

## **Pharmacotherapy in paediatric epilepsy: from trial and error to rational drug and dose selection – a long way to go**

Sven van Dijkman [van Dijkman, S.C.] [1]

Ricardo Alvarez-Jimenez [Alvarez-Jimenez, R.] [1]

Meindert Danhof [Danhof, M][1]

Oscar Della Pasqua [Della Pasqua, O.] [2,3]

[1] Division of Pharmacology. Leiden Academic Centre for Drug Research. Einsteinweg 55, 2333CC Leiden, The Netherlands

[2] Clinical Pharmacology and Discovery Medicine. GlaxoSmithKline. Stockley Park, United Kingdom

[3] Clinical Pharmacology and Therapeutics. University College London, 29-39 Brunswick Square London WC1N 1AX United Kingdom

Corresponding author:

Oscar Della Pasqua

Phone: +44 207 679 9796

Email: [o.dellapasqua@ucl.ac.uk](mailto:o.dellapasqua@ucl.ac.uk)

Address: Clinical Pharmacology & Therapeutics

BMA House

Tavistock Square

WC1 H9HX London

United Kingdom

## **Pharmacotherapy in paediatric epilepsy: from trial and error to rational drug and dose selection – a long way to go**

**Introduction:** Whereas ongoing efforts in epilepsy research focus on the underlying disease processes, the lack of a physiologically-based rationale for drug and dose selection contributes to inadequate treatment response in children. In fact, limited information on the interindividual variation in pharmacokinetics and pharmacodynamics of anti-epileptic drugs (AEDs) in children drive prescription practice, which relies primarily on dose regimens according to a mg/kg basis. Such practice has evolved despite advancements in paediatric pharmacology showing that growth and maturation processes do not correlate linearly with changes in body size. **Areas covered:** In this review we aim to provide 1) a comprehensive overview of the sources of variability in the response to AEDs, 2) insight into novel methodologies to characterise such variation and 3) recommendations for treatment personalisation. **Expert Opinion:** The use of pharmacokinetic-pharmacodynamic principles in clinical practice is hindered by the lack of biomarkers and by practical constraints in the evaluation of polytherapy. The identification of biomarkers and their validation as tools for drug development and therapeutics will require some time. Meanwhile, one should not miss the opportunity to integrate the available pharmacokinetic data with modelling and simulation concepts to prevent further delays in the development of personalised treatments for paediatric patients.

**Key words:** Antiepileptic drugs, dose rationale, epilepsy, epileptic seizures, modelling and simulation, paediatrics, personalised medicine, pharmacokinetics, pharmacokinetic-pharmacodynamic relationships, translational pharmacology.

### **Article highlights**

\* Despite the development of therapeutic guidelines for the treatment of epileptic seizures, AED selection and dose rationale for children remains empirical.

\* The use of dosing regimens in mg/kg does not correct for age-related changes in pharmacokinetics and pharmacodynamics in children, especially if one considers the use of polytherapy with two or more AEDs.

\* Inter- and intraindividual differences in pharmacokinetics and pharmacodynamics of AEDs need to be taken into account for the personalisation of treatment in paediatric epilepsy.

\* Whilst the identification of predictive biomarkers remains a challenging endeavour, quantitative clinical pharmacology methods can provide guidance for both anti-epileptic drug and dose selection. These methods allow for evidence synthesis, integration, and extrapolation of findings across different age groups, enabling better clinical decision-making and improved therapeutic response in children.

## **1. Introduction**

Epilepsy is a debilitating syndrome with an estimated 68 million people worldwide affected by it, which places the disease in the 7<sup>th</sup> position in terms of impact on disability and premature mortality among mental health, neurological, and substance-use disorders[1,2]. In addition, it takes the 19<sup>th</sup> rank out of 53 items accounting for the total costs for medical care generated in the area of neurology [3]. Whereas global figures may differ, recent prevalence data in the USA show that nearly 25% were children aged below 15 years of age [4].

Effective treatment and management of epileptic seizures has an important and direct impact on the quality of life of patients, especially those in the paediatric group. Despite the implementation and advancement of therapeutic guidelines, achieving such results remains a challenging objective. This situation prevails in the face of increasing understanding of the progression of the disease after onset in different age groups and introduction of regulatory requirements for the evaluation of efficacy and safety of AEDs in children [5,6].

### **1.1 Current drug and dose selection rationale in paediatric epilepsy**

Various guidelines exist on the diagnostic, management and treatment of epilepsies. However, only a few of them have focused on the use of antiepileptic drugs (AEDs) in children [7-9]. In fact, the British National Institute for Health and Care Excellence (NICE) guideline on epilepsy in children is the only document based on extensive review of the evidence for differences in efficacy and safety of each AED between types of epilepsy [9]. Even though recommendations are supported by evidence arising from randomised controlled trials, shortcomings are still evident. Many studies have been performed to show differences in efficacy and safety between seizure types, but no effective predictors have yet been found for differences in efficacy and safety within the same seizure type. This is likely the consequence of symptom-based criteria, which remain the foundation for diagnosis and AED treatment

selection. In addition, most paediatric trials rely on an “add-on approach”, with patients who may have more severe or refractory forms of epilepsy, which leads to inadequate evidence regarding the efficacy of monotherapy in treatment naive patients. This shortcoming is often compounded by the definition of response (clinical endpoint) in most clinical trials, which is based on a binary measure: responder (i.e., patients who show at least 50% of reduction in seizures compared to baseline) vs. non-responder. Dichotomisation of the response into two categories can be detrimental for the characterisation of dose-exposure-response relationships, especially if one considers that pharmacokinetic data are not collected systematically in efficacy trials.

Whereas limited understanding of the exposure-response relationships might be mitigated by the clinical requirement for up and down-titration or tapering of the dose. In addition to reducing side effects and withdrawal symptoms, tapering procedures offer an opportunity to factor in the effect of interindividual pharmacokinetic and pharmacodynamic variability. Yet, this information is not fully integrated to support treatment personalisation. Currently, most formularies still rely on anecdotal (empirical) evidence of efficacy and safety in children. Dose recommendations in formularies, such as the Netherlands *Kinderformularium* or the British National Formulary for Children overlook the role of covariate factors and other sources of variability in pharmacokinetics and pharmacodynamics [10,11]. Clearly, there is a substantial amount of pharmacokinetic data regarding the use of AED in children, but even when taking into account correlations with weight and age, unexplained variability appears to remain high [12-14]. Similar challenges are faced when considering the adjustment of maintenance doses of AEDs. In spite of the use of therapeutic drug monitoring (TDM), which is widely accepted in paediatric epilepsy compared to adults, AED levels are checked against a therapeutic window, which was originally determined in adults. Moreover, these therapeutic

ranges ignore known to covariate effects, which may cause variability in exposure and potentially in the exposure-response relationship.

One should also note the impact of variability in the status of the disease at the time of diagnosis and its progression, which are a hurdle for improved therapeutics and may possibly be associated with the unnecessary exposure of paediatric patients to AEDs for years after the seizures have remitted [15]. Thus, the combination of unexplained variability in pharmacokinetics, pharmacodynamic and disease leaves clinicians without a clear dosing algorithm, other than the option to taper and adjust doses based on the clinical symptoms.

The challenges a clinician faces to select the drug and dose regimen are illustrated in numerous publications on the efficacy and safety of AEDs in children [16-18]. In the next paragraphs we will highlight how dosing algorithms can be used as a valuable therapeutic tool before switching treatment or progressing to polytherapy.

## **1.2 Personalised treatment of epileptic seizures: advancing clinical practice**

The ultimate goal of a (personalised) therapeutic intervention is to ensure a positive, if not optimal, balance between the expected benefits and risks of the treatment, taking into account the costs and the inherent uncertainties about favourable and unfavourable effects [19, 20]. This concept is particularly relevant when dealing with chronic diseases such as epilepsy, but little effort has been made to evaluate the impact of a one-size fits all approach on the overall effectiveness of antiepileptic drugs. In fact, one needs to recognise that heterogeneity in the disease makes it a case for exploring treatment options beyond current guidelines. For instance, some patients may achieve complete seizure remission with higher doses before adding on a second drug, but evaluation of higher doses requires more than empirical titration. It should be guided by dosing algorithms, which take into account the role covariate factors

associated with inter- and intraindividual variability in pharmacokinetics and pharmacodynamics.

Unfortunately, formal assessment of the advantages of dosing algorithms for personalised treatment with AEDs is fraught with difficulties as it imposes the evaluation of changes in the benefit-risk balance (BRB). The determination of the BRB of a treatment requires precise, detailed information on the relationships between the dose, exposure and its favourable and unfavourable effects on the symptoms and signs of the disease. Given that the BRB of AEDs is not characterised in a quantitative manner during drug development, evidence arising from clinical practice may be too limited to allow accurate decision-making. Consequently, establishing criteria for the choice of the drug and the dose for the treatment of epileptic seizures in children cannot be performed adequately without quantifying the contribution of different sources of variability to heterogeneity in PK, PD, and disease, as discussed in previous paragraphs. Opportunities exist however to explore each of these factors (one by one and in combination) and subsequently evaluate the implications of different treatment options on the overall BRB. This can be achieved by means of model-based meta-analytical approaches including extrapolation and simulation scenarios in which patient characteristics, drug properties and disease features are integrated [19,21,22].

The aims of this review are therefore to 1) discuss the impact of known sources of variability in PK, PD, and disease and 2) explore how quantitative clinical pharmacology concepts can be used to support the development of dosing algorithms to ensure that treatment choice and dosing rationale for paediatric patients are as effective as possible. We show that some improvement may be achieved in spite of the limitations of current diagnosis criteria, lack of biomarkers and poor understanding of the mechanisms of action of AEDs. To this end, a structured literature search was performed in conjunction with supporting material from

clinical guidelines and regulatory documentation on the assessment of efficacy and safety of drugs in the paediatric population. The Pubmed search included MESH terms as well as individual and combined keywords. An overview of the initial search strategy is provided in Figure 1, where selection criteria are listed in a hierarchical manner to capture publications describing paediatric epilepsy, personalisation of treatment, pharmacokinetics, pharmacodynamics, pharmacogenetics, and biomarkers. Reviews as well as perspective papers were included in the analysis if relevant paediatric details were provided. When necessary, a separate search algorithm was used to identify publications on specific issues such as methodologies for data extrapolation and assessment of benefit-risk balance in children. If no relevant literature was retrieved, additional terms were included or excluded. The initial search resulted in a total of 145 articles, of which 56 were selected after screening the abstracts for relevance. These were complemented by an additional 70 publications, which were obtained from secondary queries and interactions with experts in paediatric clinical pharmacology.

## **2. Intrinsic sources of variability and heterogeneity in response to AEDs**

Numerous hurdles have contributed to the emphasis in current practice regarding the use of seizure reduction (i.e., clinical response) for switching treatment and monitoring of systemic drug levels as the basis for modifying or individualising the dose and dosing regimen. Sadly, the notion that plasma levels, even at steady state, may not reflect differences in target exposure or pharmacodynamics is unfamiliar to most prescribing physician. This limitation is also critical for the development of new AEDs, as the evaluation of dose-response in clinical trials relies primarily on the assumption of target plasma levels and a predefined therapeutic range. In the next sections, we will discuss the implications of variability in pharmacokinetics, pharmacodynamics and in relevant physiological factors for the personalisation of treatment.



## **2.1 Pharmacokinetics**

The pharmacokinetics of a drug is determined by up to four physiological processes, namely absorption, distribution, metabolism and excretion (ADME). Metabolism and excretion are usually summarised by systemic clearance (plasma volume being cleared of the drug per time unit; CL). Summary measures of drug disposition is often limited to the so-called secondary pharmacokinetics parameters such as peak concentration ( $C_{max}$ ), trough concentration ( $C_{min}$ ), and mean steady state ( $C_{ss}/C_{avg}$ ) concentrations, as well as the area under the concentration vs. time curve (AUC). It is important to note that secondary parameters are derived from primary PK parameters. For instance, following extravascular administration, peak concentrations depend on absorption rate, and volume of distribution, whilst  $C_{ss}$  and AUC are directly related to clearance. From a therapeutic perspective, response to AEDs is most likely explained by the average exposure or trough concentrations, with acute and some chronic side effects primarily being determined by peak concentrations. Hence, variability in the processes that determine drug disposition may affect treatment response. In this respect, one needs to consider that some of these ADME processes are incomplete or immature at birth and young age, especially in pre-term infants [23,24] (Table 1). Despite the impact of these factors on drug exposure, in most cases they are not included into the dose rationale for children. Details on the differences in the pharmacokinetics of specific AEDs in children can be found elsewhere [23, 25]. In the next paragraphs we describe the main factors determining the differences in ADME between adults and children, and overall variability in the PK of AEDs.

### **2.1.1 Drug distribution: differences between plasma and target site concentrations**

Plasma protein binding can be an important factor determining differences in pharmacokinetics, both with respect to drug distribution and clearance. In theory, only unbound drug concentrations distribute to the brain. Some authors have focused therefore on the free concentrations or free fraction of AEDs (for example carbamazepine [26], phenytoin [27], valproate [28]). In these publications, the free plasma concentration of the drug was found to better reflect the concentrations of the extracellular space and the brain's interstitial fluid. However, brain distribution can be complex and variable depending on factors related to active transport mechanisms, disease-related changes in tissue permeability and other co-morbidities. For instance, Clinkers et al. studied the influence of epileptic seizures on the concentration of oxcarbazepine in the hippocampus and in plasma in a rat model. [29]. Concentrations reached higher values in the interstitial space within the pilocarpine-induced acute seizures region and were even higher when oxcarbazepine was given in combination with a P-glycoprotein (Pgp) inhibitor. Most importantly, these differences were observed without significant changes in drug levels in plasma. These results illustrate the complex role of the functioning of the blood brain barrier (BBB) as a determinant of the target exposure. Indeed, up-regulation of the efflux transporter Pgp has been indicated as one of the possible explanations for the development of apparent tolerance [30].

Whereas active transport processes may determine tissue distribution, high variability in drug exposure can exist even between closely located areas in the brain. This was already described in 1978 in patients who had surgery after receiving carbamazepine in regular stable doses [31]. Rambeck et al. [32] analysed plasma, cerebrospinal fluid (CSF) and extracellular space (ECS) concentrations in to-be-excised live temporal brain tissue (*in vivo* with a microdialysis probe and *ex vivo* directly in the removed tissue) in patients refractory to treatment. As expected, brain extra-cellular concentrations were lower compared to plasma and CSF, which

demonstrates that the assumption of equal concentrations in CSF and ECS in one well distributed homogenous compartment is unjustified [33]. A general lack of information regarding differences in drug distribution in children, and particularly in infants and toddlers, (i.e., in the developing brain), as compared to adults renders the interpretation of treatment failure quite challenging, as lack of efficacy may not be a matter of refractoriness to therapy, but rather a pharmacokinetic problem.

