



METHOD ARTICLE

A CRISPR/Cas9-based method and primer design tool for seamless genome editing in fission yeast [version 1; referees: 1 approved, 1 approved with reservations]

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Abstract

In the fission yeast *Schizosaccharomyces pombe* the prevailing approach for gene manipulations is based on homologous recombination of a PCR product that contains genomic target sequences and a selectable marker. The CRISPR/Cas9 system has recently been implemented in fission yeast, which allows for seamless genome editing without integration of a selection marker or leaving any other genomic ‘scars’. The published method involves manual design of the single guide RNA (sgRNA), and digestion of a large plasmid with a problematic restriction enzyme to clone the sgRNA. To increase the efficiency of this approach, we have established and optimized a PCR-based system to clone the sgRNA without restriction enzymes into a plasmid with a dominant *natMX6* (nourseothricin) selection marker. We also provide a web-tool, CRISPR4P, to support the design of the sgRNAs and the primers required for the entire process of seamless DNA deletion. Moreover, we report the preparation of G1-synchronized and cryopreserved *S. pombe* cells, which greatly increases the efficiency and speed for transformations, and may also facilitate standard gene manipulations. Applying this optimized CRISPR/Cas9-based approach, we have successfully deleted over 80 different non-coding RNA genes, which are generally lowly expressed, and have inserted 7 point mutations in 4 different genomic regions.

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report



report

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Introduction

The fission yeast *Schizosaccharomyces pombe* is a potent genetic model organism. Gene deletions and other genetic manipulations in *S. pombe* are most commonly performed in a single-step by transformation of a PCR product, which includes a selectable marker gene along with flanking regions to target the genomic region to be manipulated¹. Several techniques have been developed to circumvent complications caused by selectable markers, including the LoxP-Cre recombinase system², the *rpl42* (cycloheximide resistance)-based method^{3,4}, the pop-in/pop-out methods⁵, or a recent method for scar-less gene tagging⁶. However, these methods have drawbacks that limit their applicability: they either involve two transformations or selection steps, leave ‘DNA-scars’, affect cellular physiology, or can only be used in specific genetic backgrounds.

The recent emergence of the prokaryotic CRISPR/Cas9 system for genome editing now provides the opportunity for efficient gene manipulation without any markers⁷⁻⁹. Such seamless genome editing offers several advantages: 1) it allows targeting of multiple genetic manipulations to the same strain without restrictions due to markers or any marker recycling; 2) it avoids indirect physiological effects, which accompany some markers (10; M. R.-L. and C. C., unpublished observations); and 3) it limits the perturbation of the local chromatin and transcriptional environment to the gene manipulation of interest.

The CRISPR/Cas9 genome editing system has recently been implemented in fission yeast by applying the promoter/leader sequence of K RNA (*rkl1*) and a hammerhead ribozyme to express the single guide RNA (sgRNA)¹¹. The Cas9 protein acts as an RNA-guided endonuclease that binds to a protospacer adjacent motif (PAM) site and introduces a double-strand break (DSB) three base pairs upstream of the PAM site in the spacer sequence. The cell can then repair this DSB either by non-homologous end joining, which will introduce point mutations or indels, or by efficient homologous recombination if the cell is provided with a suitable template. This approach allows for the precise editing of genomic locations without the need of any selectable marker, since cells that do not repair the DSB will die. Genome editing in *S. pombe* with CRISPR/Cas9 involves the manual identification of unique PAM and spacer sequences (sgRNA) and cloning of these sequences into an expression plasmid with a *CspCI* restriction site to produce the sgRNA. Overexpression of the Cas9 enzyme is detrimental for *S. pombe* growth, which is partially circumvented by co-expression of the sgRNA and Cas9 from the same plasmid¹¹. However, the resulting large plasmid (~11 kb) is difficult to work with, and the *CspCI* digestion required for cloning is often very inefficient. Accordingly, we and others have encountered serious problems in implementing the CRISPR/Cas9 system.

Here, we present a PCR-based, rapid and efficient method for the seamless deletion of any DNA sequence in the *S. pombe* genome, or other genome manipulations, such as point mutations, by applying modifications and optimizations of the CRISPR/Cas9 system.

We also provide the CRISPR4P web tool to design the different types of primers required for deletion of any genomic region: PCR-based sgRNA cloning, PCR-based synthesis of DNA template for deletion by homologous recombination, and checking primers to confirm the deletion. Furthermore, we have modified a protocol for the generation of cryopreserved *S. pombe* cells¹², by implementing G1 synchronization and optimizations, which substantially increases the efficiency of successful transformations, especially for regions that are difficult to delete. This protocol may also facilitate the manipulation of genomic regions using the traditional method¹.

Overview of approach

The main steps of the CRISPR/Cas9-based method to generate gene deletions are briefly highlighted below. Figure 1 provides a flow diagram of the main steps. A more detailed methodology is

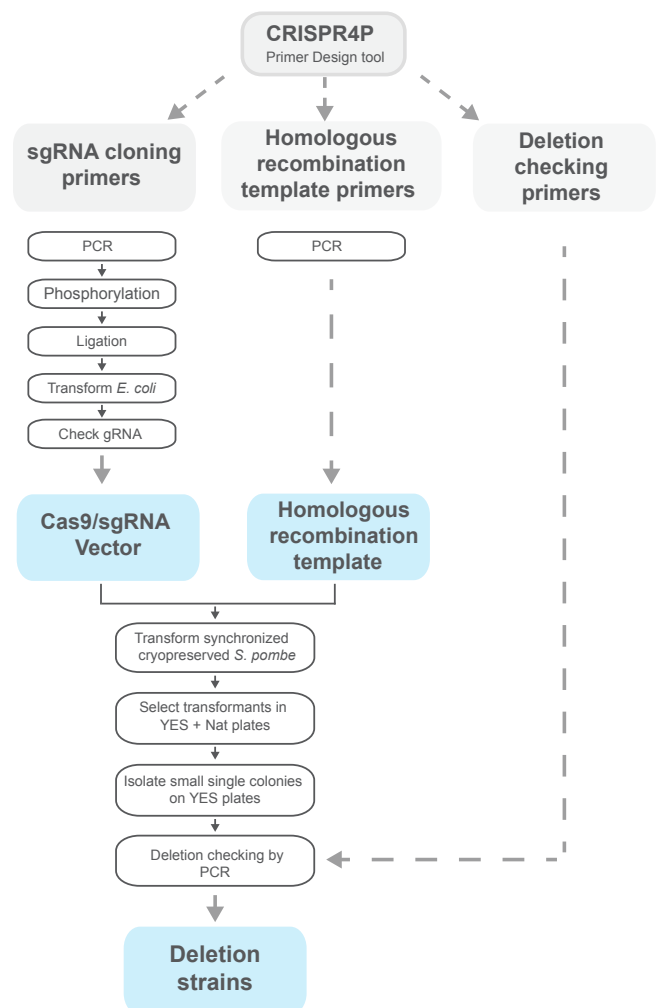


Figure 1. Flow diagram for CRISPR/Cas9-based approach for seamless genome editing in fission yeast. sgRNA, single guide RNA.

available at the end of the manuscript and as a PDF in [Supplementary File 1](#). The entire procedure takes about 8 days, including about 5 days for incubation.

1. Identify suitable sgRNAs to target region of interest using CRISPR4P tool²⁶ (bahlerlab.info/crispr4p) (Figure 2).
2. Design of primers required for whole process using CRISPR4P (Figure 3A): 1) sgRNA cloning; 2) synthesis of DNA template for homologous recombination (HR template) for gene deletion; and 3) checking primers to confirm gene deletion.
3. Clone sgRNAs into nourseothricin-selectable plasmid pMZ379 that contains Cas9 enzyme gene, the *natMX6* selection marker and the *rrk1* promoter/leader (Figure 3B; Figure 4).
4. Generate HR template by PCR using primers with sequences flanking the region of interest and overlapping at their 3' ends (Figure 3C).
5. Delete region of interest by co-transforming sgRNA/Cas9-plasmid and HR template into *S. pombe* cells that have been synchronized and cryopreserved to increase transformation efficiency (Figure 3D).
6. Select the smallest colonies from selective plate (Figure 3E) and check these colonies for deletion junction by colony PCR (Figure 3F).

Note that this approach can be adapted for applications other than gene deletions, such as insertion of point mutations or tags. The CRISPR4P tool allows the user to identify possible sgRNA sequences in any region of interest for other applications of the CRISPR/Cas9-based genome editing.

