



Expert Opinion on Drug Delivery

ISSN: 1742-5247 (Print) 1744-7593 (Online) Journal homepage: <http://www.tandfonline.com/loi/iedd20>

Protein cage nanostructure as drug delivery system: magnifying glass on apoferritin

Daniela Belletti, Francesca Pederzoli, Flavio Forni, Maria Angela Vandelli, Giovanni Tosi & Barbara Ruozi

To cite this article: Daniela Belletti, Francesca Pederzoli, Flavio Forni, Maria Angela Vandelli, Giovanni Tosi & Barbara Ruozi (2016): Protein cage nanostructure as drug delivery system: magnifying glass on apoferritin, Expert Opinion on Drug Delivery, DOI: 10.1080/17425247.2017.1243528

To link to this article: <http://dx.doi.org/10.1080/17425247.2017.1243528>



Accepted author version posted online: 30 Sep 2016.



Submit your article to this journal [↗](#)



View related articles [↗](#)



View Crossmark data [↗](#)

Full Terms & Conditions of access and use can be found at
<http://www.tandfonline.com/action/journalInformation?journalCode=iedd20>

Publisher: Taylor & Francis

Journal: *Expert Opinion on Drug Delivery*

DOI: 10.1080/17425247.2017.1243528

Protein cage nanostructure as drug delivery system: magnifying glass on apoferritin

Daniela Belletti¹, Francesca Pederzoli¹, Flavio Forni¹, Maria Angela Vandelli¹, Giovanni Tosi^{1*}, Barbara Ruozi¹

¹Laboratory of Nanomedicine, Te.Far.T.I., Department of Life Sciences, University of Modena and Reggio Emilia, Via Campi 103, 41124, Modena, Italy

*Corresponding author:

Giovanni Tosi, Tel: +39.059.2058563; Email: gtosi@unimore.it

Abstract

Introduction: New frontiers in nanomedicine are moving towards the research of new biomaterials. Apoferritin (APO), is a uniform regular self-assemblies nano-sized protein with excellent biocompatibility and a unique structure that affords it the ability to stabilize small active molecules in its inner core.

Areas covered: APO can be loaded by applying a passive process (mainly used for ions and metals) or by a unique formulative approach based on disassembly/reassembly process. In this

article, we aim to organize the experimental evidence provided by a number of studies on the loading, release and targeting. Attention is initially focused on the most investigated antineoplastic drug and contrast agents up to the most recent application in gene therapy.

Expert opinion: Various preclinical studies have demonstrated that APO improved the potency and selectivity of some chemotherapeutics. However, in order to translate the use of APO into therapy, some issues must be solved, especially regarding the reproducibility of the loading protocol used, the optimization of nanocarrier characterization, detailed understanding of the final structure of loaded APO, and the real mechanism and timing of drug release.

Keywords: Apoferritin, Chemotherapeutics, Disassembly/reassembly, Nano-cage, Passive loading, Structural characterization

Article highlights box

- APO is a self-assembled molecule highly conserved in mammalian and with a high biocompatibility.
- APO offers the possibility to guest within its inner core drugs of low molecular weight and positively charged. The mechanism of drug loading can follow passive absorption/penetration processes or pH-dependent disassembly/reassembly protocols.
- Only a limited number of drugs (mainly chemotherapeutics containing metals or anthracycline) are successfully loaded into APO, showing promising results in preclinical animal models.
- APO is characterized by long lifetime and innate affinity for tumor cells due to its ability to bind human TtR1 receptor, over-expressed on cells in rapid proliferation.
- Modern biotechnological approaches lead to the production of recombinant protein able to link molecules (active and/or targeting agents) directly to the protein structure,

thus improving the efficacy of loaded APO and expanding its applicability to different field of therapy.

ACCEPTED MANUSCRIPT

1. Introduction

The production and the development of drug delivery systems (DDS), completely biocompatible, atoxic, and biodegradable is a challenging issue. Thus, modern nanomedicine focused its interest on alternative biomaterials as proteins.

The nature offers different types of multimeric proteins able to self-assemble in protein shell surrounding a hollow interior cavity with shapes and structures within nanometer scale [1].

From a biological point of view, protein cages play numerous functions as the storage, protection and delivery of viral genomes, metal storage, and protein refolding.

Their applicability as DDS depends on the technological formulative aspects as handy, reproducible, scalable methods of formulation and drug encapsulation as well as on the efficient purification process.

In this contest, advanced competences in the field of biotechnology help a sustainable large-scale production of proteins using cell factories, making this technology highly desirable and economically advantageous for different technological applications.

One of the most investigated classes of proteins applicable in drug delivery is the ferritin superfamily and particularly Apoferritin (APO) [2,3]. In this review, starting from the analyses of the structure of APO, we focus on the mechanism of drug loading in APO cage and the versatility of this protein as DDS in therapy.

2. Ferritin superfamily

Ferritins are a superfamily of proteins which is highly conserved in eukaryotic organisms and ubiquity diffused in humans and other mammals, plants, fungi, and bacteria [4].

The role of ferritin is fundamental for life; it is the first protein involved in iron storage and seizure, thus playing a role of reservoir for metabolic iron.

The crystal structures of many Ferritins were deeply characterized since the beginning of 1990s by the combination of Transmission Electron Microscopy (TEM), small angle X ray scattering (SAXS) and proteomic techniques [4-7].

In spite of large variations in amino acid sequences from bacteria to humans, ferritins essentially show the same architecture. Focusing on mammals, the ferritin's protein cage (450-500 kDa) is composed of 24 polypeptide subunits, with different molecular weights [8] and with specific functions [9]. Heavy (H) units (molecular weight of 21 KDa) play a major role in iron oxidation while light (L) units (molecular of 19 KDa) are more involved in the efficient nucleation and mineralization of iron. Of note, H and L subunits are highly homologous, and they can form a spherical unit in any proportion [10].

The ratio between the two types of chain in Ferritin is different with respect to the species and the tissues [11 as in relation to physiological state, as the amount of H chains increases in inflammation and other pathological conditions in response to cytokine stimuli [12,13]. Moreover, as reported in the text below (Chapter 4.1), differences in the type of receptor-binding are present between H/L Ferritin/Apoferritin as only H-Ferritin or H-Apoferritin are able to bind TtR1 receptors whereas L-Ferritin and L-Apoferritin bind SCARA5 receptors.

The subunits self-assemble to form a spherical cage with outer diameter of 12 nm, a 2 nm thick protein shell and a internal cavity with diameter of 8 nm. Overall, the protein molecular mass is 450KDa and it is able to store up to 4500 Fe(III) atoms [4,14].

The inner and the outer space are connected by pores size of about 0.3-0.5 nm [15, 16]: 8 of these pores defined "3-fold channels" are hydrophilic and they lead iron to diffuse into (and out of) the core; the other 4 pores defined "4 fold channels" are hydrophobic and their function is still unclear [17,18].

Finally, it is worth to consider that during the protein reassembly, amino acidic content differs from inner to outer cavities. Generally, ferritin have a preponderance of acidic over

basic residues with high level of non apolar groups thus the isoelectric point is generally in the range 4.5-5.5).

The ferritin surface resulted highly reactive and some substances can bind to the apoferritin surface through hydrogen bonding (non-ionic molecules) or electrostatic interactions (ionic molecules) [19].

3. From Ferritin to APO the applicability as DDS

As above described, from a physiologic point of view, ferritin can be described as Fe (III) reservoir. Aiming to exploit APO as DDS, the inner cavity should be free and able to accommodate the cargo, thus the resident iron has to be removed from the cavities. Demineralization of ferritin occurred by reductive dissolution and subsequent chelation of Fe(II) producing APO [20,21] which maintains the same features of Ferritin in terms of inner and outer structures.

The octahedral scaffold of APO displays typical hydrophilic and hydrophobic channels connecting inner cavities to the external surface and which are proposed as the route for inward and outward of metals and other molecules (therapeutic agents such as imaging agents) [22].

This mechanism of accumulation into the core [23] is named “passive loading”. Unless initially investigated, being a rapid and scalable loading process, it results often less efficient and not able to stabilize therapeutic molecules with large molecular weights, which result prevalently adsorbed on the surface. Therefore, deeper studies on strategies of improving drug loading with active loading, or at least by means of strong modification in the protocols of Ferritin assembly were conducted over the last 10 years. In this contest, a number of studies mostly aimed to elucidate the permeation mechanism of organic molecules into APO and the impact of chemico-physical characteristics on drug loading efficiency. As example, NMR relaxation techniques were employed to investigate the diffusion of molecules devoid of net

charge and with 18-7500 Da MW (i.e. water, DMSO, glucose, maltose, maltotriose, maltotetraose, maltopentaose and polyethylene glycol) into the protein cavity [24]. The authors demonstrated that the channel structure is sufficiently flexible to allow the penetration of molecules even larger than their own size (3-4 Å), but with maximum dimensions typical of the maltose (Mw: 342 Da, corresponding to a diameter of 13 Å) [24].

Electron paramagnetic resonance spectroscopy analysis were employed to study the diffusion of small and differently charged nitroxide organic probes (7-9 Å of diameter) to the central cavity of the ferritin, showing that the diffusion inside the protein is not purely passive (not only linked to the concentration gradient between the inside and outside of the protein) but also strictly dependent on their respective charge and the polarity. Only positively charged probes penetrate into the protein through the 3-fold channels, while not polar molecules bind the hydrophobic regions of the protein only on its outer surface [25,26].

Aiming to extend the number of applications, the research moves through alternative methods to load drug and investigate the dynamic of disassembly and reassembly of the protein nanocage.

Thanks to a large number of intra- and inter-subunit salt bridges and hydrogen bonds, the APO structure is known for its stability: i) over heating condition (up to 85°C) [27]; ii) amongst a wide pH range (3.40-10) [28]; iii) in presence of high ionic concentrations; iv) in presence of high concentrations of denaturing agents [29,30].

Even if highly stable to chemical and physical denaturants, the APO architecture can be disassembled in response to pH variation and then restored, almost completely, by returning the pH back to the “physiological” conditions (pH 7.4). A number of hypotheses to describe the structural changes during disassembly and reassembly of protein were proposed.

One of the first theories hypothesized that APO remains stable in the range of pH 2.8-10.6. On the contrary, at lower pH (1.6-2.8) or higher pH (10.6-13.0), a drastic reduction of MW, which can be related to the presence disassembled subunits, occurs [31].

A still open question is related to the exact pH at which protein starts its disassembling process. Some authors suggest that a progressive process [32] takes place; this event consists of cleavage of non-covalent bonds as a consequence of a conformational rearrangement of the protein [30]. Along with the decrease of pH, the outer surface of APO becomes positively charged (due to the protonation of the carboxylic groups of amino acid), leading to the progressive loosening of subunits forming holes in the spherical structure which switch from the inside to the outside [31]. By combining SAXS data and 3D structural reconstruction, it was shown that more the pH decreases and more the holes become numerous and larger, finally generating rodlike structures at pH 2 [28].

In these conditions, APO is completely disassembled into its subunits and allows to molecules/drug present in solution to interact with the chains.

