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40 years of veterinary papers in JAC – what have we learnt?

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SCHOLARONE Manuscripts

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25 Abstract

This review, for the occasion of the 40th anniversary of the Journal of Antimicrobial 26 Chemotherapy (JAC), gives an overview of the manuscripts related to veterinary 27 bacteriology in the past 40 years with a focus on "One Health" aspects. From 1975 to 28 2000 the number of manuscripts related to veterinary medicine was limited, but 29 thereafter, the number steadily increased. Most manuscripts published were related 30 31 to food-producing animals, but companion animals and minor species were also 32 covered. Subjects included antimicrobial usage in animals and the consequences for 33 human medicine, new resistance genes and mechanisms, prevalence and 34 epidemiology of antimicrobial resistance and emergence of resistant bacteria in animals with zoonotic potential such as livestock-associated methicillin-resistant 35 Staphylococcus aureus (LA-MRSA), methicillin-resistant S. pseudintermedius 36 37 (MRSP) and extended-spectrum β -lactamase (ESBL)-producing Enterobacteriaceae. The manuscripts added to our knowledge on the risks of transmission of resistant 38 .en bacteria from animals to humans and the importance of prudent use of antimicrobial 39 40 agents in veterinary medicine.

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42 Introduction

43 The Journal of Antimicrobial Chemotherapy (JAC) publishes primarily articles in the 44 field of antimicrobial chemotherapy related to human medicine, but also articles from veterinary medicine, especially those likely to have an impact on public health. The 45 objective of this review is to give an overview of JAC manuscripts in the field of 46 veterinary microbiology, especially veterinary bacteriology, during the past 40 years. 47 An editorial published earlier this year marked the starting point for a series of articles 48 to celebrate the 40th anniversary of JAC.¹ The focus of this review is on bacteria from 49 50 animals, their resistance genes and mechanisms as well as antimicrobial 51 chemotherapy of bacterial diseases in animals from 1975 until 2015. Manuscripts 52 reporting antiviral, antifungal, and antiparasitic aspects related to veterinary medicine 53 were not included, neither were studies using animal models to study antimicrobials 54 for human use. Using these exclusion criteria, a total of 379 'veterinary' papers were published in JAC during 1975-2015. This corresponds to approximately 2.4% of the 55 total number of 15,584 papers published in JAC during that time period. 56

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58 **The history of veterinary papers in JAC**

During the first two years 1975 and 1976, all manuscripts published in JAC were 59 60 about human medicine and no veterinary manuscripts were published. During the 61 period 1977–2000 papers dealing with veterinary medicine and antimicrobial resistance in bacteria from animals were sporadic (usually less than 10 manuscripts 62 per year), although the total number of manuscripts published each year steadily 63 increased from 94 in 1977 to 450 in 2000 (Figure 1).¹ The first manuscript related to 64 65 veterinary microbiology was a manuscript about the relationship between antibiotics as feed additives in animals and the emergence of bacterial resistance in man by 66

Pohl in 1977.² It concluded that antibiotics as feed additives resulted in resistant 67 bacteria in the gut flora of animals and that these resistant bacteria could potentially 68 69 be transmitted to man. Pohl further concluded that many resistant bacteria in humans probably have no animal origin and therefore, the total prohibition of antibiotics in 70 animal feed would be unlikely to result in a significant decrease of resistant bacteria 71 in humans.² In 1986, Linton reviewed the evidence available at the time and 72 73 concluded "... that antibiotic resistant E. coli reach man from animal sources, and 74 colonize the human gut for a number of days, is beyond doubt" and that "carriage of 75 multiple plasmids, many of which carry multiple resistance determinants, must 76 constitute an important potential source of plasmids for indigenous E. coli in the human gut and, subsequently, to human pathogens".³ The topic of antimicrobial use 77 in livestock and pet animals and the consequences for human medicine has been the 78 79 subject of various papers in JAC during the following decades and reflects an important aspect of the "One Health" principle.4-14 80

During 2001-2015, the number of manuscripts related to veterinary microbiology 81 steadily increased exceeding 10 per year in 2003 and reaching a peak of 45 in 2014 82 83 (Figure 1). Most of the veterinary manuscripts were about food-producing animals, especially pigs, cattle and chickens. Manuscripts relating to companion animals like 84 dogs, horses and cats were less frequent. Minor numbers of manuscripts dealt with 85 86 fish, wild animals, and sheep, whereas only a few manuscripts dealt with turkeys, 87 ducks, geese, or goats (Figure 2). Many manuscripts dealt with more than one animal 88 species and most manuscripts were about Enterobacteriaceae (mainly Escherichia coli and Salmonella enterica), staphylococci, enterococci and Campylobacter spp. 89 though several studies also included animal-specific bacteria, such as Rhodococcus 90 equi from horses and Bartonella henselae from cats. 91

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93 **Topics of the veterinary papers in JAC**

Popular topics of veterinary papers in JAC reflect changing research trends over 94 time, which in turn are influenced by overall thinking in the field of antimicrobial 95 agents and the availability of new technologies such as molecular typing and DNA 96 sequencing. As mentioned above, in the 1970s and 1980s, veterinary papers in JAC 97 were rare on the whole and the topics discussed were mixed. The relationship 98 99 between antimicrobial resistance and use of antimicrobial agents in humans and 100 animals was an early theme that has steadily continued to draw the interest of 101 researchers publishing in JAC.

In the 1990s, JAC started publishing veterinary articles on topics such as 102 mechanisms of resistance and studies on the prevalence and epidemiology of 103 resistance among bacteria from animals. From 2001-2005, the number of veterinary 104 papers increased substantially, as did the number of the different topics covered with 105 larger scale surveillance and prevalence studies predominating often covering 106 several countries and animal species.¹⁵⁻²² Researchers investigated not only the 107 prevalence of phenotypic resistance but also the spread of certain resistance 108 mechanisms and genes among bacteria isolated from animals.^{17,18,20,22} 109

Prevalence studies and resistance mechanisms continued to be popular topics from 2006-2010. This period also saw a substantial increase in the number of studies carrying out detailed molecular characterization of strains, integrons,²³⁻²⁵ and/or mobile genetic elements (MGEs).²⁶⁻³¹ Detailed molecular characterizations of strains and MGEs continued to be popular in 2011-2015³²⁻³⁷, alongside the characterization of resistance mechanisms and genes³⁸⁻⁴² Prevalence and surveillance studies were still being published in 2011-2015,⁴³⁻⁴⁷ but were proportionally less popular than in preceding years. This undoubtedly reflected the increasing accessibility of molecular techniques such as whole genome sequencing, enabling molecular characterization to be performed with ease and at a relatively low cost. Between 2006-2015, there was also a substantial number of papers investigating levels of antimicrobial use in animals and its impact on resistance in both the animals themselves as well as humans.⁴⁸⁻⁵¹ This probably reflects increasing calls for better monitoring of antimicrobial use in animals and calls to reduce unnecessary antibiotics in farming.

New emerging resistant bacteria of animal origin

126 Livestock-associated methicillin-resistant Staphylococcus aureus

The first papers in JAC about the occurrence of methicillin-resistant Staphylococcus 127 aureus (MRSA) of animal origin were published in 2005^{52,53} and 2006.⁵⁴⁻⁵⁶ These 128 initial reports focused mainly on companion animals such as dogs, cats and 129 horses.⁵³⁻⁵⁶ Later on, livestock-associated MRSA (LA-MRSA) isolates of clonal 130 complex (CC) 398 from pigs were identified⁵⁷ and characterized in detail using a 131 variety of molecular methods.⁵⁸ The close similarity between the isolates from 132 humans and animals strongly suggested these LA-MRSA isolates were being 133 exchanged and, though the direction of transfer – zoonosis or humanosis – was 134 questioned,⁵⁹ proof of transfer in both directions was found. Indeed reports followed 135 about the presence of LA-MRSA CC398 in other animals (e.g. cattle, broiler 136 chickens), people in occupational contact with animals, and in food of animal 137 origin.⁶⁰⁻⁶⁷ 138

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140 Methicillin-resistant Staphylococcus pseudintermedius

Methicillin resistance also occurs in S. pseudintermedius (formerly identified as S. 141 intermedius), an opportunistic pathogen that causes infections in pet animals, 142 particularly dogs but also in cats.⁶⁸⁻⁷⁰ Since 2006, there has been a significant 143 emergence of methicillin-resistant *S. pseudintermedius* (MRSP).⁶⁸ Although 144 infections in humans with MRSP are uncommon, canine infections or carriage of 145 such organisms represent a potential hazard for people in contact with dogs.⁶⁸ MRSP 146 often display resistance to almost all classes of antimicrobial agents used in 147 veterinary medicine and several reports published in JAC have investigated the 148 occurrence of resistance and/or resistance mechanisms to various antimicrobials.⁶⁸⁻⁷² 149 Other reports focus on the evolution and clonal relationship of MRSP isolates in 150 different countries.⁶⁹⁻⁷⁴ 151

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153 Extended-spectrum β-lactamase-producing Enterobacteriaceae from animals

