

1 **3D Scanning and 3D Printing as Innovative Technologies for**
2 **Fabricating Personalized Topical Drug Delivery Systems**

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20 **Key words**

21 Three dimensional printing; fused deposition modelling; stereolithography; salicylic acid; hot
22 melt extrusion

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28 **Abstract**

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30 Acne is a multifactorial inflammatory skin disease with high prevalence. In this work, the
31 potential of 3D printing to produce flexible personalized-shape anti-acne drug loaded devices
32 was demonstrated by two different 3D printing technologies: Fused Deposition Modeling (FDM)
33 and stereolithography (SLA). 3D scanning technology was used to obtain a 3D model of a nose
34 adapted to the morphology of an individual. Salicylic acid used in the treatment of acne was
35 selected as a model drug (theoretical drug loading 2%w/w).

36 In FDM 3DP, commercially produced Flex EcoPLA™ (FPLA) and polycaprolactone (PCL)
37 filaments were loaded with salicylic acid by hot melt extrusion (HME). Drug loading in the FPLA-
38 salicylic acid and PCL-salicylic acid filaments after HME was 0.6% w/w and 1.3% w/w
39 respectively (showing significant thermal degradation of drug was observed during 3D printing).
40 Diffusion testing in Franz cells using a synthetic membrane revealed that the drug loaded
41 printed samples released less than 187 $\mu\text{g}/\text{cm}^2$ of their drug content within 3 h. FPLA-salicylic
42 acid filament was successfully printed as a nose-shape mask by FDM 3DP, but the PCL-
43 salicylic acid filament was not.

44 In the SLA printing process, the drug was dissolved in different mixtures of poly(ethylene glycol)
45 diacrylate (PEGDA) and poly(ethylene glycol) (PEG) that were solidified by the action of a laser
46 beam. SLA printing led to 3D printed devices (nose-shape) with higher resolution and higher
47 drug loading (1.9% w/w) than FDM, with no drug degradation. The results of drug diffusion tests
48 revealed that drug release was faster than with the FDM devices, 229 and 291 $\mu\text{g}/\text{cm}^2$ within 3 h
49 for the two formulations evaluated.

50 In this study, SLA printing was a more appropriate 3D printing technology to manufacture anti-
51 acne devices with salicylic acid. The combination of 3D scanning and 3D printing has the
52 potential to offer solutions to produce personalised drug loaded devices, adapted in shape and
53 size to individual patients.

54

55

56 **1. Introduction**

57 A large proportion of the population is affected by acne vulgaris (acne), particularly
58 postpubescent teenagers [1, 2]. Acne is a multifactorial chronic inflammatory skin disease
59 commonly found on the face. It originates in the pilosebaceous units and is classified based on
60 the level and severity of the inflammation as open comedones, closed comedones, papules,
61 pustules or nodules. Two of the factors that lead to acne formation are perifollicular
62 hyperkeratinization and follicular obstruction. Androgens, lipids, specific bacteria and cytokines
63 induce hyperkeratinization and hyperproliferation of keratinocytes promoting follicular
64 obstruction and formation of microcomedones [1, 3]. The disorder does not only affect patients
65 physically but also psychologically, leading in some cases to suicide attempts [1, 4].

66
67 Most of the topical treatments, generally used for mild to moderate acne, aim to eradicate the
68 pathogenic factors [1, 4]. Salicylic acid is one of the most widely used anti-acne agents; it is a
69 lipophilic β -hydroxyl acid that acts as an anti-inflammatory and exfoliating agent. It penetrates
70 through the skin and detaches corneocytes from each other, weakening the intracellular
71 cement. As a result, the skin cell turnover increases, removing the comedones. Additionally, it
72 improves the elasticity of the stratum corneum and stimulates the production of new
73 corneocytes and collagen [5]. Salicylic acid is found in topical formulations such as creams,
74 gels, cleansers and soap bars with concentrations ranging from 1 to 2% [4]. The commercial
75 treatments containing salicylic acid have high efficacy but they sometimes cause mild and
76 transient side effects (erythema, dryness, intense exfoliation and crusting) which are dependent
77 on dose. The absorption of salicylic acid via topical treatment is enhanced when the formulation
78 comprises a hydrophilic base or is kept occluded.

