic counseling of the reproductive risk, as well as disease prognosis, and patient management. © 2016 Wiley Periodicals, Inc.

Key words: chromosome 14 deletion; 14q13 microdeletion; *NKX2-1*; *PAX9*; CAHTP syndrome; genotype–phenotype

INTRODUCTION

Chromosome 14q11-q22 deletion syndrome (OMIM 613457) is a rare genomic disorder, caused by an interstitial deletion of the long arm of chromosome 14 with variable breakpoints. The deletion has been described as a contiguous gene syndrome in several reports [Shapira et al., 1994; Kamnasaran et al., 2001; Caliebe et al., 2011; Fonseca et al., 2012; Piccione et al., 2012; Santen et al., 2012]: *FOXG1*, *NKX2-1*, and *PAX9* have been identified as the candidate dosage-sensitive key genes of clinical significance, while the role of other genes (i.e., *GARNL1/TULIP1*, *NKX2-8*, *SLC25A21*) is still debated.

Based on literature reports, no recurrent breakpoints are present: the 14q deletion associated phenotype seems heterogeneous,

How to Cite this Article:

Gentile M, De Mattia D, Pansini A, Schettini F, Buonadonna AL, Capozza M, Ficarella R, Laforgia N. 2016. 14q13 distal microdeletion encompassing *NKX2-1* and *PAX9*: Patient report and refinement of the associated phenotype.

Am J Med Genet Part A 170A:1884-1888.

depending on the size, and, mostly, on the deleted region. When the deletion is proximal [chr14(GRCh37):28,000,000–35,000,000], the phenotype is severely compromised with intellectual disability, CNS malformations (i.e., agenesis of the corpus callosum, CCA), and poor prognosis. The distal 14q deletion [chr14 (GRCh37):35,000,000–40,000,000] seems associated with a milder phenotype: intelligence is normal or slightly impaired; distinctive clinical features are choreoathetosis and congenital hypothyroid-ism, with or without pulmonary dysfunction (CAHTP, OMIM 610978) [Santen et al., 2012].

We report a new patient with 14q13 distal microdeletion syndrome. The chromosome breakpoint was estimated as a 4.08 Mb sized deletion [arr14q13.2q21.1 (35,443,407–39,527,387) × 1] by array CGH, performed soon after birth because of craniofacial dysmorphisms. Our case confirms a close relationships between clinical findings and 14q deleted region, underlining the need of early genetic diagnosis for better counseling, clinical assessment, and follow up.

Mattia Gentile, M.D., Department of Medical Genetics, Hospital Di Venere, ASL BARI, via Di Venere s.n.—70131 Bari-Carbonara, Italy. E-mail: mattiagentile@libero.it Article first published online in Wiley Online Library

(wileyonlinelibrary.com): 5 May 2016 DOI 10.1002/ajmg.a.37691

^{*}Correspondence to:

CLINICAL REPORT

The proband is a female newborn, of Caucasian non-consanguineous parents: 38-year-old father and a 37-year-old mother. Family history is unremarkable; no neurological and/or psychiatric disorders have been reported.

She was born at 39 weeks by cesarean section because of fetal distress, after an uneventful pregnancy. Apgar score was 7/8, birth weight 2,620 g (3rd centile), height 46 cm (3rd centile), and head circumference 33.5 cm (>10th centile).

Soon after birth, she presented significant nasal obstruction and a Mayo cannula was applied. Craniofacial dysmorphisms were present (Fig. 1A): broad forehead with frontal bossing, large ears, down-slanted eyes, depressed and broad nasal bridge, flat nasal root, deviated nasal septum, stenosis of the left choana, small mouth, high arched palate, and central maxillary hypertrophy. Brain MRI, renal ultrasonography, echocardiography, ophthalmologic- and ORL examinations were all normal.

Genetic analysis revealed a de novo 14q13.2q21.1 interstitial microdeletion. Accordingly a clinical follow up was started. At 10 months of life, psychomotor evaluation was normal and auxological parameters were: weight 7,500 g (10th centile), height 66 cm (<3rd centile), and head circumference 43.5 cm (>10th centile). She suffered from recurrent rhinitis. A subclinical hypothyroidism was diagnosed: TSH 19.7 mIU/L (normal 0.36–3.74), fT4 1.03 ng/dl (normal 0.76–1.46), fT3 4.6 pg/ml (normal 1.7–5.2) together with a mild reduction of CD3CD8 Lymphocytes with an increased CD4/CD8 ratio (3.9, normal ratio 1.4–1.8). Complete blood cell counts, liver and renal function tests, blood glucose, IgA, IgG, and IgM levels were all within normal limits.