To lead sub units to reassemble and to form the native conformation, it is pivotal to restore the native electrostatic potential, thus, the solution needs to be buffered to the pH to “physiological” values [33]. Within this reassembly process, drug loading takes place as the drug molecules are entrapped into the spherical structure (Fig.1).

In turns, aiming to expand the drug loading possibilities, some authors chose to move to different pH values, especially in the basic range. Notwithstanding less conformational data are available regarding the basic dissociation of APO, it was supposed starting over pH 10.6 and caused by the de-protonation and re-arrangement of tyrosine residues. A high level of attention should be posed on monitoring pH values, as continuing the basification over pH 12.4 lead to an irreversible alkaline hydrolysis of peptide bonds [34], disabling any possible recovery of native conformation [31].

4. Pharmacokinetics of APO as DSS

After parenteral administration the pharmacokinetic profile of APO was investigated in tumor bearing model. Drug-loaded APO are rapidly distributed into the bloodstream reaching small

capillaries. These protein DDS do not cross normal vessels, as only molecule smaller than 3 nm are allowed to pass the endothelial cells by means of transcellular route [35], but their low diameter (less than 20 nm) is sufficient to avoid macrophages capture. This drastically reduces a-specific uptake by the reticular endothelial system (RES) [36] and assures to keep the protein longer into bloodstream circulation.

Membranes of endothelial cells of blood vessels expose receptors that recognize APO and control both iron uptake and transferrin passage from vessel to tissues. H and L subunits show some differences in receptor affinity, as the H subunit specifically binds the receptor of transferrin (TfR1) with a wild distribution in human cells while L subunit binds to the Scavenger Receptor Class A Member 5 (SCARA5) firstly localized in macrophages, retina and hepatocytes [10,37].

As evident, the protein composition is pivotal to define the APO destiny and, at the same time, the well-known over-expression of TfR1 in many cancer cells (about 100-fold respect to normal tissue) [38,39] guarantees an intrinsic site-specific targeting potential of APO nanocage for tumor.

Moreover, tumor produces much more growing factors, leading to the presence of blood capillaries which tend to be disorganized, oversized, and leaky. As consequence, a large number of drug molecules and small structure (as APO) can be effectively driven to cancer tissue and stick onto pathological cells.

Also the mechanism of APO interaction with cells and especially with tumor cells was investigated, in vitro using CACO-2 cells, showing that APO exploits clathrin-dependent endocytosis [40], accumulates into endosomes and releases the protein cargo into cytoplasm as a consequence of acidification of the compartment, which is responsible of the progressive dissociation of APO [41].

4.1 APO in drug delivery, the tumor case

A number of anticancer drugs suffer poor selectivity for neoplastic cells leading to dose-limiting side effects, which compromise clinical outcomes.

Unfortunately, only limited number of drugs were investigated as well-defined drug properties are needed: i) controlled molecular weight (typically in the range 300-600 Da); ii) positive charged at physiological pH thus able to produce electrostatic interaction with the inner negatively charged protein cavity. In this view, DOXO possesses good properties (MW= 544 Da; and pKa= 7.34 (phenol); 8.46 (amine); 9.46 (est)). Different operative conditions were screened in order to optimize its loading into APO, in particular the effect of the buffer, the ionic strength and the protein concentration during the dissociation process.

During the last decades, the major interest was dedicated to the investigation of the development of loaded APO as DDS for anticancer molecules (Table 1). Among current used chemioterapeutics with low bioavailability and selectivity some of them display chemico-physical characteristics (low MW and cationic charge at physiologic pH) suitable for stabilization into APO. Moreover, thanks to the innate affinity for tumor cells the encapsulation is supposed to enhance the cell interaction with the tumor cells thus reducing the side and not-target accumulation and side effects. Considering this particular cancer targeting, H-ferritin should be used in order to produce cages able to really and successfully target cancer human cells. Notably, mouse models of cancer should not be taken into consideration, due to the presence of different receptors.

Gefinitinib, an epidermal grown factor (EGFR) receptor tyrosine kinase inhibitor (MW 447 Da), was passively loaded in APO. Drug resulted probably distributed in the inner and outer part of the protein cage (totally 10 molecules for each APO), thus the release starts at physiological pH and maximizes at pH 2 in correspondence with the protein disassembly.

Authors demonstrated that Gefinitinib/APO induced cytotoxicity in breast cancer cell lines over-expressing both TtR1 and HER-2 receptors with a profile comparable to that of free drug [42].

A strong effort in the research was focused on strategies to force structured molecules to stably enter the APO core. In some particular cases (i.e. doxorubicin and mainly Gd), for molecules with appropriate MWs, some authors proposed a pre-complexation of the molecule with a metal in order to use the natural affinity of metal for the inner core of APO as driving force to maximize the passage through the 3-fold channels.

Doxorubicin (DOXO), a wide spectrum anticancer antibiotic with affinity for different metals was previously pre-complexed with Cu(II), and efficiently loaded by simple incubation in APO systems (Cu/DOXO/APO)[43,44]. This strategy increases the drug loading efficiency in APO compared by simple DOXO/APO (30:100 DOXO:APO w/w ratio in presence of Cu respect to 8:100 DOXO:APO w/w ratio in absence of metal) [45]. Cu/DOXO/APO keeps both the dimension and the morphology of DOXO/APO and APO alone and no aggregates were observed. The release of the drug in physiological conditions, showed a typical biphasic profile characterized by an initial burst effect (about 80% of drug) and absence of drug release over 48 h. These data could indicate that the metal-drug complex is mostly adsorbed on the surface of protein. Notwithstanding technological data seems to suggest only a limited ability to protect DOXO, *in vitro* and *in vivo* tests on tumor models showed that DOXO/APO nanocages, targeted to integrin Rv β 3 (a tumor angiogenesis biomarker) by RGD peptide anchored on APO surface, displayed a better therapeutic profile, resulting in a longer circulation half-life, higher tumor uptake, better tumor growth inhibition, and less cardiotoxicity than free DOXO [45].

A second strategy proposed to force the drug into the protein core consists of disassembly-reassembly process. The natural metal binding site present in the inner core of APO can be used as driving force to stabilize some metal-containing compounds. In this context, platinum

based anticancer drugs could be efficiently encapsulated into APO after dissociation at pH 2 and reconstitution at physiologic pH.

Cisplatin and carboplatin were loaded with the aim to reduce the systemic toxicity and high tumor resistance [46]. Through ICP-MS analysis authors demonstrated that about 2 and 5 molecules of cisplatin and carboplatin respectively became part of the protein structure, even if not clear evidences about the rate between internalization and absorption were reported. In a further study, these data were corroborated by a deeper characterization of platinum loaded APO (oxaliplatinum) and evaluating the effect on a cancer cell line overexpressing transferrin receptor. Data highlighted the low toxicity of APO on cancer cells and the rapid decreased of cellular viability as a consequence of internalization of loaded protein, thus suggesting the potentiality of APO to overcome the mechanism of resistance typical of this type of metal based anticancer drug [35].

As alternative, cisplatin was stabilized into APO after dissociation at more handy and “stable” formulative conditions, as weak alkaline pH [47]. Once tested on cancer cells cisplatin loaded into APO showed the ability to induce cytotoxicity through the same apoptosis pathway of free drug. Even if it is not clearly proved the exact localization of the drug within the APO, the evidence that APO slows down the biological response at short time respect to free drug, suggests that the protein plays an important role in controlling the drug release over the time.

Disassembly-reassembly process was also applied to load drugs in APO without pre-complexation with metal. To date, only limited number of drugs were investigated as well-defined drug properties are needed: i) controlled molecular weight (typically in the range 300-600 Da); ii) positive charged at physiological pH thus able to produce electrostatic interaction with the inner negatively charged protein cavity. In this view, DOXO possesses good properties (MW= 544 Da; and pKa= 7.34 (phenol); 8.46 (amine); 9.46 (est). Different operative conditions were screened in order to optimize its loading into APO, in particular the

effect of the buffer, the ionic strength and the protein concentration during the dissociation process.

Interestingly, the use of glycine-acetate buffer (pH 2.5) is the one more efficacious approach in disassembling APO without any effect related to the initial protein concentration [48].

Since only H chain showed affinity for cancer cells through the binding with transferrin receptor, by means of an elegant combination of chemical engineering and fermentation technology, different types of mammalian ferritin variants were formulated by self-assembling by only H subunits (H-APO) [49]. Thus, H-APO was loaded with DOXO following both the previously reported acidic [50,51] or basic dissociation protocols [52]. The complex DOXO loaded H-APO (DOXO/H-APO), tested *in vitro*, was compared in term of efficacy with free DOXO and with clinical approved liposome loaded DOXO formulation (Doxil®).

The “innate” targeting ability of the protein cage was confirmed as DOXO/H-APO showed better ability to mediate efficient and rapid interactions with cancer cells than the other control formulations [50,51]. Some conflicting data derived from *in vitro* distribution studies; DOXO/H-APO rapidly entered in a colon cancer cell line accumulating in lysosomes where DOXO is gradually released and subsequently translocated into nucleus [52]. Differently, in a liver carcinoma cellular line, DOXO/H-APO accumulated rapidly into nucleus through a specific interaction of H subunit with receptor exposed on nuclear membrane. Accordingly, DOXO intercalates DNA causing higher and more rapid cytotoxicity in cancer cells respect to controls (DOXO or Doxil) [50-51].

We could speculate on the different biological response and mechanism of cellular accumulation as consequence of multiple aspects, especially relating to the sensibility of tumor type and to the effect of the architecture and surface proprieties of loaded APO formulated applying different protocols. As evident, more efforts are needed to completely characterize these new entities.

To go further, DOXO/H-APO, obtained by means of basic dissociation protocol, was in vivo tested aiming to investigate pharmacokinetics profile and toxicity in mice bearing tumor (colon cancer cells). Namely, DOXO loaded into APO showed 10 fold higher AUC and tumor accumulation with respect to free drug and Doxil and reduced the drug concentration in healthy tissues (muscle, lung, kidney, spleen and most importantly heart). In addition, DOXO/H-APO led to a significant reduction in tumor growth two-fold higher than Doxil® [52].

The protocols for encapsulation of DOXO are now used as a template. Recently atropine and carbachol, two molecules active on muscarinic receptors, involved in the development of pancreatic tumor, was loaded into APO cage. All studied molecules show similar MW (carbachol=183 Da; atropine=290 Da) and pKa (carbachol=12.5; atropine=4.5), suggesting loading efficiencies similar to DOXO case. Authors described an efficient accumulation both in vitro and in vivo at the tumor site, a partial suppression of tumor development and a reduction of toxicity if compared with the treatment with free drugs [53].

To ameliorate the affinity of APO for cancer cells, some authors proposed the surface modification by the insertion of target moieties as antibodies or peptides.

APO has a fair versatility regarding chemical reactivity: in fact, a variety of chemical groups, e.g. primary amines, carboxylates, and thiols present on the external and internal surface, can be linked either genetically [29,45] and/or chemically [54,55]. Interestingly, after surface modification, APO diameter generally increased and the surface properties change (zeta-potential, hydrophilicity). As a consequence, the cellular interaction appeared to be more driven by the presence of ligands exposed onto the surface than by the “innate” tumor affinity of APO.