CRISPR4P primer design tool

Available primer design programs for gene targeting in *S. pombe* allow the manipulation of coding sequences using the standard PCR-based method¹³, or rely on current gene annotations to generate a database that contains primers for deletion of non-coding RNAs, 3'-UTRs or tRNAs¹⁴. We have designed an online tool, written in Python 2.7 (www.python.org/), to help with the design of all the different primers required for CRISPR/Cas9-based deletion of virtually any region in the *S. pombe* genome. This tool, named CRISPR4P (CRISPR 'for' *Pombe* or CRISPR *Pombe* PCR Primer Program), is freely available from our website (bahlerlab.info/crispr4p)²⁶. CRISPR4P designs PCR primers for sgRNA cloning and primers to generate the HR template, and also checks primers to verify gene deletions. Figure 2 provides an overview of the workflows for CRISPR4P, and Figure 3A provides an overview of the different primers that can be designed by CRISPR4P.

Design of sgRNA

The sgRNA targets the Cas9 enzyme to its recognition site to generate a DSB upstream of the PAM sequence. However, it has been reported that Cas9 can also generate DSBs in other genomic sequences that contain a few mismatches compared to the sgRNA^{15,16}, and even in sites that cannot be predicted simply by sequence homology¹⁷. Thus, it is important to choose a suitable

target region to maximize the specificity and avoid undesirable off-target effects. Mismatches within the 12 nucleotide 'seed' sequence, immediately upstream of the PAM sequence, reduce the nuclease activity of the Cas9 enzyme, and must therefore be avoided for the target sequence¹⁸. On the other hand, such mismatches in similar sequences elsewhere in the genome will reduce the likelihood of Cas9 targeting. Multiple tools are becoming available for the prediction of sgRNAs and off-target effects (see 19 for a review), but not all include the *S. pombe* genome, and there have been no studies into the issue of off-target effects in *S. pombe*. CRISPR4P facilitates the design of sgRNAs and provides basic information on the similarity of sgRNA sub-sequences to other genomic sgRNA sequences to minimize off-target effects. In the case of gene deletions, there is considerable flexibility with respect to sgRNA selection because the targeting is not limited to a narrow region.

CRISPR4P has scanned the *S. pombe* genome, downloaded from PomBase, for all possible 3-nucleotide *Streptococcus pyogenes* Cas9 PAM sites (5'-NGG-3'), and stored this information together with the sequences of the 20 nt upstream of all these PAM sites (sgRNA sequences), thus generating a database of all possible genomic sgRNAs plus PAM sites. Users can input their target regions either by gene name or genomic coordinates, with the latter providing the flexibility to delete any region of interest, such as regulatory sequences, non-coding RNAs, or specific sub-regions of genes. If the input is a gene name, the coordinates of the coding sequence are calculated based on PomBase annotation (<http://www.pombase.org/>)^{20,21}. CRISPR4P then examines the nucleotide string within the input coordinates of the target sequence for PAM sites along with the upstream 20 nt sgRNAs using the genomic database of all sgRNAs plus PAM sites. CRISPR4P is not an off-target scorer, but helps users in the selection of suitable sgRNAs, based on basic concepts of similarity to other regions. Our premise is that the chosen 20 nt sgRNA should be unique in the genome, and only unique sgRNAs will therefore be provided in the output. In addition, CRISPR4P then presents all the possible sgRNAs in the target region, ranked by similarity to other putative sgRNAs anywhere in the genome. The data to the right of each sgRNA indicates the numbers of other genomic sgRNA sequences that share a given number of nucleotides (starting from the 5' end of the PAM sequence), scanning the sgRNA from 8 bp up to 20 bp every 2bp. To minimize any off-target effects, the 12 nt 'seed' sequence immediately upstream of the PAM site should ideally be unique in the genome. Furthermore, we do recommend the use of at least two different sgRNAs for any given deletion construct and to test the phenotypes of several independent deletion strains from each transformation. Any specific off-target mutation is unlikely to occur independently in different clones, and even less likely to occur with different sgRNAs.

A specific sgRNA can be selected by clicking the round button to the left of the sequence; CRISPR4P will then provide the corresponding outputs at the bottom, including the sgRNA sequence together with its coordinates and the two primer sequences required to clone the sgRNA into the plasmid pMZ379 by PCR (Figure 3A and B). CRISPR4P also provides two other sets of PCR primers described below.

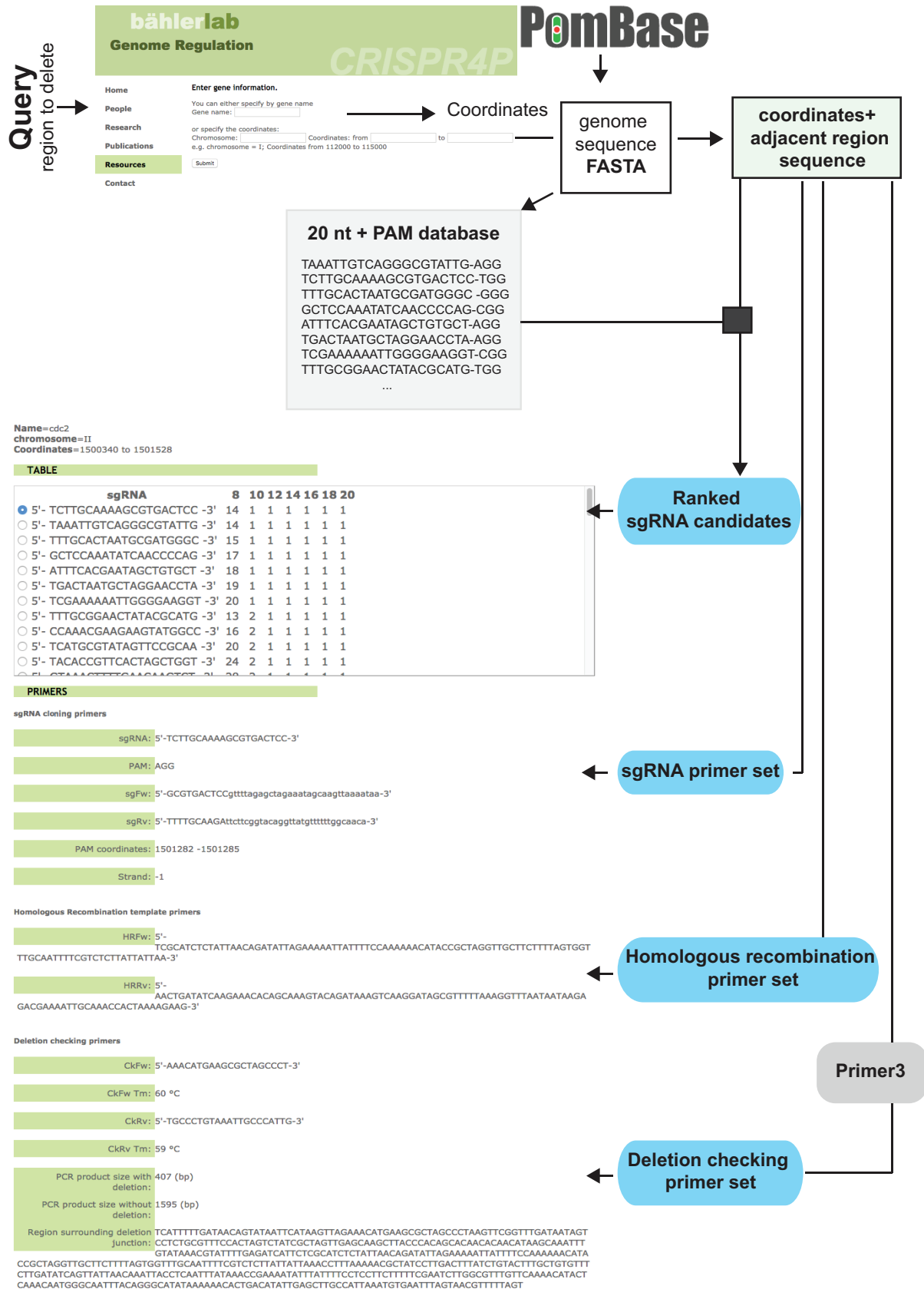


Figure 2. Overview of workflows for CRISPR4P tool for sgRNA and PCR primer design. sgRNA, single guide RNA; PAM, protospacer adjustment motif.

Primer design for HR template

CRISPR4P selects 80 nt up- and down-stream of the target sequence to be deleted and joins these sequences together into a 160 nt long HR template sequence to target the region of interest for seamless deletion by homologous recombination. This 'junction' sequence is then used to design the primers to generate the HR template DNA by PCR amplification (Figure 3A and C). The forward primer (HRfw) contains the 100 nt from the 5'-end of the HR template, and the reverse primer (HRRv) are the reverse complementary 100 nt from the 3'-end of the HR template. We have found that 20 nt of overlapping region between these two PCR primers are sufficient to generate the HR template.