154 In mid-2000s. extended-spectrum β-lactamase (ESBL)-producing 155 Enterobacteriaceae from animal sources were described. The first report published in JAC referred to ESBL-producing Salmonella isolates from poultry, poultry products 156 and human patients in The Netherlands.²² Soon thereafter, first reports about the 157 occurrence of ESBL-producing E. coli in meat and from various animal and 158 environmental sources were published.^{75,76} During the following years, ESBL-159 producing Enterobacteriaceae were reported from different countries, different food 160 161 and food animal sources, including apparently healthy animals, and also from foodborne outbreaks.^{23,27,44-46,77-88} In these studies different types of ESBL genes were 162 detected with *bla*_{CTX-M} variants, *bla*_{SHV-2} and *bla*_{TEM-52} genes being most predominant. 163 The first report about ESBL genes in *E. coli* from companion animals was published 164

in JAC in 2010,⁸⁹ followed by reports that described the presence of ESBL-producing
 E. coli and *Klebsiella pneumoniae* in dogs, cats and horses in different countries.⁹⁰⁻⁹³
 In addition, free-living birds were also identified as carriers of ESBL-producing
 Enterobacteriaceae.^{94,95} Many of the studies on ESBL-producing Enterobacteriaceae
 also provided a detailed strain characterization and a characterization of the ESBL
 gene-carrying plasmids.^{28,44-46,78,79,84,88-93,96}

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172 (Fluoro)quinolone-resistant bacteria of animal origin

Increasing levels of quinolone resistance among Enterobacteriaceae and 173 Campylobacter spp. has been a particular cause for concern since the mid-1990s.⁹⁷⁻ 174 ¹⁰⁰ In 1998, Piddock et al. demonstrated that gyrA and parC mutations were 175 responsible for quinolone resistance among veterinary isolates of Salmonella 176 enterica.¹⁰¹ Such mutations were also detected in Salmonella Typhimurium from fish 177 and E. coli from turkeys, ruminants, other food animals and food of animal 178 origin.^{18,102-106} In addition to Enterobacteriaceae, mutations in the quinolone 179 resistance determining region of the target genes were also identified in other 180 bacteria, including Campylobacter spp.,^{98,107,108} Bartonella henselae,¹⁰⁹ Pasteurella 181 multocida,³³ Haemophilus parasuis,¹¹⁰ and S. aureus.¹¹¹ Soon, it was demonstrated 182 that (i) isolates harbouring first-step mutations towards guinolone resistance also 183 184 exhibited reduced susceptibility to fluoroquinolones and (ii) fluoroquinolone exposure selects for resistant mutants.¹¹²⁻¹¹⁵ It was shown that other mechanisms, such as 185 186 active efflux, also play a role in fluoroquinolone resistance and that the mechanisms of fluoroquinolone resistance are more complex than initially thought.^{115,116} 187

188 Reports about plasmid-mediated quinolone resistance (PMQR) genes in 189 bacteria of animal origin were published in JAC from the mid-2000s on. The first

paper dated from 2006 and described a *gnrS* gene in an avian Salmonella Infantis 190 isolate.¹¹⁷ Soon thereafter, the first complete nucleotide sequence of a small *gnrS1*-191 carrying plasmid from Salmonella Typhimurium was published.¹¹⁸ The gnrS1 gene 192 was also detected in the Salmonella serovars Corvallis, Virchow, and Saintpaul, 193 whereas a gnrB5 gene was found in the Salmonella serovars Newport, Hadar, and 194 Saintpaul, all from various European countries.¹¹⁹⁻¹²¹ A study from China identified 195 the PMQR genes aac(6')-lb-cr, gepA, gnrA3, gnrB6, gnrB10 and gnrS1 among 30 196 isolates of Enterobacteriaceae. One to three mutations in the QRDRs of the genes 197 gyrA and parC were detected in all but one of the PMQR-positive isolates.¹²² Further 198 199 PMQR genes identified in bacteria of animal origin were qnrB2 in Salmonella Bredeney from poultry,¹²³ gnrB19, gnrS1 and gnrB6 together with aac(6')-lb-cr in 200 various Salmonella serovars from reptiles,¹²⁴ gnrA1, gnrB6 and aac(6')-lb-cr in H. 201 parasuis from pigs,¹¹⁰ as well as *qnrS1*, *qnrB19*, *qnrB10* and *qepA* in *E. coli* from pigs 202 and chickens.¹²⁵ A large-scale study on PMQR genes in Salmonella enterica and 203 Escherichia coli isolated from animals, humans, food and the environment in 13 204 European countries revealed the presence of gnrA1, gnrB2, gnrB4, gnrB6, gnrB7, 205 gnrB12, gnrB19, gnrS1, aac(6')-1b-cr, and gnrD genes in Salmonella enterica as well 206 as *gnrS1* and *gnrB19* in *E. coli*.¹²⁶ An additional PMQR gene *ogxAB* was detected on 207 a plasmid in *E. coli* from a chicken.³² Complete sequences of larger plasmids 208 carrying PMQR genes were also published.^{35,127} 209

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Novel and unusual resistance genes in bacteria of animal origin

212 New resistance genes in LA-MRSA and other staphylococci

The novel *mecA* homologue, initially described as $mecA_{LGA251}$, but later renamed as *mecC*, was found in MRSA isolates from a domestic dog, brown rats, a rabbit, a

common seal, sheep and a chaffinch.³⁸ This gene was also detected in a MRSA 215 isolate of a cat suffering from chronic conjunctivitis.¹²⁸ Further studies identified this 216 gene in methicillin-resistant staphylococci from wildlife, including MRSA from 217 European brown hares, an otter, and a hedgehog as well as in a methicillin-resistant 218 Staphylococcus stepanovicii from a Eurasian lynx,¹²⁹ common voles, wood mice and 219 a brown rat,¹³⁰ and from captive maras in a zoo.¹³¹ The mecC gene was also 220 detected in MRSA from cases of bovine mastitis.¹³². A new allotype, *mecC2*, was 221 identified in a methicillin-resistant Staphylococcus saprophyticus from a common 222 shrew.¹³³ 223

The multiresistance gene cfr, which confers resistance to phenicols, 224 lincosamides, oxazolidinones, pleuromutilins and streptogramin A antibiotics, was 225 initially found on plasmid pSCFS1 from a bovine *Staphylococcus sciuri* isolate.³⁹ The 226 complete sequence of this first *cfr*-carrying plasmid was published in JAC in 2004.¹³⁴ 227 Later on, cfr was also found on a small plasmid in an LA-MRSA ST9 isolate from a 228 case of bovine mastitis.³⁶ A review was published in 2013 which illustrated the wide 229 dissemination of the cfr gene in Gram-positive and Gram-negative bacteria from 230 animals and humans.³⁹ The complete sequence of the 135,615 bp *cfr*-carrying 231 plasmid pSCEC2 from *Escherichia coli* was reported in 2014.¹³⁵ 232

A multiresistance gene cluster of suspected enterococcal origin has been identified on plasmids and in the chromosomal DNA of *S. aureus* isolates from pigs and chickens, but also humans.¹³⁶ This cluster comprised the novel ABC transporter gene *Isa*(E) for combined resistance to pleuromutilins, lincosamides and streptogramin A antibiotics,^{137,138} the novel spectinomycin resistance gene *spw*,^{138,139} as well as the streptomycin resistance gene *aadE* and the lincosamide resistance gene *Inu*(B).¹³⁶⁻¹³⁹ Another novel plasmid-borne spectinomycin resistance gene, *spd*,

Journal of Antimicrobial Chemotherapy

was identified in MRSA ST398 from various animal and human sources.¹⁴⁰ This gene
was also identified in MSSA ST433 of porcine origin,¹⁴¹ and a variant of this gene
was detected in *Staphylococcus hyicus* and coagulase-negative staphylococci from
pigs.¹⁴² A variant of the pleuromutilin-lincosamide-streptogramin A-resistance gene *vga*(E), which showed only 85.7% identity to the original *vga*(E) gene from Tn*6133*,
was detected on identical plasmids in *Staphylococcus cohnii* and *Staphylococcus simulans* from pigs.¹⁴³

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248 Carbapenemase genes in bacteria of animal origin

The first carbapenemase gene bla_{VIM-1} was found in a multiresistance class 1 249 integron of an *E. coli* isolate on a pig farm in 2012.³⁴ A year later, the *bla*_{VIM-1} gene 250 was also found in Salmonella Infantis from pig and poultry farms in Germany,¹⁴⁴ and 251 the *bla*_{NDM-1} gene was found in a *Salmonella* Corvallis from a wild bird.¹⁴⁵ Other 252 carbapenemase genes found so far in animals and published in JAC include blaOXA-48 253 in *E. coli* and *K. pneumoniae* from dogs,¹⁴⁶ bla_{MP-4} in *Pseudomonas aeruginosa* from 254 a dog,¹⁴⁷ bla_{VIM-2} in P. aeruginosa from cattle and fowl as well as bla_{OXA-23} and 255 *bla*_{OXA58} in *Acinetobacter baumannii* from cattle, pig and fowl,¹⁴⁸ the *bla*_{NDM-1} gene in 256 A. baumannii of porcine origin,¹⁴⁹ and bla_{NDM-1} -producing Acinetobacter calcoaceticus 257 and Acinetobacter junii from environmental samples from livestock farms.¹⁵⁰ These 258 259 findings provoked a controversial debate about the role of animals in the 260 dissemination of carbapenemase genes which has resulted in publication of reviews and editorials.¹⁵¹⁻¹⁵³ 261

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263 Novel resistance genes in bovine and porcine Pasteurellaceae