79
80 Three-dimensional printing (3DP) is an additive manufacturing process that allows the
81 fabrication of three-dimensional solid objects of virtually any shape from a 3D model file. The 3D
82 models can be generated by computer aided design (CAD) software or obtained from 3D
83 scanners that capture images and distance information of real objects and then transfer the data
84 to a computer. The implementation of 3D printing technologies has been increasingly growing in
85 many fields. In the pharmaceutical field, 3DP has been used for the production of personalised
86 medicines, oral dosage forms, medical devices, and for tissue engineering [6].

87
88 Of the several 3D printing technologies commercially available, fused deposition modelling
89 (FDM) is perhaps the most widely used in pharmaceuticals. FDM is simple and cost effective and

90 has been shown extremely versatile in the development of drug delivery systems [7], especially
91 personalised medicines [8], and medical devices [9, 10]. In FDM an extruded polymer filament is
92 passed through a heated nozzle that softens the polymer and it is then deposited on a build
93 plate, creating one layer of the object to be printed. The build plate then lowers vertically and
94 another layer is deposited. The object is fabricated by repeating these steps in a layer-by-layer
95 manner. The main polymers used in FDM are PLA (polylactic acid or polylactide) and ABS
96 (acrylonitrile butadiene styrene), although an increasing number of polymers is becoming
97 commercially available.

98

99 Polycaprolactone (PCL) is a biocompatible polyester with many applications that has been used
100 in wound dressings, tissue engineering and drug delivery, leading to a several PCL drug-
101 delivery devices being approved by the FDA [11-13]. The use of flexible polymers would allow
102 the manufacture of more comfortable devices that are, at the same time, robust to handle.
103 NinjaFlex[®] (NF) and Flex EcoPLA[™] (FPLA) are some of the most widely used flexible filaments.
104 NF is a thermoplastic polyurethane, which is a biomaterial that due to its biocompatibility and
105 mechanical properties is currently used broadly for regeneration, bone replacement and
106 drug/gene delivery [14]. FPLA is a flexible variety of PLA, which is an aliphatic polyester that is
107 degradable in the human body and in the environment, with appropriate mechanical strength
108 and low toxicity [13].

109

110 An alternative 3D printing technology that is becoming more affordable is stereolithography
111 (SLA). In this technology, the production is based on the solidification of a liquid resin by
112 photopolymerization. A laser beam causes localized polymerization (solidification) of
113 photocrosslinkable polymers to form a solid layer and the process is repeated in a layer-by-layer
114 manner until the solid 3D object is produced. This technology has been used in the fabrication
115 of oral tablets [15], for tissue engineering [16, 17] and shows higher resolution than the FDM
116 technology [18]. Over the past few years a number of photocrosslinkable polymers have been
117 developed, such as poly(ethylene glycol) diacrylate (PEGDA) [19, 20].

118

119 The aim of this work was to evaluate the feasibility of printing anti-acne patches/masks
120 personalised to the anatomy of the patient by 3D scanning and 3D printing. Two different 3D
121 printing technologies - FDM and SLA - were evaluated in terms of manufacture capability,
122 morphological characteristics of the printed object, drug stability while printing and drug release.

123

124 **2. Materials and Methods**

125 **Materials**

126 NF (NinjaFlex[®] filament, thermoplastic polyurethane, printing temperature 220–230°C, batch no:
127 3D3071175) and FPLA (Flex EcoPLA[™] BLUE 45D filament, flexible polylactic acid, printing
128 temperature 210°C, number: 001) were purchased from iMakr, UK. PCL (Polycaprolactone, 6-
129 caprolactone polymer, (C₆H₁₀O₂)_n, MW: 80,000 Daltons, lot no: MKBR4733V), PEGDA
130 (poly(ethylene glycol) diacrylate,, MW: 700 Daltons), PEG (poly(ethylene glycol) 300, MW: 300
131 Daltons) and diphenyl(2,4,6-trimethylbenzoyl)phosphine oxide were purchased from Sigma
132 Aldrich, UK. Salicylic acid (lot no: 14G020009) was purchased from VWR International, UK.
133 Tetrahydrofuran (THF, HPLC grade) was supplied by Fisher Scientific, UK; dichloromethane
134 (DCM, >99.5% purity) was supplied by VWR International, UK; methanol (≥ 99.9% purity, HPLC
135 grade) was supplied by Sigma Aldrich, UK. The salts for preparing the buffer dissolution media
136 were purchased from VWR International Ltd., Poole, UK.