Primer design to check deletion junction

CRISPR4P also provides two PCR primers to check the seamless deletion junction. These primers are positioned up- and down-stream of the HR template region. First, CRISPR4P generates *in silico* a region surrounding the deletion junction by joining the 250 nt immediately up- and down-stream of the junction. This sequence is then used as the input for the Python implementation of Primer3 (<http://primer3.ut.ee/>)²² to design checking primers (Figure 3A and F). The output of this third module is the region surrounding the deletion junction (which can be used for verification of the junction by DNA sequencing), the two checking primers with their melting temperatures (T_m), and the expected

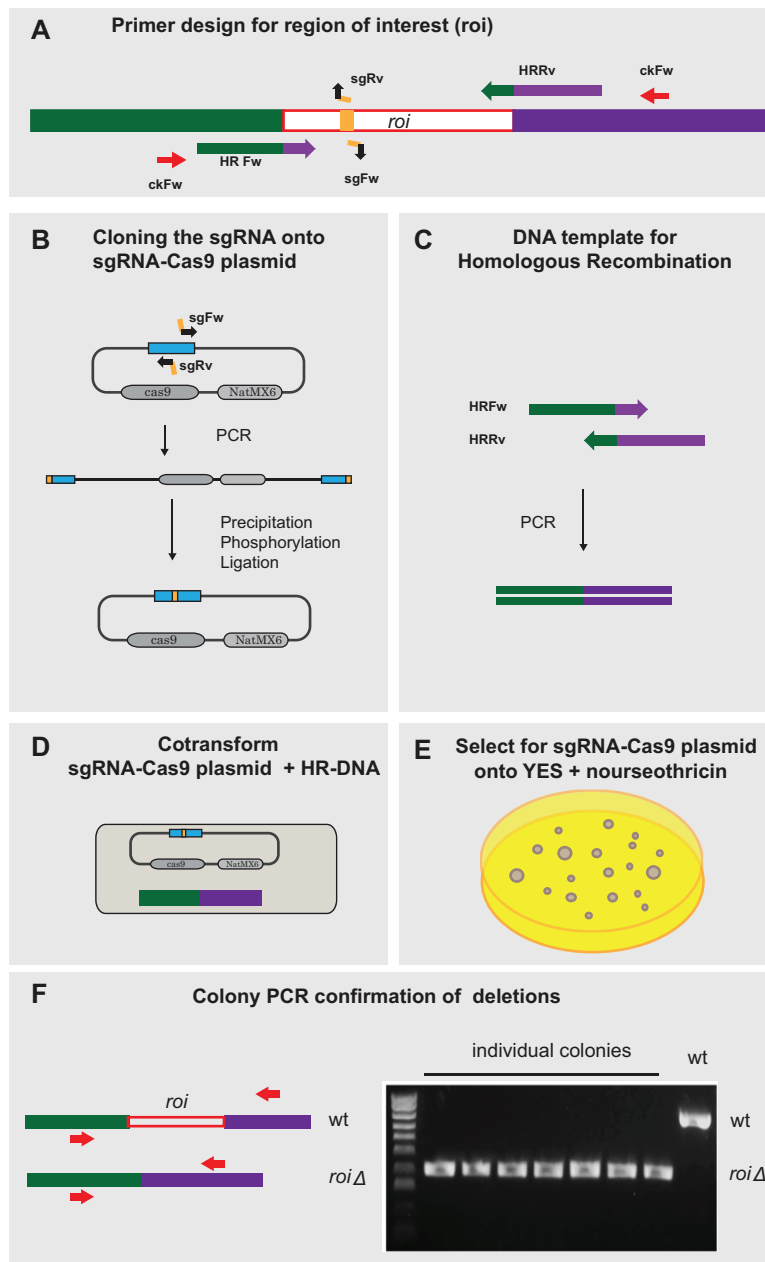


Figure 3. Scheme of key steps for CRISPR/Cas9-based method and primer design. sgRNA, single guide RNA.

sizes of the PCR products obtained for either successful deletion or without deletion (wild-type).

Rationale for optimization of experimental protocols

Cloning of sgRNA

Since the *CspCI* digestion of the plasmid containing Cas9 and the *rrk1*-guided sgRNA is often inefficient, it can be very difficult to clone sgRNAs into the plasmid optimized for CRISPR/Cas9 gene editing in *S. pombe*. We therefore devised alternative approaches for the introduction of the sgRNA into the pMZ379 plasmid (available through Addgene; plasmid no., 74215). The new Cas9-sgRNA plasmid pMZ379 contains a dominant selection marker that does not rely on auxotrophy (Figure 4). This plasmid enables the application of the CRISPR-Cas9 technique in any genetic background. Moreover, we have observed that the use of auxotrophic markers, such as *ura4*, can lead to undesirable physiological side effects (M.R.-L. and J.W., unpublished observations), as also observed for *S. cerevisiae*¹⁰.

The first method introduces the sgRNA sequence via the 5' ends of the primers used for PCR amplification of the pMZ379 plasmid sequence, followed by phosphorylation and ligation of the PCR product to reconstruct a new circular plasmid containing the desired sgRNA. We provide detailed PCR optimizations and other methods to deal with the large (~11kb) pMZ379 plasmid, which are critical for the success of the approach. Moreover, we currently optimize a second, alternative method that involves a generic PCR reaction to amplify the pMZ379 plasmid without introducing the sgRNA, followed by cloning of the sgRNA into the plasmid by *cloneEZ* or *Gibson Assembly* with annealed primers that contain the sgRNA sequence and flanking regions homologous to the ends of the PCR product. We are also planning to introduce a *NotI* restriction site for efficient linearization of the pMZ379 plasmid

without PCR reactions to facilitate large-scale cloning. These methods will be added to updated versions of the paper as they are developed.

G1 synchronization and cryopreserved competent cells

The activity of the prokaryotic Cas9 enzyme is likely increased in sites with more accessible chromatin and lower nucleosome occupancy. In mammalian cells, for example, Cas9 is more effective for sgRNAs that target coding sequences where chromatin is more open compared with other regions²³. This could cause problems for deleting or editing poorly transcribed and inaccessible regions, as we have observed for several non-coding RNAs (M.R.-L., C.C. and N.B.B., unpublished data). Moreover, proliferating *S. pombe* cells spend most of their time in G2 phase with a 2C DNA content. In these cells, two genomic copies need to be successfully modified by CRISPR/Cas9, and if only one copy is modified, the wild-type copy could be used as a template for homologous repair of the DSB. Therefore, it is likely that the efficiency of CRISPR/Cas9 genome editing is increased in G1 cells that contain a 1C of DNA.

Having these issues in mind when encountering low efficiencies for the CRISPR/Cas9 system, we implemented the synchronization of *S. pombe* cells in G1 using a simple nitrogen starvation protocol. This treatment not only greatly increases the proportion of cells with a 1C DNA content, but also substantially remodels the transcriptional programme^{24,25}, which can render many genomic regions more accessible. Moreover, we optimized a protocol for cryopreservation of competent, G1-synchronized cells¹², which greatly improves transformation and deletion efficiencies. Accordingly, we observed dramatically enhanced transformation rates when using G1-synchronized and cryopreserved cells (Figure 5).

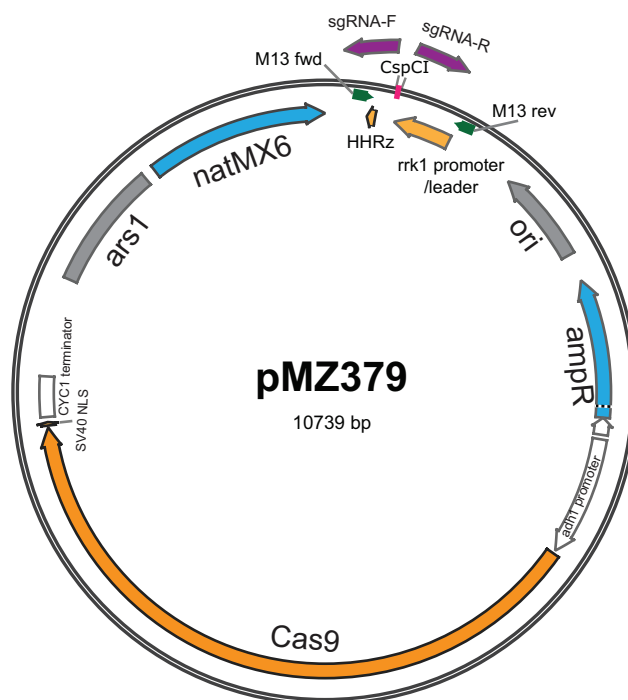


Figure 4. Map of pMZ379 plasmid. The primers to clone sgRNA are indicated by purple arrows. The primers for sequencing insertion of sgRNA are indicated by green arrows. Image adapted from *snapGene* viewer. sgRNA, single guide RNA.

Application of CRISPR/Cas9-based approach

Using this optimized approach, we have deleted over 80 non-coding RNA genes, 36 of which were deleted using 2 different sgRNAs (all primers for the deletions can be found in [Supplementary Table 1](#)). The efficiencies for successful deletions vary considerably for different genes ([Figure 6A](#)) and for different sgRNAs targeting the same gene ([Figure 6B](#)), with success rates ranging from 3% to 100%. For example, for the deletion of SPNCRNA.745, we obtained 5% positive colonies with one sgRNA (sgRNA.745.2) and 64% positive colonies with another one (sgRNA.745.3). Thus, using at least 2 different sgRNAs per

deletion not only minimizes the risk of being misled by phenotypes from off-target effects but it also maximizes the chance of successful deletion.