The first description of florfenicol resistance in a target bacterium was published in 264 2005.¹⁵⁴ This report described the presence of the phenicol exporter gene floR on a 265 plasmid in bovine Pasteurella multocida from the UK. A few years later, the floR gene 266 was also detected on plasmids in bovine Pasteurella trehalosi (meanwhile renamed 267 as Bibersteinia trehalosi) from France and in porcine and bovine P. multocida from 268 Germany.^{31,155} The *floR* gene has also been detected on a small plasmid in porcine 269 H. parasuis.¹⁵⁶ This phenicol resistance gene was also part of the multiresistance 270 271 integrative and conjugative element ICEPmu1 from bovine P. multocida, which carried a total of twelve resistance genes, including the novel macrolide resistance 272 genes erm(42), msr(E) and mph(E).³³ Macrolide resistance in Mannheimia 273 haemolytica was shown to be caused by the mutation A2058G in the the 23S rRNA 274 and in *P. multocida* by the mutation A2059G in 23S rRNA.⁴¹ The tetracycline 275 resistance gene tet(L), which is widespread among Gram-positive bacteria, was 276 identified on plasmids and in the chromosomal DNA of *M. haemolytica*, Mannheimia 277 glucosida and P. multocida.¹⁵⁷ Another tet gene, tet(H) was detected on novel 278 plasmids in Actinobacillus pleuropneumoniae.¹⁵⁸ The trimethoprim resistance gene 279 280 dfrA1 was found in a partially truncated class 2 integron in a porcine Pasteurella aerogenes isolate.¹⁵⁹ The trimethoprim resistance gene dfrA14 was identified on 281 different plasmids in *A. pleuropneumoniae*.³⁷ 282

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284 New resistance genes and resistance-mediating mutations in other bacteria of

285 animal origin

A novel chloramphenicol exporter gene, *cmlB1*, has been identified in the porcine respiratory tract pathogen *Bordetella bronchiseptica*.¹⁶⁰ Another novel chloramphenicol/florfenicol exporter gene, designated *fexB*, was identified in

Enterococcus faecium and Enterococcus hirae of porcine origin.¹⁶¹ Both, CmIB and 289 FexB belong to the Major Facilitator Superfamily of exporters. A novel floR gene 290 variant was detected as part of a multiresistance genomic island in a porcine 291 Stenotrophomonas maltophila isolate. This FloRv protein showed only 84.1%-91.8% 292 amino acid identity to the various described FloR proteins.¹⁶² A novel ABC 293 transporter, OptrA, that confers combined resistance to phenicols and oxazolidinones 294 was identified in *E. faecalis* and *E. faecium* from humans, pigs and chickens.⁴² Other 295 new resistance genes reported in JAC include the aminoglycoside resistance genes 296 armA and rtmB. Gonzalez-Zorn et al. investigated the genetic environment of the 297 298 armA gene in an E. coli isolate from a pig and showed that it was embedded in a novel transposon composite facilitating spread between Enterobacteriaceae of 299 human and animal origin.¹⁶³ The *rtmB* gene was isolated from porcine *E. coli* and 300 Enterobacter isolates as well as from *E. coli* chicken isolates in China.¹⁶⁴⁻¹⁶⁶ A novel 301 macrolide efflux gene mef(B) was found in porcine E. coli isolates.¹⁶⁷ Wang et al. 302 reported the novel *fosX^{CC}* gene conferring fosfomycin resistance in *Campylobacter* 303 coli from swine faeces.¹⁶⁸ Novel gentamicin resistance genes were found in 304 *Campylobacter* from humans and retail food.¹⁶⁹ 305

Novel mutations in the *rpoB* gene responsible for rifampicin resistance have been identified in canine MRSP and in equine *Rhodococcus equi*.^{170,171}

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310 Relationship between antimicrobial use and resistance in animals

311 and humans

312 It is a generally accepted fact that the use of antimicrobial agents in both humans

313 and animals results in a selective pressure under which bacteria can either develop

Journal of Antimicrobial Chemotherapy: under review

resistance mediating mutations or acquire resistance genes. Indeed the use of 314 antimicrobial agents is perhaps the major driving force in resistance development and 315 dissemination. Consequently, several studies have analysed sales patterns of 316 antimicrobial agents used in veterinary medicine in Europe as well as their 317 consumption in various animal species.^{50,51,172,173} The latest data from 2011, 318 published in 2014, showed that in all 25 EU countries analysed, tetracyclines, 319 penicillins and sulphonamides accounted for more than half (53%-88%) of the total 320 amount of antimicrobial agents sold by country.⁵⁰ Another study also evaluated the 321 appropriateness of use compared with prudent use guidelines in Switzerland.¹⁷⁴ The 322 323 authors concluded that most prescriptions corresponded well to guidelines on prudent use of antimicrobials. However, there was a wide variation in prescriptions 324 between different veterinarians which might indicate that the usage and amount of 325 antimicrobials used for group medication lacking a specific indication could be further 326 reduced.¹⁷⁴ Several papers investigated the levels of antimicrobial use in animals and 327 its impact on resistance in both the animals themselves as well as humans.^{14,49,175-180} 328 Some studies demonstrated a correlation between antimicrobial usage in animals 329 and the occurrence of resistant bacteria in animals.^{14,49,175-177} Others also found a 330 331 relationship between the antimicrobial usage and the occurrence of antimicrobial resistant bacteria not only in animals, but also in humans with close contact to 332 animals.¹⁷⁸⁻¹⁷⁹ For instance, a study on enterococci revealed that the overall 333 334 resistance in broiler isolates corresponded with resistance in the isolates of broiler farmers and poultry slaughterers.¹⁷⁸ A more recent study also showed that MRSA of 335 the same MLST, spa and dru types with very similar resistance patterns were seen 336 among chickens at slaughter and abattoir workers and underlines the exchange of 337 resistant isolates between animals and people in occupational contact with them.⁶⁶ A 338

study of Danish pigs and their farmers and families showed that ESBL-producing *E*. *coli* was detected in pigs on 79% of the farms with a high consumption of
cephalosporins compared to 20% of the pigs on farms that did not use these drugs.
At four farms ESBL-producing *E. coli* isolates with the same CTX-M enzyme,
phylotype, PFGE type and MLST type were detected in both pigs and farmers.¹⁷⁹
These examples underline the interrelationship of antimicrobial use and
dissemination of resistant bacteria.

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348 **Conclusion**

Despite the fact that veterinary papers in JAC represent only a minority of all 349 manuscripts published in JAC during 1975-2015 (Fig. 1), they address important 350 aspects of antimicrobial usage and antimicrobial resistance in various animal 351 352 species. Antimicrobial resistance remains an important public health issue and that needs an integrated global perspective as bacteria do not respect geographical or 353 354 species borders. The 'One Health' concept states that human health, animal health, environmental health, agriculture as well as food safety and security are closely 355 linked. The focus of the veterinary papers in JAC often included links to human 356 health, but also to food safety and security, especially when dealing with bacteria of 357 358 zoonotic importance. Clearly, veterinary papers are indispensable in helping provide a more complete picture of the complex interactions between humans and animals in 359 the field of antimicrobial chemotherapy. Consequently, their continued publication in 360 361 the JAC is assured for the foreseeable future. 362

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364 **Transparency declarations**

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- 366
- 367
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- 370
- 371
- 372 **References**
- 373 **1** Donnelly JP. Celebrating 40 years of the Journal. J Antimicrob Chemother
- 374 2016; **71**: 1-2.

Pohl P. Relationship between antibiotic feeding in animals and emergence of bacterial resistance in man. *J Antimicrob Chemother* 1977; **3 Suppl C**: 67-72.

377 **3** Linton AH. Flow of resistance genes in the environment and from animals to 378 man. *J Antimicrob Chemother* 1986; **18 Suppl C**: 189-97.

Walton JR. Therapeutic antibiotics in veterinary practice. J Antimicrob *Chemother* 1981; 7:114-5.

5 Piddock LJV. Does the use of antimicrobial agents in veterinary medicine and animal husbandry select antibiotic-resistant bacteria that infect man and compromise antimicrobial chemotherapy? *J Antimicrob Chemother* 1996; **38**: 1-3.

Cook RR. Antimicrobial resistance--use in veterinary and human medicine. J
 Antimicrob Chemother 1997; 39: 435.

7 Bywater RJ, Casewell MW. An assessment of the impact of antibiotic resistance in different bacterial species and of the contribution of animal sources to resistance in human infections. *J Antimicrob Chemother* 2000; **46**: 643-5. Phillips I, Casewell M, Cox T *et al.* Does the use of antibiotics in food animals
pose a risk to human health? A critical review of published data. *J Antimicrob Chemother* 2004; **53**: 28-52.

Phillips I, Casewell M, Cox T *et al*. Antibiotic use in animals. *J Antimicrob Chemother* 2004; **53**: 885.

Turnidge J. Antibiotic use in animals--prejudices, perceptions and realities. J
 Antimicrob Chemother 2004; 53: 26-7.

Casewell M *et al.* The European ban on growth-promoting antibiotics and
 emerging consequences for human and animal health. *J Antimicrob Chemother* 2003; **52**: 159-61.

399 **12** Guardabassi L, Schwarz S, Lloyd DH. Pet animals as reservoirs of 400 antimicrobial-resistant bacteria. *J Antimicrob Chemother* 2004; **54**: 321-32.

401 **13** Lord Soulsby of Swaffham Prior. The 2008 Garrod Lecture: antimicrobial
 402 resistance--animals and the environment. *J Antimicrob Chemother* 2008; **62**: 229-33.

403 14 Chantziaras I, Boyen F, Callens B *et al.* Correlation between veterinary
404 antimicrobial use and antimicrobial resistance in food-producing animals: a report on
405 seven countries. *J Antimicrob Chemother* 2014; **69**: 827-34.

406 **15** Cruchaga S, Echeita A, Aladueña A *et al.* Antimicrobial resistance in
407 salmonellae from humans, food and animals in Spain in 1998. *J Antimicrob*408 *Chemother* 2001; **47**: 315-21.

Marie J, Morvan H, Berthelot-Hérault F *et al*. Antimicrobial susceptibility of
 Streptococcus suis isolated from swine in France and from humans in different
 countries between 1996 and 2000. *J Antimicrob Chemother* 2002; **50**: 201-9.