### **2.1.2 Clearance: influence of genotype, size, and maturation**

Inter-individual, and intra-individual variability in drug elimination processes mostly results from differences in the availability of the drug at the clearing organ, changes in the clearing capacity due to varying intrinsic clearance, and the size of the organ.

Although it is known that organ perfusion varies with age [34], specific quantitative information regarding hepatic and renal changes are still sparse in some groups of the paediatric population. Consequently, it is unclear to what degree variability in organ perfusion determines the changes in clearance between adults and children. Similarly, very limited information is available regarding AED protein binding in young children and its implications for differences in systemic clearance between adults and children [35,36].

Intrinsic clearance can also be influenced by polymorphisms in genes coding for metabolising enzymes which may lead to significant differences in hepatic clearance of many AEDs [37], with increase or reduction in metabolic capacity resulting in different phenotypes [38]. Similarly, renal clearance can be affected by differences in the expression level of renal transporters [39,40]. Whilst the impact of such genetic differences can be accounted for when defining the dose and dosing regimen, genotyping or phenotyping are not used in standard practice when initiating or changing therapy, and is most probably not encouraged in children. Apart from the differences in the genetic make-up of the clearing organ, age-dependent

changes also affect the amount of drug that can be cleared. As a child grows organs develop both in terms of size and metabolic capacity (i.e., enzyme activity). It has been postulated that the influence of increasing size on clearance can, at least in part, be accounted for by adjusting for body weight. However, the relation between size (e.g. body weight) and elimination rate has been demonstrated to be non-linear. This implies that dosing in mg/kg does not accurately correct for the underlying differences [41]. In fact, unless explicit differences have been identified in the underlying pharmacokinetic-pharmacodynamic relationship, dose adjustment in children should aim at achieving comparable exposure or similar PK profile across the target population, irrespective of body weight or age. One needs to be aware that whereas the use of weight-banded dosing regimens may be necessary to compensate for such nonlinearity, drug-drug interactions may have a higher impact on clearance than the effect of body size (Figure 2) [42–45].

## **2.2 Pathophysiology and pharmacodynamics**

Every brain is unique in its structure, connectivity, plasticity, and neurotransmitter homeostasis. As a result, wide intra- and inter-individual variation is observed in the response to CNS active drugs. Differences in physiology, whether genetic, congenital or acquired, can both give rise to epileptic seizures and affect one's ability to respond to treatment. In fact, over the course of the disease, these differences as well as the progression of the underlying (patho-)physiological processes can change the way the brain responds to seizures, and consequently to therapy. In other words, variability in physiology begets variability in disease progression and treatment response, which in turn beget changes in physiology. Disentangling this circular web of interactions is perhaps the most challenging of the issues plaguing the field of AED therapy. Whereas characterising such interactions on an individual patient level may be unrealistic in the foreseeable future, personalisation of treatment may be achieved by

identifying disease-specific factors that are age-related or common to subgroups in the population. The impact of such concepts has been illustrated in a recent investigation by Pellock and collaborators who showed that evidence of efficacy in partial-onset seizures (POS) in adults can be used to predict drug response in children [5]. Yet, in other childhood epilepsies that persist or evolve to adulthood, changes in pathophysiology are not yet understood well enough to allow individual prediction of outcome.

Another challenging aspect in the characterisation of interindividual differences is the nature of the interaction between drug and receptor or target. From a pharmacological point of view, pharmacodynamics (PD) describes the interaction between a drug and its target or receptor and the transduction mechanisms leading to a change in function. PD processes are a major determinant of the efficacy/safety profile of AEDs, but little is known about their (molecular) mechanisms. This is partly due to the fact that most AEDs have been discovered on the basis of phenotypic screening at a time when brain imaging and other innovative functional protocols were not available. Moreover, drug development in epilepsy has traditionally aimed at evaluating efficacy in adults. Only recently, changes in regulatory requirements have defined the need to characterise the efficacy and safety of AEDs in children. Such a sequential approach may however be inappropriate to address childhood-specific epilepsies.

### **2.2.1 Assessment of anti-epileptic drug response: symptoms *versus* functional measures of brain activity**

In spite of the advances in imaging technologies, the evaluation of brain physiology *in vivo* remains a challenging undertaking. Although EEG is regularly used to identify pathological signs and confirm diagnosis, patients are not routinely subjected to a long-term biochemical and/or electrophysiological evaluation throughout the course of the disease and its treatment. Medical history (i.e. occurrence of seizures) rather than measurement of physiological

endpoints is used to support clinical evaluation and decision making regarding the choice of drug and dosing regimens.

Clearly, the lack of data regarding the correlation between AED exposure, pharmacological effects (i.e. biomarkers) and therapeutic response (i.e. seizure reduction or suppression) makes it difficult for a physician to predict which treatment, and which exposure level, will work best for an individual patient or group. Close monitoring of the variation in response between patients over the course of treatment time is required to understand the role of differences in brain physiology. Such a monitoring imposes the availability of biomarkers which are sufficiently sensitive to detect variations in response as well as to predict treatment failure or toxicity. To date, the only known valid antiepileptic drug biomarker is HLA-B\*1502, which is a strong predictor of Stevens-Johnson syndrome in patients of specific Asian backgrounds taking carbamazepine [46]. No other parameters exist with sufficient predictive performance for efficacy.

Another point to consider in paediatric epilepsy is the role of neuronal maturation in the progression of epilepsy. Maturation and neurological development are processes that take place during growth. Changes in the expression of voltage gate dependent ion channels as well as structural changes associated with growth can have an impact on the sensitivity of the brain to a drug and consequently on the magnitude of drug effects [47]. Similarly, the time of diagnosis and initiation of AED therapy are potential causes of variability in treatment response. For example, the clinical management of seizures in the new-born has remained unchanged in spite of evidence that “classic” medications (phenobarbital and phenytoin) are largely ineffective (with more than half of the population being non-responders for both drugs) and potentially neurotoxic [48]. Most symptomatic seizures in neonates are due to hypoxic-ischemic encephalopathy and do not persist beyond the first few days of life. Due to

this natural improvement, any prompt intervention would appear effective and even curative. Such an apparent efficacy, which is wrongly attributed to the drug could be relevant across many types of epilepsy and result in AEDs being used more often than necessary, especially in the case of the developing brain of a new-born infant. This is particularly worrying if one takes into account the effect of AEDs on cognitive development and growth [49–53].

### **2.2.2 Disease progression and maturation**

In paediatric epilepsy, it is clearly the natural progression of disease varies not only between patients, but also between and within epilepsy subtypes and syndromes [54,55]. For instance, benign epilepsy with centrotemporal spikes (BECST) typically occurs between the age of 3 – 14 years of age and resolves by age 17 despite the incidence of cognitive and behavioural disorders [56]. By contrast, Lennox-Gastaut syndrome begins between the age of 1-6, with seizures that generally do not respond well to treatment [57] Schmidt *et al.* estimated that without intervention, 20-44% of untreated epilepsies remit within one to two years [58]. Of the remaining patients, around 60% will respond favourably to therapy and the rest will present an insidious or recurrent syndrome in which approximately half of this subpopulation will not respond to treatment. Unfortunately, the authors seem to pay little attention to the differences between types of epilepsy and their aetiology [59,60]. Even more controversial are the prognostic factors for response to treatment, as only around 11% of patients with lack of efficacy to the first AED will respond to the second treatment option [15]. Without relevant biomarkers it is impossible to predict disease progression and/or treatment response. Consequently, clinical decisions regarding treatment choice and dose selection are determined by the disease status at time of the diagnosis or intervention.

### **2.2.3 Target receptor polymorphisms, density, and adaptation**

Many AEDs are believed to share a common mechanism of action through the interaction at the receptor level, usually an ion channel on the surface of the target neurons [61]. In addition, it can be assumed that *caeteris paribus* the higher the target engagement the stronger the signal being transmitted or blocked. Consequently, the exposure-response curve of an AED *in vivo* will vary depending on the availability (density) of receptors [62]. Additional variability may arise from polymorphisms of target receptors (which can be caused by differences in the aetiology of epilepsy) as well as from variable binding kinetics at the target. Indeed, changes to binding kinetics can alter drug potency, which in turn affects the dosing requirements [63]. From a clinical perspective, it should be highlighted that epileptic patients often experience a decreased drug effect over the course of treatment, which cannot be explained by the aforementioned processes or mechanisms. This reduction may be a gradual process, but often occurs suddenly, possibly after discontinuation and reinstatement of drug therapy. One of the potential causes of pharmacoresistance is down/up regulation of the target receptors [64–66]. In these circumstances, whereas increases in the dose may off-set the effects of down-regulation, higher drug exposure may lead to side effects, preventing achievement of satisfactory response levels. Pharmacoresistance has been reported to affect about 23% of paediatric patients [67], whom respond better to surgical intervention than adults. [68].

### **3. Extrinsic sources of variability and heterogeneity in response to AEDs**

Apart from the biological factors implicated in previous sections, some extrinsic factors limit our understanding of the PKPD relationships of AEDs and consequently may affect treatment choice and dose selection for the paediatric population. Here we focus on the implications of food-drug and drug-drug interactions, as well as on the impact of variable treatment adherence.



### **3.1 Drug-food interaction and formulation variability**

Most used AEDs have been off-patent for some time and thus generic versions exist in all kinds of formulations. Although the pharmacologically active substance is the same, and bioequivalence studies should provide evidence for similar exposure to the drug, different formulations have been introduced, which are intended to modify drug release profile and as such can lead to faster or slower absorption possibly resulting in different peak concentrations [69] and consequently in a different safety profile [70]. This issue can be compounded by small differences in the bioavailability (fraction of the dose that is absorbed and reaches the systemic circulation) of AEDs (Figure 4)[71]. For example, the bioavailability of carbamazepine is considered to be 80% on average, but ranges considerably [72]. In the case of gabapentin, bioavailability is inversely proportional to the taken dose, resulting in reduced increases in exposure with increasing doses [73]. Finally, absorption and first pass metabolism can be influenced by food intake and beverages, such as grapefruit juice [74]. These factors are difficult to control but can contribute to overall variability in the exposure to AEDs. Thus, to minimise the influence of absorption kinetics on the disposition of AEDs, many extended-release formulations have been developed for adult patients, which reduce peak/trough concentration ratios while maintaining similar overall exposure. By contrast, extended release tablet formulations are not always an option in children, as swallowing such tablets can be too difficult for younger patients. This limitation could be overcome by specially designed liquid extended-release formulations [75].

### **3.2. Drug combinations and drug-drug interactions**

Current clinical guidelines recommend drug combination or polytherapy only in those cases in which monotherapy is proven to be insufficiently effective. In the case of effective polytherapy, it is suggested to taper off the previous treatment to achieve monotherapy over a

longer time interval. Monotherapy is therefore assumed to be the best treatment choice, but this practice does not take into account the possibility of pharmacodynamic interactions, and in particular, synergy, for which some evidence exists [76–78]. Combining drugs with a different mechanism of action may offer the best chance of achieving synergistic interactions, although there is scarce evidence for this concept from clinical trials [79]. These claims occur despite the lack of consensus on whether patients might benefit of an alternative drug or multiple AEDs [80]. On the other hand, pharmacokinetic drug-drug interactions (DDI) have been identified for many AEDs. Consequently, it may be challenging to disentangle changes in drug effects due to a pharmacodynamic interaction from the effects associated with changes in the exposure due to the primary AED. Given safety and ethical constraints, the characterisation of possible pharmacodynamic interactions remains difficult in a clinical setting.

### **3.2 Adherence to treatment**

Treatment with AEDs often leads to cognitive, behavioural and physical adverse effects [81]. When such effects are experienced as burdensome, it is likely that a patient will not comply with the prescribed regimen and take short or longer drug holidays, leading to poor persistence and eventually discontinuation of treatment [82]. Whereas some of these adverse effects can be prevented or reversed by adjusting the dose correctly for the individual patient or group, limited information is available on the impact that drug holidays have both on the efficacy and safety profile of AEDs. This issue is further compounded in paediatric epilepsy, as adherence does not involve on the patients themselves, but parents or caregivers who can also interfere with drug intake. In fact, random missingness of the dose during a single day of treatment can already decrease exposure levels significantly. A recent study has found that approximately a quarter of the paediatric patients are nonpersistent in taking their prescribed

AED therapy, but the impact of variable adherence on treatment outcome was not evaluated [83].

Given that poor adherence is often not disclosed by patients, physicians may attribute a potential loss of efficacy to disease or pharmacodynamic factors, rather than to variation in drug exposure due to variable patterns of drug intake. In this case, patients may be recommended a dose increase or an alternative treatment, which may result in increased incidence of adverse effects [82]. Open, honest communication between physician, patients and parents when necessary is therefore critical to minimise the risk of inaccurate treatment decisions [84].

#### **4. Conclusion**

Children are not small adults and it is known that syndromes in paediatric epilepsy undergo variable progression and changes in the natural course of the disease due to neurodevelopment. Changes in pharmacokinetic, pharmacodynamic and physiological processes associated with maturation and developmental growth determine the differences in response to AED treatment in this population. Many of these changes occur concurrently, preventing accurate prediction of the response (and prognosis) at an individual patient level. An integrated approach, supported by potential biomarkers and dosing algorithms is needed to ensure appropriate selection of drug(s) and dose for a specific patient or group of patients. Regardless the large amount of data collected on existing and new AEDs, knowledge is not sufficiently integrated to support the implementation of treatment personalisation. This lack of integration prevails, despite efforts by health technology assessment organisations to establish the effectiveness of available medicines. Guidelines such as NICE rely on published evidence, which may lag considerably behind the introduction of a new medicinal product into clinical

practice. Moreover, such guidelines are not fit-for-purpose, i.e., do not specifically focus on subgroups in such a way that fully supports the use of personalised treatments in children.

To allow paediatricians to better decide on which AED(s) to prescribe and at which dose, a novel approach is required that takes into account the aforementioned complexities of epilepsy [85]. A promising, readily available methodology for the selection of a drug and dosing regimen is PKPD and disease modelling [86]. However, to be an effective resource for treatment personalisation, biomarkers must be identified that are sensitive to the disease state and progression, so that efficacy and toxicity of drugs can be better characterised in clinical practice. Undoubtedly, the availability of biomarkers would also represent an advancement to diagnosis, minimising the need for a trial-and-error approach to pharmacotherapy [87–90]. In our expert opinion, we explore how the application of model-based algorithms may achieve these goals.