We have also successfully and efficiently introduced 7 point mutations in 4 different genomic regions using the CRISPR/Cas9 system, without leaving any other scars in the genome. Sites can only be mutated if they are located within the first 8–10 nt upstream of the PAM site. If no PAM site is available within this distance, a workaround could be applied by inserting two mutations as follows: use the HR template to introduce a synonymous change

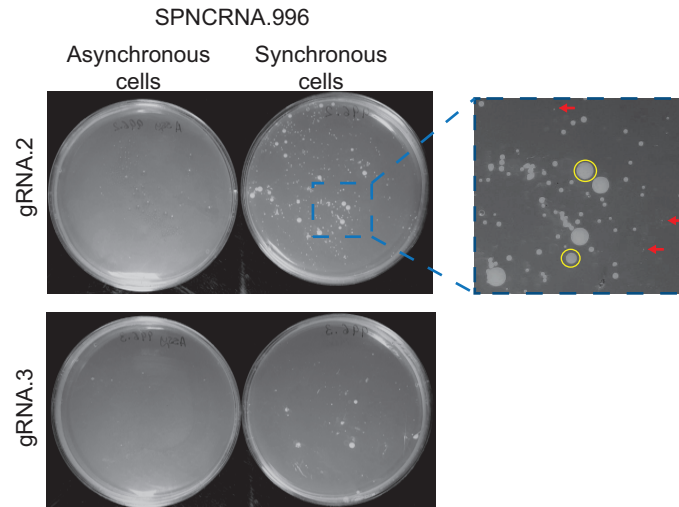


Figure 5. Improved transformation using G1-synchronized, cryopreserved cells. Wild-type cells were cultured in EMM medium and either subjected to nitrogen starvation for 2 hrs (synchronous) or maintained on normal EMM medium (asynchronous) at 25°C. The same number of cells were then made competent and frozen for synchronous and asynchronous cultures. Cryopreserved synchronous and asynchronous cells were transformed with the same amount of DNA for two different sgRNAs, as indicated at top and left. Cells were incubated at 32°C for 4 days, revealing a greatly increased transformation efficiency of the synchronous cells, with no colonies present for unsynchronized cells. The enlarged image indicates the smallest colonies that are much more likely to contain successful deletions (red arrows) than the large colonies (yellow circles).

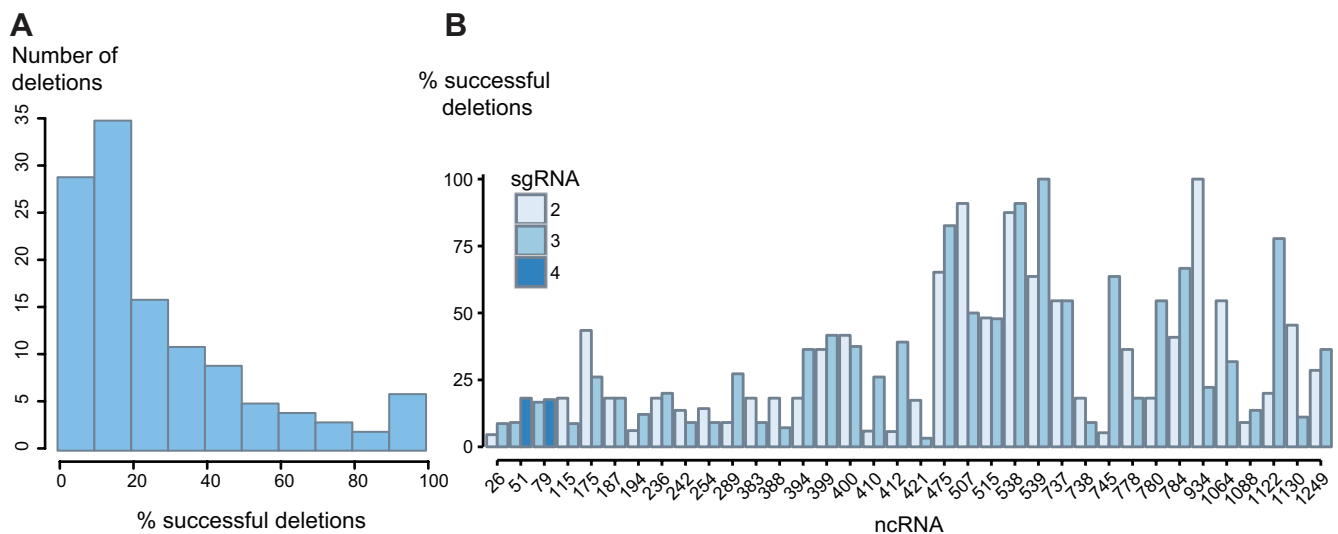


Figure 6. Deleting non-coding RNAs using CRISPR/Cas9-based approach. (A) Percentage of successful deletions among all colonies assayed for different non-coding RNA gene deletions, aggregated data of 120 different deletions. **(B)** Data on efficiencies of successful deletion for 36 non-coding RNA genes, for transformation with different sgRNAs (color-coded 2-4). In these cases, sgRNA1 were designed manually and did not yield successful deletions, whereas the sgRNAs 2-4 designed by CRISPR4P proved to be largely successful, and sgRNA4 were used for the two genes where sgRNA2 did not work. Note the greatly varying success rate for different genomic loci and for different sgRNAs.

8–10 nt upstream of the nearest PAM site, to prevent re-cutting by Cas9, plus the desired mutation where required. Mutagenesis is a particularly attractive application of the CRISPR/Cas9 method. In principle, the CRISPR/Cas9 system could also be adapted to tag genes, but we have not yet tried this out.

The CRISPR4P tool greatly facilitates the design of the sgRNAs and the different sets of primers required for the entire approach. The sgRNAs designed by CRISPR4P generally showed much higher success rates than manually designed sgRNAs. The current version of CRISPR4P only provides primers for deletion of genomic regions using the CRISPR/Cas9 system. However, CRISPR4P can be used to design sgRNAs to generate point mutations by inputting the coordinates of the region of interest. In future work, we are assembling a database with all sgRNAs used, whether they worked or not, to help with learning the principles for successful sgRNAs in *S. pombe*.

Help box 1. How to use the CRISPR4P tool

- Input region to be deleted, either by Gene name (e.g., *cdc2* or *SPNCRNA.01*) or by Coordinates (e.g., Chromosome: *II*; Coordinates: from *1500340* to *1501528*).
- Output list of sgRNAs is ordered by similarity to other genomic sgRNAs, with most unique sgRNA on top; select sgRNA by clicking on radial button at left.
- Output provides 3 types of primers
 - Primers for sgRNA cloning depending on selected sgRNA.
 - Primers to generate HR template.
 - Primers for checking the deletion construct, along with melting temperatures and expected sizes (in nt) of PCR products for successful deletion or without deletion, and along with sequence surrounding the deletion junction.

Help box 2. Important steps for successful application of CRISPR/Cas9

- Select at least 2 sgRNAs per construct with low similarity to other genomic regions.
- The large pMZ379 plasmid is unstable: aliquot and store at -80°C (do not thaw and re-freeze aliquots).
- To avoid generation of partial plasmids during PCR reaction, optimized PCR conditions are required: *Phusion High Fidelity* mastermix, 60°C annealing temperature, 2% DMSO, 1 ng (40 fg/μl) of pMZ379 template, 25 PCR cycles.
- Use high-fidelity polymerase to amplify plasmid and HR templates.
- After bacterial transformation of sgRNA plasmid, perform 'E. coli micro-cultures' to reduce number of minipreps.
- Check for correct sgRNA by sequencing Cas9-sgRNA plasmid with M13F primer.
- Synchronize *S. pombe* cells for 2 hrs in EMM without nitrogen before making them competent to increase efficiency of transformation, reduce incubation times, and facilitate deletion of difficult genomic regions.
- To support homologous recombination of HR template, incubate cells for 16 hours in EMM without nitrogen after transformation which avoids need for first plating cells onto YES before replica-plating on selective media (common practice for antibiotic markers selection) and thus prevents cells from proliferating.
- Select smallest colonies from transformation as these are most likely to contain correct deletions. Positive colonies typically appear only 4 days or later after plating, while colonies growing faster are typically negative for the deletion.