Aarestrup FM, Lertworapreecha M, Evans MC *et al*. Antimicrobial susceptibility
and occurrence of resistance genes among *Salmonella enterica* serovar Weltevreden
from different countries. *J Antimicrob Chemother* 2003; **52**: 715-8.

415 **18** Guerra B, Junker E, Schroeter A, *et al.* Phenotypic and genotypic
416 characterization of antimicrobial resistance in German *Escherichia coli* isolates from
417 cattle, swine and poultry. *J Antimicrob Chemother* 2003; **52**: 489-92.

19 Bywater R, Deluyker H, Deroover E *et al*. A European survey of antimicrobial
susceptibility among zoonotic and commensal bacteria isolated from food-producing
animals. *J Antimicrob Chemother* 2004; **54**: 744-54.

Gebreyes WA, Thakur S, Davies PR *et al.* Trends in antimicrobial resistance,
 phage types and integrons among *Salmonella* serotypes from pigs, 1997-2000. *J Antimicrob Chemother.* 2004; **53**: 997-1003.

Esaki H, Morioka A, Ishihara K *et al.* Antimicrobial susceptibility of *Salmonella*isolated from cattle, swine and poultry (2001-2002): report from the Japanese
Veterinary Antimicrobial Resistance Monitoring Program. *J Antimicrob Chemother*2004; **53**: 266-70.

Hasman H, Mevius D, Veldman K *et al.* β-Lactamases among extendedspectrum β-lactamase (ESBL)-resistant *Salmonella* from poultry, poultry products
and human patients in The Netherlands. J. Antimicrob. Chemother 2005; **56**: 115-21.

Vo AT, van Duijkeren E, Fluit AC *et al*. A novel *Salmonella* genomic island 1
and rare integron types in *Salmonella* Typhimurium isolates from horses in The
Netherlands. *J Antimicrob Chemother* 2007; 59: 594-9.

434 24 van Essen-Zandbergen A, Smith H, Veldman K *et al.* Occurrence and
435 characteristics of class 1, 2 and 3 integrons in *Escherichia coli*, *Salmonella* and
436 *Campylobacter* spp. in the Netherlands. *J Antimicrob Chemother* 2007; 59: 746-50.

Page 19 of 41

25 Kadlec K, Schwarz S. Analysis and distribution of class 1 and class 2 437 438 integrons and associated gene cassettes among *Escherichia coli* isolates from swine, 439 horses, cats and dogs collected in the BfT-GermVet monitoring study. J Antimicrob Chemother 2008; 62: 469-73. 440 441 26 Mesa RJ, Blanc V, Blanch AR et al. Extended-spectrum beta-lactamaseproducing Enterobacteriaceae in different environments (humans, food, animal farms 442 443 and sewage). J Antimicrob Chemother 2006; 58: 211-5. 27 Jouini A, Vinué L, Slama KB et al. Characterization of CTX-M and SHV 444 445 extended-spectrum beta-lactamases and associated resistance genes in Escherichia 446 coli strains of food samples in Tunisia. J Antimicrob Chemother 2007; 60: 1137-41. 447 28 García-Fernández A, Chiaretto G, Bertini A et al. Multilocus sequence typing of Incl1 plasmids carrying extended-spectrum beta-lactamases in Escherichia coli 448 449 and Salmonella of human and animal origin. J Antimicrob Chemother 2008; 61: 1229-33. 450 451 29 Hauschild T, Lüthje P, Schwarz S. Staphylococcal tetracycline-MLS_B resistance plasmid pSTE2 is the product of an RS_A-mediated in vivo recombination. J 452 453 Antimicrob Chemother 2005; 56: 399-402. 30 Garcia-Migura L, Hasman H, Svendsen C, Jensen LB. Relevance of hot spots 454 in the evolution and transmission of Tn1546 in glycopeptide-resistant Enterococcus 455 456 faecium (GREF) from broiler origin. J Antimicrob Chemother 2008; 62: 681-7. 457 31 Kehrenberg C, Wallmann J, Schwarz S. Molecular analysis of florfenicol-

resistant *Pasteurella multocida* isolates in Germany. *J Antimicrob Chemother* 2008;
62: 951-5.

460 **32** Liu BT, Wang XM, Liao XP *et al.* Plasmid-mediated quinolone resistance 461 determinants *oqxAB* and *aac(6')-lb-cr* and extended-spectrum β-lactamase gene *bla*_{CTX-M-24} co-located on the same plasmid in one *Escherichia coli* strain from China. *J Antimicrob Chemother* 2011; **66**: 1638-9.

464 **33** Michael GB, Kadlec K, Sweeney MT *et al.* ICEPmu1, an integrative 465 conjugative element (ICE) of *Pasteurella multocida*: analysis of the regions that 466 comprise 12 antimicrobial resistance genes. *J Antimicrob Chemother* 2012; **67**: 84-467 90.

468 **34** Fischer J, Rodríguez I, Schmoger S *et al. Escherichia coli* producing VIM-1 469 carbapenemase isolated on a pig farm. *J Antimicrob Chemother* 2012; **67**:1793-5.

35 Schink AK, Kadlec K, Schwarz S. Detection of *qnr* genes among *Escherichia coli* isolates of animal origin and complete sequence of the conjugative *qnrB19*carrying plasmid pQNR2078. *J Antimicrob Chemother* 2012; **67**:1099-102.

Wang XM, Zhang WJ, Schwarz S *et al.* Methicillin-resistant *Staphylococcus aureus* ST9 from a case of bovine mastitis carries the genes *cfr* and *erm*(A) on a
small plasmid. *J Antimicrob Chemother* 2012; 67:1287-9.

37 Bossé JT, Li Y, Walker S. Identification of *dfrA14* in two distinct plasmids
conferring trimethoprim resistance in *Actinobacillus pleuropneumoniae*. *J Antimicrob Chemother* 2015; **70**: 2217-22.

38 Paterson GK, Larsen AR, Robb A *et al.* The newly described *mecA*homologue, *mecA*LGA251, is present in methicillin-resistant *Staphylococcus aureus*isolates from a diverse range of host species. *J Antimicrob Chemother* 2012; 67:
2809-13.

39 Shen J, Wang Y, Schwarz S. Presence and dissemination of the
multiresistance gene *cfr* in Gram-positive and Gram-negative bacteria. J *Antimicrob Chemother* 2013; 68:1697-706.

508

486	40 Harmer CJ, Holt KE, Hall RM. A type 2 A/C2 plasmid carrying the aacC4		
487	apramycin resistance gene and the erm(42) erythromycin resistance gene recovered		
488	from two Salmonella enterica serovars. J Antimicrob Chemother 2015; 70: 1021-5.		
489	41 Olsen AS, Warrass R, Douthwaite S. Macrolide resistance conferred by rRNA		
490	mutations in field isolates of Mannheimia haemolytica and Pasteurella multocida.		
491	Antimicrob Chemother 2015; 70 : 420-3.		
492	42 Wang Y, Lv Y, Cai J et al. A novel gene, optrA, that confers transferable		
493	resistance to oxazolidinones and phenicols and its presence in Enterococcus faecalis		
494	and Enterococcus faecium of human and animal origin. J Antimicrob Chemother		
495	2015; 70 : 2182-90.		
496	43 Dierikx CM, van Duijkeren E, Schoormans AH et al. Occurrence and		
497	characteristics of extended-spectrum-β-lactamase- and AmpC-producing clinical		
497 498	characteristics of extended-spectrum-β-lactamase- and AmpC-producing clinical isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012;		
498	isolates derived from companion animals and horses. J Antimicrob Chemother 2012;		
498 499	isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012; 67: 1368-74.		
498 499 500	 isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012; 67: 1368-74. 44 Schink AK, Kadlec K, Kaspar H <i>et al.</i> Analysis of extended-spectrum-β- 		
498 499 500 501	 isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012; 67: 1368-74. 44 Schink AK, Kadlec K, Kaspar H <i>et al.</i> Analysis of extended-spectrum-β-lactamase-producing isolates collected in the GE<i>RM</i>-Vet monitoring programme. <i>J</i> 		
498 499 500 501 502	 isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012; 67: 1368-74. 44 Schink AK, Kadlec K, Kaspar H <i>et al.</i> Analysis of extended-spectrum-β-lactamase-producing isolates collected in the GE<i>RM</i>-Vet monitoring programme. <i>J Antimicrob Chemother</i> 2013; 68: 1741-9. 		
498 499 500 501 502 503	 isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012; 67: 1368-74. 44 Schink AK, Kadlec K, Kaspar H <i>et al.</i> Analysis of extended-spectrum-β-lactamase-producing isolates collected in the GE<i>RM</i>-Vet monitoring programme. <i>J Antimicrob Chemother</i> 2013; 68: 1741-9. 45 Doublet B, Praud K, Nguyen-Ho-Bao T <i>et al.</i> Extended-spectrum β-lactamase- 		
498 499 500 501 502 503 504	 isolates derived from companion animals and horses. <i>J Antimicrob Chemother</i> 2012; 67: 1368-74. 44 Schink AK, Kadlec K, Kaspar H <i>et al.</i> Analysis of extended-spectrum-β-lactamase-producing isolates collected in the GE<i>RM</i>-Vet monitoring programme. <i>J Antimicrob Chemother</i> 2013; 68: 1741-9. 45 Doublet B, Praud K, Nguyen-Ho-Bao T <i>et al.</i> Extended-spectrum β-lactamase-and AmpC β-lactamase-producing D-tartrate-positive <i>Salmonella enterica</i> serovar 		

and Wales, 2010-12. *J Antimicrob Chemother* 2014; **69**: 977-81.

cephalosporins in human non-typhoidal Salmonella enterica isolates from England

Al Bayssari C, Dabboussi F, Hamze M *et al.* Emergence of carbapenemaseproducing *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in livestock animals in Lebanon. *J Antimicrob Chemother* 2015; **70**: 950-1.