137

138 **Methods**

139 *2.1 3D scanning*

140 3D scanning of the face of a volunteer was performed with a commercial scanner (Sense[™] 3D
141 scanner, 3D Systems Inc., USA) at a distance of 40 cm in accordance with the manufacturer's
142 instructions. The 3D scanner was panned 360° around the head of the subject to capture a 3D
143 image that was exported to Meshmixer (v.10.9.332, Autodesk Inc., USA) to extract the final 3D
144 template for 3DP. A nose-shaped mask (41.2 mm length x 34.5 mm width x 22.7 mm height)
145 adapted to the physical characteristics of an individual was used as 3D model in order to the
146 intricate shape of the design allowed the evaluation of the resolution of the different 3D printing
147 technologies.

148

149 *2.2 Fused deposition modelling*

150 *2.2.1 Preparation of the drug loaded polymers for HME by solvent casting*

151 Before the HME process, polymer (29.4 g) - NF, FPLA or PCL - was dissolved in organic
152 solvent (200 mL) with salicylic acid (0.6 g) using an overhead stirrer. For NF and PCL, THF was
153 used as the solvent while DCM was used for FPLA. The solution obtained after overnight stirring
154 was transferred to Teflon Petri dishes and maintained in a fume cupboard until evaporation of
155 the solvent was accomplished (room temperature, for 2 days). The casted films obtained were
156 stored in an oven (40°C, for 3 days) to completely remove the solvent and, finally, cut into small

157 pieces to make them suitable for extrusion. The theoretical loading of salicylic acid in the
158 polymer was 2% w/w. Blank filaments were prepared following the same process without
159 addition of the drug.

160

161 2.2.2 Hot melt extrusion (HME)

162 The previously prepared pieces of polymer-salicylic acid were loaded into a single-screw
163 filament extruder (Filabot®, USA) to obtain 1.75 ±0.1 mm diameter polymer filaments for 3D
164 printing. The extruding temperatures were 170°C for NF- salicylic acid, 190°C for FPLA-salicylic
165 acid and 60°C for PCL-salicylic acid. The diameter of the filament was randomly measured with
166 a ProMax Electronic Calliper (Fowler High Precision, USA) at different positions along the
167 filament. Drug loading of the filaments was determined by HPLC analysis (see below).

168

169 2.2.3 FDM 3D printing

170 Devices were fabricated from the drug-loaded filaments using a commercial fused-deposition
171 modelling 3D printer, MakerBot Replicator 2X (MakerBot Inc, USA). The templates used to print
172 the devices obtained by 3D scanning were modified with AutoCAD 2014® (Autodesk Inc., USA)
173 and exported as a stereolithography file (.stl) into the 3D printer software (MakerWare v. 2.2.2,
174 MakerBot Inc., USA). The .stl format encodes only the surface data of the object to be printed
175 and requires the thickness of the surface, the infill and the temperature to be defined in order to
176 print the desired object. The printing parameter settings are shown in **Table 1**.

177

Table 1

FDM printing settings

Parameters	Polymer	
	FPLA	PCL
Printing temperature (°C)	230	170
Infill (%)	100	100
Number of shell	2	2
Layer height (mm)	0.2	0.1
Speed while extruding (mm/s)	90	50
Speed while travelling (mm/s)	150	50
Raft	No	No
Support	No	No

178

179 2.3 Stereolithography (SLA) 3D printing

180 Two different formulations were printed by SLA 3D Printing: PEGDA/PEG (4:6) and
181 PEGDA/PEG (8:2). To do so, two photopolymer solutions were prepared mixing PEGDA and
182 PEG 300 in different ratios to make a total volume of 40 mL. The photoinitiator, diphenyl(2,4,6-
183 trimethylbenzoyl) phosphine oxide was then added to the mixture solution to a concentration of
184 1% w/v. Finally, salicylic acid was added into the solution to a concentration of 2% w/w.

185 Devices were fabricated from the drug-loaded solutions using a commercial SLA 3D printer
186 (Form 1+ Stereolithography 3D printer, Formlabs, UK) equipped with a 405nm laser. The
187 templates used to print the devices were the same as for FDM. They were exported as a
188 stereolithography file (.stl) into the 3D printer software (Preform Software v. 1.9.1, Formlabs,
189 UK). In the settings of the 3D printer, the layer thickness was 0.1mm and the material selection
190 was flexible.