Methods

Reagents and equipment

- pMZ379 plasmid (Addgene, plasmid # 74215)
- Phusion® High-Fidelity PCR Master Mix with HF Buffer (NEB, cat. no. M0531S)
CAUTION: this product contains DMSO which is known to be harmful for aquatic life, discard appropriately.
- TopTaq Polymerase (QIAGEN, cat. no. 200201)
- dNTPmix 10 μM (Bioline, cat. no. BIO-39044)
- 5× DNA Loading Buffer Blue (Bioline, BIO-37045)
- ExoSAP-IT PCR Product Cleanup (Affymetrix, cat. no. 78200 200 UL)
CAUTION: this product is irritant and can cause severe eye damage, wear protective eye wear.
- Sodium acetate 3M (Thermo Fisher Scientific, cat. no. AM9740)
- Ethanol (MERCK, cat. no. 1.08543.0250)
- T4 Polynucleotide Kinase (NEB, cat. no. M0201S)
- T4 DNA ligase (NEB cat. no. M0202S)
- Mix & Go Competent Cells - Strain DH5α, 96 × 50 μl (Cambridge Bioscience, cat. no. T3009)
- QIAprep Spin Miniprep Kit (QIAGEN, cat. no. 27104)
CAUTION: Buffers P2 and N3 are corrosive and causes skin and eye damage/irritation, wear eye protection and gloves. Buffer PB is highly flammable and causes skin and eye irritation, and possibly dizziness. Keep away from fire, wear protective clothing and eye wear. Avoid inhalation of RNase A.
- Herring sperm DNA 10 μg/μl (Promega, cat. no. D1811)
- Lithium Acetate dihydrate (Sigma-Aldrich, cat. no. L-4158-250g)
- Glycerol (Fisher Scientific, cat. no. BP229-1)
CAUTION: irritant, wear protective eye wear gloves and clothing.
- Polyethylene glycol 4000 (PEG4000) (VWR cat. no. 26606.293)
- Nourseothricin-dihydrogen sulfate (Werner BioAgents cat. no. 5.002.000)
CAUTION: this product is harmful if swallowed, wear protective clothing and eye wear.
- LB Broth Base w/o Trace elements (Formedium, cat. no. AIMLB0110)
- Agar powder (VWR, cat. no.20767.298)
- Ampicillin (Sigma, cat. no.A9518-25g)
CAUTION: might cause skin irritation and respiratory problems, wear protective eye wear, gloves and avoid inhalation.
- YES BROTH (Formedium, cat. no.PMCUCL1000)
- EMM BROTH W/O NITROGEN (Formedium, cat. no. PMD1310)
- NH₄Cl (Sigma, cat. no. 09718/1kg)
CAUTION: NH₄Cl is toxic if swallowed, can cause eye irritation, wear protective clothing and eye wear.

- Ultrapure Agarose (THERMO FISHER, cat. no. 16500500)
- 10X Ultrapure TBE buffer (Life tech, cat. no. 15581044)
CAUTION: can cause damage in the unborn child, may cause fertility problems, wear protective clothing and eye wear.

HyperLadder 1kb (Bioline, cat. no. BIO-33053)

- Freeze 'N Squeeze™ DNA Gel Extraction Spin Columns (Bio-Rad, cat. no. 7326166)

Primer M13F: tgtaaacgacggccagt

Step-by-step procedure (can be downloaded as [Supplemental File 1](#))

Selection of sgRNAs and primers to delete region of interest

1. Use CRISPR4P (bahlerlab.info/crispr4p) to input desired deletion target by gene name or by coordinates as chromosome (in roman numeral), start and end sites. Select sgRNA by clicking button at left to display primers required for this sgRNA. We have found that there is no need for HPLC-purified oligos, desalted oligonucleotides synthesized by our usual provider (Life Technologies) work well for the entire procedure, substantially reducing the cost of the deletions.

CRITICAL STEP: CRISPR4P allows selection from all unique sgRNAs present within the input target region. The sgRNAs are ranked from least likely to most likely to have off-target effects, based on similarity of sub-sequences to other genomic sgRNAs. It is recommended to choose at least two sgRNAs from the top of the list.

Cloning sgRNA into pMZ379 plasmid (TIME: ~9 hours)

2. Prepare master mix for PCR to clone sgRNA into pMZ379 plasmid as in table below.

Use sgRNA cloning primers designed by CRISPR4P

	Final concentration	Volume per reaction (25 µl)
pMZ379 DNA (1 ng/µl)	1 ng (40 fg/µl)	1 µl
Primer mix (10 µM per primer)	0.4 nM/primer	1 µl
Phusion HF-buffer (2X)	1×	12.5 µl
DMSO (100%)	2%	0.5 µl
H ₂ O		10 µl

CRITICAL STEP: pMZ379 is unstable and should be stored in 1 ng/µl aliquots at -80°C, discard after thawing.

3. Perform PCR following protocol below.

Number of cycles	Temperature	Duration
1	98°C	2 min
25	98°C	10 sec
	60°C	30 sec
	72°C	5 min 30 sec
1	72°C	5 min

CRITICAL STEP: PCR conditions have been adjusted for Phusion High-Fidelity Polymerase to minimize number of unspecific PCR bands ([Figure 7A, B](#)).

4. Check PCR products by running 5 µl on 0.7% agarose TBE gel ([Figure 7A](#)).
5. **Optional step:** Add 8 µl of ExoSAP-IT PCR Product Cleanup to 20 µl of PCR reaction. Incubate in PCR machine for 15 min at 37°C and for 15 min at 80°C.
6. Precipitate DNA by adding 60 µl of 100% ethanol and 6 µl of 3M sodium acetate.
7. Incubate 30 min at -20°C.
8. Centrifuge for 20 min at 20,000g, 4°C to precipitate PCR product.
9. Remove supernatant.
10. Add 50 µl of 70 % ethanol (do **not** resuspend pellet).
11. Centrifuge for 10 min at 20,000g, 4°C.
12. Remove supernatant and air dry pellet.
13. Resuspend pellet in 20 µl of H₂O.

PAUSE POINT: PCR product can be stored at -20°C until further processing.

14. Phosphorylate the 5' ends of PCR product by preparing master mix as below.

	µl per reaction in 30 µl final volume
PCR product	20
T4 DNA ligase buffer	3
T4 PNK	1
H ₂ O	6

CRITICAL STEP: DNA ligase buffer is used because it provides the ATP required for the phosphorylation reaction (as recommended by manufacturer), and this enzyme exerts 100% activity in this buffer.

15. Incubate 30 min at 37°C.
16. Inactivate the enzyme by incubating for 20 min at 65°C.

PAUSE POINT: Phosphorylated DNA can be stored at -20°C until further processing.

17. Ligate plasmid ends by preparing master mix below

	µl per reaction in 10 µl final volume
Phosphorylated DNA	8
T4 DNA ligase buffer	1
T4 DNA ligase	1