At subsequent follow up, at age of 15 months, no psychomotor development delay was present, weight and head circumference were at 25th centile, while height <3rd centile. Only 4 teeth had

erupted, with a single upper central incisor. Because of still elevated TSH, according to the endocrinologist tyroxine supplementation was started. She had a serious staphylococcal infection at the age of 1 year (Fig. 1B) for which hospitalization and intravenous antibiotic therapy were required.

GENETIC INVESTIGATION

Cytogenetic analysis, performed on QFQ-banded metaphases (550-band level), showed a normal female karyotype (46,XX). Genomic DNA was extracted from peripheral blood. DNA concentration was measured by fluorimeter (Amersham, Piscataway, NJ) using the Hoechst reagent and adjusted to 400 ng/ml. Array-CGH analysis was performed using the Cytochip oligo ISCA 4 × 44 K (TechnoGenetics Srl, Italy). This array is composed of 60-mer oligonucleotides spaced at about 75 Kb density across the genome. Labeling and hybridization were carried out following the manufacturer's protocol. Slides were scanned on the InnoScan 710 (Innopsys), and analyzed using the BlueFuse Multi v3.1 (Bluegnome). The analysis revealed an interstitial deletion of the long arm of chromosome 14, with a loss in copy number at the 14q13.2q21.1 region [chr14:35,443,407–39,527,387 (GRCh37), size 4,083,981 bp] (Fig. 2A). Result of array CGH analysis were confirmed by FISH. BAC probes RP11-561B11 (proximal 14q13.2), RP11-481F8, RP11-537P22, and RP11-506K19 (distal 14q21.1) (Bluegnome), mapping in the deleted region, produced hybridization signals only on the normal chromosome 14 (Fig. 2B). The deletion was not present in the parents. The 4.08 Mb deleted region contained 16 OMIM genes: SRP54, KIAA0391 (2), PSMA6, NFKBIA, INSM2, GARNL1, MBIP, NKX2-1, NKX2-8, PAX9, SLC25A21, MIPOL1, FOXA1, SSTR1, and SEC23A. The final karyotype of the patient was designed as (ISCN 2013): 46, XX.arr 14q13.2q21.1(35,443,407–39,527,387) × 1 dn.



FIG. 1. Facial characteristics of our patient at 6 months (A); staphylococcal infection at age 1 year (B). [Color figure can be seen in the online version of this article, available at http://wileyonlinelibrary.com/journal/ajmga].

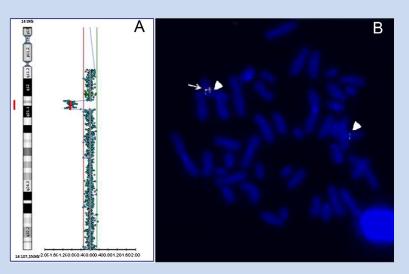


FIG. 2. (A) Oligo array CGH profile of chromosome 14, showing decreased log2 Ratio intensity values within the 14q13 microdeletion; (B) FISH analysis: RP11-481F8 (14q13.2) (green, arrow), and RP11-506K19 (14q21.1) (red, arrow) BAC clones gave signals only on the normal chromosome 14; the RP11-388G3 BAC clone (14q12) (arrowheads) gave signals of equal intensity on both chromosomes 14. [Color figure can be seen in the online version of this article, available at http://wileyonlinelibrary.com/journal/ajmga].

DISCUSSION

We report the clinical and molecular characterization of a newborn female with a mild phenotype, characterized by dysmorphic facial features, and poor fetal growth (weight and height \leq 3rd centile). Using array-CGH, we identified a 4.08 de novo interstitial deletion of the 14q13.2q21.1 region, which includes 16 OMIM genes.

The comparative evaluation of our case with other 14q deletion reported cases is not easy: the size and breakpoints of the deleted regions are highly variable, with different phenotypes. Santen et al. [2012], presenting the molecular karyotyping and the phenotypic description of seven patients with partially overlapping 14q13 deletions, assumes that the impact on the phenotype of the deletion of the 14q13 region is mainly due to haploinsufficiency of three genes: *FOXG1*, *NKX2-1*, and *PAX9*. When *FOXG1* is deleted, the development is severely delayed and the prognosis is poor, if haploinsufficiency is limited to *NXKX2-1* and *PAX9*, the phenotype is milder with quite normal psychomotor development [Santen et al., 2012].