That way, a complex of daunorubicin (DAUNO) with negatively charged polypeptide PLLA (poly-L-aspartic acid) [24,56] was incorporated into APO after a partial disassembling at pH 5 and reassembly at physiological pH. To target solid tumor cells, the surface of APO was

engineered (by means of NHS/EDC protocol) with hyaluronic acid able to specifically bind CD44. The efficacy of this engineered nanocage was in vitro tested on a lung cancer cells overexpressing CD44 and a control cell line, not expressing any antigen. Modified nanocages were safe and tolerable for both cell lines, but it was clearly demonstrated that the cellular uptake was strongly driven by the binding with the cellular antigen. Briefly, DAUNO/APO accumulated only in CD44 positive cell line and both the cellular uptake and cytotoxicity were sensible to the pre-incubation with free antigen (which means inhibiting the cell specific binding) [56].

In a second example, APO loaded with cisplatin proposed by Xing and collaborators [35] was conjugated through a NHS-PEG-Mal cross-linker with an antibody selective to a proteoglycan expressed on the surface of melanoma cells (CSPG4). The new modified vectors showed a significant increase in both their molar mass and diameters (33 nm Vs 13 nm of native APO). In vitro studies confirmed that modified APO specifically bound melanoma cells expressing CSPG4(+), but not CSPG4(-) breast cancer cells and, accordingly, in vivo study showed a delayed tumor growth in model mice of melanoma. On the contrary, the tumor size of breast tumor, which does not over-expresses the specific antigen, was only marginally affected [57]. Recently APO was proposed as non-viral system to stabilize and protect small interfering RNA (siRNA). SiRNA are characterized by high MW and negatively charges, features that hampered a stable internalization into the protein core. Taking advantage of the protein recombinant technology, APO was modified aiming to orderly expose cationic peptides on its surface. Thus, protamine-derived peptides were inserted as moieties able to complex and capture siRNA; besides, APO was also modified with penetrating peptides to enhance the target delivery to tumor cells.

The experiments clearly highlighted that after modified-APO/siRNA complexation, the targeting peptides exposed on the protein surface drove the complex-cellular interaction and

siRNA accumulated into the cell cytoplasm, thus opening to a great potential for further applications of APO in gene therapy applied to cancer treatment (Fig. 2; Table 1) [58].

4.2 Application of APO in diagnosis

Imaging agents are among those compounds that can be successfully loaded into the interior cavity of APO protein (Table 2).

Magnetic Resonance Imaging (MRI) is a non-invasive imaging method for tumor diagnosis, however most of the diagnostic agents suffers from a lack of selectivity, low proton relaxivity and higher toxicity after administration [59].

APO could act as dual-functional tools allowing both the use of protein shell to target tumors and the ability in loading probes within the internal core to image tumors.

Gadolinium (GD) is one of the most used contrast agent in clinical and experimental settings; thus a number of authors proposed the formulation of water suspended APO-GD formulation with high NMR longitudinal relaxivity similar to clinically approved GD-chelates. [59-62].

The technology employed in order to stabilize this contrast agent, described for the first time more than 10 years ago, was based on a well-described acidic protocol (dissociation at pH 2) and the use of a neutral chelates (GD-HPDO3A) [59].

To improve tumor targeting, the surface of APO/GD-HPDO3A was engineered via streptavidin-biotin technology with an antibody directed to neural cell adhesion molecules (NCAM), overexpressed during the angiogenesis in endothelial cells. After in vivo administration, this new contrast agent was stable for few hours with a relaxation efficiency 5-fold higher with respect to the free probe, showing an increase in selectivity for tumor vessels [62].

In a more recent work, in order to maximize the loading efficiency, GD-HPDO3A was replaced with a positively charged GD-Me2-DO2A. With the aim to assure a stable insertion

into the cavity, the process was modified by adding GD complex during the acidification stage (at pH 4). Finally, the surface decoration of GD loaded APO with dextran lead to the accumulation of APO in transplanted tumor region in mice due to the exploitation of EPR effect [60].

Also Mn ions could be used as contrast agent in MRI. Through the stabilization into APO it has been demonstrated the improving in relaxivities [63, 64]. These non invasive imaging tools were applied as MRI sensor for different kinds of tumor. Taking advantage of the reduced expression of SCARA5 (one of the APO cellular receptor) into hepatoma cells respect to health hepatocytes, Geninatti and co-workers demonstrated the possibility to discriminate between tumor lesion and monitor the hepatic tumor progression with a sensitivity similar to commercial probe (Gd-BOTA) [65]. A similar structure was also developed for the early diagnoses and imaging of melanoma cells. The increased melanin production in cancer cells correspond to an increased reduction of Mn (III) to Mn (II) (by the oxidation of L-DOPA to melanin) thus producing an high relaxivities and contrast during MRI investigation [66].

Also iron is another optimal contrast agent for MRI. Autologous ferritins have been successfully investigated as a MRI contrast agent. One of the first example reported described the use of a cationized ferritins for MR imaging of kidneys (obtained after coupling with N, N dimethyl-1,3 propanediamine (DMPA) that after iv administration accumulated in the kidneys because the glomerular basement membrane is negatively charged [67]. More recently ferritin was investigated as MRI contrast agent in neurodegenerative disorders because of the documented altered iron metabolism in many neurodegeneration (particularly Alzheimer's disease and Huntington's disease [68]. As example of this application in vivo experiments in mice demonstrated that it is possible to increase the iron expression (thus the MRI signal) in nerve cells of mice after transfection of nerve cells using lentiviral and adenovirus vectors expressing transferrin [69] (for a complete review on this point see ref. 70). Unless some application reported, generally endogenous ferritin shows low relativity index if compared to

synthetic iron oxide nanoparticles [71]. Human recombinant H-APO offers the possibility to act as suitable platform for synthesis and encapsulation of magnetite nanoparticles (Fe_3O_4), leading to the obtainment of optimal contrast agent for MRI for tumor diagnoses. Starting from the first studies on APO formulation containing an iron core in the form of magnetite (Fe_3O_4) [72-73], the research moved to the surface engineering aiming to achieve improved tumor selectivity. That way, magnetoferritin was modified using ligand able to target tumor markers such as RGD-4C that can specifically target tumor angiogenesis via binding to integrin molecules on vascular endothelium [74] melanocyte-stimulating hormone (MSH), or epidermal growth factor (EGF) [75,76]. Besides the surface modification to increase the tumor detection, other imaging modalities, such as fluorescence imaging and positron emission tomography (PET) were investigated [77]. Recently, APO loaded with Cu^{64} and Cy5 was modified with RGD4C peptide anchored to the ferritin surface by conjugation with the surface amines. This structure was able to target the tumor, by means of RGD-integrin interaction and the enhanced permeability and retention (EPR) effect [78].

4.3 Application of Apo as theranostic tool

The growing knowledge and advances in nanomedicine drive the recent research in formulating systems containing simultaneously both drugs and imaging agents. The combination of therapy and diagnoses in a single drug delivery system is now emerging as theranostic approach.

This strategy can find applications for various different purposes, as to monitor the biodistribution and the target site accumulation of nanomedicines, to visualize and quantify drug release, and to assess the efficacy of the therapeutic treatment [79,80].

The design of a theranostic nanosystems could be obtained by following different protocols in terms of drug loading processes and insertion of the diagnostic agents (Table 3).

The DOXO/APO systems characterized by a natural affinity for tumor were largely studied and recently were modified onto their surface with magnetic particles by means of streptavidin-biotin technology. This approach preserved the natural loading ability of APO and it avoided any possible incompatibility between drug and diagnostic agent. Authors demonstrated that modified DOXO/APO was sensible to the application of a magnetic field, thus enabling also a pH-dependent release of the drug and opening the pave to a possible in vivo application in the treatment and diagnoses of tumors [54].

More recently a DOXO/APO complex was specifically designed as theranostic agent for prostate cancer. Briefly, the surface was double engineered with an antibody direct to anti PMSA (prostate specific membrane antigen) and with gold nanoparticles. The complex coverage of protein seems to protect the cargo from undesired premature release in the bloodstream; moreover, once tested in vitro, this new carrier assured a nice selectivity for prostate cancer cells preserving healthy cells from the drug toxicity [81].

A different approach is based on a contemporary loading into the inner cavity of the protein of diagnostic and drug molecules.

The cytostatic drug 5-fluorouracil (5-FU), being characterized by a low molecular weight, rapidly leak from the APO cavity; thus to better stabilize the encapsulation, [82] proposed a simultaneous loading with gold nanoparticles (AU-NPs, 3nm of diameter) known for their optical and photothermal proprieties. 5-FU enters trough hydrophilic protein pores and once into the protein core binds AU-NPs via electrostatic interaction. This strategy avoided the leaking of 5-FU under physiological condition and promotes the drug release only after accumulation of APO inside the intracellular acidic compartments. In a very complete and worthy in vitro study, the complex was demonstrated to be able to increase the cytotoxic

profile of the drug in different tumor cell lines, pointing out the wide applicability of the new therapeutic entities.

Similarly, curcumin, a multi-target drug with potent antioxidant proprieties, was loaded into APO simultaneously with GD through an acidic dissociation protocol, obtaining about 9 and 0.4 molecules of curcumin and GD respectively loaded into APO, with a consequent increase in APO diameter of about 66% (from 12 to 19 nm). In presence of curcumin, the gadolinium complex conserves its typical relaxivities and rapidly accumulates into liver (passive targeting). Based on this peculiar distribution, authors proposed the vector as a theranostic agent for a variety of liver diseases (Fig. 3) [83].

4.4 A look over cancer, other possible applications of Apo as DDS

The versatility of APO was recently exploited for the potentiality of antigen presentation onto the surface and immune stimulation in vaccines formulation.

Generally, vaccines for immunization standard schedules are stably prepared and protocols of production are well standardized, but, for rapidly mutating-pathogens (as the influenza virus), it is necessary to prepare new complex vaccines on an annual basis to avoid possible circumvention of the immune system by mutated pathogen.

The subunits of APO offer different regions tolerant to peptide insertion; these regions were therefore modified to obtain proteins expressing exogenous components that are able to conserve the ability to re-organize into spherical nanocage [84].

Following this approach, APO was synthesized in a controlled manner in terms of quantity and location of substituent. By selecting the site for protein insertion, it is possible to expose the antigens on the outer surface [77-78] or both inner and outer [85]. From a technological point of view, new vaccines were characterized by using PCS and TEM to prove spherical shape and presence of the exposed antigen.

To evaluate the real efficacy of these new entities in comparison with licensed inactivated vaccines, different in vitro and in vivo tests were performed. Generally, after administration in mice, those APO exposing the antigen protein onto the outer surface induced a higher CD8+ and CD4+ T cell proliferative response and therefore their transformation in functional T cells [86].