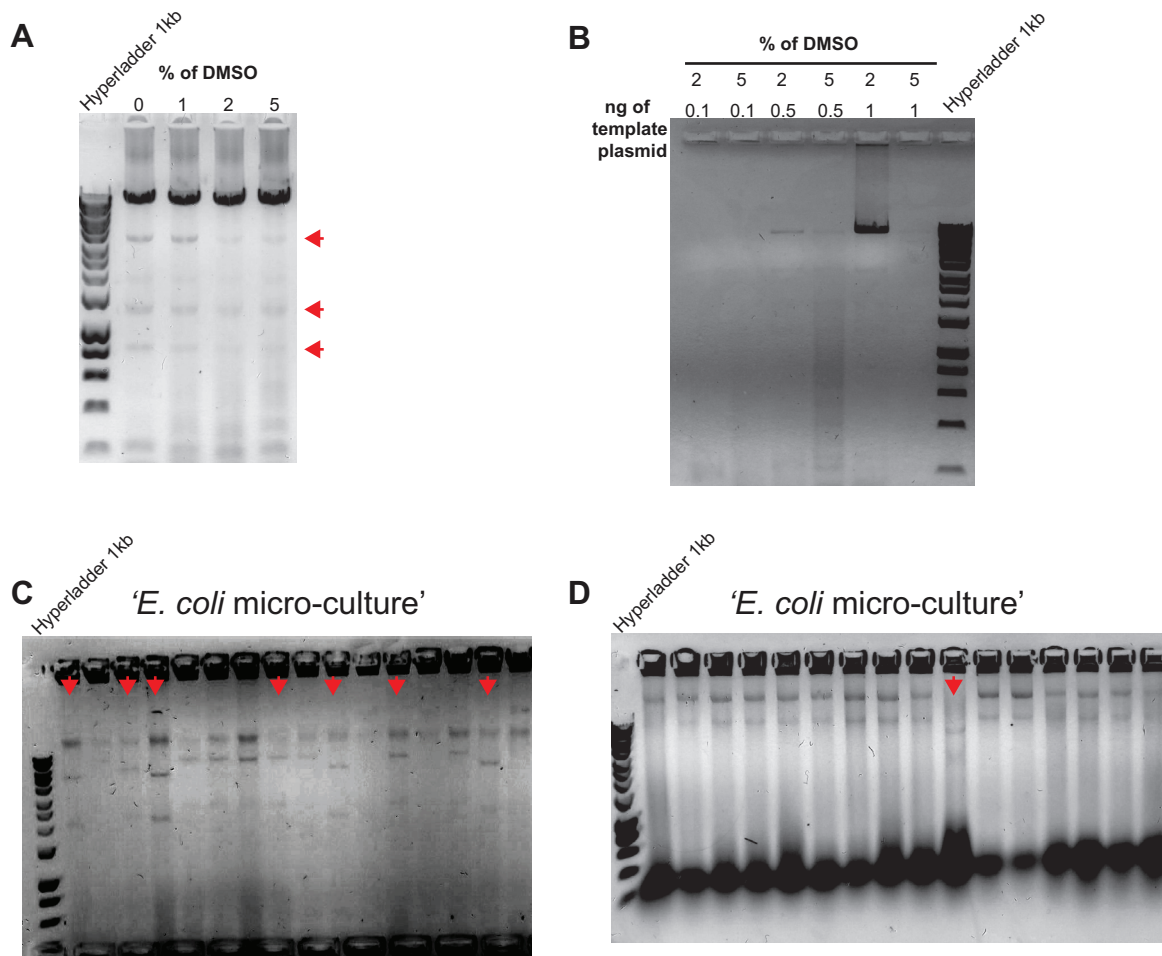


Figure 7. PCR cloning of sgRNA into pMZ379 plasmid and 'E. coli mini cultures'. (A) Gel of non-optimized PCR reactions for sgRNA cloning. Red arrows indicate shorter, unspecific products that need to be removed to avoid faulty plasmids. (B) As in (A), but with optimized PCR reaction which eliminates unspecific products (see protocol for details). (C) Gel of sgRNA-pMZ379 plasmid using non-optimized PCR reactions for sgRNA cloning. Each well contains DNA from different colony of same transformation. Red arrows indicate faulty cloning reactions containing plasmids of different sizes. (D) As in (C), but with optimized PCR reaction used for cloning which largely prevents faulty cloning reactions.

18. Ligate for 16 hrs at 16°C.

PAUSE POINT: Ligated DNA can be stored at -20°C until further processing.

Transformation of chemically competent E. coli cells (TIME: 30 min)

Cambridge Bioscience Mix & Go Competent Cells can be transformed using a short protocol provided by manufacturer.

19. Prepare LB-agar plates containing 75 µg/ml of ampicillin, and incubate them at 37°C for 15 min.

20. Thaw one aliquot (per transformation) of Mix & Go Competent Cells (DH5α strain) on ice.

21. Add 5 µl of ligated plasmid to cells, mix gently by tapping with finger.

CRITICAL STEP: Do not pipette competent cells.

22. Incubate cells on ice for 5 min.

23. Plate whole mixture of cells and DNA onto pre-warmed (37°C) LB-ampicillin plates.

24. Incubate plates for 20 hrs at 37°C.

Confirmation of sgRNA cloning (TIME: ~2 days)

Smaller, unspecific products during PCR amplification can lead to cloning mistakes (Figure 7A). But even in the absence of such unspecific PCR products, the plasmid can recombine during cloning, which results in aberrant sizes. To confirm that the sgRNA has been cloned correctly and that there are no mutations or rearrangements, we recommend to test about 12 colonies for each transformation by performing an 'E. coli micro-cultures', as follows.

25. Prepare '*E. coli* micro-culture' plate by adding 30 μ l of LB + 75 μ g/ml of ampicillin onto each well of a sterile 96-well plate.
26. Inoculate different transformant in each well.
27. Close plate with adhesive seal.
28. Incubate cells for 20 hrs at 37°C.
29. Prepare new 96-well plate with 30 μ l of LB + 75 μ g/ml of ampicillin on each well.
30. Make replica of bacterial micro-culture plate onto the new plate by inoculating 5 μ l of original micro-culture into new plate, seal and incubate it at 37°C.
31. Boil original bacterial micro-culture plate for 10 min at 98°C in PCR machine.
32. Let it rest for 2 min at 4°C.
33. Mix with appropriate volume of loading buffer for each well (e.g. 5 μ l of 5 \times loading buffer).
34. Run 20 μ l of bacteria-loading buffer mix on 0.7% agarose TBE gel to check for appropriate plasmid size.

CRITICAL STEP: This step will allow to identify plasmids of the wrong size, so that only plasmids of the correct size are selected to test by sequencing (Figure 7C, D).

35. Prepare 5 ml inoculums of bacteria containing clones of correct size on LB + 75 μ g/ml of ampicillin. Cells are taken from replica plate of '*E. coli* micro-culture'. We normally check 1–2 transformants by sequencing.
36. Incubate for 20 hrs at 37°C.
37. Prepare glycerol stock of bacteria by mixing 500 μ l of bacterial culture with 500 μ l of 50% sterile glycerol, and store at -20°C.
38. With remaining 4.5 ml of bacterial culture perform a 'mini prep' with QIAprep Spin Miniprep Kit.
39. Quantify plasmid DNA and send for Sanger sequencing to confirm that correct sgRNA has been cloned. Use primer M13F (TGTAACGACGGCCAGT) for sequencing.

PAUSE POINT: Plasmids and glycerol stocks can be stored at -20°C until sequence has been confirmed.

CRITICAL STEP: In case of ligation-mediated sgRNA cloning, the last base pair next to the junction point is deleted in rare cases, leading to 19 nt sgRNA, so it is advisable to check 1–2 colonies for each sgRNA clone to ensure the appropriate sequence of the sgRNA.

Generation of HR template (TIME: ~ 2.5 hrs)

40. Prepare PCR mastermix as in table below.

	final concentration	Volume to add for 1 reaction (50 μ l)
HR primer mix (10 μ M per primer)	0.4 nM/primer	2 μ l
Phusion HF-buffer (2X)	1x	25 μ l
H ₂ O		23 μ l

41. Perform PCR following protocol below.

Cycles	Temperature	Duration
1 cycle	98°C	2 min
30 cycles	98°C	10 sec
	55°C	10 sec
	72°C	30 sec
1 cycle	72°C	5 min

42. Check PCR products by running 5 μ l on 1.5 % agarose TBE gel.

PAUSE POINT: HR template can be stored at -20°C.

*Preparation of synchronized competent cryopreserved *S. pombe* cells (TIME: ~1.5 days)*

This protocol is a modification of a previously described method¹² to prepare 200 ml of competent cells that allows for 40 transformations.

43. Prepare 20 ml preculture in EMM and grow cells by shaking at 32°C for 8–16 hrs.
44. Dilute cells in 200 ml EMM and grow cells until they reach mid-exponential phase (~2 \times 10⁹ cells in total).
45. Centrifuge cells for 3 min at 1800g, room temperature.
46. Remove supernatant.
47. Wash in one volume (200 ml) of EMM without nitrogen (EMM-N).
48. Repeat steps 45 and 46 once more.
49. Resuspend cells in 200 ml of EMM-N and transfer to sterile flask.
50. Incubate for 2 hrs at 25°C with shaking.
51. Check that cells have become smaller and rounder, under light microscope.
52. Place cell culture on ice for 15 min.

CRITICAL STEP: To maintain integrity and competency of cells, they must be kept at 4°C from this moment.

53. Centrifuge cells for 5 min at 1600g, 4°C and remove supernatant.
54. Resuspend cells on **ice-cold**, sterile water.
55. Centrifuge for 5 min at 1600g, 4°C and remove supernatant.
56. Repeat steps 54 and 55 twice more.
57. Resuspend cells in 2 ml of **ice-cold**, filter-sterilized 30% Glycerol, 0.1M Lithium acetate (pH 4.9), which gives 10⁹ cells/ml.
58. Prepare 50 μ l cell aliquots in 1.5 ml sterile Eppendorf tubes, place aliquots on ice for 2 min. Each aliquot is for one transformation.
59. Store aliquots at -80°C immediately.

PAUSE POINT: Cryopreserved cells can be stored at -80°C for at least 2 months.

Transformation of cryopreserved S. pombe cells (TIME: 30 min)

60. Thaw aliquots of synchronized, cryopreserved cells in dry block at 40°C for 2 min.
61. For each transformation, add 2 µl of 10 µg/µl denatured herring sperm DNA, 10 µl of HR template, and 2 µg of gRNA plasmid (~10 µl of standard mini-prep yield of 200ng/µl).
62. Add 145 µl of 50% PEG4000, mix well and immediately incubate mix for 15 min at 43°C.
63. Centrifuge cells for 3 min at 1600g, room temperature.
64. Remove supernatant and resuspend cells in 1 ml of EMM-N.

CRITICAL STEP: In the case of using auxotrophic mutant, we recommend the addition of 1/10 of usual concentration of relevant supplement. If using an *h⁹⁰* strain, use EMM with nitrogen to prevent sporulation.