We have found only two patients with a deletion similar to our case: patient 1, reported by Santen et al. [2012], and patient 3, described by Dale et al. [2012], while all other cases showed larger or smaller deletions [Shapira et al., 1994; Kamnasaran et al., 2001; Caliebe et al., 2011; Torgyekes et al., 2011; Fonseca et al., 2012; Piccione et al., 2012; Santen et al., 2012; Hayashi et al., 2015]. The two patients, respectively 7 and 1.5 years old, have hypothyroidism, movement disorders (chorea), hypo/oligodontia, and normal growth. Mild developmental delay was present with joint laxity and gait disturbances in the patient described by Dale et al. [2012] (patient 3), while motor delay was reported in the Santen

et al. case [2012] (patient 1). In both cases, no lung involvement was reported [Dale et al., 2012; Santen et al., 2012]; this data is of prognostic relevance because pulmonary disease is responsible for significant mortality in patients with *NKX2-1* anomalies [Carré et al., 2009].

These reports, together with our case, suggest that when the 14q13 deletion is distal, the phenotype is milder, more benign, mainly due to the haploinsufficiency of the *NKX2-1*, *PAX9*, and, probably, *NFKBIA* genes.

The most important pathogenetic role is that of *NKX2-1 gene* (or *TITF1*, thyroid transcription factor 1, OMIM 600635), which encodes for a NK-2 family transcription factor, expressed in thyroid gland, lung, and the forebrain. In the lung, NKX2-1 contributes to branching of epithelium, alveolarization and surfactant production [Barnett et al., 2012], while, in the brain, it is involved in neuronal migration, and in the development of the basal forebrain, pituitary, hypothalamus, and basal ganglia [Goulburn et al., 2011]. *NKX2-1* mutations are associated with a rare disease (CAHTP, OMIM 610978), characterized by a highly variable penetrance and expressivity, with manifestations ranging from benign hereditary chorea (BHC), the hallmark of NKX2.1-related disorders, to choreoathetosis, congenital hypothyroidism, and pulmonary disease (neonatal respiratory distress and/ or interstitial lung disease).

According to Williamson et al. [2014], brain and/or thyroid involvement is very common, while lung is involved in less than 50% of cases. This phenotypic variability can be explained considering that *NKX2-1* is an haplosensitive gene (DECIPHER HI% of 1.49): the positional effect of one or more deleted genes [Lupski and Stankiewicz, 2005], the tissue-specific gene expression regulation, as well as the extent of the deletion and the role of putative modifier

genes, not present in the critical region, all contribute to the phenotype [Barnett et al., 2012]. In our case, the respiratory distress at birth was due to the deviated nasal septum with left choana stenosis. The subclinical hypothyroidism could be due to *NKX2-1* deletion. At last examination psychomotor development was normal, but our patient is still too young (15 months) to exclude the possible onset of movement disorders (with a median onset age of 2 years), as well as of mild/moderate intellectual disability.

PAX9 is a transcription factor involved in the odontogenesis and craniofacial skeletogenesis, especially of the palate. Deletions or mutations of PAX9 have been previously described in patients with oligodontia, including families with autosomal dominant inheritance [Stockton et al., 2000; Guala et al., 2008]. Based on literature findings, the PAX9-related oligodontia is associated to the agenesis of permanent maxillary and mandibular second molars, while the primary dentition is often normal [Haldeman-Englert et al., 2012]. Recently, Zhou et al. [2013] have also demonstrated that Pax9 regulates a molecular network, involving the Bmp4, Fgf10, Shh signaling and the Osr2 transcription pathways, to control palate morphogenesis. At 15 months of age, our patient shows a very tight and arched palate with only four incisor teeth erupted, so that oligodontia is a very likely possibility. The presence of a single upper central incisor could indicate a midline development defect, due to PAX9 deletion, as suggested by the stenosis of the left choana [Hall, 2006].

Finally, the Nuclear Factor of Kappa light chain gene enhancer in B cells Inhibitor (*NFKBIA*) encodes for a master transcription factor required for normal activation of the immune response. Impaired *NFKB* signaling, due to mutations of this gene, causes an anhidrotic ectodermal dysplasia with T-immunodeficiency (EDA-ID, OMIM 612132), firstly described by Courtois et al. [2003]. However, the genotype–phenotype correlation are still not clear: Janssen et al. [2004] have described two related subjects with the same *NFKBIA* mutation, but completely different clinical syndromes. In our patient, *NFKBIA* haploinsufficiency could be responsible of recurrent rhinitis, severe staphylococcal infection, and mild reduction in CD3/CD8 Lymphocytes with an increased CD4/CD8 ratio.