In view of expanding the applicability of APO in therapy, Zhang and coworkers exploited the applicability of APO in enzyme stabilization. Considering the large MW and steric hindrance of the molecule to be loaded, the internalization into the protein core appeared difficult, thus the enzyme glucose oxidase (GOx) (chosen as model enzyme) was immobilized onto the protein surface. Authors demonstrated that GOx linked on the surface through a biotin-streptavidin approach retained most of its activity up to two weeks and resulted stable to exposition to high temperatures (up to 50°C). This study presents a novel approach to enhance stability and activity of enzyme with promising application in different therapy but also in biological assay [87].

Finally, in a recent paper aiming to explore the applicability of APO in brain disease therapy, H-Apo was tested on a cerebellar organotypic culture demonstrating high affinity for astrocyte cell and a rapid and massive accumulation into nuclei [50].

Table 3: APO utilization in therapy

5. Conclusion

APO, featured by high stability, special and reproductive structure, auto-assembly ability and biocompatibility, is certainly one of the most investigated and interesting structures proposed in the modern scenario of nanomedicines.

From the beginning of its history (less than 15 years) as material for drug delivery, its applicability was investigated within the field of cancer medicine due to its natural affinity for iron receptor, largely distributed on cells in rapid division. Unfortunately, based on its specific

conformation and small internal spaces, only a limited number of chemotherapeutics (mainly belonging to anthracycline family as daunomycin and doxorubicin or contrast agent for tumor diagnoses and resulted suitable for successful loading into APO cage. Aiming to broaden the field of APO application, two main technological innovations were proposed: i) the engineering the protein surface by means of linkage of molecules able to drive the destiny of the protein cage; ii) the use of protein reactivity to link the drug or the active molecules directly on the surface. That way, APO became object of investigations in other fields of nanomedicine as gene therapy, immunology or liver pathology.

6. Expert opinion

Despite extensive promises, much work is needed before clinical translation of APO. As example, analyzing literature data, even if interestingly biological data are reported supporting the efficacy of this DDS, the study of the architecture and the structure of loaded APO are still at early stages. This situation, which represents the major limitation is translatability of APO to become DDS, is principally caused by unclear protocols for APO formulation and un-clear chemico-physical, morphological and technological characterization and it frequently generates conflicting outputs.

Points to be focused with major attention are related to the nature and narrow size of APO complex and to the evidence that the analytical technique commonly used to characterize synthetic DDS as liposomes or nanoparticles (TEM, AFM, SEM, PCS, DSC etc) are unable to completely demonstrate structure, architecture and, in particular, the presence of drug or ligands onto APO surface.

These data relating to chemico-physical and morphological characterization of APO formulations are essential to determine the fate and the perspective of any DDS, as any change in surface proprieties, even small differences in chains re-arrangements could strongly

affect the interaction with serum proteins, recognition by immune systems, impacting on the cage biocompatibility and also on the ability to transport drug to the site of action.

Currently, within their experimental plan only few studies are considering the evaluation of the effective recovery of conformation after the loading and disassembly process. Generally, when investigated, this issue on the final APO structure is approached by applying far-UV circular dichroism (CD). CD permits to evaluate the presence and the distribution of helical structures into the new protein entities. Despite these data are surely reliable, the results are not representative of the real integrity of protein after loading or disassembly/reassembly process.

To overcome these limits, X-ray diffraction characterization, even if expensive and sometimes difficult to be rendered, could be considered as the best choice for the characterization of protein structural conformation. As an example, a study performed by SAXS technique showed clearly that protein reassembly process couldn't be totally complete after strong acidification; under pH 2.4 the protein completely loses its spherical structure that cannot be recovered. The pH limit for pseudo-reversible restoration of the quaternary structure was set at 2.66 [28].

One of the best example of an almost complete study of APO formulation is represented by a recent work [88] in which cisplatin loaded APO were formulated by applying the same method proposed by Ji and coworkers [47] and were submitted to a deep structural characterization by means of X-ray analyses and elaboration of data. By comparing loaded and unloaded samples with native APO, the authors confirmed that the overall structure (shape, polarity/hydrophobicity, volume and electrostatic potential of the surface) remained stable after the formulation process. Notably, comparing native and cisplatin-loaded APO, the most relevant structural differences were detected close to the site of drug loading and in particular at the His 132, found to be the metal-binding site.

This work represents an important keystone for further investigations aiming to confirm the maintenance of the structural integrity of the protein, in particular regarding the encapsulation of non-metal drug with large MW.

Furthermore, another point to be investigated aiming to improve the chances of translatability of APO as drug delivery systems is related to the drug release and in particular to the changes in morphology and architecture of APO cage during and after the release of the drugs. To our knowledge, poor information are given on this topic which obviously impacts on several technological key-features of APO drug delivery systems, as the efficiency in drug release, the possibility of governing/modifying the drug release and also the biocompatibility of APO after the release of the drugs. These aspects are still far from an overall evaluation and therefore need to be considered with a major interest.

As final consideration, it seems almost clear that the improvement and the consecration of potentialities of APO application in different fields of drug targeting must be obtained by an extensive collaboration among researchers in the field of technology, chemical-physical characterization and physicians.

Funding

This work was partially financed by FAR UNIMORE grants (MA Vandelli and GTosi) by University of Modena and Reggio Emilia.

Declaration of interest

The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

References

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

[1] Ma-Ham,A, Wu H, Wang J, et al. Apo-based nanomedicine platform for drug delivery: equilibrium binding study of daunomycin with DNA. *Journal of Materials Chemistry* 2011; 21: 8700–08

[2] Yildiz I, Shukla S, Steinmetz NF. Applications of viral nanoparticles in medicine. *Biochem J* 2011;22:901–08

[3] Uchida M, Terashima M, Cunningham CH, et al. A human ferritin iron oxide nano-composite magnetic resonance contrast agent. *Magn Reson Med* 2008;60:1073–81

[4] Harrison PM, Arosio P. The ferritins: molecular properties, iron storage function and cellular regulation. *Biochim Biophys Acta* 1996;1275:161-203

•One of the first study describing the structure and function of APO

[5] Harrison PM. The structure of Apo: Molecular size, shape and symmetry from X-ray data. *J Mol Biol* 1963; 6:404-22

[6] Rice DW, Ford GC, White JL, et al. The spatial structure of horse spleen Apo. *Adv Inorg Biochem* 1983;5:39–50

- [7] Ha Y, Shi D, Small GW, et al. Crystal structure of bullfrog M ferritin at 2.8 Å resolution: analysis of subunit interactions and the binuclear metal center. *J Biol Inorg Chem* 1999;4:243-56
- [8] Arosio A, Adelman G, Drysdale JW. On Ferritin Heterogeneity. *J Biol Chem* 1978; 253:4451-58
- [9] Lawson DM, Artymiuk PJ, Yewdall SJ, et al. Solving the structure of human H ferritin by genetically engineering intermolecular crystal contacts. *Nature* 1991; 349:541-4
- [10] Li L, Fang CJ, Ryan JC, et al. Binding and uptake of H-ferritin are mediated by human transferrin receptor-1. *Proc Natl Acad Sci U S A* 2010;107: 3505–10.
- [11] Levi S, Salfeld J, Franceschinelli F et al. Expression and structural and functional properties of human ferritin L-chain from *Escherichia coli*. *Biochemistry* 1989; 28:5179-84.
- [12] Masover WH. Ultrastructure of ferritin and Apo: A review. *Micron* 1993;24:389–437.
- [13] Douglas T, Allen M, Young M. Self-assembling Protein Cage Systems and Applications in Nanotechnology. *Polyamides and Complex Proteinaceous Materials* 2005
DOI: 10.1002/3527600035.bpol8015
- [14] Ford GC, Harrison PM, Rice DW et al. Ferritin: design and formation of an iron-storage molecule. *Philos Trans R Soc Lond B Biol Sci* 1984;304: 551–65
- [15] Zhao G. Phytoferritin and its implications for human health and nutrition. . *BiochimBiophysActa* 2010;1800:815-23
- [16] Takahashi T, Kuyucak S. Functional properties of threefold and fourfold channels in ferritin deduced from electrostatic calculations. *Biophysical Journal* 2003;84:2256–63.
- [17] Bou-Abdallah F. The iron redox and hydrolysis chemistry of the ferritins. *Biochim. Biophys Acta* 2010;1800:719–31
- [18] Theil EC. Ferritin protein nanocages use ion channels, catalytic sites, and nucleation channels to manage iron/oxygen chemistry. *Curr Opin Chem Biol* 2011;15:304–11.

[19] Liu F, Du BJ, Chai Z et al. Binding properties of apoferritin to nicotinamide and calcium. *Eur Food Res Technol* 2012; 235: 893-99

[20] Ponka P, Richardson DR. Can ferritin provide iron for hemoglobin synthesis? *Blood* 1997;89:2611-3

[21] Gelvan D, Fibach E, Meyron-Holtz EG et al. Ferritin uptake by human erythroid precursors is a regulated iron uptake pathway. *Blood* 1996;88:3200-07

[22] Maham A, Tang Z, Wu H, et al. Protein-based nanomedicine platforms for drug delivery. *Small* 2009;5:1706-21

•• An interesting review summarizing the versatility of protein as drug delivery systems

[23] Linder MC. Mobilization of Stored Iron in Mammals: A Review. *Nutrients* 2013; 5: 4022-50

[24] Yang D, Nagayama K. Permeation of small molecules into the cavity of ferritin as revealed by proton nuclear magnetic resonance relaxation. *Biochem J* 1995; 307:253-56

[25] Yang X, Chasteen ND. Molecular diffusion into horse spleen ferritin: a nitroxide radical spin probe study. *Biophysical Journal* 1996;71:1587-95

[26] Yang X, Arosio P, Chasteen ND. Molecular diffusion into ferritin: pathways, temperature dependence, incubation time, and concentration effects. *Biophysical Journal* 2000;78:2049-59

[27] Sengonul M, Ruzicka J, Attygalle AB, et al. Surface Modification of Protein Nanocontainers and Their Self-Directing Character in Polymer Blends. *Polymer (Guildf)* 2007;48:3632-40

[28] Kim M, Rho Y, Jin KS, et al. pH-Dependent Structures of Ferritin and Apo in Solution: Disassembly and Reassembly. *Biomacromolecules* 2011; 12:1629-40

[29] Arosio P, Ingrassia, R, Cavadini, et al. A Family of Molecules for Iron Storage, Antioxidation and More. *Biochim Biophys Acta* 2009; 1790: 589-99