## **5. Expert Opinion**

### **5.1. Definition of treatment response and assessment of efficacy and safety**

Seizure frequency or similar continuous measures be considered as primary endpoints for the assessment of efficacy. The use of number of responders, i.e., patients achieving a decrease in seizure count of at least 50% at the end of the study relative to baseline and the percentage of the population that achieves such “seizure control” compared to placebo or a control treatment are not sufficiently informative. Such a dichotomisation of the response results in a loss of information, as it does not allow the characterisation of the drug effect at the individual patient level. As a result, personalisation of treatment, including dosing recommendations cannot be derived unless a broad dose range is tested and stratified for. Such a requirement is unrealistic as more patients would be required for adequate evaluation of response in a

clinical trial. This limitation is further compounded by bias in the comparison between experimental and control treatments when applying the aforementioned response criteria [91]. In addition to the use an endpoint which offers more granularity to the evaluation of efficacy, experimental protocols need to be revisited. Typically, the efficacy of new AEDs is tested in a so-called “add on” trial design, in which patients who are refractory to treatment receive the new drug. This complicates the interpretation of the results for a variety of reasons. First, it introduces selection bias in drug potency and on the required dose recommendations. In patients who are refractory to treatment, response is expected to be less than in non-refractory patients. Moreover, the observed response is the result of a combination of the direct effect of the drug and/or an interaction with the background treatment. As a result, interactions must be taken into account to establish the magnitude of the effect of the new drug in the absence of other AEDs. These limitations apply *a fortiori* in children. Ethical considerations make it virtually impossible to evaluate efficacy and safety in children according to typical Phase IIb dose ranging studies.

## **5.2 Understanding and predicting variability**

L.B. Sheiner envisioned a learning-confirming paradigm [92] in which available prior information is first used to *learn* by prediction or extrapolation using modelling and simulation techniques (evidence synthesis), where possible taking into account multiple sources of information (integration). An experiment can then be optimised to address the gaps in knowledge (evidence generation), the outcome of which is then used to *confirm* the predictions and build new theories and models (Figure 4). More specifically with regard to the use of AEDs in paediatric epilepsy, accurate predictions of treatment response may be achieved as a result of systematic integration of data on pharmacokinetics, pharmacodynamics

and disease [93]. Such an approach may have direct implications for the implementation of personalised treatments, including dosing algorithms for paediatric patients.

The use of PKPD and disease models relies on current understanding of the disease and pharmacology. Usually, one endeavours to describe the biological system of interest with sufficient detail to ensure accurate predictions for a range of possible interventions. This process relies on a set of assumptions is often referred to as parameterisation and is aimed at identifying descriptors of the physiological or pharmacological effects in a simple, but yet robust manner. For instance, using a PK model instead of collecting and summarising drug concentrations only, it is possible to predict the time course of the drug concentrations following drug administration of different doses and dosing regimens, as well as better account for the impact of covariates such as body weight or age. Similarly, PKPD and disease models provide the basis for the assessment of the interaction between drug and biological system, taking into account the progression or changes associated with the disease itself. Such parameterisation also allows one to quantify the impact of influential factors on parameter values and describe them as covariates. The incorporation of covariates into a PKPD or disease model has an important advantage in that it enhances the prediction of response for specific groups of patients [94–96]. In conjunction with clinical trial simulations, model-based techniques offer an excellent opportunity for the evaluation of novel therapies [97] as well as personalisation of the dosing regimen for children [98].

### **5.2.1 Personalised treatment**

Clinical guidelines for epilepsy [99] still rely on diagnostic criteria which are primarily determined by symptoms, Consequently, AED treatment selection is based on the underlying epileptic syndrome, as defined by the type of epileptic seizures (e.g. partial, primary or

secondary generalised, absence, etc.) and age (adults, children, etc.), with aetiology playing only a minor role. For each syndrome group, multiple lines of treatment are considered. Given the heterogeneity in the aetiology of the disease within each group, it is likely that the different treatment options simply reflect the uncertainty about the interindividual differences in response.

A more mechanistic approach is required for the classification of seizures, as it would facilitate the distinction between AEDs which can modify the disease from those which act on symptoms [100]. The use of disease modelling can also contribute to another pressing issue, i.e., the nature and magnitude of the effect of drug-drug interactions. It has been proposed that combining AEDs with different mechanisms of action might have a synergistic effect compared to combining those with a similar mechanisms of action, but no research has conclusively supported this idea [101]. By contrast, others have suggested a more practical approach of exploring doses and combinations in difficult refractory cases [102]. A more aggressive pre-emptive intervention may very well be the answer to treatment resistant epilepsy, but no systematic studies are available to support this hypothesis. Despite concerns about the use polytherapy, the concept is appealing especially in children if evidence can be gathered of the implications of early interventions with multiple AEDs. Advancements will only become tangible after sensitive biomarkers have been identified. In conjunction with disease modelling, biomarkers may also allow one to discriminate the contribution of one of more compounds to the overall response and determine whether AEDs affect disease progression.

In the absence of biomarkers, long term longitudinal (observational) studies represent an important step to further characterise the pros and cons of a given intervention. It is regrettable that no attempts have been made to apply disease modelling concepts to

(pharmaco)epidemiological studies. Despite the retrospective nature of such an approach, important insight may be gained about predictors and determinants of response in children.

### **5.2.2 Personalised dose and dosing regimen**

As previously stated, 10-20% of refractory patients can benefit from dose adjustments [15], but little discussion exists in the literature regarding appropriate dosing in non-refractory patients. In fact, it is likely that in numerous cases the lack of response to AEDs may occur due to inadequate dosing, whereas other patients may experience adverse events due to overexposure. Efforts from therapeutic drug monitoring have not addressed this issue and caused PK considerations to be misinterpreted during clinical decision about the dose and dosing regimen of AED. Most importantly, limited attention is given to the role of covariates that are known affect PK and potentially alter the efficacy and safety profile of an AED.

Since therapeutic concentration ranges for each AED are available in literature, such data can be used with PK models, including the contribution of covariates to identify suitable titration and maintenance dosing algorithms. Unfortunately, these therapeutic concentration ranges were generally determined in the adult population, making their relevance for the different epilepsy subtypes in the paediatric population questionable. The development of dosing algorithms is particularly important for the paediatric population, irrespective of the lack of further data on exposure-response and exposure-toxicity relationships. A major benefit from this approach is the opportunity to provide recommendations for dosing adjustment taking into account complex drug-drug interactions in a strictly quantitative manner; this issue is poorly addressed by current therapeutic guidelines. In this context, simulation scenarios can also be explored to predict the response to drug combinations also in refractory patients. Whilst one needs to acknowledge the role of disease progression over time in paediatric



epilepsy, efforts to ensure comparable exposure to drugs, irrespective of their age or body weight, represent a more robust approach than trial and error in a vulnerable patient population.

We also note that despite the considerable number of publications aimed at PK modelling of AED, most authors offer this as a somewhat technical description of ADME properties of the drugs. Most publications lack insight into core clinical pharmacology issues and do not expand their analysis and interpretation to meet clinical needs such as dose rationale and implications for prescription practice. In summary, the information available is not being integrated and most importantly, the lack of a “big picture” regarding core clinical pharmacology principles seems to perpetuate the gaps in data generation, i.e., missing information is not being generated. Figure 5 depicts the steps required to ensure personalised treatment, with a stronger rationale for drug and dose selection. Clinical dosing could be enhanced by algorithms, which are more efficient than typical titration procedures and therapeutic drug monitoring (TDM). Combined with dried blood spot or saliva analysis techniques, the burden of TDM on the paediatric patient could be minimised. [103,104] The benefits of a model-based approach are illustrated in a simulation study [online supplement 1], using published data as an example of what dosing algorithms can represent to clinical practice in paediatric epilepsy [105]. Clearly, effective implementation of dosing algorithms imposes further integration of existing and new evidence on the efficacy and safety of AEDs. It also demands for extrapolation tools and evidence generation based on more informative experimental protocols. The potential impact to such efforts is highlight in the following paragraphs.

## **5.3 Evidence synthesis**

### **5.3.1 Integration of historical and new evidence**

One of the most powerful characteristics of model-based approaches is the possibility of integrating information from different sources and combining them with statistical concepts to make predictions about new scenarios, beyond the experimental evidence available from the data itself. Given the complexity of epilepsy's many interacting factors, these techniques represent a valuable research tool in this field. Currently, its use remains, however, limited to pharmacokinetic data analysis.

### **5.3.2 Extrapolations**

Translational medicine can be defined as extrapolating findings from basic science and quickly making them useful for practical applications that enhance human health [106]. Whilst its implementation is often limited to stand-alone experimental protocols, translational steps can be achieved by the use of model-based extrapolations [107,108]. The use of extrapolations based on clinically and biologically plausible assumptions can make translational medicine a valid and powerful tool. The approach involves appropriate scrutiny by simulation exercises enabling the integration of different types of data, such as pre-clinical *in vitro* (cell lines, tissue, organs), *in vivo* (mice, rats, dogs, etc.) and clinical data [109, 110]. Of interest is the role that extrapolations can have to characterise differences and similarities between paediatric and adult patients [111-113]. As recently defined by the European Medicines Agency (EMA), extrapolation may be generally defined as: "Extending information and conclusions available from studies in one or more subgroups of the patient population (source population), or in related conditions or with related medicinal products, to make inferences for another subgroup of the population (target population), or condition or product, thus reducing the need to generate additional information (types of studies, design modifications, number of patients required) to reach conclusions for the target population, or condition or medicinal product" [6].

It should be clear that the primary rationale for extrapolation is to avoid unnecessary studies in children. However, extrapolations are not generally acceptable as a default approach [Table 2]. As discussed previously, an interesting finding in epilepsy is the extrapolation of efficacy results in adults to predict a similar adjunctive treatment response in 2- to 18-year-old children with partial onset seizure [5].

#### **5.4 Evidence generation**

An important shortcoming of the primary measure of efficacy is the fact that seizure reduction from baseline does not reflect changes in epileptic activity in the brain in a strictly quantitative manner nor does it relate to the mechanism of the drug on such processes. In fact, a more careful evaluation of this criterion may not be comparable across all subpopulations [114]. Clearly, early, sensitive biomarkers and endpoints are essential to accurately characterise interactions of drug(s) and disease. One needs to establish how drug effects interact with the underlying disease and explore whether longitudinal changes in such endpoints can be used to predict long term response to treatment. So far, very few attempts have been made to identify predictors of response or treatment failure; such investigations have however relied on seizure reduction or establish the potential prognostic rather than predictive value of the variables of interest (Figure 3) [115]. Therefore we strongly support the views that clinical research protocols need to integrate clinical measures to markers of physiological and pharmacological effects of AEDs. In this context, imaging techniques need to be coupled to the evaluation of efficacy in clinical trials. Functional magnetic resonance imaging (fMRI) and positron emission tomography (PET) represent promising opportunities, but their evaluation as biomarkers in epilepsy has not yet been fully explored [116–118] and may be too burdensome to use in paediatric epilepsy.

A final point to consider in evidence generation is the informative value of data, which should include, rather than exclude relevant covariates and influential factors on exposure-response relationships. Numerous examples exist where early adoption of modelling and simulation has led to better trial design, in particular with regard to the dose selection and characterisation of influential factors on PK , PD and response [121,122]. Although successful studies have been conducted to derive paediatric dosing based on empirical designs, others failed and possibly could have been successful based on modelling and simulation [123–125]. In summary, clinical researchers and regulators need to acknowledge the limitations of traditional protocols to evaluate efficacy and safety of AEDs in children [126–128]. Effective implementation of personalised treatment for the paediatric population requires concerted efforts to ensure that experimental data are generated and integrated beyond traditional statistical hypothesis testing. Lessons can be learned from recent developments in oncology [129], where clinical trials, treatment and dose selection have undergone major advancements both conceptually and clinically over the last decade.

## References

1. Ngugi AK, Bottomley C, Kleinschmidt I, et al. Estimation of the burden of active and life-time epilepsy: a meta-analytic approach. *Epilepsia* 2010;51:883–90.
2. England MJ, Liverman CT, Schultz AM, et al. Epilepsy across the spectrum: Promoting health and understanding. Committee on the Public Health Dimensions of the Epilepsies. Institute of Medicine (ed)., Washington, DC, The National Academies Press, 2012, p 19-47.
3. Olesen J, Gustavsson A, Svensson M, et al. The economic cost of brain disorders in Europe. *Eur. J. Neurol.* 2012;19:155–62.
4. Cardarelli WJ, Smith BJ. The Burden of Epilepsy to Patients and Payers. *Am. J. Manag. Care* 2010;16:331–336.
5. Pellock JM, Carman WJ, Thyagarajan V, et al. Efficacy of antiepileptic drugs in adults predicts efficacy in children: a systematic review. *Neurology* 2012;79:1482–9.
6. European Medicines Agency. Concept paper on extrapolation of efficacy and safety in medicine development. EMA/129698/2012 2012;
7. Krumholz A, Wiebe S, Gronseth GS, et al. Evidence-based guideline: Management of an unprovoked first seizure in adults. Report of the Guideline Development Subcommittee of the American Academy of Neurology and the American Epilepsy Society. *Neurology* 2015; 84: 1705-1713.
8. French JA, Kanner AM, Bautista J, et al. Efficacy and tolerability of the new antiepileptic drugs I: Treatment of new onset epilepsy. Report of the Therapeutics and Technology Assessment Subcommittee and Quality Standards Subcommittee of the American Academy of Neurology and the American Epilepsy Society. *Neurology* 2004; 62:1252-1260
9. National Institute for Health and Clinical Excellence (NICE). The epilepsies: the diagnosis and management of the epilepsies in adults and children in primary and secondary care. London (UK): National Institute for Health and Clinical Excellence (NICE); 2012 Jan. 117 p. (Clinical guideline; no. 137).
10. Kinderformularium. Nederlands Kenniscentrum voor Farmacotherapie bij Kinderen (NKFK), 2016 (<https://www.kinderformularium.nl/>, last accessed on 15 May 2016).
11. Paediatric Formulary Committee. BNF for Children. London: BMJ Group, Pharmaceutical Press, and RCPCH Publications; 2016
12. Cella M, Knibbe C, de Wildt SN, et al. Scaling of pharmacokinetics across paediatric populations: the lack of interpolative power of allometric models. *Br. J. Clin. Pharmacol.* 2012;74:525–35.
13. Cella M, Zhao W, Jacqz-Aigrain E, et al. Paediatric drug development: Are population models predictive of pharmacokinetics across paediatric populations? *Br. J. Clin. Pharmacol.* 2011;72:454–464.6.