65. Incubate at room temperature for 16 hours, without shaking.
66. Centrifuge cells for 3 min 1600g, room temperature.
67. Remove supernatant and plate all cells on YES plates containing 100 µg/ml of Noursethracin.
68. Incubate plates at 32°C for at least 4 days. In case not getting successful deletions, longer incubations are sometimes required to allow small colonies to grow up.
69. Re-streak smallest colonies onto YES plates. Cas9 expression is deleterious for cells, and re-streaking onto non-selective YES allows for elimination of the Cas9 plasmid.

CRITICAL STEP: It is important to select the smallest colonies present (Figure 5): Large colonies are likely to emerge from transformants with mutations or rearrangements of Cas9¹¹, and this problem is compounded in the large and unstable Cas9-sgRNA plasmid. Unpublished data suggest that the mutations happen during *E. coli* growth (large colony counts fluctuate between miniprep cultures but are quite stable within one culture), and may get worse with freeze-thaws of the plasmid (more negative large colonies in freeze-thawed plasmids).

Checking of deletions by colony PCR (TIME: 4 hrs)

70. Prepare master mix for PCR reactions following table below:

Components	Final concentration	Volume per reaction (25 µl total)
Cell colony	--	Scoop cells with tip of pipette
TopTaq PCR Buffer 10x	1x	2.5 µl
MgCl ₂ 25 mM	0.5 mM	0.5 µl
Primer mix (10 µM per primer)	0.4 nM/each primer	1 µl
dNTPs mix (10 µmol per dNTP)	0.4 nmol/each	1 µl
Q Solution 5x	1x	5 µl
H ₂ O	-	14.75 µl
TopTaq DNA Polymerase (5 U/µl)		0.25 µl

71. For each colony take a little biomass with 10 µl pipette tip and resuspend in PCR mix (with this polymerase there is no need to boil cells prior to PCR).

72. Perform PCR following protocol below.

Cycles	Temperature	Duration
1 cycle	94°C	2 min
35 cycles	94°C	30 sec
	52°C	30 sec
	72°C	2min 30 sec
1 cycle	72°C	7 min

73. Add 5 µl of loading dye and load 10 µl of PCR product with loading dye mix on 0.7% agarose gel. The sizes of expected products are indicated in the output of CRISPR4P. (For successful deletions, this will be ~200 bp and for wild-type, the size of target region to delete plus ~200 bp of flanking regions). OPTIONAL: add 0.5 µl of 10 mg/ml RNase A solution to mix before loading to remove RNA that might complicate band visualization on gel. To confirm the deletion junction, the PCR products can be sent for Sanger sequencing (the expected sequence surrounding the deletion junction is provided by CRISPR4P). Figure 3F provides an example of an agarose gel showing successful deletions.

Data and software availability

CRISPR4P software available from: bahlerlab.info/crispr4p

Latest source code: <https://github.com/Bahler-Lab/crispr4p>

Archived source code: DOI: [10.5281/zenodo.16468326](https://doi.org/10.5281/zenodo.16468326)

License: MIT

Raw data are deposited in OSF (<https://osf.io/5de22/>) DOI: [10.17605/OSF.IO/5DE227](https://doi.org/10.17605/OSF.IO/5DE227).

Author contributions

M.R.-L. and J.B. conceived the study. O.F.S. wrote the CRISPR4P web tool. M.R.-L. and C.C. developed the methodology. M.R.-L., C.C., N.B.B., R.O., H.A., D.U., M.H. and J.W. performed the experimental work. M.Z. created the pMZ379 plasmid. M.R.-L. and J.B. drafted the manuscript.

Competing interests

No competing interests were disclosed.

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The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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Supplementary material

Supplementary Table 1: Single guide RNA sequences and all primers used.

[Click here to access the data.](#)

Supplementary File 1: Step-by-step protocol.

[Click here to access the data.](#)

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Damien Hermand, Olivier Finet, Carlo Yague-Sanz

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The implementation of the CRISPR/Cas9 system in fission yeast by the Zaratiegui laboratory created a large interest within the community and many laboratories have tried to set up the method with apparently low success. Therefore, the present work by the Zaratiegui and Bahler groups is important and will help to solve some issues. It is expected that the improvements presented here will spread within the community. A new selection is proposed for the Cas9 vector together with a new transformation method. A straightforward web tool is added to the already long list of very useful resources developed over the years by the Bahler lab.

The manuscript is well written and most of the time easy to read. Solving the following issues will improve clarity.

- In Figure 2, in « Table », the « numbers on the right » may be explained with a simple sentence. Also, the naming « Table » seems a bit odd.
- The colour code used in Figures 3 and 4 is different for the sg primers, which may be misleading to some readers.
- In Figure 5, the red arrows do not seem to point to anything while supposed to highlight small colonies. Maybe using red circles will be better.
- Figure 6 is not easy to understand. According to Figure 6A, there are 29 deletions with a percentage of successful deletions between 0 and 10% while the legend of Figure 6B suggests that 38 sgRNAs (36 sgRNAs #1 and 2 sgRNAs #2) did not yield to successful deletions. How is this possible? A table in the supplementary data may be easier to read if showing for every ncRNAs: the number of sgRNA tried and for each sgRNAs, the succes rate. The abscissa axis in Figure 6B is especially very hard to read.
- It would also be useful to discuss if the genome position matters and if the list of the targeted ncRNAs roughly covers the whole genome.
- The paragraph related to point mutations could provide more details or be removed and inserted later on when more data are available.

Other comments.

- It may be useful to mention that commercial kits are available to introduce the sgDNA into the vector, for example the BioLabs Q5® Site-Directed Mutagenesis Kit that is inexpensive and efficient.
- The code on GitHub should be at least minimally documented.
- The version of Pombase used to build the database should be indicated.

- There are still quite a few typos and mistakes, for examples:

on page 3 : The data to the right of each sgRNA indicates the numbers of other genomic sgRNA sequences that share a given number of nucleotides (starting from the 5' end of the PAM sequence), isn't « the number of genomic sequences » rather than the « the number of OTHER genomic sequences »?

on page 6: rkk1-guided sgRNA should be rrk1-guided sgRNA

We have read this submission. We believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however we have significant reservations, as outlined above.

Competing Interests: No competing interests were disclosed.

Author Response (*Member of the F1000 Faculty and F1000Research Advisory Board Member*) 21 Dec 2016

Jürg Bahler, Department of Genetics, Evolution and Environment, University College London, UK

We thank the reviewers for their helpful and constructive comments. Below we provide a point-by-point response to the specific issues raised (pasted in italic).

- *In Figure 2, in « Table », the « numbers on the right » may be explained with a simple sentence. Also, the naming « Table » seems a bit odd.*

We have changed the title of this Table to 'Suggested sgRNAs' in both the web tool and in Figure 2. In the figure legend, we have also provided an explanation for the numbers on the right of the table.

- *The colour code used in Figures 3 and 4 is different for the sg primers, which may be misleading to some readers.*

We have now modified Figure 4 to match the color of the sgRNA primers with the ones in Figure 3.

- *In Figure 5, the red arrows do not seem to point to anything while supposed to highlight small colonies. Maybe using red circles will be better.*

We want to highlight the very smallest colonies because these are the most likely to be correct. In the screen version of the figure, these colonies are visible. We have increased the contrast to better visualize these colonies and now highlight small colonies using red circles as suggested.

- *Figure 6 is not easy to understand. According to Figure 6A, there are 29 deletions with a percentage of successful deletions between 0 and 10% while the legend of Figure 6B suggests that 38 sgRNAs (36 sgRNAs #1 and 2 sgRNAs #2) did not yield to successful deletions. How is this possible? A table in the supplementary data may be easier to read if showing for every ncRNAs: the number of sgRNA tried and for each sgRNAs, the success rate. The abscissa axis in Figure 6B is especially very hard to read.*

We understand that this was confusing. We have now corrected the x-axis of Figure 6A to indicate that the lowest bin contains 1-10% of successful deletions (instead of 0-10%). We have also clarified this in the figure legend. Here we only show the percentages of successful deletions. We did not add numbers for any unsuccessful deletions, because we cannot be certain whether they failed because of the sgRNA sequence, mutations in the plasmid, or any other reason. We now also provide a Supplementary Table 2 showing the data from Figures 6A and 6B, which is cited in the legend of Figure 6.

- *It would also be useful to discuss if the genome position matters and if the list of the targeted ncRNAs roughly covers the whole genome*

We have deleted ncRNA genes spread across all 3 chromosomes. We have now added an additional section C to Figure 6 to show the genomic positions of all annotated ncRNA genes (grey

dots) and the ncRNA genes that we have successfully deleted (red dots). As for all genome manipulations, there may of course be genomic regions which are less amenable to changes, e.g. due to inaccessible chromatin.

- *The paragraph related to point mutations could provide more details or be removed and inserted later on when more data are available.*

We think that it is helpful to report at this point that it is also possible to get point mutations using our CRISPR/Cas9 method. We will expand on this in a future update of the paper.