- [30] Gerl M, Jaenicke R. Assembly of Apo from horse spleen: comparison of the protein in its native and reassembled state. *Biol Chem Hoppe Seyler* 1987; 368:387-96
- [31] Crichton RR, Bryce CFA. Subunit interactions in horse spleen Apo. Dissociation by extremes of pH. *Biochemical Journal* 1973;133:289–99
- [32] Imai N, Arata Y, Fujiwara S. ¹H NMR study of dissociation and re-association of apoferritin and ferritin. *Bulletin of the Chemical Society of Japan* 1981;54:1243–44
- [33] Chen L, Bai G, Yang R, et al. Encapsulation of β -carotene within ferritin nanocages greatly increases its water-solubility and thermal stability. *Food Chem* 2014; 149:30712
- [34] Xing R, Wang X, Zhang C, et al. Characterization and cellular uptake of platinum anticancer drugs encapsulated in apoferritin. *J Inorg Biochem* 2009;103:1039-44
- [35] Azzi S, Hebda JK, Gavard J, et al. Vascular Permeability and Drug Delivery in Cancers. *Front Oncol* 2013;3:211
- [36] Molino NM, Wang SW. Caged protein nanoparticles for drug delivery. *Curr Opin Biotechnol* 2014;28:75–82.
- [37] Huang J, Zheng DL, Qin FS et al. Genetic and epigenetic silencing of SCARA5 may contribute to human hepatocellular carcinoma by activating FAK signaling. *J Clin Invest.* 2010; 120:223-41
- [38] Daniels TR, Delgado T, Rodriguez JA, et al. The transferrin receptor part I: Biology and targeting with cytotoxic antibodies for the treatment of cancer. *Clin Immunol* 2006;121:144-58.
- [39] Bretscher MS, Thomson JN. Distribution of ferritin receptors and coated pits on giant HeLa cells. *EMBO J* 1983; 2: 599–603
- [40] Oh N, Park JH. Endocytosis and exocytosis of nanoparticles in mammalian cells. *Int J Nanomedicine* 2014; 9:51-63
- [41] Liu X, Wei W, Yuan Q. Apo ceO₂ nano truffle that has excellent redox enzyme activity. *Chem Commun (Camb)* 2012;48:3155-7

[42] Kuruppu AI, Zhang L, Collins H, et al. An Apo based drug delivery system for the tyrosine kinase inhibitor Gefitinib. *Adv Healthc Mater* 2015;4:2816-21

[43] Feng M, Yang Y, He P, et al, Spectroscopic studies of copper(II) and iron(II) complexes of adriamycin. *Spectrochim Acta A Mol Biomol Spectrosc* 2000;56:581-7

[44] May PM, Williams GK, Williams DR. Speciation studies of adriamycin, quelamycin and their metal complexes. *InorgChim A-bioinorg* 1980;46:221-228

[45] Zhen Z, Tang W, Chen H, et al. RGD-Modified Apo Nanoparticles for Efficient Drug Delivery to Tumors. *ACS Nano* 2013;7:4830–37

[46] Yang Z, Wang X, Diao H, et al. Encapsulation of platinum anticancer drugs by apoferritin. *Chem Commun (Camb)* 2007 Sep 7;(33):3453-5

[47] Ji XT, Huang L, Huang HQ. Construction of nanometer cisplatin core-ferritin (NCC-F) and proteomic analysis of gastric cancer cell Apoptosis induced with cisplatin released from the NCC-F. *J Proteomics* 2012;75:3145-57

[48] Kilic MA, Ozlu E, Calis S. A novel protein-based anticancer drug encapsulatingnanosphere: Apo-doxorubicin complex. *J Biomed Nanotechnol* 2012; 8:508-14

[49] Grace JE, Van Eden ME, Aust SD. Production of recombinant human Apo heteromers. *Arch Biochem Biophys* 2000;384:116-22

[50] Zhang L, Li L, Di Penta A, et al. H-Chain Ferritin: A Natural Nuclei Targeting and Bioactive Delivery Nanovector. *Adv Healthc Mater* 2015;4:1305-10

• An interesting paper describing the intracellular localization of loaded APO.

[51] Bellini M, Mazzucchelli S, Galbiati E, et al. Protein nanocages for self-triggered nuclear delivery of DNA-targeted chemotherapeutics in Cancer Cells. *J Control Release* 2014; 196:184-96

[52] Liang M, Fan K, Zhou M, et al. H-ferritin-nanocaged doxorubicin nanoparticles specifically target and kill tumors with a single-dose injection. *Proc Natl Acad Sci U S A.* 2014; 111:14900-5

•• A very interesting paper describing the efficacy of recombinant protein as effective vehicle for doxorubicin.

[53] Lei Y, Hamada Y, Lia J, et al. Targeted tumor delivery and controlled release of neuronal drugs with ferritin nanoparticles to regulate pancreatic cancer progression. *J Control Release* 2016; 28;232:131-42.

[54] Blazkova I, Nguyen HV, Dostalova S, et al. Apoferritin modified magnetic particles as doxorubicin carriers for anticancer drug delivery. *Int J Mol Sci* 2013; 14:13391-402

[55] Liu, G., Wang, J., Lea, S.A., et al. Bioassay Labels Based on Apoferritin Nanovehicles. *ChemBioChem* 2006;9:1315–19

[56] Luo Y, Wang X, Du D, et al. Hyaluronic acid-conjugated apoferritin nanocages for lung cancer targeted drug delivery. *Biomater Sci* 2015;3:1386-94

[57] Falvo E, Tremante E, Fraioli R, et al, Antibody-drug conjugates: targeting melanoma with cisplatin encapsulated in protein-cage nanoparticles based on human ferritin. *Nanoscale* 2013;5:12278-85

[58] Lee EJ, Lee SJ, Kang YS, et al. Engineered proteinticles for targeted delivery of siRNA to cancer cells. *Adv Funct Mater* 2015;25:1279–86

[59] Aime S, Cabella C, Colombatto S, et al. Insights into the use of paramagnetic Gd(III) complexes in MR-molecular imaging investigations. *J Magn Reson Imaging* 2002;16:394-406

[60] Makino A, Harada H, Okada T, et al. Effective encapsulation of a new cationic gadolinium chelate into Apo and its evaluation as an MRI contrast agent. *Nanomedicine* 2011;7:638-46

[61] Sánchez P, Valero E, Gálvez N, et al. MRI relaxation properties of water-soluble Apo-encapsulated gadolinium oxide-hydroxide nanoparticles. *Dalton Trans* 2009;7:800-4

[62] Geninatti Crich S, Bussolati B, Tei L, et al. Magnetic resonance visualization of tumor angiogenesis by targeting neural cell adhesion molecules with the highly sensitive gadolinium-loaded apoferritin probe. *Cancer Res* 2006;66:9196-201

- [63] Kálmán FK, Geninatti-Crich S, Aime S. Reduction/dissolution of a beta-MnOOH nanophase in the ferritin cavity to yield a highly sensitive, biologically compatible magnetic resonance imaging agent. *Angew Chem Int Ed Engl* 2010;49:612-5
- [64] Meldrum FC, Douglas T, Levi S et al. Reconstitution of manganese oxide cores in horse spleen and recombinant ferritins. *Inorg Biochem.* 1995; 58:59-68
- [65] Geninatti Crich S, Cutrin JC, Lanzardo S et al. Mn-loaded apoferritin: a highly sensitive MRI imaging probe for the detection and characterization of hepatocarcinoma lesions in a transgenic mouse model. *Contrast Media Mol Imaging* 2012; 7:281-8
- [66] Szabó I, Crich SG, Alberti Dzabó I et al. Mn loaded apoferritin as an MRI sensor of melanin formation in melanoma cells. *Chem Commun (Camb).* 2012; 48:2436-8
- [67] Danon D, Goldstein L, Marikovsky Y et al., Use of cationized ferritin as a label of negative charges on cell surfaces. *J Ultrastruct Res* 1972; 38:500-10
- [68] Bartzokis G, Tishler TA. MRI evaluation of basal ganglia ferritin iron and neurotoxicity in Alzheimer's and Huntington's disease. *Cell Mol Biol (Noisy-le-grand)* 2000; 46:821-33
- [69] He D, Marles-Wright J. Ferritin family proteins and their use in bionanotechnology. *N Biotechnol* 2015;32:651-7
- [70] Zhen Z, Tang W, Todd T et al. Ferritins as Nanoplatfoms for Imaging and Drug Delivery. *Expert Opin Drug Deliv.* 2014; 11:1913-22.
- [71] Gossuin Y, Muller RN, Gillis P, et al. Relaxivities of human liver and spleen ferritin. *Magn Reson Imaging* 2005; 23:1001-4
- [72] Uchida M, Klem MT, Allen P et al. Biological Containers: Protein Cages as Multifunctional Nanoplatfoms. *Advanced Materials* 2007;19:1025-42
- [73] Uchida M, Terashima M, Cunningham CH, et al. A human ferritin iron oxide nanocomposite magnetic resonance contrast agent. *Magn Reson Med* 2008;60:1073-81

[74] Kitagawa T, Kosuge H, Uchida M, et al. RGD-conjugated human ferritin nanoparticles for imaging vascular inflammation and angiogenesis in experimental carotid and aortic disease. *Mol Imaging Biol* 2012;14:315–24

[75] Vannucci L, Falvo E, Fornara M, et al. Selective targeting of melanoma by PEG-masked protein-based multifunctional nanoparticles. *Int J Nanomedicine* 2012;7:1489-509

[76] Li X, Qiu LH, Zhu P, et al., Epidermal growth factor-ferritin h-chain protein nanoparticles for tumor active targeting. *Small* 2012;8:2505–14

[77] Fan K, Gao L, Yan X. Human ferritin for tumor detection and therapy. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2013;5:287-98

- An interesting review summarizing the versatility of APO in imaging.

[78] Lin X, Xie J, Niu G, et al. Chimeric ferritin nanocages for multiple function loading and multimodal imaging. *Nano Lett* 2011;11:814–19

[79] Lammers T, Kiessling G, Hennick WE, et al. Nanotheranostics and image-guided drug delivery: current concepts and future directions. *Nanomedicine* 2008;3:137-40

[80] Janib SM, Moses AS, MacKay JA. Imaging and drug delivery using theranostic nanoparticles. *Adv Drug Deliv Rev* 2010;62:1052-63

[81] Dostalova S, Cerna T, Hynek D, et al. Site-Directed Conjugation of Antibodies to Apoferritin Nanocarrier for Targeted Drug Delivery to Prostate Cancer Cells. *ACS Appl Mater Interfaces* 2016; 8:14430-41

- A very complete and well done study describing the formulation of an engineered APO structure applicable in theranostic.

[82] Liu XY, Wei W, Huang SJ et al. Bio-inspired protein–gold nanoconstruct with core–void–shell structure: beyond a chemo drug carrier. *J Mat Chem B* 2013;1:3136–43

[83] Cutrin JC, Crich SG, Burghilea, et al. Curcumin/Gd Loaded Apo: A Novel “Theranostic” Agent To Prevent Hepatocellular Damage in Toxic Induced Acute Hepatitis. *Mol Pharm* 2013;10:2079-85

[84] Kang HJ, Kang YJ, Lee YM, et al. Developing an antibody-binding protein cage as a molecular recognition drug modular nanoplatform. *Biomaterials* 2012; 33:5423-30

[85] Han JA, Kang YJ, Shin C. Ferritin protein cage nanoparticles as versatile antigen delivery nanoplatforms for dendritic cell (DC)-based vaccine development. *Nanomedicine* 2014; 10:561-9

[86] Kanekiyo M, Wei C-J, Yassine HM, et al. Self-assembling influenza nanoparticle vaccines elicit broadly neutralizing H1N1 antibodies. *Nature* 2013; 499:102–106

•• An excellent paper exploiting the applicability of APO for vaccine development.