14. Anderson BJ, Holford NHG. Mechanism-based concepts of size and maturity in pharmacokinetics. *Annu. Rev. Pharmacol. Toxicol.* 2008;48:303–32.
15. Stagi S, Lasorella S, Piccorossi A, et al. Cessation of epilepsy therapy in children. *Expert Rev. Neurother.* 2016;16:549–559.
16. Min K, Hur Y, Yu H, et al. Initial response to antiepileptic drugs in patients with newly diagnosed epilepsy. *J. Clin. Neurosci.* 2014;21:923–926.
17. Kwan P, Brodie MJ. Epilepsy after the first drug fails: substitution or add-on? *Seizure* 2000;9:464–8.
18. Sillanpää M, Schmidt D. Predicting antiepileptic drug response in children with epilepsy. *Expert Rev. Neurother.* 2011;11:877–85; quiz 886.
19. Bellanti F, van Wijk RC, Danhof M, Della Pasqua O. Integration of PKPD relationships into benefit-risk analysis. *Br. J. Clin. Pharmacol.* 2015;80:979–991.
20. EMA. Benefit-risk methodology project tools and processes for regulatory benefit-risk. 2011;44:1–33.
21. Holmes E, Plumpton C, Duerden M, Marson T, Hughes D. New advice on switching antiepileptic drugs might be a false economy. *BMJ.* 2013; 347:f7471.
22. Plumpton CO, Yip VL, Alfirevic A, Marson AG, Pirmohamed M, Hughes DA. Cost-effectiveness of screening for HLA-A\*31:01 prior to initiation of carbamazepine in epilepsy. *Epilepsia.* 2015; 56: 556-63.
23. Perucca E. Clinical pharmacokinetics of new-generation antiepileptic drugs at the extremes of age. *Clin. Pharmacokinet.* 2006;45:351–363.
24. Johannessen SI, Tomson T. Pharmacokinetic variability of newer antiepileptic drugs: when is monitoring needed? *Clin. Pharmacokinet.* 2006;45:1061–75.
25. Italiano D, Perucca E. Clinical pharmacokinetics of new-generation antiepileptic drugs at the extremes of age: an update. *Clin Pharmacokinet.* 2013; 52: 627-45.
26. Deleu D, Aarons L, Ahmed IA. Population pharmacokinetics of free carbamazepine in adult Omani epileptic patients. *Eur. J. Clin. Pharmacol.* 2001;57:243–248.
27. Deleu D, Aarons L, Ahmed IA. Estimation of population pharmacokinetic parameters of free-phenytoin in adult epileptic patients. *Arch. Med. Res.* 2005;36:49–53.
28. Ueshima S, Aiba T, Makita T, et al. Characterization of non-linear relationship between total and unbound serum concentrations of valproic acid in epileptic children. *J. Clin. Pharm. Ther.* 2008;33:31–8.
29. Clinckers R, Smolders I, Michotte Y, et al. Impact of efflux transporters and of seizures on the pharmacokinetics of oxcarbazepine metabolite in the rat brain. *Br. J. Pharmacol.* 2008;155:1127–1138.
30. Löscher W, Schmidt D. Experimental and clinical evidence for loss of effect (tolerance) during prolonged treatment with antiepileptic drugs. *Epilepsia* 2006;47:1253–84.

31. Friis ML, Christiansen J, Hvidberg EF. Brain concentrations of carbamazepine and carbamazepine-10,11-epoxide in epileptic patients. *Eur. J. Clin. Pharmacol.* 1978;14:47–51.
32. Rambeck B, Jürgens UH, May TW, et al. Comparison of brain extracellular fluid, brain tissue, cerebrospinal fluid, and serum concentrations of antiepileptic drugs measured intraoperatively in patients with intractable epilepsy. *Epilepsia* 2006;47:681–694.
33. Christensen J, Højskov CS, Dam M, Poulsen JH. Plasma concentration of topiramate correlates with cerebrospinal fluid concentration. *Ther. Drug Monit.* 2001;23:529–35.
34. Wu C, Honarmand AR, Schnell S, et al. Age-Related Changes of Normal Cerebral and Cardiac Blood Flow in Children and Adults Aged 7 Months to 61 Years. *J. Am. Heart Assoc.* 2016;5:e002657.
35. Koyama H, Sugioka N, Uno A, et al. Age-related alteration of carbamazepine-serum protein binding in man. *J. Pharm. Pharmacol.* 1999;51:1009–1014.
36. Heine R Ter, van Maarseveen EM, van der Westerlaken MML, et al. The Quantitative Effect of Serum Albumin, Serum Urea, and Valproic Acid on Unbound Phenytoin Concentrations in Children. *J. Child Neurol.* 2013;29:803–810.
37. Lopez-Garcia MA, Feria-Romero IA, Fernando-Serrano H, et al. Genetic polymorphisms associated with antiepileptic metabolism. *Front. Biosci. (Elite Ed).* 2014;June:377–386.
38. Chang Y, Yang L, Zhang M, Liu S-Y. Correlation of the UGT1A4 gene polymorphism with serum concentration and therapeutic efficacy of lamotrigine in Han Chinese of Northern China. *Eur. J. Clin. Pharmacol.* 2014;941–946.
39. Urban TJ, Brown C, Castro R, et al. Effects of genetic variation in the novel organic cation transporter, OCTN1, on the renal clearance of gabapentin. *Clin. Pharmacol. Ther.* 2008;83:416–421.
40. Piana C, Antunes NDJ, Pasqua O Della. Implications of pharmacogenetics for the therapeutic use of antiepileptic drugs. *Expert Opin. Drug Metab. Toxicol.* 2014;10:341–358.
41. Anderson BJ, Holford NHG. Mechanistic Basis of Using Body Size and Maturation to Predict Clearance in Humans. *Drug Metab. Pharmacokinet.* 2009;24:25–36.
42. Takeuchi T, Natsume J, Kidokoro H, Ishihara N. The effects of co-medications on lamotrigine clearance in Japanese children with epilepsy. *Brain Dev.* 2016;
43. Mikaeloff Y, Rey E, Soufflet C, et al. Topiramate pharmacokinetics in children with epilepsy aged from 6 months to 4 years. *Epilepsia* 2004;45:1448–52.
44. Bouillon-Pichault M, Jullien V, Bazzoli C, et al. Pharmacokinetic design optimization in children and estimation of maturation parameters: example of cytochrome P450 3A4. *J. Pharmacokinet. Pharmacodyn.* 2011;38:25–40.
45. Vovk T, Jakovljević MB, Kos MK, et al. A nonlinear mixed effects modelling analysis of topiramate pharmacokinetics in patients with epilepsy. *Biol. Pharm. Bull.*

- 2010;33:1176–82.
46. Glauser TA. Biomarkers for antiepileptic drug response. *Biomark. Med.* 2011;5:635–641.
  47. Kwan P, Brodie MJ. Early identification of refractory epilepsy. *N. Engl. J. Med.* 2000;342:314–319.
  48. Painter M, Scher M, Stein A, et al. Phenobarbital compared with Phenytoin for the Treatment of Neonatal Seizures. *N. Engl. J. Med.* 1999;341:485–489.
  49. Bittigau P, Sifringer M, Genz K, et al. Antiepileptic drugs and apoptotic neurodegeneration in the developing brain. *Proc. Natl. Acad. Sci. U. S. A.* 2002;99:15089–94.
  50. Meador KJ, Baker G a., Browning N, et al. Fetal antiepileptic drug exposure and cognitive outcomes at age 6 years (NEAD study): A prospective observational study. *Lancet Neurol.* 2013;12:244–252.
  51. Lee H-S, Wang S-Y, Salter DM, et al. The impact of the use of antiepileptic drugs on the growth of children. *BMC Pediatr.* 2013;13:211.
  52. Banach R, Boskovic R, Einarson T, Koren G. Long-term developmental outcome of children of women with epilepsy, unexposed or exposed prenatally to antiepileptic drugs: a meta-analysis of cohort studies. *Drug Saf.* 2010;33:73–9.
  53. Farwell J, Lee Y, Hirtz DG, et al. Phenobarbital for febrile seizures—effects on intelligence and on seizure recurrence. *N. Engl. J. Med.* 1990;322:364–369.
  54. Nicoletti A, Sofia V, Vitale G, et al. Natural history and mortality of chronic epilepsy in an untreated population of rural Bolivia: A follow-up after 10 years. *Epilepsia* 2009;50:2199–2206.
  55. Okuma T, Kumashiro H. Natural History and Prognosis of Epilepsy : Report of a Multi-institutional Study in Japan. *Epilepsia* 1981;22:35–53.
  56. Vannest J, Tenney JR, Gelineau-morel R, et al. Cognitive and behavioral outcomes in benign childhood epilepsy with centrotemporal spikes. *Epilepsy Behav.* 2015;45:85–91.
  57. Montouris GD, Wheless JW, Glauser TA. The efficacy and tolerability of pharmacologic treatment options for Lennox-Gastaut syndrome FDA-Approved Medications. 55:10–20.
  58. Schmidt D, Sillanpää M. Evidence-based review on the natural history of the epilepsies. *Curr. Opin. Neurol.* 2012;25:159–63.
  59. Sloviter RS. The neurobiology of temporal lobe epilepsy: too much information, not enough knowledge. *C. R. Biol.* 2005;328:143–153.
  60. Bender RA, Baram TZ. Epileptogenesis in the developing brain: what can we learn from animal models? *Epilepsia* 2007;48 Suppl 5:2–6.
  61. Kwan P, Sills GJ, Brodie MJ. The mechanisms of action of commonly used antiepileptic drugs. *Pharmacol. Ther.* 2001;90:21–34.



62. Ploeger BA, van der Graaf PH, Danhof M. Incorporating Receptor Theory in Mechanism-Based Pharmacokinetic-Pharmacodynamic (PK-PD) Modeling. *Drug Metab. Pharmacokinet.* 2009;24:3–15.
63. Hung C-C, Chen C-C, Lin C-J, Liou H-H. Functional evaluation of polymorphisms in the human ABCB1 gene and the impact on clinical responses of antiepileptic drugs. *Pharmacogenet. Genomics* 2008;18:390–402.
64. Weaver DF, Pohlmann-Eden B. Pharmacoresistant epilepsy: unmet needs in solving the puzzle(s). *Epilepsia* 2013;54 Suppl 2:80–5.
65. Vega-hern A, Felix R. Down-Regulation of N-Type Voltage-Activated Ca<sup>2+</sup> Channels by Gabapentin. 2002;2002:185–190.
66. Byrnes JJ, Miller LG, Greenblatt DJ, Shader RI. Chronic benzodiazepine administration. *Biochem. Pharmacol.* 1991;42:S99–S104.
67. Berg AT, Rychlik K. The course of childhood-onset epilepsy over the first two decades : A prospective , longitudinal study. 2014;56:40–48.
68. Francione S, Liava A, Mai R, et al. Drug-resistant parietal epilepsy : polymorphic ictal semiology does not preclude good post-surgical outcome. 2015;17:32–46.
69. Yamada M, Welty TE. Generic Substitution of Antiepileptic Drugs: A Systematic Review of Prospective and Retrospective Studies. *Ann. Pharmacother.* 2012;46:304–304.
70. Crawford P, Feely M, Guberman A, Kraemer G. Are there potential problems with generic substitution of antiepileptic drugs? A review of issues. *Seizure* 2006;15:165–176.
71. Johannessen C, Beiske G, Baftiu A, et al. Experience from therapeutic drug monitoring and gender aspects of gabapentin and pregabalin in clinical practice. *Seizure Eur. J. Epilepsy* 2015;28:88–91.
72. Marino SE, Birnbaum AK, Leppik IE, et al. Steady-state carbamazepine pharmacokinetics following oral and stable-labeled intravenous administration in epilepsy patients: effects of race and sex. *Clin. Pharmacol. Ther.* 2012;91:483–8.
73. Chen C. Meta-analyses of dose-exposure relationships for gabapentin following oral administration of gabapentin and gabapentin enacarbil. *Eur. J. Clin. Pharmacol.* 2013;69:1809–1817.
74. Krauss GL, Caffo B, Chang YT, et al. Assessing bioequivalence of generic antiepilepsy drugs. *Ann. Neurol.* 2011;70:221–228.
75. Mishra B, Sahoo BL, Mishra M, et al. Design of a controlled release liquid formulation of lamotrigine. *Daru* 2011;19:126–37.
76. Jonker DM, Voskuyl RA, Danhof M. Synergistic combinations of anticonvulsant agents: what is the evidence from animal experiments? *Epilepsia* 2007;48:412–34.
77. Lee JW, Dworetzky B. Rational Polytherapy with Antiepileptic Drugs. *Pharmaceuticals* 2010;3:2362–2379.
78. Stafstrom CE. Mechanisms of action of antiepileptic drugs: the search for synergy.