191 2.4 Characterisation of the filaments and 3D printed objects

192

193 2.4.1 Mechanical evaluation of the filaments

194 An Instron[®] 5900 Series (Instron, UK) equipped with a 100 N load cell was utilized to carry out
195 tensile tests of standardized filaments (in terms of diameter and length). The equipment was
196 controlled by BlueHill software selecting the default tensile test method. Prior to the test, the
197 system was calibrated for balance length of extension (mm) and load (N).

198 The filaments were fixed vertically to the clamps of the equipment and stretched gradually
199 during the test until a rupture point or irregular deformation was observed (n=4). The distance
200 between the two clamps was adjusted to 30mm and the diameter and length of tested filament
201 was introduced in the software for the calculation of Young's modulus from the strain-stress
202 curve automatically plotted by BlueHill software. Young's modulus is calculated in the linear
203 region as stress divided by strain.

204

205 2.4.2 Thermal analysis

206 DSC measurements were performed with a Q2000 DSC (TA instruments, Waters, LLC, USA) at
207 a heating rate of 10°C/min. Calibration for cell constant and enthalpy was performed with indium
208 ($T_m = 156.6^\circ\text{C}$, $\Delta H_f = 28.71 \text{ J/g}$) according to the manufacturer's instructions. Nitrogen was used
209 as a purge gas with a flow rate of 50 mL/min for all the experiments. Data were collected with

210 TA Advantage software for Q series (version 2.8.394), and analysed using TA Instruments
211 Universal Analysis 2000. All melting temperatures are reported as extrapolated onset unless
212 otherwise stated. TA aluminium pans and pin-holed hermetic lids (T_{zero}) were used with an
213 average sample mass of 8-10 mg.

214
215 For TGA analysis, samples (average weight: 3-5 mg) were heated at 10°C/min in open
216 aluminium pans with a Discovery TGA (TA instruments, Waters, LLC, USA). Nitrogen was used
217 as a purge gas with a flow rate of 25 mL/min. Data collection and analysis were performed using
218 TA Instruments Trios software and % mass loss and/or onset temperature were calculated.

219
220 *2.4.3 X-ray powder diffraction (XRPD)*
221 Discs (23.78 mm diameter x 1.00 mm height) made from pure polymers or drug-loaded
222 polymers were printed and analysed. A sample of pure salicylic acid was also analysed. The X-
223 ray powder diffraction patterns were obtained in a Rigaku MiniFlex 600 (Rigaku, USA) using a
224 Cu K α X-ray source ($\lambda=1.5418\text{\AA}$). The intensity and voltage applied were 15 mA and 40 kV. The
225 angular range of data acquisition was 3–60° 2 θ , with a stepwise size of 0.02° at a speed of
226 5°/min.

227
228 *2.4.4 Scanning Electron Microscopy (SEM)*
229 The Surface and cross-section images of the filaments and the printed devices were captured
230 with an FEI Quanta 200F Scanning Electron Microscope (FEI, UK). The voltage and working
231 distance were set at 5 V and 50 mm, respectively. Filament samples for SEM imaging were
232 previously coated with gold. Pictures of the 3D printed devices were taken with a Nikon
233 CoolpixS6150 with the macro option of the menu.

234
235 *2.4.5 Determination of the drug content*
236 For filaments and FDM printed devices, a section of the drug-loaded filament and the 3D printed
237 objects (approx. 0.50 g) was placed in a volumetric flask (25mL) with the appropriate organic
238 solvent (THF was used for NF and PCL while DCM was used for FPLA) under magnetic stirring
239 until complete dissolution. Aliquots (10 mL) were transferred to a volumetric flask with methanol
240 (40 mL) to precipitate the polymer. The solution was then filtered through a 0.22 μm filter
241 (Millipore Ltd., Ireland) and the concentration of drug in the filtrate was determined by HPLC.

242

243 For SLA printed devices, a section of the drug-loaded 3D printed mask (approx. 0.50 g) was
244 milled with mortar and pestles and placed into a volumetric flask with methanol (25mL) under
245 magnetic stirring until complete dissolution of the drug. The solution was then filtered through a
246 0.22 µm filter (Millipore Ltd., Ireland) and the concentration of drug in the filtrate was determined
247 by HPLC (see below).