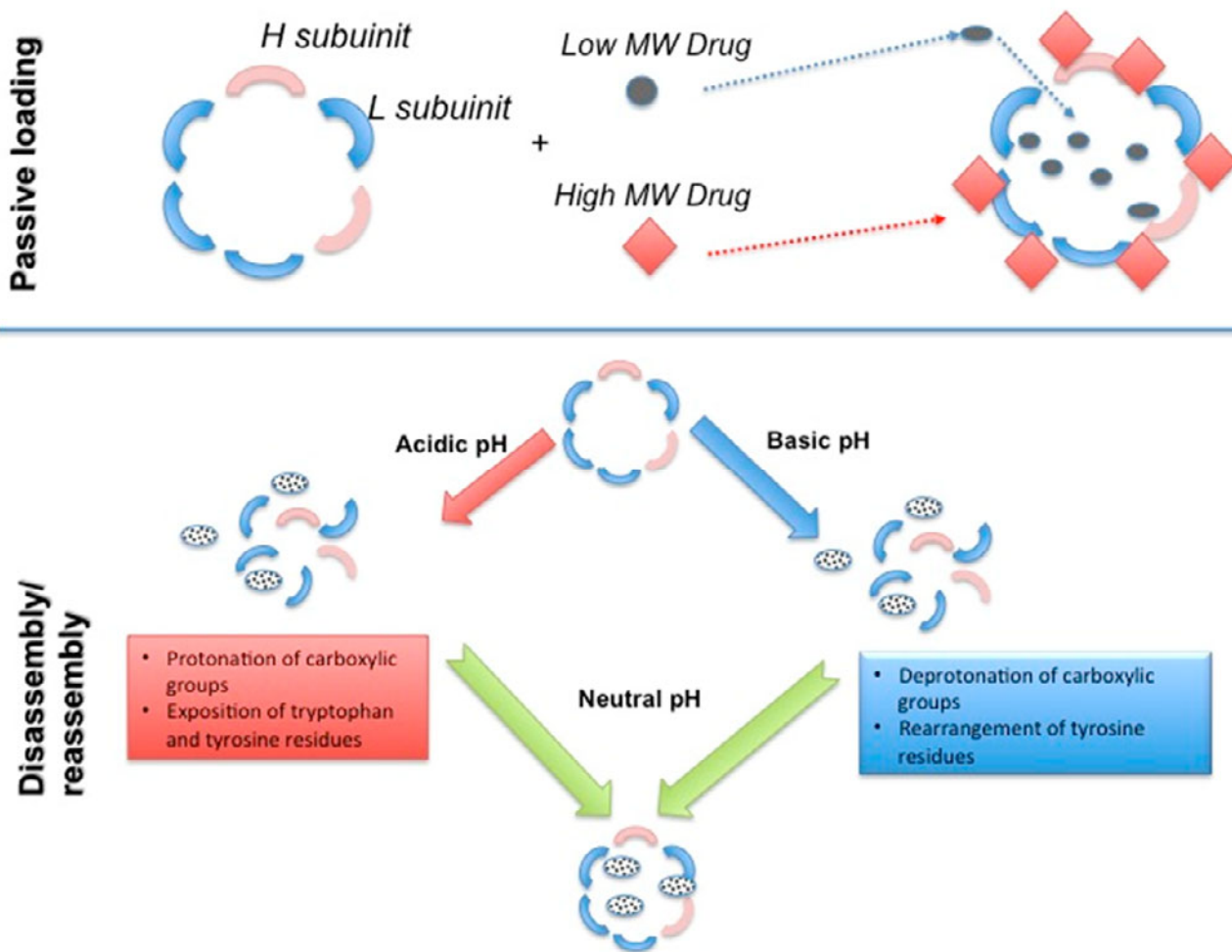
[87] Zhang Y, Tang Z, Wang J, et al. Apoferritin nanoparticle: a novel and biocompatible carrier for enzyme immobilization with enhanced activity and stability. *J Mater Chem* 2011;21:17468-75

[88] Pontillo N, Pane F, Messori L, et al. Cisplatin encapsulation within a ferritin nanocage: a high-resolution crystallographic study. *Chem Commun (Camb)*. 2016; 52:4136-9.

•• The first complete chemical physical and crystallographic characterization of APO loaded with a metal drug.

Fig 1: Graphical representation of the mechanisms for drug loading into APO.

Fig 1: Graphical representation of the mechanisms for drug loading into APO. A) APO formed by H/L subunit can be passively loaded by simple incubation with a drug solution. In function of the MW of the drug, different loading dynamics could be supposed. B) APO structure can be broken down to subunits at low or high pH, incubated with drug and finally the structure can be reassembled once the pH is tuned back to neutral lead the drug molecules to be trapped into the protein inner core.



ACCEPTED

Fig.2: Schematic illustration of APO application in therapy.

Fig.2: Schematic illustration of APO application in therapy. A) Different kind of active molecules were loaded or chemically linked on the surface of APO. These systems were applied for drug delivery and targeting, imaging and theranostic applications. B) Due to the intrinsic affinity for tumor cells loaded APO was mainly applied in cancer treatment, however recently this technology is applied in other field of research (immunotherapy, brain disease, enzyme replacement therapy).

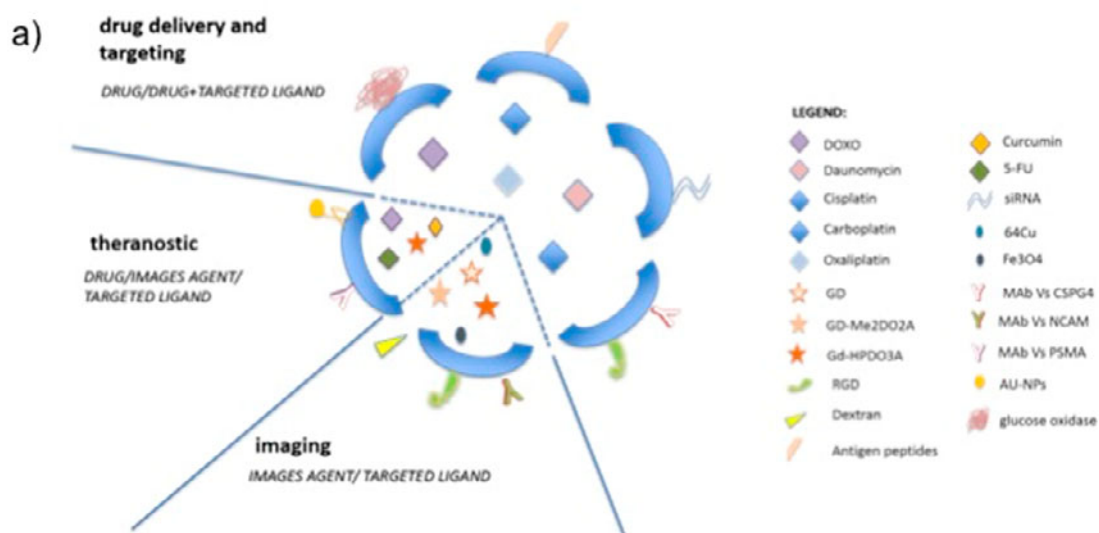
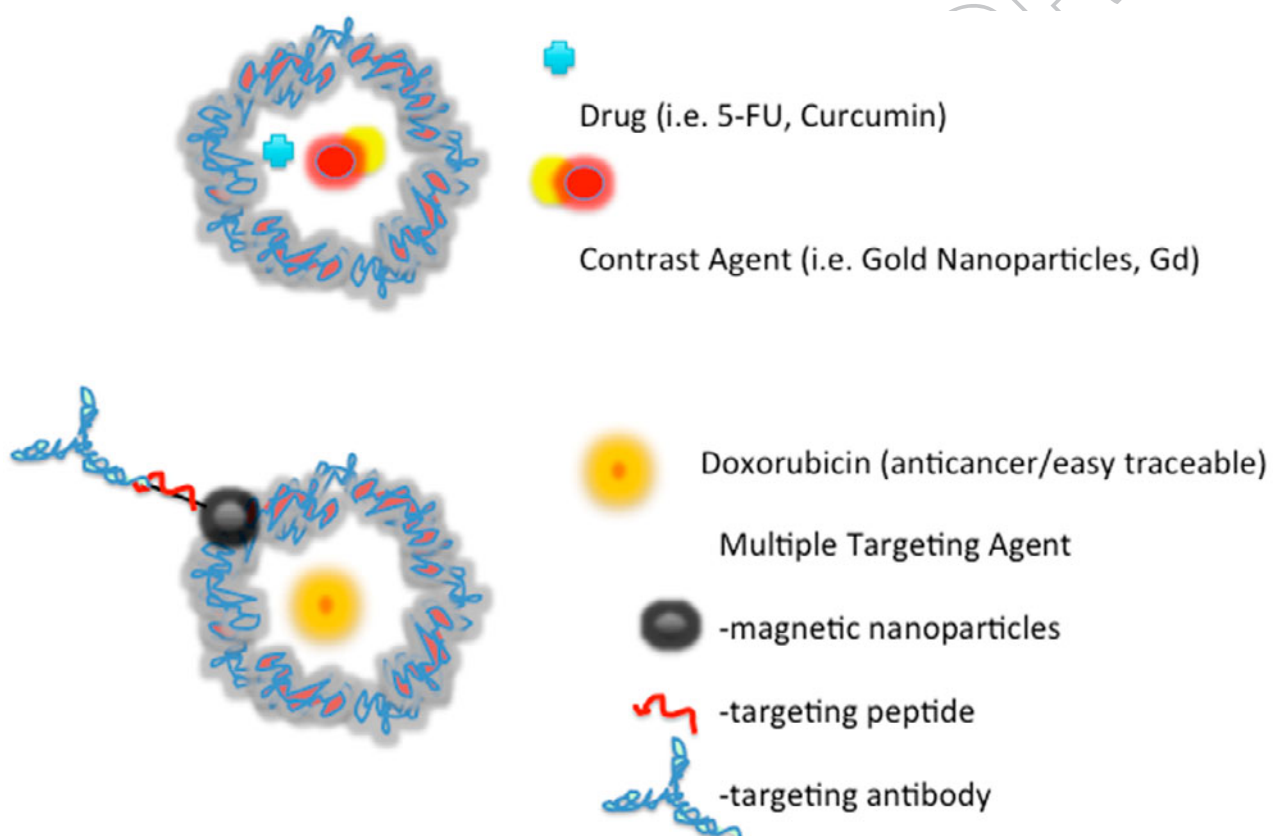


Fig.3. Theranostic Applications:

Figure 3. **Theranostic Applications:** a) co-encapsulation of active molecules (i.e. anticancer 5-FU, anti-oxidant curcumin) and contrast agents (i.e. gold nanoparticles, Gd) into APO cage; b) encapsulation of anticancer molecule (i.e. doxorubicin) and surface engineering with multiple ligands for targeting and detection