Moulin G, Cavalie P, Pellanne I, *et al.* A comparison of antimicrobial usage in human and veterinary medicine in France from 1999 to 2005. *J Antimicrob Chemother* 2008; **62**: 617-25.

Agerso Y, Aarestrup FM. Voluntary ban on cephalosporin use in Danish pig
 production has effectively reduced extended-spectrum cephalosporinase-producing
 Escherichia coli in slaughter pigs. *J Antimicrob Chemother* 2013; 68: 569-72.

519 **50** Grave K, Torren-Edo J, Muller A *et al*. Variations in the sales and sales 520 patterns of veterinary antimicrobial agents in 25 European countries. *J Antimicrob* 521 *Chemother* 2014; **69**: 2284-91.

522 **51** Grave K, Greko C, Kvaale MK *et al.* Sales of veterinary antibacterial agents in 523 nine European countries during 2005-09: trends and patterns. *J Antimicrob* 524 *Chemother* 2012; **67**: 3001-8.

525 **52** Kwon NH, Park KT, Moon JS *et al.* Staphylococcal cassette chromosome *mec* 526 (SCC*mec*) characterization and molecular analysis for methicillin-resistant 527 *Staphylococcus aureus* and novel SCC*mec* subtype IVg isolated from bovine milk in 528 Korea. *J Antimicrob Chemother* 2005; **56**: 624-32.

529 **53** Loeffler A, Boag AK, Sung J *et al.* Prevalence of methicillin-resistant 530 *Staphylococcus aureus* among staff and pets in a small animal referral hospital in the 531 UK. *J Antimicrob Chemother* 2005; **56**: 692-7.

532 **54** Strommenger B, Kehrenberg C, Kettlitz C *et al.* Molecular characterization of 533 methicillin-resistant *Staphylococcus aureus* strains from pet animals and their 534 relationship to human isolates. *J Antimicrob Chemother* 2006; **57**: 461-5. 535 **55** Malik S, Coombs GW, O'Brien FG *et al*. Molecular typing of methicillin-536 resistant staphylococci isolated from cats and dogs. *J Antimicrob Chemother* 2006; 537 **58**: 428-31.

538 **56** Moodley A, Stegger M, Bagcigil AF *et al. spa* typing of methicillin-resistant 539 *Staphylococcus aureus* isolated from domestic animals and veterinary staff in the UK 540 and Ireland. *J Antimicrob Chemother* 2006; **58**: 1118-23.

541 **57** Schwarz S, Kadlec K, Strommenger B. Methicillin-resistant *Staphylococcus* 542 *aureus* and *Staphylococcus pseudintermedius* detected in the BfT-GermVet 543 monitoring programme 2004-2006 in Germany. *J Antimicrob Chemother* 2008; **61**: 544 282-5.

545 **58** Kadlec K, Ehricht R, Monecke S *et al.* Diversity of antimicrobial resistance 546 pheno- and genotypes of methicillin-resistant *Staphylococcus aureus* ST398 from 547 diseased swine. *J Antimicrob Chemother* 2009; **64**: 1156-64.

548 **59** Morgan M. Methicillin-resistant *Staphylococcus aureus* and animals: zoonosis 549 or humanosis? *J Antimicrob Chemother* 2008; **62**: 1181-7.

550 **60** Cui S, Li J, Hu C *et al.* Isolation and characterization of methicillin-resistant 551 *Staphylococcus aureus* from swine and workers in China. *J Antimicrob Chemother* 552 2009; 64: 680-3.

553 **61** Pomba C, Baptista FM, Couto N *et al.* Methicillin-resistant *Staphylococcus* 554 *aureus* CC398 isolates with indistinguishable Apal restriction patterns in colonized 555 and infected pigs and humans. *J Antimicrob Chemother* 2010; **65**: 2479-81.

556 **62** Nienhoff U, Kadlec K, Chaberny IF *et al.* Transmission of methicillin-resistant 557 *Staphylococcus aureus* strains between humans and dogs: two case reports. *J* 558 *Antimicrob Chemother* 2009; **64**: 660-2. Haenni M, Saras E, Châtre P *et al.* A USA300 variant and other human-related
methicillin-resistant *Staphylococcus aureus* strains infecting cats and dogs in France. *J Antimicrob Chemother* 2012; **67**: 326-9.

562 64 Feßler A, Scott C, Kadlec K. Characterization of methicillin-resistant
563 Staphylococcus aureus ST398 from cases of bovine mastitis. J Antimicrob
564 Chemother 2010; 65: 619-25.

Groves MD, O'Sullivan MV, Brouwers HJ *et al. Staphylococcus aureus* ST398
 detected in pigs in Australia. *J Antimicrob Chemother* 2014; **69**: 1426-8.

567 **66** Wendlandt S, Kadlec K, Feßler AT *et al.* Resistance phenotypes and 568 genotypes of methicillin-resistant *Staphylococcus aureus* isolates from broiler 569 chickens at slaughter and abattoir workers. *J Antimicrob Chemother* 2013; **68**: 2458-570 63.

67 Lozano C, López M, Gómez-Sanz E *et al.* Detection of methicillin-resistant *Staphylococcus aureus* ST398 in food samples of animal origin in Spain. *J Antimicrob Chemother* 2009; **64**: 1325-6.

574 **68** van Duijkeren E, Catry B, Greko C *et al.* Review on methicillin-resistant 575 *Staphylococcus pseudintermedius. J Antimicrob Chemother* 2011; **66**: 2705-14.

69 Perreten V, Kadlec K, Schwarz S *et al.* Clonal spread of methicillin-resistant
577 *Staphylococcus pseudintermedius* in Europe and North America: an international
578 multicentre study. *J Antimicrob Chemother* 2010; **65**: 1145-54.

70 Kadlec K, Schwarz S, Perreten V *et al*. Molecular analysis of methicillinresistant feline *Staphylococcus pseudintermedius* from different European countries

and North America. *J Antimicrob Chemother* 2010; **65**: 1826-8.

582 **71** Loeffler A, Baines SJ, Toleman MS *et al* In vitro activity of fusidic acid and 583 mupirocin against coagulase-positive staphylococci from pets. *J Antimicrob* 584 *Chemother* 2008; **62**:1301-4.

Clark SM, Loeffler A, Bond R. Susceptibility in vitro of canine methicillin resistant and -susceptible staphylococcal isolates to fusidic acid, chlorhexidine and
 miconazole: opportunities for topical therapy of canine superficial pyoderma. *J Antimicrob Chemother* 2015; **70**: 2048-52.

73 Osland AM, Vestby LK, Fanuelsen H *et al.* Clonal diversity and biofilm-forming
ability of methicillin-resistant *Staphylococcus pseudintermedius*. *J Antimicrob Chemother* 2012; **67**: 841-8.

592 **74** McCarthy AJ, Harrison EM, Stanczak-Mrozek K *et al.* Genomic insights into 593 the rapid emergence and evolution of MDR in *Staphylococcus pseudintermedius*. *J* 594 *Antimicrob Chemother* 2015; **70**: 997-1007.

595 **75** Jensen LB, Hasman H, Agersø Y *et al.* First description of an oxyimino-596 cephalosporin-resistant, ESBL-carrying *Escherichia coli i*solated from meat sold in 597 Denmark. *J Antimicrob Chemother* 2006; **57**: 793-4.

598 **76** Mesa RJ, Blanc V, Blanch AR *et al.* Extended-spectrum beta-lactamase-599 producing Enterobacteriaceae in different environments (humans, food, animal farms 600 and sewage). *J Antimicrob Chemother* 2006; **58**: 211-5.

77 Riaño I, Moreno MA, Teshager T *et al.* Detection and characterization of
extended-spectrum β-lactamases in *Salmonella enterica* strains of healthy food
animals in Spain. *J Antimicrob Chemother* 2006; **58**: 844-7.

- 604 **78** Machado E, Coque TM, Cantón R *et al.* Antibiotic resistance integrons and
- 605 extended-spectrum β-lactamases among Enterobacteriaceae isolates recovered from
- chickens and swine in Portugal. *J Antimicrob Chemother*. 2008; **62**: 296-302.

Rodríguez I, Barownick W, Helmuth R *et al.* Extended-spectrum β-lactamases
and AmpC β-lactamases in ceftiofur-resistant *Salmonella enterica* isolates from food
and livestock obtained in Germany during 2003-07. *J Antimicrob Chemother* 2009;
610 64: 301-9.

611 **80** Lavilla S, González-López JJ, Miró E *et al.* Dissemination of extended-612 spectrum β-lactamase-producing bacteria: the food-borne outbreak lesson. *J* 613 Antimicrob Chemother 2008; **61**:1244-51.

614 **81** Dhanji H, Murphy NM, Doumith M *et al.* Cephalosporin resistance mechanisms 615 in *Escherichia coli* isolated from raw chicken imported into the UK. *J Antimicrob* 616 *Chemother* 2010; **65**: 2534-7.

617 **82** Randall LP, Clouting C, Horton RA *et al.* Prevalence of *Escherichia coli* 618 carrying extended-spectrum β-lactamases (CTX-M and TEM-52) from broiler 619 chickens and turkeys in Great Britain between 2006 and 2009. *J Antimicrob* 620 *Chemother* 2011; **66**: 86-95.