248
249 The HPLC (Hewlett Packard 1050 Series HPLC system, Agilent Technologies, UK) assay
250 entailed injecting samples (20.0 µL) into a mobile phase consisting of methanol (70%) and 0.1%
251 trifluoroacetic acid (TFA) in water (30%), through a reverse phase column (Ascentis® C18, 5 µm
252 particle size, 4.6 × 150 mm) connected with a pre-column all maintained at 25°C. The mobile
253 phase was pumped at a flow rate of 1.0 mL/min and the eluent was screened at the wavelength
254 of 234 nm.

255

256 *2.4.6 Diffusion studies*

257

258 Drug diffusion experiments from circular-shaped 3D printed devices (16 mm diameter and 1 mm
259 thickness) were conducted in vertical glass Franz cells with an effective diffusion area of $98.5 \pm$
260 4.8 mm^2 and a receptor volume of 4.6 mL (n=3).

261

262 The 3D printed patches were mounted between the donor and receptor compartments,
263 separated from the receptor by a cellulose nitrate membrane (pore size 0.45µm, cat no. 7184-
264 002, Whatman, UK) previously soaked in receptor fluid for at least 12 hours before starting the
265 test was started. The receptor compartment of the diffusion cell was filled with phosphate-
266 buffered saline (Dulbecco A, Thermo Scientific, UK) at pH 7.3

267

268 The whole assembly was incubated at 32°C in a water bath in order to mimic the skin
269 temperature and the solution in the receptor compartment was constantly stirred at 400 rpm
270 using a magnetic stirrer. Donor (with no solution) and receptor compartments were occluded
271 with Parafilm to prevent evaporation.

272

273 At different intervals, 200 µL aliquots were withdrawn from each cell and replaced with an equal
274 amount of phosphate-buffered saline. The drug concentration was then determined by HPLC.
275 The cumulative percentages of drug permeated per square centimetre from the 3D printed
276 patches were plotted against time.

277 **3. Results and discussion**

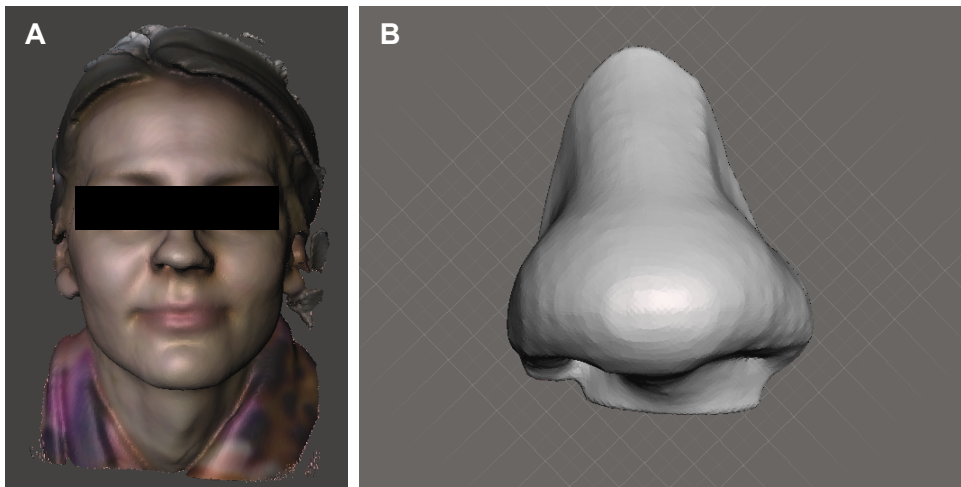
278 **3D scanning**

279 The 3D scanning process performed with the commercial Sense™ 3D scanner showed good
280 resolution (according to the manufacturer, the point-to-point spacing is around 0.65mm),
281 capturing the object with the real size without the need of calibration (**Fig. 1A**). The quality of the
282 scanning process is dependent on the experience of the operator and the light conditions, but
283 the system is easy to operate with basic training. The scanning process can be performed
284 without contact in a few seconds, so it could be useful for people who have difficulties remaining
285 still and as a fast/routine technique.