Drug	Method of encapsulation	Surface ligand	Source of APO	Outcomes/Shortcomings				
				1.Chemico-physical and technological studies	2. In vitro (VT)/in vivo (VV) model <i>Test adopted</i>	Structural/technological end point <i>Focus to solve</i>	Biological end points	Ref
Gefitinib	Passive loading at pH 7.2		Recombinant	TEM, UV-VIS, MALDI, gel electrophoreses	VT: breast cancer cells (SKBR3 and MDA-MB-231). <i>LSCM, FACS, MTT test</i>	Spherical cage of 12nm, stabilizing 10 molecules of drug. <i>To solve: localization of the drug.</i>	VT: cellular internalization mediated by interaction with the TrR1 receptor; selective antitumor activity in HER2 overexpressing cells.	[42]
Doxorubicin	Pre-complexation with Cu and incubation with intact APO	RGD	Recombinant (bacterial)	AFM, DLS, gel filtration chromatography, Fluorescence spectrometry	VT: Human glioblastoma cell line (U87MG). <i>CLFM, MTT test</i> VV: WT and mice bearing U87MG cells. <i>Pharmacokinetics; tissue distribution; histology; tumor volume</i>	Cage of 18.7 nm with a loading efficiency of 73%. <i>To solve: localization of the drug.</i>	VT: cellular internalization and cytotoxicity concentration dependent. VV: increased T(1/2) with respect to free drug. Accumulation into tumor tissue mediated	[45]

							by RGD-integrin interaction. Tumor growth inhibition respect controls (APO Vs saline or free dug).	
Doxorubicin	Disassembly in glycin buffer(pH 2.5) followed by reassembly at physiological pH		Horse spleen	gel electrophoreses, Fluorescence spectrometry		Drug loading of 23 molecules of drug. <i>To solve: structural conformation</i>		[48]
Doxorubicin	Disassembly at pH 2 followed by reassembly at physiological pH		Recombinant (H-Human)	UV-VIS, gel electrophoreses	VT: human colon adenocarcinoma cell line (Caco-2), human liver carcinoma cell line (HepG2). <i>LSCM and WB.</i>		VT: Rapid cellular uptake and nuclear translocation of loaded APO	[50]
Doxorubicin	Disassembly at acidic pH followed by reassembly at physiological pH		Recombinant (H-Human)	Fluorescence spectrometry	VT: Hela cells and breast cancer cells (MDA MB468).	Cage stabilizing 28 molecules of drug; long term controlled of drug release (up to 72 h) in PBS pH 7. <i>To solve: structural</i>	VT: Rapid cellular internalization and cytotoxicity (Apo Vs free dug)	[51]

					<i>LSCM, MTT</i>	<i>conformation</i>		
Doxorubicin	Disassembly in urea buffer followed by reassembly at physiological pH		Recombinant (bacterial)	TEM, DLS, UV-VIS, HPLC-UV.	<p>VT: human colon cancer cell line (HT-29). <i>CLFM</i>.</p> <p>VV: mice bearing tumor cells (HT-29 or A375 or MDA-MB-231).</p> <p><i>Pharmacokinetics ; tumor volume</i></p>	<p>Spherical cage of 14nm, stabilizing 33 molecules of drug. pH-dependent drug release.</p> <p><i>To solve: structural conformation</i></p>	<p>VT: cellular internalization and accumulation into lysosomes.</p> <p>VV: Increased plasma half-life. Preferential accumulation in tumor tissue respect to healthy one. Tumor growth inhibition. (APO Vs free drug or Doxil)</p>	[52]
Doxorubicin Atropine Carbachol	Disassembly in urea buffer followed by reassembly at physiological pH		Not reported	TEM, DLS, UV-VIS, HPLC-UV	<p>VT: pancreatic cancer cells (PANC-1). <i>LCSM</i>.</p> <p>VV: WT and mice bearing PANC-1.</p> <p><i>Pharmacokinetics ; tumor volume</i></p>	<p>Spherical cage of 11nm, stabilizing 33 (DOXO), 47 (Atropine) or 58 (Carbachol) molecules of drug. pH-dependent drug release.</p> <p><i>To solve: structural conformation</i></p>	<p>VT : Rapid accumulation into tumoral cell through interaction with TfR1 receptors.</p> <p>VV: long term circulation. Accumulation</p>	[53]

							into tumor site. Reduction of tumor volume and increased survival rate. (loaded APO Vs free drug)	
Daunomycin	Pre-complexation with PLLA. Disassembly at pH 5 followed by reassembly at physiological pH		Not reported	Fluorescence spectroscopy		Increased drug stabilization (4 times higher) respect to non complexed drug. <i>To solve: chemical and structural characterization.</i>		[24]
Daunomycin	Pre-complexation with PLLA. Disassembly at pH 5 followed by reassembly at physiological pH	HA	Not reported	TEM, UV-VIS-UV, FTIR, Fluorescence spectroscopy	VT: Human lung carcinoma cell line (A549) and human lung fibroblast (MRC-5). <i>MTT, LCSM</i>	Spherical cage of 28 nm. pH dependent drug release. <i>To solve: localization of the drug</i>	VT: Rapid accumulation into cancer cell expressing CD44 and cytotoxicity (APO Vs free drug).	[56]
Cisplatin Carboplatin Oxaliplatin	Disassembly at pH 2 followed by reassembly at physiological pH		Not reported	DLS, UV-VIS, CD, ICP-MS	VT: rat pheochromocytoma cell line (PC12).	Spherical cage of 13 nm encapsulating 45 (cisplatin), 17 (carboplatin), 23 (oxaliplatin) molecule of drug.	VT: Increased uptake and cytotoxicity of actives after loading (respect to free	[35]

					<i>MTT, ICP-MS</i>	<i>To solve: localization of the drug</i>	drug).	
Cisplatin Carboplatin	Disassembly at pH 2 followed by reassembly at physiological pH/Passive loading by pre incubation with Pt		Not reported	TEM, UV-VIS, NMR, ICP-AES, gel electrophoreses	VT: rat pheochromocytoma cell line (PC12). <i>MTT</i>	Cage encapsulating 15 of drug <i>To solve: localization of the drug, structural characterization</i>	VT: Decreased viability of cancer cells (loaded Vs unloaded APO)	[46]
Cisplatin	Disassembly at pH 13 followed by reassembly at physiological pH		Pig pancreas	TEM, UV-VIS CD, ICP-MS.	VT: gastric cancer cells (BGC823) and Hela. <i>FACS, MALDI-TOF/TOF, RT-PCR.</i>	Spherical cage encapsulating 12 Molecules of drug. <i>To solve: localization of the drug</i>	VT: Induction of apoptosis.	[47]
Cisplatin	Disassembly at pH 2 followed by reassembly at physiological pH	Antibody direct to CSPG4	Recombinant (H-bacterial)	DLS, AUC, SEC	VT: melanoma cell line (CSPG4+) and breast cancer cells (CSPG4-). <i>Proliferation test</i> VV: CD1 nude mice bearing tumor.	Protein cage of 33 nm with a drug/protein molar ratio of 50 <i>To solve: localization of the drug, structural</i>	VT: increased antiproliferative effect (modified Vs not modified APO). VV: reduction of tumor size in mice bearing melanoma.	[57]

					<i>Evaluation of tumor size</i>	<i>characterization and quantification of ligand</i>		
siRNA (fluorescent model sequence)	Incubation of modified APO with siRNA	CAP and CTP	Recombinant (H-Human)	DLS, TEM, polyacrylamide gel	VT: human breast cancer cells (MDA-MB-468) and vimentin expressing murine melanoma (B16F10). <i>CLSM</i>	Complex of 53 nm stabilizing siRNA <i>To solve: structural characterization</i>	VT: Rapid accumulation into cytoplasm of tumor cells	[58]

Table 1: APO utilization in drug delivery to cancer

LEGEND: AUC: analytical ultracentrifugation; CAP: Cationic peptides; CD: circular dichroism; CSPG4: Chondroitin sulfate proteoglycan 4; CTP: cell penetrating peptide; DLS: dynamic light scattering; FACS: fluorescence activated cells sorting; HA: Hyaluronic acid; HER2: Epidermal growth factor receptor 2; HPLC-UV: High-performance liquid chromatography- UV; ICP-EAS: Inductively coupled plasma atomic emission spectroscopy; ICP-MS: inductively coupled plasma mass spectrometer; LSCM: Laser-scanning Confocal Microscopy; MALDI TOF/TOF: Matrix-assisted laser desorption/ionization; MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium; NMR: Nuclear magnetic resonance; PLLA: poly-L-aspartic acid; RGD: peptide with affinity for integrin receptor upregulated on tumor endothelial cells; RT-PCR: Reverse transcriptase-polymerase chain reaction; SEC: size exclusion chromatography; TEM: transmission electron microscopy; TfR1: transferrin receptor; UV-FTIR: Fourier transform infrared spectroscopy; UV-VIS: Ultraviolet-visible spectroscopy; WB: western blot.

Drug	Method of encapsulation	Surface modification	Source of APO	Outcomes/Shortcomings				
				1. Chemico-physical and technological studies	2. In vitro (VT)/in vivo (VV) model <i>test adopted</i>	Structural/technological end point <i>Focus to solve</i>	Biological end points	REF
Gadolinium	Passive loading		Not reported	IR, TEM, UV-Vis, ICP, SDS-PAGE		Small cage stabilizing GD (III); increase GD longitudinal and transverse relaxivities <i>To solve: structural characterization; localization of the drug.</i>		[61]
GD-Me ₂ DO ₂ A	Disassembly at pH 2 followed by reassembly at physiological pH. Drug was added during the basification process.	dextran	Not reported	TEM, UV-VIS, ICP-EAS	VV: mice bearing HeLa cells. <i>MRI, NIRF</i>	Spherical 30 nm modified cage encapsulating 36 molecules of contrast agent. <i>To solve: structural characterization; localization of the drug.</i>	VV: massive accumulation in tumor site and high proton relaxivity.	[60]
GD-HPDO ₃ A	Disassembly at pH 2 followed by reassembly at physiological pH	NCAM	Not reported		VT: renal cell carcinoma cells (TEC). <i>ICP-MS, MRI</i> VV: SCID mice bearing TEC cells. <i>MRI, histology,</i>	Modified cage encapsulating 8-10 molecules of contrast agent with high relaxivity (5 times higher respect to free molecule)	VT: efficient cellular internalization (loaded/modified APO Vs loaded APO) VV: accumulation	[62]

					<i>immunofluorescence.</i>	<i>To solve: structural characterization; localization of the drug.</i>	into tumor site (modified/loaded APO>loaded APO)	
GD-HPDO3A	Disassembly at pH 2 followed by reassembly at physiological pH		Not reported			Cage encapsulating 10 molecules of contrast agent. Relaxivity constant for several days. <i>To solve: structural characterization; localization of the drug.</i>		[59]
⁶⁴ Cu+Cy5	Dissociation at pH 2 followed by reconstruction at physiological pH	RGD4C	Not reported	DLS , TEM	VV: tumor bearing U87MG glioma. <i>NIRF, PET, LCSM</i>	<i>Absence of structural analyses and quantification of exposed ligands</i>	VV: massive tumor accumulation mediated by RGD-integrin interaction.	[78]
Fe ₃ O ₄	Mineralization procedure	None	Recombinant (H-Human)	DLS, TEM, MR	VT: murine macrophage cells. <i>MR, ICP-MS</i>	Cage of 12-13 nm encapsulating 5000 Fe. <i>To solve: structural characterization.</i>	VT: uptake of Apo into cells and accumulation of 116 ug Fe/cell within 72 h	[72]
Fe ₃ O ₄	Mineralization procedure	RGD4G	Recombinant (H-Human)	DLS, TEM, magnetic measurement	VT: human melanoma cell line (C32).	Spherical surface modified cage of 14 nm encapsulating 5000 molecules of Fe	VT: specific cellular uptake mediated by interaction with integrin.	[73]