B3 Ho PL, Chow KH, Lai EL *et al.* Extensive dissemination of CTX-M-producing *Escherichia coli* with multidrug resistance to 'critically important' antibiotics among
food animals in Hong Kong, 2008-10. *J Antimicrob Chemother* 2011; 66: 765-8.

B4 Jiang HX, Tang D, Liu YH *et al.* Prevalence and characteristics of β-lactamase
and plasmid-mediated quinolone resistance genes in *Escherichia coli* isolated from
farmed fish in China. *J Antimicrob Chemother* 2012; 67: 2350-3.

85 Hordijk J, Wagenaar JA, van de Giessen A *et al.* Increasing prevalence and
diversity of ESBL/AmpC-type β-lactamase genes in *Escherichia coli* isolated from
veal calves from 1997 to 2010. *J Antimicrob Chemother* 2013; 68: 1970-3.

86 Eller C, Simon S, Miller T *et al.* Presence of β-lactamases in extended spectrum-cephalosporin-resistant *Salmonella enterica* of 30 different serovars in
 Germany 2005-11. *J Antimicrob Chemother* 2013; 68:1978-81.

87 Randall LP, Lemma F, Rogers JP *et al.* Prevalence of extended-spectrum-β lactamase-producing *Escherichia coli* from pigs at slaughter in the UK in 2013. *J* Antimicrob Chemother 2014; 69: 2947-50.

B8 Huijbers PM, Graat EA, Haenen AP *et al.* Extended-spectrum and AmpC βlactamase-producing *Escherichia coli* in broilers and people living and/or working on
broiler farms: prevalence, risk factors and molecular characteristics. *J Antimicrob Chemother* 2014; **69**: 2669-75.

Ewers C, Grobbel M, Stamm I *et al.* Emergence of human pandemic O25:H4ST131 CTX-M-15 extended-spectrum-beta-lactamase-producing *Escherichia coli*among companion animals. *J Antimicrob Chemother* 2010; **65**: 651-60.

Dolejska M, Duskova E, Rybarikova J *et al.* Plasmids carrying *bla*_{CTX-M-1} and *qnr* genes in *Escherichia coli* isolates from an equine clinic and a horseback riding
centre. *J Antimicrob Chemother* 2011; **66**: 757-64.

646 **91** Ewers C, Bethe A, Stamm I *et al.* CTX-M-15-D-ST648 *Escherichia coli* from
647 companion animals and horses: another pandemic clone combining multiresistance
648 and extraintestinal virulence? *J Antimicrob Chemother* 2014; **69**: 1224-30.

Dahmen S, Haenni M, Châtre P *et al.* Characterization of *bla*CTX-M IncFII
plasmids and clones of *Escherichia coli* from pets in France. J *Antimicrob Chemother*2013; **68**: 2797-801.

652 93 Ewers C, Stamm I, Pfeifer Y *et al.* Clonal spread of highly successful ST15653 CTX-M-15 *Klebsiella pneumoniae* in companion animals and horses. *J Antimicrob*654 *Chemother* 2014; 69: 2676-80.

Bonnedahl J, Drobni P, Johansson A *et al.* Characterization, and comparison,
of human clinical and black-headed gull (Larus ridibundus) extended-spectrum betalactamase-producing bacterial isolates from Kalmar, on the southeast coast of
Sweden. *J Antimicrob Chemother* 2010; **65**: 1939-44.

95 Tausova D, Dolejska M, Cizek A *et al. Escherichia coli* with extended spectrum β-lactamase and plasmid-mediated quinolone resistance genes in great
 cormorants and mallards in Central Europe. *J Antimicrob Chemother* 2012; **67**:1103 7.

Wang J, Stephan R, Power K *et al.* Nucleotide sequences of 16 transmissible
plasmids identified in nine multidrug-resistant *Escherichia coli* isolates expressing an
ESBL phenotype isolated from food-producing animals and healthy humans. *J Antimicrob Chemother* 2014; **69**: 2658-68.

667 **97** Griggs DJ, Hall MC, Jin YF *et al*. Quinolone resistance in veterinary isolates of 668 Salmonella. *J Antimicrob Chemother* 1994; **33**: 1173-89.

669 98 Piddock LJ. Quinolone resistance and *Campylobacter* spp. J Antimicrob
670 Chemother 1995; 36: 891-8.

99 Frost JA, Kelleher A, Rowe B. Increasing ciprofloxacin resistance in
salmonellas in England and Wales 1991-1994. *J Antimicrob Chemother* 1996; 37: 8591.

100 Seyfarth AM, Wegener HC, Frimodt-Moller N. Antimicrobial resistance in *Salmonella enterica* subsp. *enterica* serovar Typhimurium from humans and
production animals. *J Antimicrob Chemother* 1997; **40**: 67-75.

101 Piddock LJV, Ricci V, McLaren I *et al.* Role of mutation in the *gyrA* and *parC*genes of nalidixic-acid-resistant *Salmonella* serotypes isolated from animals in the
United Kingdom. *J Antimicrob Chemother* 1998; 41: 635-41.

Journal of Antimicrobial Chemotherapy

Ruiz J, Capitano L, Nuñez L *et al.* Mechanisms of resistance to ampicillin,
 chloramphenicol and quinolones in multiresistant *Salmonella* Typhimurium strains
 isolated from fish. *J Antimicrob Chemother* 1999; **43**: 699-702.

Giraud E, Leroy-Sétrin S, Flaujac G *et al*. Characterization of high-level
 fluoroquinolone resistance in *Escherichia coli* O78:K80 isolated from turkeys. *J Antimicrob Chemother* 2001; **47**: 341-3.

- Orden JA, Ruiz-Santa-Quiteria JA, Cid D *et al.* Quinolone resistance in
 potentially pathogenic and non-pathogenic *Escherichia coli* strains isolated from
 healthy ruminants. *J Antimicrob Chemother* 2001; **48**: 421-4.
- Gorman R, Adley CC. Nalidixic acid-resistant strains of *Salmonella* showing
 decreased susceptibility to fluoroquinolones in the mid-west region of the Republic of
 Ireland. *J Antimicrob Chemother* 2003; **51**:1047-9.
- 106 Sáenz Y, Zarazaga M, Briñas L *et al*. Mutations in *gyrA* and *parC* genes in
 nalidixic acid-resistant *Escherichia coli* strains from food products, humans and
 animals. *J Antimicrob Chemother* 2003; 51:1001-5.
- 107 Van Looveren M, Daube G, De Zutter L *et al.* Antimicrobial susceptibilities of *Campylobacter* strains isolated from food animals in Belgium. *J Antimicrob Chemother* 2001; 48: 235-40.
- 108 Piddock LJ, Ricci V, Pumbwe L *et al.* Fluoroquinolone resistance in *Campylobacter* species from man and animals: detection of mutations in
 topoisomerase genes. *J Antimicrob Chemother* 2003; **51**:19-26.
- 109 Biswas S, Maggi RG, Papich MG *et al*. Molecular mechanisms of *Bartonella henselae* resistance to azithromycin, pradofloxacin and enrofloxacin. *J Antimicrob*

703 *Chemother* 2010; **65**: 581-2.

110 Guo L, Zhang J, Xu C *et al*. Molecular characterization of fluoroquinolone
resistance in *Haemophilus parasuis* isolated from pigs in South China. *J Antimicrob Chemother* 2011; **66**: 539-42.

111 Hauschild T, Feßler AT, Billerbeck C *et al.* Target gene mutations among methicillin-resistant *Staphylococcus aureus* and methicillin-susceptible *S. aureus* with elevated MICs of enrofloxacin obtained from diseased food-producing animals or food of animal origin. *J Antimicrob Chemother* 2012; **67**: 1791-3.

112 van Boven M, Veldman KT, de Jong MC *et al* Rapid selection of quinolone
resistance in *Campylobacter jejuni* but not in *Escherichia coli* in individually housed
broilers. *J Antimicrob Chemother* 2003; 52: 719-23.

113 Delsol AA, Woodward MJ, Roe JM. Effect of a 5 day enrofloxacin treatment on *Salmonella enterica* serotype Typhimurium DT104 in the pig. *J Antimicrob Chemother* 2004; **53**: 396-8.

114 Delsol AA, Sunderland J, Woodward MJ *et al.* Emergence of fluoroquinolone
 resistance in the native *Campylobacter coli* population of pigs exposed to
 enrofloxacin. *J Antimicrob Chemother* 2004; **53**: 872-4.

115 Randall LP, Eaves DJ, Cooles SW *et al.* Fluoroquinolone treatment of
experimental *Salmonella enterica* serovar Typhimurium DT104 infections in chickens
selects for both *gyrA* mutations and changes in efflux pump gene expression. *J Antimicrob Chemother* 2005; **56**: 297-306.

116 Kehrenberg C, de Jong A, Friederichs S *et al.* Molecular mechanisms of
 decreased susceptibility to fluoroquinolones in avian *Salmonella* serovars and their
 mutants selected during the determination of mutant prevention concentrations. *J Antimicrob Chemother* 2007; **59**: 886-92.

Journal of Antimicrobial Chemotherapy

117 Kehrenberg C, Friederichs S, de Jong A *et al.* Identification of the plasmidborne quinolone resistance gene *qnrS* in *Salmonella enterica* serovar Infantis. *J Antimicrob Chemother* 2006; **58**: 18-22.

118 Kehrenberg C, Hopkins KL, Threlfall EJ *et al.* Complete nucleotide sequence
 of a small *qnrS1*-carrying plasmid from *Salmonella enterica* subsp. enterica
 Typhimurium DT193. *J Antimicrob Chemother* 2007; **60**: 903-5.