The role of other genes is still debated (i.e., *GARNL1/TULIP1*, *NKX2-8*, *SLC25A21*). For example, *SLC25A21*, the solute carrier family 25 (mitochondrial oxodicarboxylate carrier), was previously described as a possible cause of 2-oxoadipate acidemia, an inborn error of metabolism of lysine, tryptophan and hydroxylysine, associated to mental delay, hypotonia, and seizures [Fiermonte et al., 2001]. Meyertholen et al. [2012] suggested an association of 14q13.3 deletion, involving the *SLC25A21*, with a familial synpolydactyly, thorough a possible interaction between *SLC25A21*, *PAX9*, and *MIPOL1*. *MIPOL1* is a candidate gene for mirror-image polydactyly [Kondoh et al., 2002], but, until now, the clinical significance of copy number variations of this gene is unclear.

In conclusion, our report shows that, despite the extreme rarity, the 14q13 distal deletion encompassing *NKX2-1*, but not *FOXG1* gene and/or *HPE8* region, identifies a well defined, more benign, microdeletion syndrome: except for growth retardation, all the signs and symptoms of our patient can be explained on the basis of the deleted genes. An accurate nosological classification is of great help for the patient and the family. It allows to make a proper

genetic counseling about the reproductive risk, and the prognosis, enabling to reassure the family about the recurrence, the severity of phenotype, and the possible occurrence of associated clinical manifestations (i.e., neurological movement disorders). In addition, an appropriate targeted follow up can help to avoid unnecessary exams, often expensive and/or invasive, with the possibility of early diagnosis and prompt medical therapy, that is, for thyroid abnormalities, or respiratory infections due to the association of the deletion with chronic interstitial lung disease.

ACKNOWLEDGMENTS

We are grateful to the family who participated in this study. We thank C. Nanna, E. Tangari, and P. Zambetti for technical assistance.

REFERENCES

- Barnett CP, Mencel JJ, Gecz J, Waters W, Kirwin SM, Vinette KMB, Uppill M, Nicholl J. 2012. Choreoathetosis, congenital hypothyroidism, and neonatal respiratory distress syndrome with intact NKX2-1. Am J Med Genet Part A 158A:3168–3173.
- Caliebe A, Martin Subero JI, Muhle H, Gesk S, Jänig U, Kraus M, Plendl H, Stephani U, Siebert R, Eckmann-Scholz C. 2011. A 2 Mb deletion in 14q13 associated with severe developmental delay and hemophagocytic lymphohistiocytosis. Eur J Med Genet 54:e505–e509.
- Carré A, Szinnai G, Castanet M, Sura-Trueba S, Tron E, Broutin-L'Hermite I, Barat P, Goizet C, Lacombe D, Moutard M-L, Raybaud C, Raynaud-Ravni C, Romana S, Ythier H, Léger J, Polak M. 2009. Five new TTF1/NKX2.1 mutations in brain-lung-thyroid syndrome: Rescue by PAX8 synergism in one case. Hum Mol Genet 18:2266–2276.
- Courtois G, Smahi A, Reichenbach J, Döffinger R, Cancrini C, Bonnet M, Puel A, Chable-Bessia C, Yamaoka S, Feinberg J, Dupuis-Girod S, Bodemer C, Livadiotti S, Novelli F, Rossi P, Fischer A, Israël A, Munnich A, Le Deist F, Casanova J-L. 2003. A hypermorphic IkappaBalpha mutation is associated with autosomal dominant anhidrotic ectodermal dysplasia and T cell immunodeficiency. J Clin Invest 112:1108–1115.
- Dale RC, Grattan-Smith P, Nicholson M, Peters GB. 2012. Microdeletions detected using chromosome microarray in children with suspected genetic movement disorders: A single-centre study. Dev Med Child Neurol 54:618–623.
- Fiermonte G, Dolce V, Palmieri L, Ventura M, Runswick MJ, Palmieri F, Walker JE. 2001. Identification of the human mitochondrial oxodicarboxylate carrier. Bacterial expression, reconstitution, functional characterization, tissue distribution, and chromosomal location. J Biol Chem 276:8225–8230.
- Fonseca DJ, Prada CF, Siza LM, Angel D, Gomez YM, Restrepo CM, Douben H, Rivadeneira F, de Klein A, Laissue P. 2012. A de novo 14q12q13.3 interstitial deletion in a patient affected by a severe neurodevelopmental disorder of unknown origin. Am J Med Genet Part A 158A:689–693.
- Goulburn AL, Alden D, Davis RP, Micallef SJ, Ng ES, Yu QC Lim SM, Soh C-L, Elliott DA, Hatzistavrou T, Bourke J, Watmuff B, Lang RJ, Haynes JM, Pouton CW, Giudice A, Trounson AO, Anderson SA, Stanley EG, Elefanty AG. 2011. A targeted NKX2.1 human embryonic stem cell reporter line enables identification of human basal forebrain derivatives. Stem Cells 29:462–473.
- Guala A, Falco V, Breedveld G, De Filippi P, Danesino C. 2008. Deletion of PAX9 and oligodontia: A third family and review of the literature. Int J Paediatr Dent 18:441–445.