					<i>FACS, TEM</i>			
Fe ₃ O ₄	Mineralization procedure	RGD	Recombinant (H-Human)		<p>VT: bovine aortic endothelial cells.</p> <p><i>Fluorescence microscopy</i></p> <p>VV: WT mice.</p> <p><i>Fluorescence microscopy, histology</i></p>		<p>VT: Enhanced cellular uptake (modified vs not modified APO)</p> <p>VV: High accumulation in carotid arteries (modified vs not modified APO)</p>	[74]
Fe ₃ O ₄	Mineralization procedure	EGF (ligand for EGFR)	Recombinant (H-Human)	DLS, TEM.	<p>VT: breast epithelial cells (MCF-10A cells), breast cancer cells (MCF-7).</p> <p><i>FACS, MTT</i></p> <p>VV: nude mice bearing MDA.MB-231 tumor cells.</p> <p><i>Weight and tumor size, fluorescence microscopy</i></p>	Spherical cage of 19 nm	<p>VT: absence of toxicity at high APO concentration. Good targeting through cancer cell respect to normal cells (interaction with EGFR)</p> <p>VV: no influence in body weight and tumor size (modified/loaded APO Vs saline). Accumulation into tumor tissue.</p>	[76]

Mn	Incubation with MnCl ₂ at pH 9 followed by reduction	None	Horse spleen	TEM, UV-VIS, size exclusion chromatography, NMR		Protein shell with a mineral core. Encapsulation of about 1090 Mn atoms. <i>To solve: structural characterization and localization of the contrast agent</i>		[63]
Mn	Incubation with MnCl ₂ at pH 9 followed by reduction	None	Horse spleen and recombinant (H and L Human)	TEM, UV-VIS, EDXA		Protein shell encapsulating mineral core <i>To solve: structural characterization.</i>		[64]
Mn	Incubation with MnCl ₂ at pH 9 followed by reduction	None	Horse spleen	ICP-MS, Bradford quantification	VT: HTC and hepatocytes <i>ICP-MS</i> VV: WT and HBV-Tg mice <i>Biodistribution, MRI</i>	Protein structure encapsulating 3000 Mn(II) ions <i>To solve: structural conformation</i>	VT: high accumulation in healthy cells respect to hepatoma VV: detection of tumor lesion and improved sensitivity respect to commercial probe for MRI	[65]
Mn	Incubation with MnCl ₂ at pH 9	None	Not reported	UV-VIS, NMR	VT: melanogenic cells (B16-F10m) and non		VT: discrimination between	[66]

	followed by reduction				melanogenic cells (B16-F10non-m) <i>UV-VIS; ICP-MS</i> VV: mouse xenograft model bearing B16-F10m <i>MR</i>		melanogenic and non melanogenic cells; high cellular uptake in melanogenic cells. VV: APO accumulation into tumor site	
--	-----------------------	--	--	--	---	--	--	--

Table 2: APO utilization in diagnosis

LEGEND: EDXA: Energy Dispersive X-ray Analysis; LSCM: Laser-scanning Confocal Microscopy; ICP-MS: Inductively coupled plasma mass spectrometry; NMR: nuclear magnetic resonance; MR: magnetic resonance; NCAM: neural cell adhesion molecule; NIRF: near infrared fluorescence; PET: positron emission tomography; IR: Infrared spectroscopy; TEM: transmission electron microscopy; UV-VIS: Ultraviolet-visible spectroscopy; SDS-PAGE: Sodium Dodecyl Sulphate - PolyAcrylamide Gel Electrophoresis; ICP-EAS: Inductively coupled plasma atomic emission spectroscopy; DLS: : dynamic light scattering; MRI: Magnetic resonance imaging; ICP-MS: inductively coupled plasma mass spectrometer; PET: Positron Emission Tomography; MTT: 3-(4,5-dimethyliazol-2-il)-2,5-difeniltetrazolio; FACS: fluorescence activated cells sorting; RGD: Arginylglycylaspartic acid; EGF: Epidermal growth factor;

Drug	Method of encapsulation	Surface modification	Source of APO	Outcomes/Shortcomings				
				1. Chemico-physical and technological studies	2. In vitro (VT)/in vivo (VV) model test adopted	Structural/technological end point <i>Focus to solve</i>	Biological end points	REF
Curcumin + Gd-HPDO3A	Disassembly at pH 2 followed by reassembly at physiological pH.		Horse spleen	SEC, UV-VIS, ICP-MS	VV: male mice (C57BL), mice model of liver injury. <i>MR, histology</i>	Protein cage of 19 nm encapsulating 9.5 and 0.4 molecules of curcumin and Gd-HPDO3A respectively. Ability to protect drug from degradation. <i>To solve: structural characterization</i>	VV: preferential uptake by the liver (about 19% of injected dose). Reduction of ALT, leucocyte infiltration and hepatocyte apoptosis levels. (loaded Vs native APO).	[83]
AU-NP and 5-FU	Syntheses of AU-NPs inside the protein cage (AU reduction) followed by a passive loading of 5-FU		Not reported	TEM, EDXA, PAGE, CD, UV-VIS, ¹⁹ F-NMR.	VT: non cancer cells (HKC); cancer cells (HeLa, HepG2, Caco-2, human hepatic L02). <i>CCK-8, CLSM, TEM, ICP-MS, FACS, BrdU, WB</i>	Regular protein cage encapsulation 45 molecules of drug. pH dependent drug release <i>To solve: localization of the drug</i>	VT: Accumulation of APO into lysosomes; low toxicity in control cells; decreasing of IC50 value (15 times Vs free drug) and increase of the antiproliferative effect in cancer cells.	[84]
Doxorubicin	Disassembly at pH 2	Magnetic nanoparticles	Not reported	Fluorimetric analysis, CE		Partial encapsulation of drug into protein cage		[54]

	followed by reassembly at physiological pH					and pH triggered drug release. <i>To solve: structural characterization; localization of the drug</i>		
Doxorubicin	Disassembly at pH 2 followed by reassembly at physiological pH	HWR peptide and anti PSMA antibodies; AU-NP	Horse spleen	DLS, FT-IR, PAGE, ICP-MS	VT: Human umbilical vein endothelial cells (HUVEC), human prostate adenocarcinoma cells (LNCCaP). <i>FACS</i>	Complex protein cage of 22 nm, pH dependent drug release.	VT: specific uptake in tumoral cells and time dependent inhibition of cell growth. Excellent hemocompatibility.	[81]

Table 3: APO utilization in theranostic applications

LEGEND: AU-NPs: gold nanoparticles; BrdU: Bromodeoxyuridine; CCK-8: cell counting kit; CD= circular dichroism; CE: Capillary electrophoresis; CLSM: confocal laser scanning microscopy; DLS: dynamic light scattering; EDXA: energy dispersive X-ray analysis; FACS: fluorescence activated cells sorting; FACS: fluorescence activated cells sorting; FT-IR: Fourier transform infrared spectroscopy; ICP-MS: inductively coupled plasma mass spectrometer; MR: magnetic resonance; NMR: nuclear magnetic resonance; PMSA: prostate specific membrane antigen; SEC: size exclusion chromatography; TEM: transmission electron microscopy; UV-VIS: Ultraviolet-visible spectroscopy; WB: western blot; 5-FU: 5-Fluorouracil.

Drug	Method of encapsulation	Surface modification	Source of APO	Outcomes/Short comings				
				1.Chemico-physical and technological studies	2. In vitro/in vivo test	Structural/technological end point	Biological end points	REF
		OT-1 and OT-2	Recombinant (bacterial)	DLS, TEM, MS	VT: splenocyte from C57BL/6 mice. <i>FACS.</i> VV: C57BL/6 mice. <i>Cytokine assay</i>	Spherical cage of 13 nm exposing antigen in the inner or outer surface. <i>To solve: structural characterization</i>	VT: increased of T cell proliferation after APO exposure. VV: effective differentiation of activated T cells in functional effector T cells	[85]
	Incubation of recombinant protein in TRIS buffer at physiologic pH and purification	HA	Recombinant (bacterial)	DLS, TEM, SEC.	VV: female BALB/c mice	Smooth spherical cage exposing antigens <i>To solve: structural characterization</i>	VV: inhibition of hemoagglutination	[86]
glucose oxidase	Surface attachment through biotin-streptavidin technology		Not reported	TEM, UV-VIS		Cage of 16 nm with an irregular morphology and largely aggregated stabilizing 8 molecules of enzyme. <i>To solve: structural characterization</i>		[50]

Table 4: APO utilization in therapy

LEGEND: DLS: dynamic light scattering; FACS: fluorescence activated cells sorting; HA: Influenza virus hemagglutinin; MS: mass spectrometry; OT1/OT²: Antigen peptides derived from ovalbumin; SEC: size exclusion chromatography; TEM: transmission electron microscopy; UV-VIS: Ultraviolet-visible spectroscopy;

ACCEPTED MANUSCRIPT

	Category	Drug	Note	Ref
Anticancer	Tyrosine kinase inhibitor	Gefinitinib	10 molecules/APO loading	42
	Anthracyclin	Doxorubicin	-pre-complexation with Cu ²⁺ increase the drug loading	43-45
	Anthracyclin	Daunorubicin	-acidic protocol -surface conjugation for targeting -pre-complexation with negatively charged peptides	22, 56
	Alkylating agents	Cisplatin	-strong acidic protocol -2 molecules/APO	46
	Alkylating agents	Carboplatin	-strong acidic protocol -5 molecules/APO	46
	Alkylating agents	Cisplatin	-alkaline protocol	47
	Alkylating agents	Cisplatin	-alkaline protocol -surface modification	34,57
	Antimuscarinic	Atropine	/	53
	Cholinomimetic	Carbachol	/	53
		SiRNA	-modification with protamine-derived peptides for complexation	58
	Diagnostic	Gadolinium	-acidic protocol -neutral chelates	59-62
		Gadolinium	-neutral chelates -surface modification for tumor targeting	62

	Gadolinium	- acidic dissociation -positively charged complexation	60
	Magnetic Fe ₃ O ₄	-iron core formation	72-73
	Magnetic Fe ₃ O ₄	-surface engineering for tumor targeting	74-75
	Cu ⁶⁴ Cy5	-surface modification for tumor targeting	78
	Mn	-alkaline protocol -accumulation in tumor	63,64,65,66

Table 5: Summary of molecules and active agents loaded into APO nanocages.