119 Cavaco LM, Hendriksen RS, Aarestrup FM. Plasmid-mediated quinolone
 resistance determinant *qnrS1* detected in *Salmonella enterica* serovar Corvallis
 strains isolated in Denmark and Thailand. *J Antimicrob Chemother* 2007; 60: 704-6.

Avsaroglu MD, Helmuth R, Junker E *et al.* Plasmid-mediated quinolone
resistance conferred by *qnrS1* in *Salmonella enterica* serovar Virchow isolated from
Turkish food of avian origin. *J Antimicrob Chemother.* 2007; **60**:1146-50.

121 Cavaco LM, Korsgaard H, Sørensen G *et al.* Plasmid-mediated quinolone
resistance due to *qnrB5* and *qnrS1* genes in *Salmonella enterica* serovars Newport,
Hadar and Saintpaul isolated from turkey meat in Denmark. *J Antimicrob Chemother*2008; **62**: 632-4.

Wu CM, Wang Y, Cao XY *et al.* Emergence of plasmid-mediated quinolone
 resistance genes in Enterobacteriaceae isolated from chickens in China. *J Antimicrob Chemother* 2009; **63**: 408-11.

Fortini D, García-Fernández A, Veldman K *et al.* Novel genetic environment of
plasmid-mediated quinolone resistance gene *qnrB2* in *Salmonella* Bredeney from
poultry. *J Antimicrob Chemother* 2009; **64**: 1332-4.

Guerra B, Helmuth R, Thomas K *et al.* Plasmid-mediated quinolone resistance
determinants in *Salmonella* spp. isolates from reptiles in Germany. *J Antimicrob Chemother* 2010; **65**: 2043-5.

125 Fortini D, Fashae K, García-Fernández A *et al.* Plasmid-mediated quinolone
 resistance and β-lactamases in *Escherichia coli* from healthy animals from Nigeria. *J Antimicrob Chemother* 2011; **66**: 1269-72.

Veldman K, Cavaco LM, Mevius D *et al.* International collaborative study on
 the occurrence of plasmid-mediated quinolone resistance in *Salmonella enterica* and
 Escherichia coli isolated from animals, humans, food and the environment in 13
 European countries. *J Antimicrob Chemother* 2011; **66**: 1278-86.

Dolejska M, Villa L, Minoia M *et al.* Complete sequences of IncHI1 plasmids
 carrying *bla*_{CTX-M-1} and *qnrS1* in equine *Escherichia coli* provide new insights into
 plasmid evolution. *J Antimicrob Chemother* 2014; **69**: 2388-93.

128 Medhus A, Slettemeås JS, Marstein L *et al.* Methicillin-resistant
 Staphylococcus aureus with the novel *mecC* gene variant isolated from a cat
 suffering from chronic conjunctivitis. *J Antimicrob Chemother* 2013; 68: 968-9.

129 Loncaric I, Kübber-Heiss A, Posautz A *et al.* Characterization of methicillin resistant *Staphylococcus* spp. carrying the *mecC* gene, isolated from wildlife. *J* Antimicrob Chemother 2013; 68: 2222-5.

769 **130** Gómez P, González-Barrio D, Benito D *et al*. Detection of methicillin-resistant

570 Staphylococcus aureus (MRSA) carrying the mecC gene in wild small mammals in

771 Spain. *J Antimicrob Chemother* 2014; **69**: 2061-4.

131 Espinosa-Gongora C, Harrison EM, Moodley A *et al*. MRSA carrying *mecC* in
captive mara. *J Antimicrob Chemother* 2015; **70**:1622-4.

132 Haenni M, Châtre P, Tasse J et al. Geographical clustering of mecC-positive

775 Staphylococcus aureus from bovine mastitis in France. J Antimicrob Chemother

776 2014; **69**: 2292-3.

133 Małyszko I, Schwarz S, Hauschild T. Detection of a new *mecC* allotype,
 mecC2, in methicillin-resistant *Staphylococcus saprophyticus*. *J Antimicrob Chemother* 2014; 69: 2003-5.

134 Kehrenberg C, Ojo KK, Schwarz S. Nucleotide sequence and organization of
 the multiresistance plasmid pSCFS1 from *Staphylococcus sciuri*. *J Antimicrob Chemother* 2004; 54: 936-9.

135 Zhang WJ, Xu XR, Schwarz S *et al.* Characterization of the IncA/C plasmid
 pSCEC2 from *Escherichia coli* of swine origin that harbours the multiresistance gene
 cfr. J Antimicrob Chemother 2014; 69: 385-9.

Wendlandt S, Li J, Ho J *et al.* Enterococcal multiresistance gene cluster in
 methicillin-resistant *Staphylococcus aureus* from various origins and geographical
 locations. *J Antimicrob Chemother* 2014; 69: 2573-5.

137 Wendlandt S, Lozano C, Kadlec K *et al.* The enterococcal ABC transporter
 gene *Isa*(E) confers combined resistance to lincosamides, pleuromutilins and
 streptogramin A antibiotics in methicillin-susceptible and methicillin-resistant
 Staphylococcus aureus. J Antimicrob Chemother 2013; **68**: 473-5.

138 Li B, Wendlandt S, Yao J *et al.* Detection and new genetic environment of the
pleuromutilin-lincosamide-streptogramin A resistance gene *lsa*(E) in methicillinresistant *Staphylococcus aureus* of swine origin. J *Antimicrob Chemother* 2013;
68:1251-5.

Wendlandt S, Li B, Lozano C *et al.* Identification of the novel spectinomycin
resistance gene *spw* in methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* of human and animal origin. *J Antimicrob Chemother* 2013;
68:1679-80.

140 Jamrozy DM, Coldham NG, Butaye P *et al.* Identification of a novel plasmidassociated spectinomycin adenyltransferase gene *spd* in methicillin-resistant *Staphylococcus aureus* ST398 isolated from animal and human sources. *J Antimicrob Chemother* 2014; **69**:1193-6.

141 Wendlandt S, Feßler AT, Kadlec K *et al.* Identification of the novel
spectinomycin resistance gene *spd* in a different plasmid background among
methicillin-resistant *Staphylococcus aureus* CC398 and methicillin-susceptible *S. aureus* ST433. *J Antimicrob Chemother* 2014; **69**: 2000-3.

Wendlandt S, Kadlec K, Schwarz S. Four novel plasmids from *Staphylococcus hyicus* and CoNS that carry a variant of the spectinomycin resistance gene *spd*. *J Antimicrob Chemother* 2015; **70**: 948-9.

Li J, Li B, Wendlandt S et al. Identification of a novel *vga*(E) gene variant that
confers resistance to pleuromutilins, lincosamides and streptogramin A antibiotics in
staphylococci of porcine origin. *J Antimicrob Chemother* 2014; **69**: 919-23.

Fischer J, Rodríguez I, Schmoger S *et al. Salmonella enterica* subsp. enterica
producing VIM-1 carbapenemase isolated from livestock farms. *J Antimicrob Chemother* 2013; **68**: 478-80.

Fischer J, Schmoger S, Jahn S *et al.* NDM-1 carbapenemase-producing *Salmonella enterica* subsp. enterica serovar Corvallis isolated from a wild bird in
Germany. *J Antimicrob Chemother* 2013; **68**: 2954-6.

146 Stolle I, Prenger-Berninghoff E, Stamm I *et al.* Emergence of OXA-48
carbapenemase-producing *Escherichia coli* and *Klebsiella pneumoniae* in dogs. *J Antimicrob Chemother* 2013; 68: 2802-8.

- 147 Wang Y, Wang X, Schwarz S *et al.* IMP-45-producing multidrug-resistant *Pseudomonas aeruginosa* of canine origin. *J Antimicrob Chemother* 2014; 69: 257981.
- 148 Al Bayssari C, Dabboussi F, Hamze M *et al.* Emergence of carbapenemaseproducing *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in livestock
 animals in Lebanon. *J Antimicrob Chemother* 2015; **70**: 950-1.
- 830 **149** Zhang WJ, Lu Z, Schwarz S *et al*. Complete sequence of the *bla*_{NDM-1}-carrying
- plasmid pNDM-AB from Acinetobacter baumannii of food animal origin. J Antimicrob
- 832 Chemother 2013; 68:1681-2.
- **150** Wang B, Sun D. Detection of NDM-1 carbapenemase-producing *Acinetobacter*
- *calcoaceticus* and *Acinetobacter junii* in environmental samples from livestock farms.
- 835 *J Antimicrob Chemother* 2015; **70**: 611-3.
- 151 Woodford N, Wareham DW, Guerra B *et al.* Carbapenemase-producing
 Enterobacteriaceae and non-Enterobacteriaceae from animals and the environment:
 an emerging public health risk of our own making? J *Antimicrob Chemother* 2014; 69:
 287-91.
- 840 **152** Abraham S, Wong HS, Turnidge J et al. Carbapenemase-producing bacteria
- in companion animals: a public health concern on the horizon. *J Antimicrob Chemother* 2014; **69**:1155-7.
- 843 **153** Poirel L, Stephan R, Perreten V *et al*. The carbapenemase threat in the animal
 844 world: the wrong culprit. *J Antimicrob Chemother* 2014; **69**: 2007-8.
- 154 Kehrenberg C, Schwarz S. Plasmid-borne florfenicol resistance in *Pasteurella multocida*. *J Antimicrob Chemother* 2005; 55: 773-5.
- 847 155 Kehrenberg C, Meunier D, Targant H et al. Plasmid-mediated florfenicol
- resistance in *Pasteurella trehalosi*. J Antimicrob Chemother 2006; **58**:13-7.