286

287 A section of the model obtained from the 3D scan (the nose of the volunteer) was selected as a
288 model for 3D printing due to the prevalence of acne localized in that region and the challenge of
289 printing intricate shapes (**Fig. 1B**). The use of CAD software (Meshmixer) easily allowed the
290 selection of specific parts to be printed. In this case, the internal part of the nose design was
291 designed hollow, to fit perfectly on the nose of the individual.

292



293

294 **Fig. 1.** A) Volunteer scanning image and B) 3D model nose used for 3DP.

295

296 **Fused Deposition Modelling (FDM) 3DP**

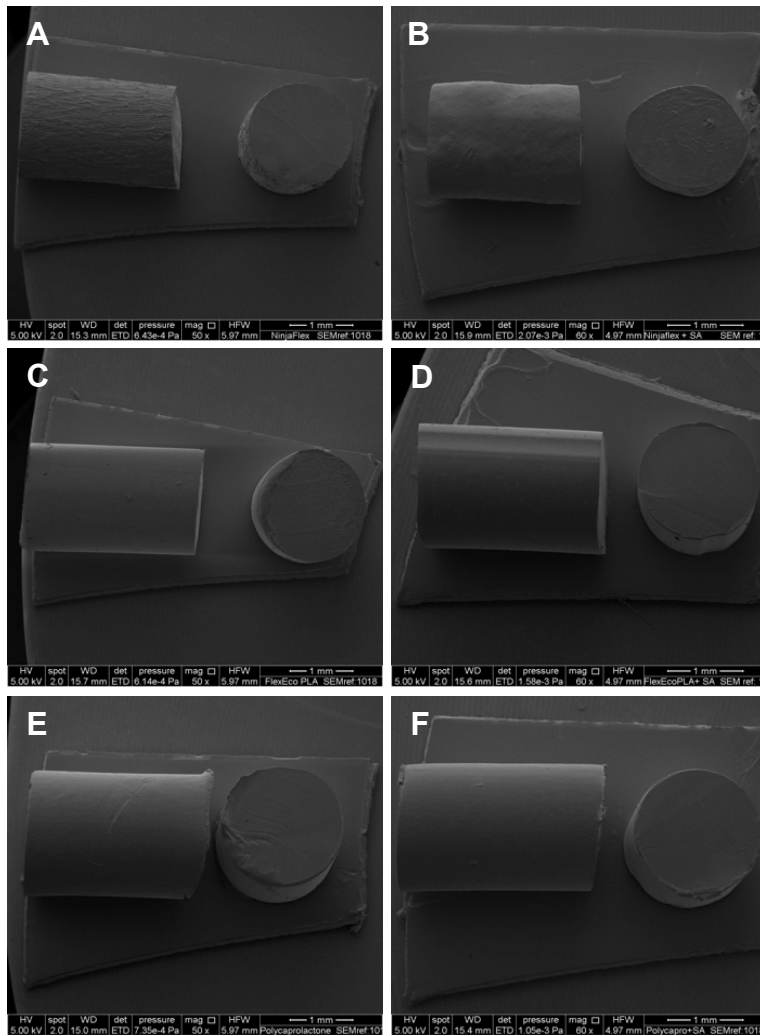
297 It was possible to produce drug-loaded filaments by HME incorporating salicylic acid into the
298 polymers; however, for the commercial filaments the characteristic of the filaments changed
299 considerably by inclusion of the drug and were significantly different from the commercial
300 filament in terms of size, physical appearance and mechanical behaviour.

301
302 The NF-salicylic acid filament obtained from HME became red-brown with a rough surface and
303 brittle after being extruded. It was so brittle that it was not possible to hold it in the Instron
304 equipment to determine Young's modulus. The SEM images show that the NF-salicylic acid
305 filament had an irregular surface and inconsistent morphology (cross section not circular)
306 compared with commercial NF (**Fig. 2A and 2B**). According to the manufacturer's material
307 safety data sheet (MSDS), NF is made of polyurethanes and its properties are highly affected by
308 solvents and acids [21]. It has also been reported that salicylic acid is incompatible with
309 polyurethanes, affecting physical properties of the polymer, such as tensile strength, hardness
310 and elongation [22]. Consequently, the NF-salicylic acid filament was significantly different from
311 the NF filament in terms of flexibility and colour, which made the filament not suitable for 3D
312 printing.