- Curr. Opin. Neurol. 2010;23:157–63.
79. Sake J-K, Hebert D, Isojärvi J, et al. A pooled analysis of lacosamide clinical trial data grouped by mechanism of action of concomitant antiepileptic drugs. *CNS Drugs* 2010;24:1055–68.
  80. Stephen LJ, Brodie MJ. Antiepileptic drug monotherapy versus polytherapy: pursuing seizure freedom and tolerability in adults. *Curr. Opin. Neurol.* 2012;25:164–72.
  81. Zaccara G, Giovannelli F, Giorgi FS, et al. Analysis of placebo effects of antiepileptic drugs across different conditions. *J. Neurol.* 2016;
  82. Faught E. Adherence to antiepilepsy drug therapy. *Epilepsy Behav.* 2012;25:297–302.
  83. Aylward BS, Rausch JR, Modi AC. An Examination of 1-Year Adherence and Persistence Rates to Antiepileptic Medication in Children With Newly Diagnosed Epilepsy. 2015;40:66–74.
  84. Smithson WH, Hukins D, Colwell B, Mathers N. Developing a method to identify medicines non-adherence in a community sample of adults with epilepsy. *Epilepsy Behav.* 2012;24:49–53.
  85. Castro FA De, Piana C, Simões BP, et al. Busulfan dosing algorithm and sampling strategy in stem cell transplantation patients. *Br. J. Clin. Pharmacol.* 2015;80:618–629.
  86. Margineanu DG. Systems biology impact on antiepileptic drug discovery. *Epilepsy Res.* 2012;98:104–15.
  87. Danhof M, Alvan G, Dahl SG, et al. Mechanism-based pharmacokinetic-pharmacodynamic modeling—a new classification of biomarkers. *Pharm. Res.* 2005;22:1432–7.
  88. Engel J, Pitkänen A, Loeb JA, et al. Epilepsy biomarkers. *Epilepsia* 2013;54 Suppl 4:61–9.
  89. Chen X, de Haas S, de Kam M, van Gerven J. An Overview of the CNS-Pharmacodynamic Profiles of Nonselective and Selective GABA Agonists. *Adv. Pharmacol. Sci.* 2012;2012:134523.
  90. Santen G, van Zwet E, Bettica P, et al. From trial and error to trial simulation III: a framework for interim analysis in efficacy trials with antidepressant drugs. *Clin. Pharmacol. Ther.* 2011;89:602–7.
  91. Schobben F, Hekster Y, van Zwieten-Boot B. Outcome measures for the assessment of new antiepileptic drugs. *Pharm. World Sci.* 1997;19:223–226.
  92. Sheiner LB. Learning versus confirming in clinical drug development. *Clin. Pharmacol. Ther.* 1997;61:275–291.
  93. Pennell PB, Peng L, Newport DJ, Ritchie JC, et al. Lamotrigine in pregnancy: clearance, therapeutic drug monitoring, and seizure frequency. *Neurology.* 2008 May 27;70(22 Pt 2):2130-6.

94. Csajka C, Verotta D. Pharmacokinetic-pharmacodynamic modelling: History and perspectives. *J. Pharmacokinet. Pharmacodyn.* 2006;33:227–279.
95. Lalonde RL, Kowalski KG, Hutmacher MM, et al. Model-based drug development. *Clin. Pharmacol. Ther.* 2007;82:21–32.
96. Danhof M, de Jongh J, De Lange ECM, et al. Mechanism-based pharmacokinetic-pharmacodynamic modeling: biophase distribution, receptor theory, and dynamical systems analysis. *Annu. Rev. Pharmacol. Toxicol.* 2007;47:357–400.
97. Lee JY, Garnett CE, Gobburu JVS, et al. Impact of pharmacometric analyses on new drug approval and labelling decisions: a review of 198 submissions between 2000 and 2008. *Clin. Pharmacokinet.* 2011;50:627–35.
98. Knibbe C a J, Danhof M. Individualized dosing regimens in children based on population PKPD modelling: are we ready for it? *Int. J. Pharm.* 2011;415:9–14.
99. National Institute for Health and Care Excellence. Epilepsies: diagnosis and management (CG137). Available at: <http://www.nice.org.uk/guidance/cg137>.
100. Coombes S, Terry JR. The dynamics of neurological disease: Integrating computational, experimental and clinical neuroscience. *Eur. J. Neurosci.* 2012;36:2118–2120.
101. Deckers CL, Hekster YA, Keyser A, et al. Reappraisal of polytherapy in epilepsy: a critical review of drug load and adverse effects. *Epilepsia* 1997;38:570–5.
102. French JA, Faught E. Rational polytherapy. *Epilepsia* 2009;50 Suppl 8:63–8.
103. Milosheska D, Grabnar I, Vovk T. Dried blood spots for monitoring and individualization of antiepileptic drug treatment. *Eur. J. Pharm. Sci.* 2015;75:25–39.
104. Patsalos PN, Berry DJ. Therapeutic drug monitoring of antiepileptic drugs by use of saliva. *Ther. Drug Monit.* 2013;35:5–29.
105. Girgis IG, Nandy P, Nye JS, et al. Pharmacokinetic-pharmacodynamic assessment of topiramate dosing regimens for children with epilepsy 2 to 10 years of age. *Epilepsia* 2010;51:1954–1962.
106. Woolf SH. The meaning of translational research and why it matters. *JAMA* 2008;299:211–3.
107. Danhof M, de Lange ECM, Della Pasqua OE, et al. Mechanism-based pharmacokinetic-pharmacodynamic (PK-PD) modeling in translational drug research. *Trends Pharmacol. Sci.* 2008;29:186–91.
108. Donovan MD, Boylan GB, Murray DM, Cryan JF, Griffin BT. Treating disorders of the neonatal central nervous system: pharmacokinetic and pharmacodynamic considerations with a focus on antiepileptics. *Br J Clin Pharmacol.* 2016; 81:62-77.
109. Brochot A, Zamacona M, Stockis A. Physiologically based pharmacokinetic/pharmacodynamic animal-to-man prediction of therapeutic dose in a model of epilepsy. *Basic Clin. Pharmacol. Toxicol.* 2010;106:256–62.
110. Guillemain I, Kahane P, Depaulis A. Progress in Epileptic Disorders Workshop on

- AED trials Animal models to study aetiopathology of epilepsy: what are the features to model? *Epileptic Disord.* 2012;14:217–225.
111. Dunne J, Rodriguez WJ, Murphy MD, et al. Extrapolation of adult data and other data in pediatric drug-development programs. *Pediatrics* 2011;128:e1242–9.
  112. Chiron C, Dulac O, Pons G. Antiepileptic drug development in children: considerations for a revisited strategy. *Drugs* 2008;68:17–25.
  113. Kang HE, Lee MG. Approaches for predicting human pharmacokinetics using interspecies pharmacokinetic scaling. *Arch. Pharm. Res.* 2011;34:1779–88.
  114. Stefan H, Lopes da Silva FH, Löscher W, et al. Epileptogenesis and rational therapeutic strategies. *Acta Neurol. Scand.* 2006;113:139–55.
  115. Bonnett L, Smith CT, Smith D, et al. Prognostic factors for time to treatment failure and time to 12 months of remission for patients with focal epilepsy: post-hoc, subgroup analyses of data from the SANAD trial. *Lancet Neurol.* 11:331–340.
  116. Ronan L, Alhusaini S, Scanlon C, et al. Widespread cortical morphologic changes in juvenile myoclonic epilepsy: evidence from structural MRI. *Epilepsia* 2012;53:651–8.
  117. Laufs H, Duncan JS. Electroencephalography/functional MRI in human epilepsy: what it currently can and cannot do. *Curr. Opin. Neurol.* 2007;20:417–23.
  118. Kim S, Salamon N, Jackson HA, et al. PET imaging in pediatric neuroradiology: current and future applications. *Pediatr. Radiol.* 2010;40:82–96.
  119. Wang Y, Bhattaram VA, Jadhav PR, et al. Leveraging prior quantitative knowledge to guide drug development decisions and regulatory science recommendations: impact of FDA pharmacometrics during 2004-2006. *J. Clin. Pharmacol.* 2008;48:146–56.
  120. Mid E, Sf W, Burghaus R, et al. Good Practices in Model-Informed Drug Discovery and Development: Practice, Application, and Documentation. 2016;93–122.
  121. Piana C, Surh L, Furst-recktenwald S. Integration of Pharmacogenetics and Pharmacogenomics in Drug Development: Implications for Regulatory and Medical Decision Making in Pediatric Diseases. 2012;704–716.
  122. Cohen AF. Developing drug prototypes: pharmacology replaces safety and tolerability? *Nat. Rev. Drug Discov.* 2010;9:856–65.
  123. Harnisch L, Shepard T, Pons G, Della Pasqua O. Modeling and Simulation as a Tool to Bridge Efficacy and Safety Data in Special Populations. *CPT Pharmacometrics Syst. Pharmacol.* 2013;2:e28.
  124. Jadhav PR, Kern SE. The need for modeling and simulation to design clinical investigations in children. *J. Clin. Pharmacol.* 2010;50:121S–129S.
  125. Wilby J, Kainth A, Hawkins N, et al. Clinical effectiveness, tolerability and cost-effectiveness of newer drugs for epilepsy in adults: a systematic review and economic evaluation. *Health Technol. Assess. (Rockv).* 2005;9:Executive summary.
  126. Glauser T, Ben-menachem E, Bourgeois B, et al. ILAE treatment guidelines:

- evidence-based analysis of antiepileptic drug efficacy and effectiveness as initial monotherapy for epileptic seizures and syndromes. *Epilepsia* 2006;47:1094–120.
127. Perucca E. Designing clinical trials to assess antiepileptic drugs as monotherapy: difficulties and solutions. *CNS Drugs* 2008;22:917–38.
  128. Perucca E. When clinical trials make history: demonstrating efficacy of new antiepileptic drugs as monotherapy. *Epilepsia* 2010;51:1933–5.
  129. Della Pasqua O. PKPD and Disease Modeling: Concepts and Applications to Oncology. In: Kimko HH, Peck CC, eds. *Clinical Trial Simulations: Applications and Trends*. Springer New York, : 281–310.

Table 1. Pharmacokinetic characteristics of commonly used antiepileptic drugs (*adapted from [24]*).

Table 2. Acceptability of different extrapolation approaches for the prediction of disease progression, pharmacodynamics and pharmacokinetics between and within species.

Figure 1. The diagram depicts the search strategy, including MESH terms and keywords used to select the publications included in this review.

Figure 2. An example of the complex interaction between multiple covariates on the clearance of lamotrigine. In this diagram lamotrigin dose-corrected concentrations (DCC) are stratified by groups: Group 1, samples with VPA co-medication; Group 2, samples with LTG metabolic inducers (inducers) (CBZ, PHT, or PB); Group 3, samples with antiepileptic drugs other than VPA and inducers (CBZ, PHT, or PB); Group 4, samples with VPA and inducers (CBZ, PHT, or PB); and Group 5, samples with LTG monotherapy. The bottom and top of each box show the 25th and 75th percentiles, respectively. The horizontal line in each box indicates the median. The groups are indicated by the dotted lines. The horizontal lines in the upper part of the figure indicate significant differences between groups (\* $p < 0.001$ , \*\* $p = 0.01$ ). Among patients with VPA (Group 1) and inducers (Group 2), the DCC of LTG is lower in cases under 6 years old (*adapted from [42]*)

Figure 3. In this example, plots show the relative hazard ratio for age and total number of seizures before randomisation for the time to treatment failure. Hazard ratio estimates with 95% CIs are shown for overall time to treatment failure, for age (A) and total number of

seizures (B), and for time to treatment failure because of inadequate seizure control and because of unacceptable adverse events, for age (C) and total number of seizures (D). Ideally, biomarkers should be identified that can be used as predictors of response or failure without the need to measure the reduction in seizure frequency.

Figure 4. (a) Dose and concentration relationship of (a) gabapentin (n = 189), ref. range (70–120 mmol/L) and (b) pregabalin (n = 167), ref. range (10–30 mmol/L) (*with permission from [71]*).

Figure 5. Information on disease processes, pharmacodynamics and pharmacokinetics must be integrated to ensure accurate personalisation of AED treatment and rational dose selection in children. Whereas interindividual differences in disease and pharmacodynamics of AEDs play an important role in treatment selection, understanding of the effect of developmental growth and maturation processes is essential for the selection of the paediatric dosing regimen.

Table 1. Pharmacokinetic characteristics of commonly used antiepileptic drugs (*adapted from [24]*).

Drug	Time to steady state (d)	Half-life (h)	Tentative therapeutic range <sup>a</sup>		Major route of elimination
			( $\mu\text{mol/L}$ )	( $\mu\text{g/mL}$ )	
Felbamate	3-5	14-22	125-250	30-60	Oxidation and renal excretion
Gabapentin	2	5-7	70-120	12-20	Renal excretion
Lamotrigine	3-15	8-33	10-60	2.5-15	Glucuronide conjugation
Levetiracetam	2	7-8	35-120	8-26	Renal excretion and hydrolysis
Oxcarbazepine	2-3	8-15	50-140 <sup>b</sup>	12-35	Keto-reduction, then glucuronide conjugation of MHD
Pregabalin	2	6-7	NE	2.8-8.2	Renal excretion
Tiagabine	2	7-9	50-250 <sup>c</sup>	20-100 <sup>d</sup>	Oxidation
Topiramate	4-6	20-30	15-60	5-20	Renal excretion, oxidation
Vigabatrin	1-2	5-8	NA	NA	Renal excretion
Zonisamide	5-12	50-70	45-180	10-38	Glucuronide conjugation, acetylation, oxidation and renal excretion

a The lower limit of the therapeutic range is of limited value, because many patients do well at serum concentrations below this limit.  
b Monohydroxy derivative.  
c nmol/L.  
d ng/mL.  
**MHD** = monohydroxy metabolite; **NA** = not applicable; **NE** = not established



Table 2. Acceptability of different extrapolation approaches for the prediction of disease progression, pharmacodynamics and pharmacokinetics between and within species.