Li B, Zhang Y, Wei J *et al.* Characterization of a novel small plasmid carrying
the florfenicol resistance gene *floR* in *Haemophilus parasuis*. *J Antimicrob Chemother* 2015; **70**: 3159-61.

157 Kehrenberg C, Catry B, Haesebrouck F *et al. tet*(L)-mediated tetracycline
resistance in bovine *Mannheimia* and *Pasteurella* isolates. *J Antimicrob Chemother*2005; 56: 403-6.

Blanco M, Kadlec K, Gutiérrez Martín CB *et al.* Nucleotide sequence and
transfer properties of two novel types of *Actinobacillus pleuropneumoniae* plasmids
carrying the tetracycline resistance gene tet(H). *J Antimicrob Chemother* 2007; **60**:
864-7.

Kehrenberg C, Schwarz S. Trimethoprim resistance in a porcine *Pasteurella aerogenes* isolate is based on a *dfrA1* gene cassette located in a partially truncated
class 2 integron. *J Antimicrob Chemother* 2011; **66**: 450-2.

160 Kadlec K, Kehrenberg C, Schwarz S. Efflux-mediated resistance to florfenicol
and/or chloramphenicol in *Bordetella bronchiseptica*: identification of a novel
chloramphenicol exporter. *J Antimicrob Chemother* 2007; 59: 191-6.

161 Liu H, Wang Y, Wu C *et al.* A novel phenicol exporter gene, *fexB*, found in
enterococci of animal origin. *J Antimicrob Chemother* 2012; 67: 322-5.

162 He T, Shen J, Schwarz S *et al.* Characterization of a genomic island in
Stenotrophomonas maltophilia that carries a novel *floR* gene variant. *J Antimicrob*Chemother 2015; **70**:1031-6.

163 González-Zorn B, Catalan A, Escudero JA *et al.* Genetic basis for
dissemination of *armA*. *J Antimicrob Chemother* 2005; **56**: 583-5.

872	164 Chen L, Chen ZL, Liu JH et al. Emergence of RmtB methylase-producing
873	Escherichia coli and Enterobacter cloacae isolates from pigs in China. J Antimicrol
874	<i>Chemother</i> 2007; 59 : 880-5.

Yao Q, Zeng Z, Hou J *et al.* Dissemination of the *rmtB* gene carried on IncF
and IncN plasmids among Enterobacteriaceae in a pig farm and its environment. J *Antimicrob Chemother* 2011; 66: 2475-9.

166 Du XD, Wu CM, Liu HB *et al.* Plasmid-mediated ArmA and RmtB 16S rRNA
methylases in *Escherichia coli* isolated from chickens. *J Antimicrob Chemother* 2009;
64, 1328-30.

167 Liu J, Keelan P, Bennett PM *et al.* Characterization of a novel macrolide efflux
gene, *mef*(B), found linked to *sul3* in porcine *Escherichia coli*. *J Antimicrob Chemother* 2009; 63: 423-6.

168 Wang Y, Yao H, Deng F. Identification of a novel *fosXCC* gene conferring
fosfomycin resistance in *Campylobacter*. *J Antimicrob Chemother* 2015; **70**:1261-3.

169 Zhao S, Mukherjee S, Chen Y *et al.* Novel gentamicin resistance genes in *Campylobacter* isolated from humans and retail meats in the USA. *J Antimicrob Chemother* 2015; **70**: 1314-21.

170 Kadlec K, van Duijkeren E, Wagenaar JA *et al.* Molecular basis of rifampicin
resistance in methicillin-resistant *Staphylococcus pseudintermedius* isolates from
dogs. *J Antimicrob Chemother* 2011; 66:1236-42.

Riesenberg A, Feßler AT, Erol E *et al.* MICs of 32 antimicrobial agents for *Rhodococcus equi* isolates of animal origin. *J Antimicrob Chemother*. 2014; 69: 10459.

Grave K, Lingaas E, Bangen M *et al.* Surveillance of the overall consumption
of antibacterial drugs in humans, domestic animals and farmed fish in Norway in
1992 and 1996. *J Antimicrob Chemother* 1999; **43**: 243 -52.

Pardon B, Catry B, Dewulf J *et al.* Prospective study on quantitative and
qualitative antimicrobial and anti-inflammatory drug use in white veal calves. *J Antimicrob Chemother* 2012; **67**:1027-38.

901 **174** Regula G, Torriani K, Gassner B *et al.* Prescription patterns of antimicrobials in
 902 veterinary practices in Switzerland. *J Antimicrob Chemother* 2009; **63**: 805-11.

903 **175** van den Bogaard AE, London N, Stobberingh EE. Antimicrobial resistance in
904 pig faecal samples from the Netherlands (five abattoirs) and Sweden. *J Antimicrob*905 *Chemother* 2000; **45**: 663-71.

906 **176** Jensen VF, Jakobsen L, Emborg HD *et al.* Correlation between apramycin and
907 gentamicin use in pigs and an increasing reservoir of gentamicin-resistant
908 *Escherichia coli. J Antimicrob Chemother* 2006; **58**:101-7.

177 Nguyen VT, Carrique-Mas JJ, Ngo TH *et al.* Prevalence and risk factors for
carriage of antimicrobial-resistant *Escherichia coli* on household and small-scale
chicken farms in the Mekong Delta of Vietnam. *J Antimicrob Chemother* 2015; 70:
2144-52.

913 **178** van den Bogaard AE, Willems R, London N *et al.* Antibiotic resistance of
914 faecal enterococci in poultry, poultry farmers and poultry slaughterers. *J Antimicrob*915 *Chemother* 2002; **49**: 497-505.

916 **179** Hammerum AM, Larsen J, Andersen VD *et al.* Characterization of extended-

917 spectrum β-lactamase (ESBL)-producing *Escherichia coli* obtained from Danish pigs,

pig farmers and their families from farms with high or no consumption of third- or

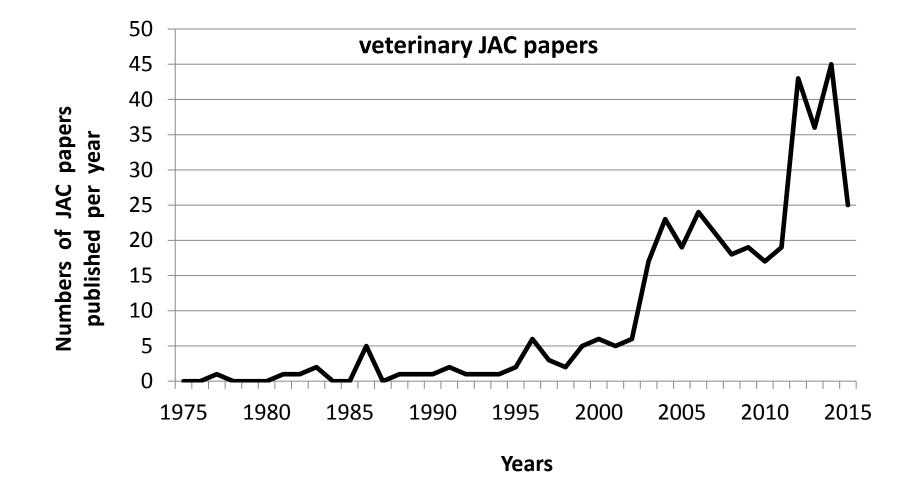
fourth-generation cephalosporins. *J Antimicrob Chemother* 2014; **69**: 2650-7.

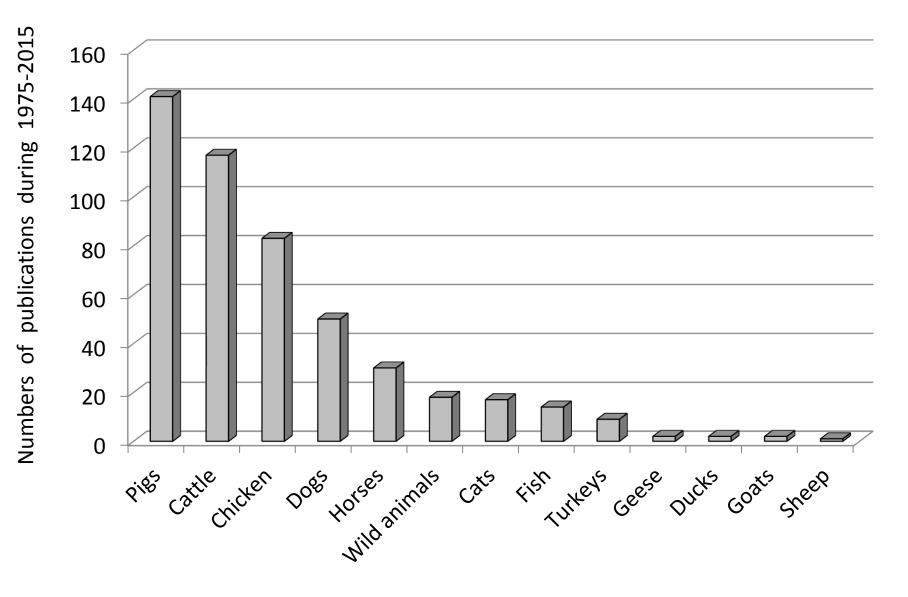
920 Figure 1. Numbers of 'veterinary' papers published in JAC during 1975-2015.

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925 Figure 2. Numbers of veterinary papers published in JAC during 1975-2015 according to the animals species involved. It should be noted that papers which dealt 926 with more than one animal species, are separately listed for each animal species 927 involved. Consequently, the total number of the papers listed in Fig. 2 exceeds the 928 a actual total number of the 'veterinary' papers published in JAC during 1975-2015. 929





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