- *It may be useful to mention that commercial kits are available to introduce the sgDNA into the vector, for example the BioLabs Q5® Site-Directed Mutagenesis Kit that is inexpensive and efficient.*

We know this kits, but have optimized the conditions for the polymerase indicated. A different polymerase may require optimization of the initial PCR reaction, and the specific protocol provided may no longer be valid in all details. Naturally, other users can experiment with different procedures or kits, and we would be interested to hear of any alternatives that have been implemented.

- *The code on GitHub should be at least minimally documented.*

We now provide basic documentation of the code in GitHub.

The version of Pombase used to build the database should be indicated.

The genome assembly is ASM294v2 and the annotation version 55, which is now specified in the text.

- *There are still quite a few typos and mistakes, for examples:*

on page 3 : The data to the right of each sgRNA indicates the numbers of other genomic sgRNA sequences that share a given number of nucleotides (starting from the 5' end of the PAM sequence), isn't « the number of genomic sequences » rather than the « the number of OTHER genomic sequences »?

- *on page 6: rkk1-guided sgRNA should be rrk1-guided sgRNA*

We have corrected these typos as suggested.

Competing Interests: No competing interests were disclosed.

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Rodriguez-Lopez and colleagues report improvements on CRISPR/Cas9-mediated genome editing in *S. pombe* (fission yeast). Their work builds on a previous paper by Jacobs, Zaratiegui *et al.* (2014). In this earlier paper, Jacobs *et al.* describe an expression vector for Cas9 and sgRNA that functions in *S. pombe* and demonstrate that mutagenesis and epitope-tagging is possible with this system.

Rodriguez-Lopez *et al.* now make several changes to improve the efficiency and the ease of use.

1. The authors change the auxotrophic selection marker *ura4+* in the Cas9/sgRNA plasmid to a nourseothricin-resistance, which does not require a specific genetic background and allows selection on rich medium.

Another group has recently implemented a fluoride export channel as another marker replacing

ura4+ on this plasmid, which also allows selection on rich medium and accelerates Cas9-mediated genome editing (Fernandez and Berro 2016). This also works well in our hands. I think it would be great if the authors cited this paper, so that readers are aware of all the different possibilities. Non-specific double-strand breaks created by Cas9 are always a concern, and it is important that the plasmid is efficiently lost after successful genome modification. With the *ura4+* version, 5-FOA could be used for counterselection. Since expression of Cas9 impairs growth, I am assuming that loss of the plasmid is very efficient, even without counterselection. However, if the authors happen to have data on this, it would be nice to mention it. (For example, how many clones lost the plasmid after one passage on non-selective medium?)

2. The authors have written a program to select specific sgRNA target regions. The program then suggests primers for sgRNA cloning, as well as primers to delete a gene of interest and to check for successful deletion.

This is generally very useful. When I tried the online version, I had no problem specifying a gene by name, but finding sgRNA targets by entering specific coordinates did not work for me (using two different browsers). My input caused an "Internal Server Error". It would be great if the authors could look into this.

The current implementation of the program suggests sgRNA targets based on specificity, but - as the authors show - efficiency can be highly variable. For *S. pombe*, there is no data available to indicate which target regions may be particularly efficient (and even in other organisms, information is still scarce). I was wondering whether it could be useful to extend the program to allow community feedback (i.e. when a researcher is using one of the suggested sgRNAs, she/he could input how well this site worked). This would (a) avoid that several people try using target regions that are not efficient, and (b) in the long run maybe allow it to determine which factors influence efficiency. Obviously, some sort of quality control on the user input would be required (e.g. number of successful genome modifications per how many clones tested, and a gel picture to support this), which may make it too time-consuming for the Bähler Lab to curate.

3. The previous system needed digest of the Cas9/sgRNA plasmid with the restriction enzyme *Csp* CI, which sometimes is inefficient. The authors have now solved this problem by amplification of the entire 11 kb plasmid with primers that contain the specificity region, followed by ligation. This seems more inconvenient than is necessary. However, the authors already mention that they work on other improved strategies for the sgRNA cloning step that they will add to the paper as they are implemented. I agree that this will be highly useful.
4. The authors have further improved an existing protocol for *S. pombe* transformation to increase transformation and deletion efficiency. Figure 5 shows that G1 synchronization greatly improves transformation efficiency. If the authors happen to have data to which extent the cryopreservation affects this result, it would be great if they could add it.

Overall, these are all highly useful changes - a clear improvement over the previous protocol. Motivation and results are well described, and the step-by-step procedure is detailed and easy to follow.

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and application to fast genome editing with CRISPR/Cas9. *Yeast*. 2016; **33** (10): 549-557 [PubMed Abstract](#) | [Publisher Full Text](#)

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

Author Response (*Member of the F1000 Faculty and F1000Research Advisory Board Member*) 21 Dec 2016

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We thank the reviewer for her helpful, constructive comments. Our response to the specific issues raised (pasted in italic) is presented below.

- *The authors change the auxotrophic selection marker $ura4+$ in the Cas9/sgRNA plasmid to a nourseothricin-resistance, which does not require a specific genetic background and allows selection on rich medium. Another group has recently implemented a fluoride export channel as another marker replacing $ura4+$ on this plasmid, which also allows selection on rich medium and accelerates Cas9-mediated genome editing (Fernandez and Berro, 2016). This also works well in our hands. I think it would be great if the authors cited this paper, so that readers are aware of all the different possibilities.*

We missed this paper, thank you. We have now cited it in the introduction. A drawback of is that fluoride selection requires a specific strain background.

- *Non-specific double-strand breaks created by Cas9 are always a concern, and it is important that the plasmid is efficiently lost after successful genome modification. With the $ura4+$ version, 5-FOA could be used for counterselection. Since expression of Cas9 impairs growth, I am assuming that loss of the plasmid is very efficient, even without counterselection. However, if the authors happen to have data on this, it would be nice to mention it. (For example, how many clones lost the plasmid after one passage on non-selective medium?)*

We have checked 106 colonies after one pass onto non-selective media, and 88 (83%) of these colonies have lost the plasmid passively.

- *The authors have written a program to select specific sgRNA target regions. The program then suggests primers for sgRNA cloning, as well as primers to delete a gene of interest and to check for successful deletion. This is generally very useful. When I tried the online version, I had no problem specifying a gene by name, but finding sgRNA targets by entering specific coordinates did not work for me (using two different browsers). My input caused an "Internal Server Error". It would be great if the authors could look into this.*

This bug has been corrected.

- *The current implementation of the program suggests sgRNA targets based on specificity, but - as the authors show - efficiency can be highly variable. For *S. pombe*, there is no data available to indicate which target regions may be particularly efficient (and even in other organisms, information is still scarce). I was wondering whether it could be useful to extend the program to allow community feedback (i.e. when a researcher is using one of the suggested sgRNAs, she/he could input how well this site worked). This would (a) avoid that several people try using target regions that are not efficient, and (b) in the long run maybe allow it to determine which factors influence efficiency. Obviously, some sort of quality*

control on the user input would be required (e.g. number of successful genome modifications per how many clones tested, and a gel picture to support this), which may make it too time-consuming for the Bähler Lab to curate.

Yes, we agree this is a good idea. As already mentioned in the paper, we are assembling a database with all sgRNAs used, whether they worked or not, to help with learning the principles for successful sgRNAs in *S. pombe*. We may implement such a community system in a future update of the paper.

- *The previous system needed digest of the Cas9/sgRNA plasmid with the restriction enzyme CspCI, which sometimes is inefficient. The authors have now solved this problem by amplification of the entire 11 kb plasmid with primers that contain the specificity region, followed by ligation. This seems more inconvenient than is necessary. However, the authors already mention that they work on other improved strategies for the sgRNA cloning step that they will add to the paper as they are implemented. I agree that this will be highly useful.*

We are working on a method to avoid the PCR cloning, and once this is implemented will report it in a future update of the paper.

- *The authors have further improved an existing protocol for *S. pombe* transformation to increase transformation and deletion efficiency. Figure 5 shows that G1 synchronization greatly improves transformation efficiency. If the authors happen to have data to which extent the cryopreservation affects this result, it would be great if they could add it.*

We have not systematically checked to what extent cryopreservation helps, but it improves the procedure in our hands. In the original paper describing the protocol, Suga et al. report that the solution containing glycerol used as cryoprotectant improves efficiency: *“These permeating agents have an ability not only to cryoprotect cells but also to improve transformation efficiency, and glycerol was a more effective agent for *Sz. pombe* cells. Thus, the thawed competent cells could be used directly for transformation without removing the glycerol because the presence of glycerol in the transformation mixture was important.”*

But the G1 synchronization seems to make an even a bigger difference, and we have further specified this in the legend of Figure 5. Transformation of synchronous cells consistently resulted in 3-fold to over 1000-fold higher numbers of colonies than transformation of asynchronous cells.

Competing Interests: No competing interests were disclosed.