<b>Extrapolation of</b>	<b>From</b>	<b>To</b>	<b>Acceptability</b>	<b>References</b>
Disease mechanisms and PD	animals	humans	Unclear	[59,60,109,113,115]
Disease progression and PD with similar aetiology	adults	children	Possibly	[5,116]
Disease progression and PD with different aetiologies	adults	children	Not acceptable	[117]
Pharmacokinetics (allometrically)	animals	humans	Possibly	[118]
Pharmacokinetics (allometrically)	adults	children >3yo	Probably	[13,119,120]

>3yo: older than 3 years

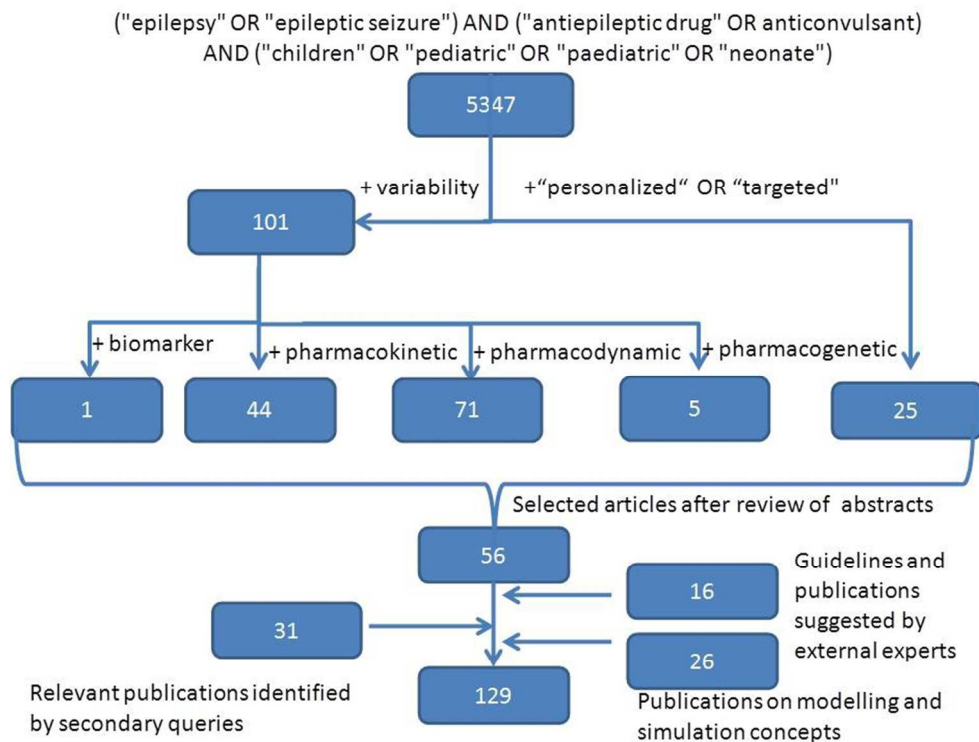


Figure 1. The diagram depicts the search strategy, including MESH terms and keywords used to select the publications included in this review.  
255x192mm (96 x 96 DPI)

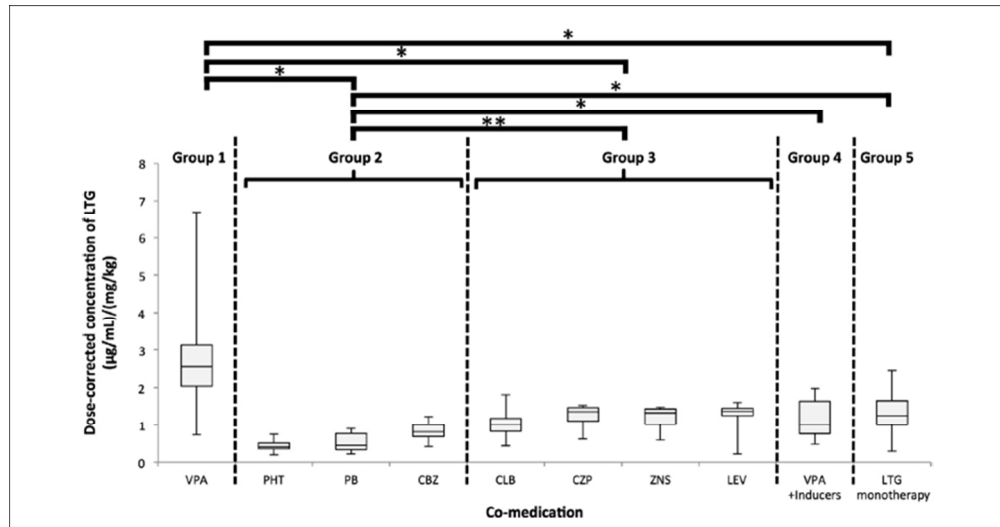


Figure 2. An example of the complex interaction between multiple covariates on the clearance of lamotrigine. In this diagram lamotrigine dose-corrected concentrations (DCC) are stratified by groups: Group 1, samples with VPA co-medication; Group 2, samples with LTG metabolic inducers (inducers) (CBZ, PHT, or PB); Group 3, samples with antiepileptic drugs other than VPA and inducers (CBZ, PHT, or PB); Group 4, samples with VPA and inducers (CBZ, PHT, or PB); and Group 5, samples with LTG monotherapy. The bottom and top of each box show the 25th and 75th percentiles, respectively. The horizontal line in each box indicates the median. The groups are indicated by the dotted lines. The horizontal lines in the upper part of the figure indicate significant differences between groups (\* $p < 0.001$ , \*\* $p = 0.01$ ). Among patients with VPA (Group 1) and inducers (Group 2), the DCC of LTG is lower in cases under 6 years old (adapted from [42])

326x171mm (72 x 72 DPI)

new Only

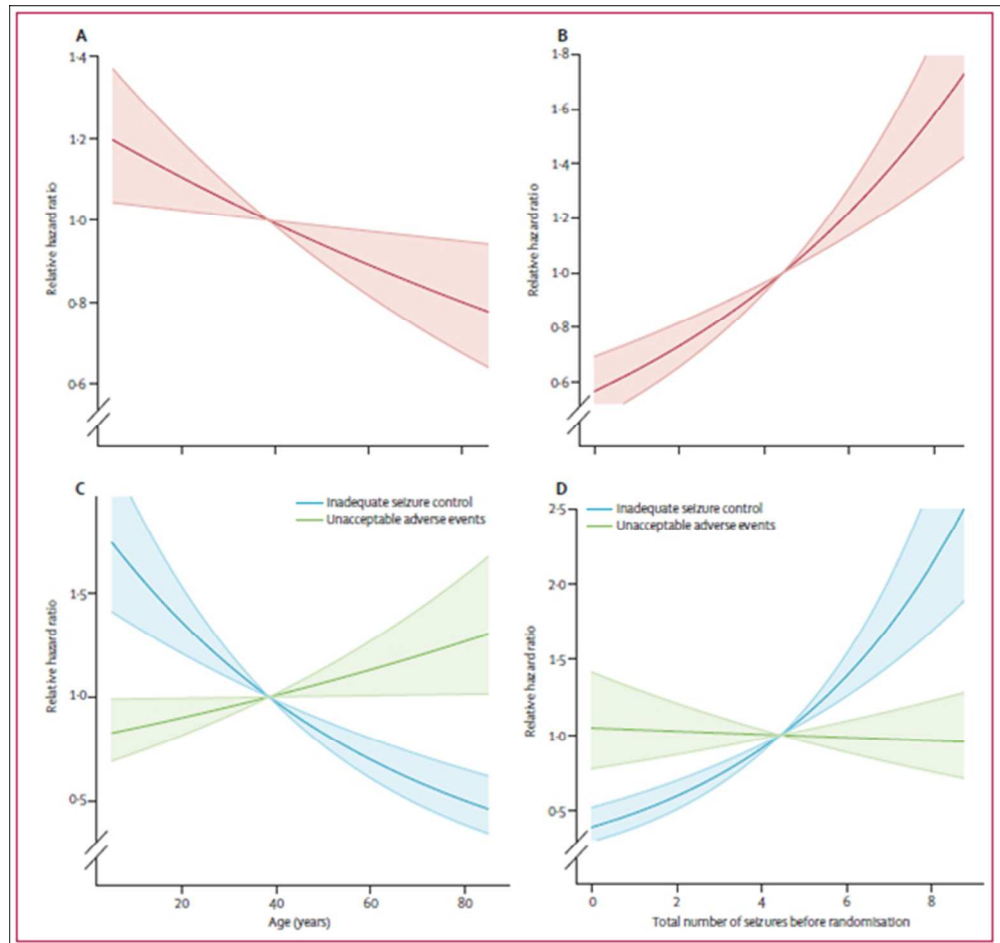


Figure 3: In this example, plots show the relative hazard ratio for age and total number of seizures before randomisation for the time to treatment failure. Hazard ratio estimates with 95% CIs are shown for overall time to treatment failure, for age (A) and total number of seizures (B), and for time to treatment failure because of inadequate seizure control and because of unacceptable adverse events, for age (C) and total number of seizures (D). Ideally, biomarkers should be identified that can be used as predictors of response or failure without the need to measure the reduction in seizure frequency.

228x215mm (73 x 73 DPI)



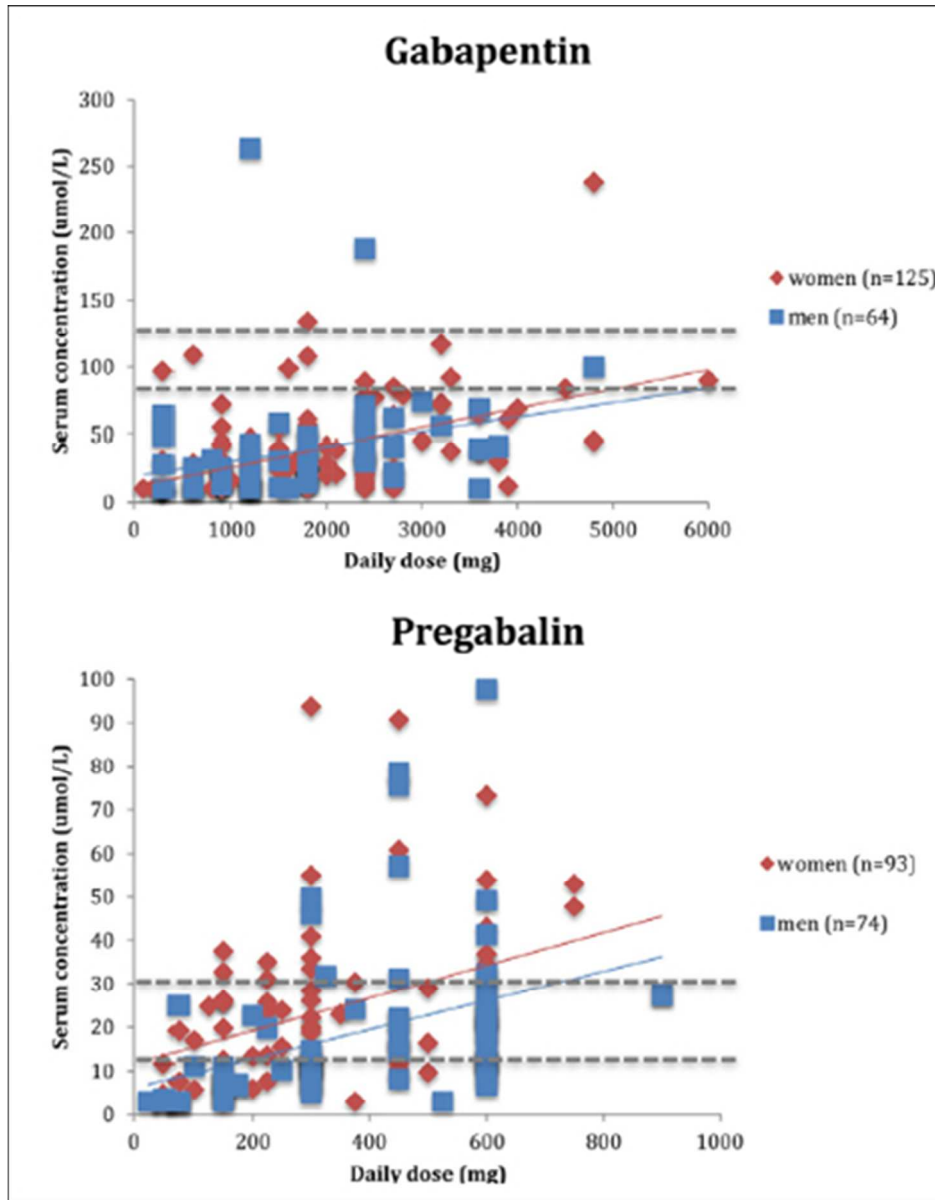


Figure 4. (a) Dose and concentration relationship of (a) gabapentin ( $n = 189$ ), ref. range (70–120  $\mu\text{mol/L}$ ) and (b) pregabalin ( $n = 167$ ), ref. range (10–30  $\mu\text{mol/L}$ ) (with permission from [71]).  
156x200mm (76 x 76 DPI)

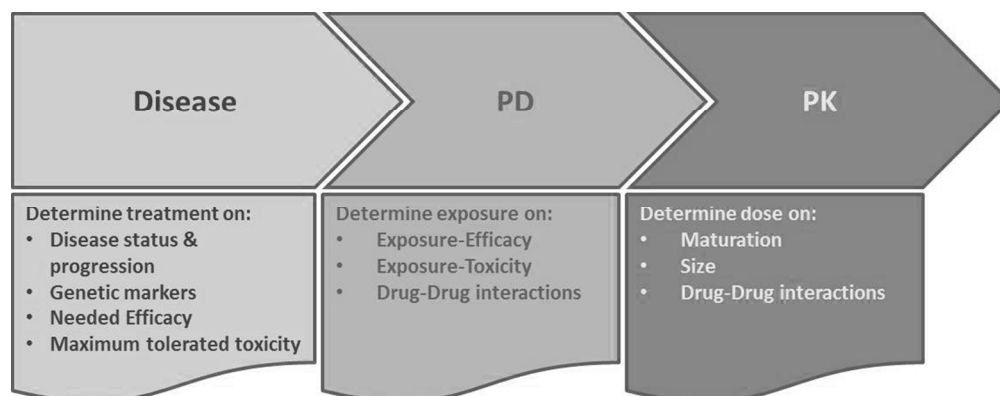


Figure 5. Information on disease processes, pharmacodynamics and pharmacokinetics must be integrated to ensure accurate personalisation of AED treatment and rational dose selection in children. Whereas interindividual differences in disease and pharmacodynamics of AEDs play an important role in treatment selection, understanding of the effect of developmental growth and maturation processes is essential for the selection of the paediatric dosing regimen.

97x38mm (300 x 300 DPI)

Review Only

## Case study: PKPD of topiramate in children – Does dose adjustment help?

### 1. Introduction

Many patients do not respond to first line anti-epileptic drug (AED) treatment and then have to go through cycles of trial and error to find the treatment that works for them. In case the first line treatment fails, a TDM sample can be taken and compared to the therapeutic window of the drug. If the sample AED concentration is below the therapeutic window, the dose is increased, but in case it's within the therapeutic window, the drug is often tapered off and substituted by another one. It is unreasonable to think that efficacy is homogenous across the range of the therapeutic window. Instead, increases in dose can result in improvement in efficacy even when the TDM sample was within the therapeutic window[1]. In fact, for topiramate (TPM), the efficacy has been related to the trough concentrations in a literature PKPD model[2]. In this exercise, we aim to use simulations of topiramate PKPD to investigate how choices in dosing can alter the apparent efficacy of the AED, and determine how model-based treatment choices can improve dosing rationale and subsequent clinical response.

### 2. Methods

#### 2.1. Population PK and PD

The TPM pharmacokinetic/pharmacodynamic (PK/PD) model developed by Girgis *et al.* [2] is one of the few PK/PD models available for AEDs in literature. They describe its PK using a typical two compartment model with first order absorption and elimination (equations 1.1-1.5 & 2.1-2.5, figure 1, table 1), which can be used to predict the concentration over time for each day for each individual, after which the minimum concentration for each day for each individual can be determined (C<sub>min</sub>). This C<sub>min</sub> is then coupled to response (odds of the individual not having had a seizure since inclusion) using a logarithmic function (equation 4) that depends on the baseline number of seizures per 3 months.