- Haldeman-Englert CR, Biser A, Zackai EH, Ming JE. 2012. Revision of "A 223-kb de novo deletion of PAX9 in a patient with oligodontia". J Craniofac Surg 23:e149–e151.
- Hall RK. 2006. Solitary median maxillary central incisor (SMMCI) syndrome. Orphanet J Rare Dis 1:12.
- Hayashi S, Yagi M, Morisaki I, Inazawa J. 2015. Identical deletion at 14q13.3 including PAX9 and NKX2-1 in siblings from mosaicism of unaffected parent. J Hum Genet 60:203–206.
- Janssen R, van Wengen A, Hoeve MA, ten Dam M, van der Burg M, van Dongen J, van de Vosse E, van Tol M, Bredius R, Ottenhoff TH, Weemaes C, van Dissel JT, Lankester A. 2004. The same IkappaBalpha mutation in two related individuals leads to completely different clinical syndromes. J Exp Med 200:559–568.
- Kamnasaran D, O'Brien PC, Schuffenhauer S, Quarrell O, Lupski JR, Grammatico P, Ferguson-Smith MA, Cox DW. 2001. Defining the breakpoints of proximal chromosome 14q rearrangements in nine patients using flow-sorted chromosomes. Am J Med Genet 102:173–182.
- Kondoh S, Sugawara H, Harada N, Matsumoto N, Ohashi H, Sato M, Kantaputra PN, Ogino T, Tomita H, Ohta T, Kishino T, Fukushima Y, Niikawa N, Yoshiura K. 2002. A novel gene is disrupted at a 14q13 breakpoint of t(2;14) in a patient with mirror-image polydactyly of hands and feet. J Hum Genet 47:136–139.
- Lupski JR, Stankiewicz P. 2005. Genomic disorders: Molecular mechanisms for rearrangements and conveyed phenotypes. PLoS Genet 1:e49.
- Meyertholen K, Ravnan JB, Matalon R. 2012. Identification of a novel 14q13.3 deletion involving the *SLC25A21* gene associated with familial synpolydactyly. Mol Syndromol 3:25–29.

- Piccione M, Serra G, Consiglio V, Di Fiore A, Cavani S, Grasso M, Malacarne M, Pierluigi M, Viaggi C, Corsello G. 2012. 14q13.1–21.1 deletion encompassing the HPE8 locus in an adolescent with intellectual disability and bilateral microphthalmia, but without holoprosencephaly. Am J Med Genet Part A 158A:1427–1433.
- Santen GW, Sun Y, Gijsbers AC, Carré A, Holvoet M, Haeringen A van, Lesnik Oberstein SAJ, Tomoda A, Mabe H, Polak M, Devriendt K, Ruivenkamp CA, Bijlsma EK. 2012. Further delineation of the phenotype of chromosome 14q13 deletions: (Positional) involvement of FOXG1 appears the main determinant of phenotype severity, with no evidence for a holoprosencephaly locus. J Med Genet 49:366–472.
- Shapira SK, Anderson KL, Orr-Urtregar A, Craigen WJ, Lupski JR, Shaffer LG. 1994. De novo proximal interstitial deletions of 14q: Cytogenetic and molecular investigations. Am J Med Genet 52:44–50.
- Stockton DW, Das P, Goldenberg M, D'Souza RN, Patel PI. 2000. Mutation of PAX9 is associated with oligodontia. Nat Genet 24:18–19.
- Torgyekes E, Shanske AL, Anyane-Yeboa K, Nahum O, Pirzadeh S, Blumfield E, Jobanputra V, Warburton D, Levy B. 2011. The proximal chromosome 14q microdeletion syndrome: Delineation of the phenotype using high resolution SNP oligonucleotide microarray analysis (SOMA) and review of the literature. Am J Med Genet Part A 155A:1884–1896.
- Williamson S, Kirkpatrick M, Greene S, Goudie D. 2014. A novel mutation of NKX2-1 affecting 2 generations with hypothyroidism and choreoa-thetosis: Part of the spectrum of brain-thyroid-lung syndrome. J Child Neurol 29:666–669.
- Zhou J, Gao Y, Lan Y, Jia S, Jiang R. Pax9 regulates a molecular network involving Bmp4, Fgf10, Shh signaling and the Osr2 transcription factor to control palate morphogenesis. Development 140:4709–4718.