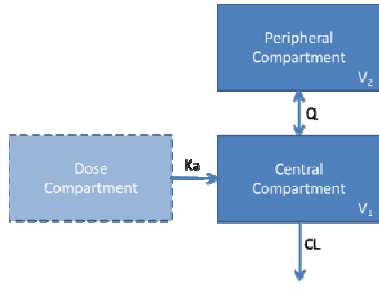


Figure 1. 2 compartmental structure of the Girgis TPM PK model

$$CL = \theta_1 * (1 + ADJ * \theta_2) * \left(\frac{WT}{69.9}\right)^{\theta_3} * e^{\theta_4 * (Age - 31.4)} * \theta_5^{INMD} * \theta_6^{VPA} * \theta_7^{NEMD} * e^{\eta_{CL,i}} \quad (1.1)$$

$$V_1 = \theta_8 * \left(\frac{WT}{69.9}\right)^{\theta_9} * e^{\eta_{V1,i}} \quad (1.2)$$

$$k_a = \theta_{10} * e^{\eta_{ka,i}} \quad (1.3)$$

$$k_{12} = \theta_{11} \quad (1.4)$$

$$k_{21} = \theta_{12} \quad (1.5)$$

Ka: absorption rate constant, CL=clearance, V1: volume of central compartment, K12: rate constant of distribution from central to peripheral compartment, K21: rate constant of distribution from peripheral to central compartment, ADJ=TPM given as adjunctive therapy (1=yes,0=no), WT=weight in kg, Age=age in years, INMD=inducing medication given (such as carbamazepine, phenobarbital or phenytoin, 1=yes, 0=no), VPA=valproate/valproic acid given (1=yes,0=no), NEMD=no effect medication given (such as zonisamide, 1=yes, 0=no).

$$\alpha = \frac{k_{21} \frac{CL}{V_1}}{\beta} \quad (2.1)$$

$$\beta = \frac{1}{2} \left( k_{12} + k_{21} + \frac{CL}{V_1} - \sqrt{\left( k_{12} + k_{21} + \frac{CL}{V_1} \right)^2 - 4k_{21} \frac{CL}{V_1}} \right) \quad (2.2)$$

$$A = \frac{k_a}{V_1} \frac{k_{21} - \alpha}{(k_a - \alpha)(\beta - \alpha)} \quad (2.3)$$

$$B = \frac{k_a}{V_1} \frac{k_{21} - \beta}{(k_a - \beta)(\alpha - \beta)} \quad (2.4)$$

$$C_t = \sum_{i=1}^n D_i \left( A e^{-\alpha(t-t_{D_i})} + B e^{-\beta(t-t_{D_i})} - (A + B) e^{-k_a(t-t_{D_i})} \right) \quad (2.5)$$

i:  $i^{th}$  dose, n=number of doses. t=time,  $t_{D_i}$ : time of dose  $i$ .



Table 1. Parameter values and variance (CV%) of the Girgis TPM PK model

Parameter (unit)	Value	CV%
<b>CL: Clearance (L/h)</b>		
$\theta_1$	1.21	27.28
$\theta_2$	0.479	
$\theta_3$	0.453	
$\theta_4$	-0.00306	
$\theta_5$	1.94	
$\theta_6$	0.686	
$\theta_7$	0.635	
<b>V1: Central volume of distribution (L)</b>		
$\theta_8$	4.61	116.2
$\theta_9$	1.14	
<b>Ka (h)</b>		
$\theta_{10}$	0.105	22.34
<b>K12: Distribution rate constant to the peripheral compartment (/h)</b>		
$\theta_{11}$	0.577	-
<b>K21: Distribution rate constant from the peripheral compartment (/h)</b>		
$\theta_{12}$	0.0586	-

$$\lambda_0 = \theta_{13} \quad (3.1)$$

$$\lambda_t = \theta_{14} \quad (3.2)$$

$$\lambda_{C_{min}} = \theta_{15} \quad (3.3)$$

$$\lambda_{BS3-10} = \theta_{16} \quad (3.4)$$

$$\lambda_{BS10} = \theta_{17} \quad (3.5)$$

$$\log(\lambda_i) = \lambda_0 + \lambda_t * t + \lambda_{C_{min}} * C_{min,i} + \lambda_{BS3-10} * BS_{3-10,i} + \lambda_{BS10} * BS_{10,i} \quad (3.5)$$

$\lambda_i$ : hazard of individual  $i$ ,  $t$ : time (weeks),  $C_{min,i}$ : minimum TPM concentration in individual  $i$ ,  $BS_{3-10,i}$ : baseline between 3-10 seizures per 3 months (1=yes,0=no),  $BS_{10,i}$ : baseline more than 10 seizures per 3 months (1=yes,0=no).

Table 2. Parameter values of the Girgis TPM PD model

Parameter	Value
$\theta_{13} (\lambda_0)$	-3.130
$\theta_{14} (\lambda_t)$	-0.051
$\theta_{15} (\lambda_{C_{min}})$	-0.112
$\theta_{16} (\lambda_{BS3-10})$	1.048
$\theta_{17} (\lambda_{BS10})$	2.411

## 2.2. Exploratory pharmacokinetic simulations

TPM concentrations were simulated for virtual paediatric patients receiving 2.5, 5 or 7.5 mg/kg bi-daily (5, 10, 15 mg/kg/day respectively) of TPM doses for 31 consecutive days, each dose level simulated for 100 patients per dose level. Patients had ages ranging (uniformly sampled) between 4-14 years, and corresponding weights sampled from a normal distribution with mean  $3 \times \text{Age} + 7$  and a coefficient of variance of 25%.

## 2.3. Exploratory pharmacodynamic simulations

Clinical response (number of patients not having any seizures during 1 month) was simulated for virtual paediatric patients with different baseline seizure levels (0-3, 3-10, or more than 10 per month) receiving 2.5, 5 or 7.5 mg/kg bi-daily (5, 10, 15 mg/kg/day respectively) of TPM doses for 31 consecutive days, each dose level simulated for 100 patients per baseline seizure rate level. In total, 9 scenarios were simulated (3 dose levels \* 3 baseline seizure levels), with 100 patients per scenario. Patients had ages ranging (uniformly sampled) between 4-14 years, and corresponding weights sampled from a normal distribution with mean  $3 \times \text{Age} + 7$  and a coefficient of variance of 25%.

## 2.4. Simulation scenarios

Two scenarios were selected, scenario 1 simulating an approximation of current clinical practice, and scenario 2 simulating model-based individualized medicine. One hypothetical population of 1000 typical 4-14 year old patients with corresponding weights (as above) and baseline seizures per 3 months as in Girgis *et al.* were simulated, which were kept identical for both scenarios. The impact of treatment regimens was simulated and at the end of every treatment period of 4 weeks the percentage of treatment success and failure was calculated for each scenario. For the sake of simplicity, up-titration was not simulated, i.e. patients were directly initiated on their target dose.

### 2.4.1. Scenario 1

After treatment with the initial target dose of 5 mg/kg/day for 28 days, a decision was made based on whether the patient had seizures in those 28 days or not. Patients who had a seizure had their dose increased to 10 mg/kg/day. Those receiving 10 mg/kg/day still having had a seizure after another 28 days, had their dose

increased one last time to 15 mg/kg/day. If after another 28 days seizures still had occurred, the treatment was believed to have failed and the patient was switched to another AED.

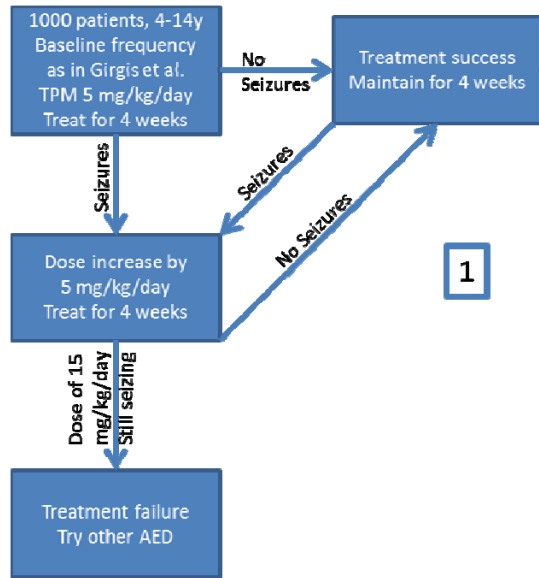


Figure 2. Representation of scenario 1

#### 2.4.2. Scenario 2

In the model-based treatment scenario (Figure 3) information on the relationship between  $C_{min}$ , baseline seizure rate and efficacy is used to determine the treatment plan for each individual. Depending on the baseline of 1-3, 3-10, or >10 seizures per 3 months, patients' target  $C_{min}$ s were set to 10, 12.5, or 15 mg/L respectively. Target doses (mg/dosing interval) were calculated that should result in  $C_{min}$  levels at the target  $C_{min}$  using equation 4. Patients were initiated with those doses for four weeks, after which a trough sample was taken regardless of the occurrence of seizures. If the trough concentration was above the threshold, treatment was maintained. If the trough concentration was below the threshold, the dose was increased using equation 5. After another 4 weeks, the patient was re-evaluated. If seizures had occurred, the treatment was deemed unsuccessful, if not, the treatment was deemed successful.

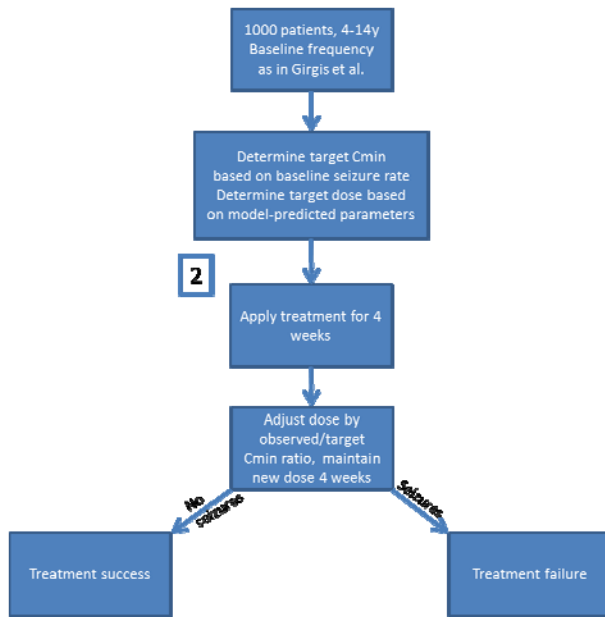


Figure 3. Representation of scenario 2

$$initial\ dose\ \left(\frac{mg}{\tau}\right) = \frac{Target\ Cmin}{\left(\frac{A}{1-e^{-\alpha\tau}} + \frac{B}{1-e^{-\beta\tau}} + \frac{(A+B)}{1-e^{-k_a\tau}}\right)} \quad (4)$$

$$new\ dose\ \left(\frac{mg}{day}\right) = old\ dose\ \left(\frac{mg}{day}\right) * \left(\frac{Target\ Cmin}{Measured\ Cmin}\right) \quad (5)$$

Where  $\tau$  is the dosing interval. A, B, alpha ( $\alpha$ ), beta ( $\beta$ ), and  $K_a$  are here the model-predicted values (not taking into account random variability).

### 3. Results

#### 3.1.1. Exploratory pharmacokinetic simulations

The individual pharmacokinetic profile of a single patient, with corresponding  $C_{min}$  values is plotted in figure 4. From this graph we can see that daily fluctuations of TPM are fairly large, even when dosing bi-daily. As a consequence, attaining adequate  $C_{min}$  levels while ensuring peak levels below toxic levels could be difficult. This is corroborated by figure 5, in which median and 95% prediction intervals of concentrations over time resulting from three dose levels (5, 10 and 15 mg/kg/day) are shown. Here we see that variability in the overall population is large, which points to the need for more individualised dosing.

### 5 mg/kg bid Topiramate

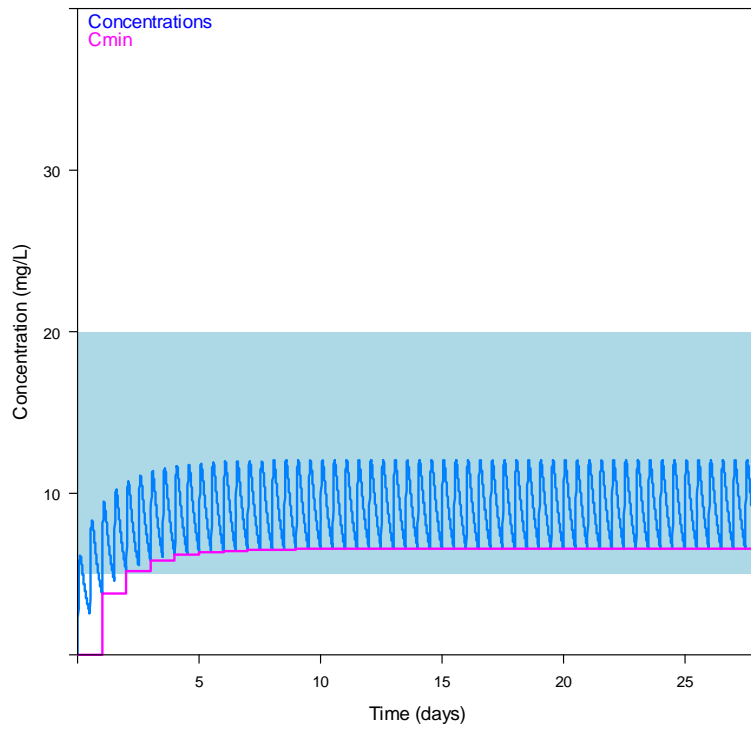


Figure 4. Concentrations over time (blue line) and Cmin of each day (magenta line) for a typical patient after administration of 5 mg/kg twice daily (10 mg/kg/day)

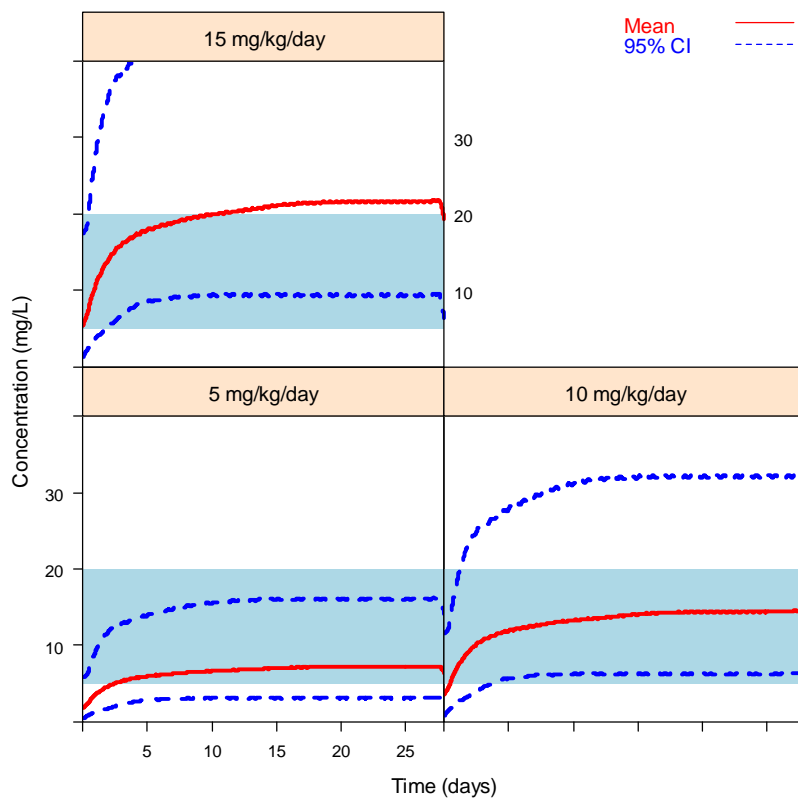


Figure 5. Median and 95% prediction interval of exposures ( $C_{min}$ ) per dose level in the overall population

### 3.1.2. Exploratory pharmacodynamic simulations

Figures 6 and 7 show the median and 95% prediction interval of the seizure free percentage of the simulated population depending on  $C_{min}$  or dose level respectively. It is clear from these graphs that either dose or  $C_{min}$  levels have a profound impact on the percentage of the population having had at least one seizure, and thus on the probability of any such patient in the population having a seizure. It is also evident that the higher the baseline level of seizures, the higher dose or  $C_{min}$  level is required to counteract it. Based on these graphs, we can surmise that target  $C_{min}$  levels should be tailored to the individual patient's baseline instead of treating all patients equally.

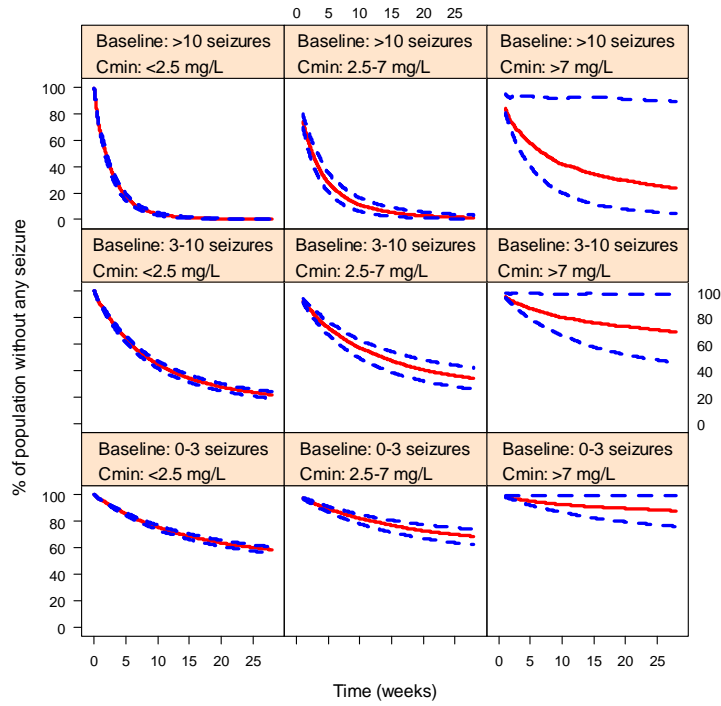


Figure 6. Median and 95% prediction interval of the seizure free percentage of the population over time, per Cmin level

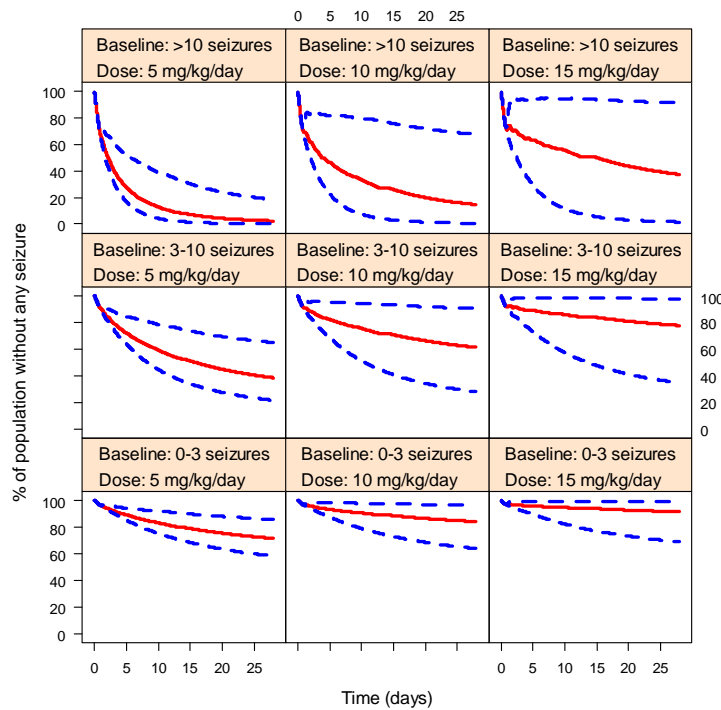


Figure 7. Median and 95% prediction interval of the seizure free percentage of the population over time, per dose level

### 3.2. Simulation scenarios

#### 3.2.1 Clinical scenario 1

Concentrations over time resulting from scenario 1 are shown in figure 8 and give an idea of the change in exposure due to increases in dose for those patients with seizures. Given the relatively low (but clinically typical) starting dose, these patients do not achieve adequate exposure in the first weeks of treatment, and subsequently show that the percentage of the population not having had a seizure drops fairly rapidly (figure 9). The delay in adequate exposure by slow dosing increases results in many patients having to wait for adequate effect, but also may result in patients requesting to have a different drug, as the efficacy is perceived to be inadequate.

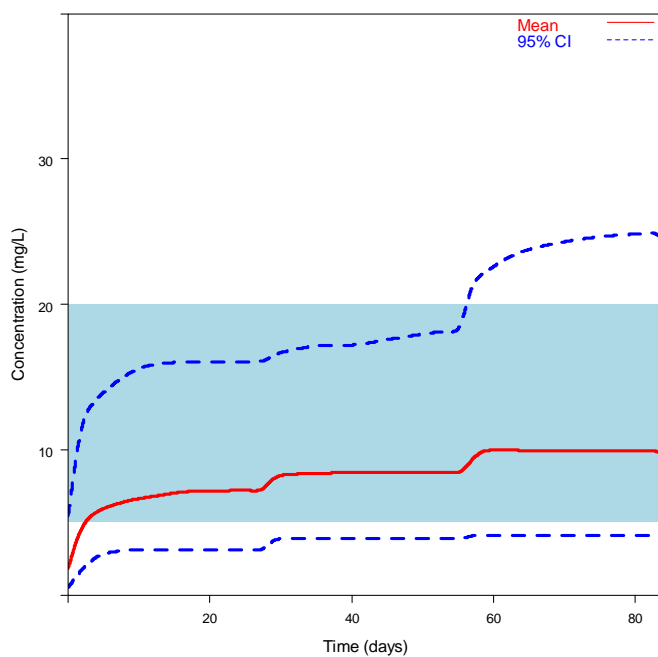


Figure 8. Median and 95% prediction interval of exposures (Cmin) in scenario 1



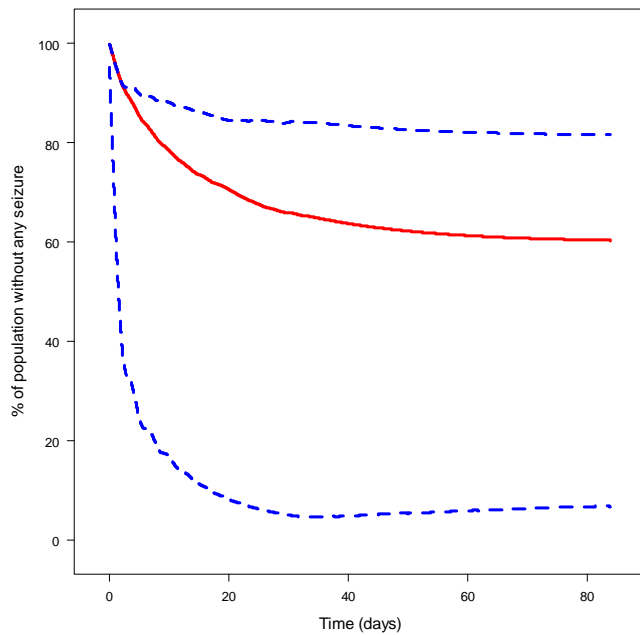


Figure 9. Median and 95% prediction interval of the seizure free percentage of the population over time, for scenario 1

### 3.2.2 Clinical scenario 2

Figure 10 shows the median and 95% prediction interval of concentrations over time resulting from scenario 2. Although initially exposure may be a little more excessive than achieved at the end of scenario 1, the adjustment made after 28 days largely improves on this. Exposure at the end of the scenario is achieved at a higher median level, with a smaller fraction of patients having exposures above the therapeutic window. The higher median concentration levels also translate into efficacy, considering that the percentage of patients not having had a seizure is roughly 75%, while at this same point in time (56 days) this level is roughly 60% in scenario 1.

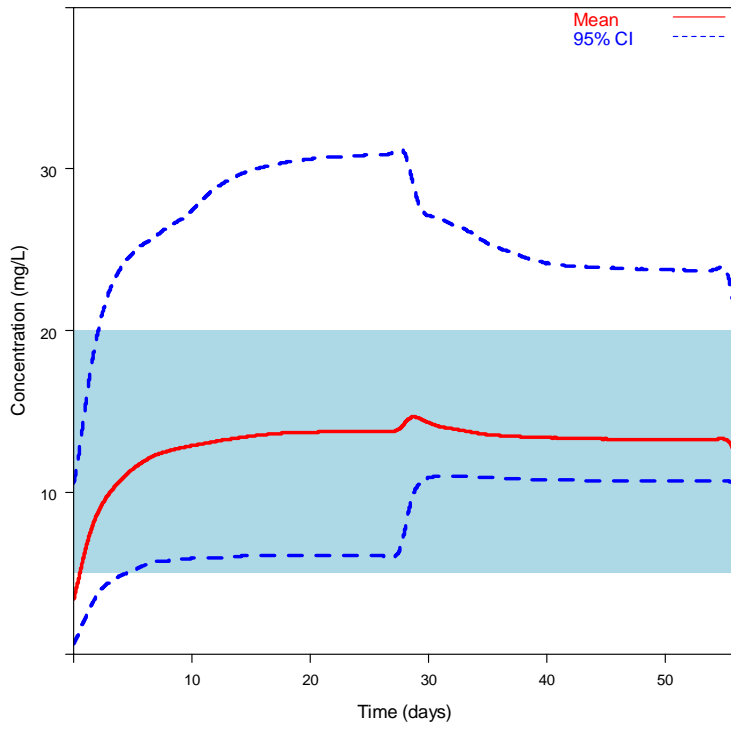


Figure 10. Median and 95% prediction interval of exposures (Cmin) in scenario 2

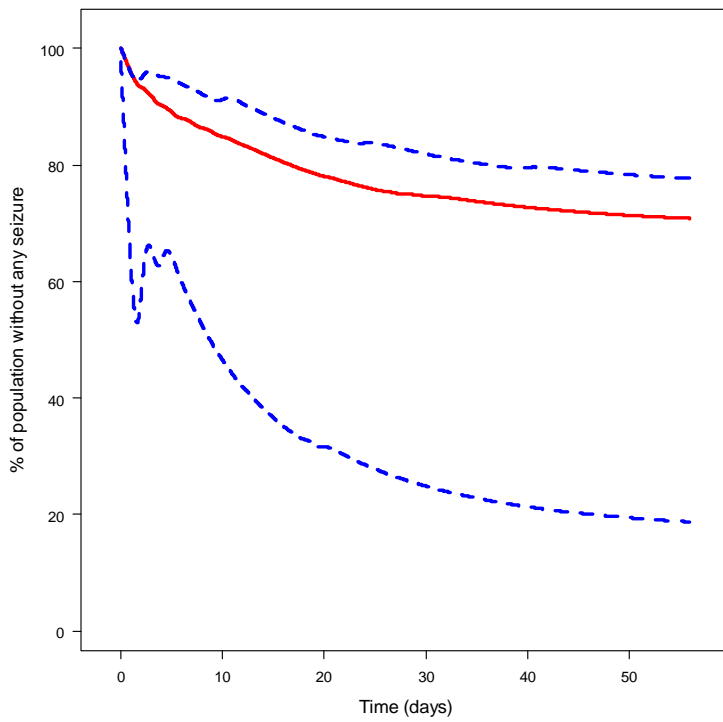


Figure 11. Median and 95% prediction interval of the seizure free percentage of the population over time, for scenario 2

## **Discussion & conclusion**

In this brief exploratory simulation exercise we set out to understand how the implementation of a more rational, model-based dosing guidance would impact clinical practice both from a practical, and an efficacy point of view. These preliminary results show that, assuming all other things being equal, the model-based dosing rationale helps to i. better determine the required level of exposure, ii. achieve efficacious drug levels faster, and iii. take into account inter-individual variability to a significant degree by only requiring one TDM sample. Obviously the results shown here are only based on a simplified view of clinical practice, we dare not hope to simulate the immense variability that occurs in clinical practice to any decent degree in the nearby future. Instead, the current exercise is an example of how clinical practice could be made more rationale, using basic PKPD concepts and possibly PKPD model information. It is our view that such an application could improve response in this highly variable population, expressly because of its variability.

## **References**

1. Kwan P, Brodie MJ. Epilepsy after the first drug fails: substitution or add-on? *Seizure*. 2000 Oct;9(7):464–8.
2. Girgis IG, Nandy P, Nye JS, Ford L, Mohanty S, Wang S, et al. Pharmacokinetic-pharmacodynamic assessment of topiramate dosing regimens for children with epilepsy 2 to 10 years of age. *Epilepsia*. 2010 Oct;51(10):1954–62.