313 The FPLA-salicylic acid filament (1.67 ± 0.16 mm diameter) showed a surface as smooth as the
314 commercial FPLA filament (1.75 ± 0.02 mm diameter) (**Fig. 2C and 2D**). The colour of the FPLA-
315 salicylic acid filament was slightly darker compared with the commercial FPLA, however the
316 flexibility of the filaments was comparable. Young's modulus of FPLA-salicylic acid and
317 commercial FPLA filament was 93.53 ± 4.34 MPa and 93.17 ± 3.72 MPa respectively.

318
319 The PCL-salicylic acid filament was white, smooth and with good morphology (uniform diameter,
320 1.65 ± 0.11 mm), comparable to that of the PCL filament obtained from PCL pellets without drug
321 by HME (**Fig. 2E and F**). Young's modulus of PCL-salicylic acid (286.43 ± 25.23 MPa) and not
322 drug-loaded PCL filament (280.39 ± 25.92 MPa) are similar, but stiffer than FPLA filaments
323 (Young's modulus ~ 93 MPa).

324



325

326

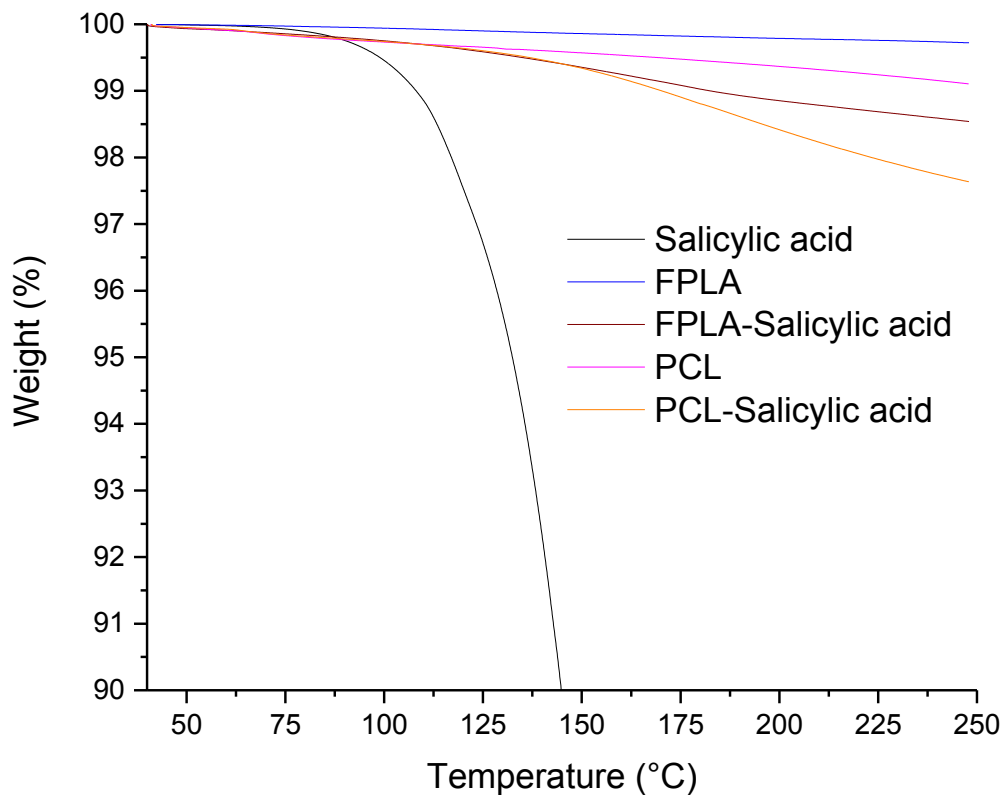
327 **Fig. 2.** SEM images of A) commercial NF filament, B) the NF-salicylic acid filament, C)
 328 commercial FPLA filament, D) FPLA-salicylic acid filament, E) plain PCL filament prepared from
 329 PCL pellets and F) PCL-salicylic acid filament.

330

331 The drug loading of the FPLA-salicylic acid filament was $0.63 \pm 0.10\%$ w/w. It is evident that the
 332 extrusion process causes degradation of salicylic acid (theoretical drug loading 2% w/w), which
 333 was a result of the high extruding temperature (190°C). The drug loading in the PCL-salicylic
 334 acid filament was $1.34 \pm 0.01\%$ w/w. The amount of salicylic acid in the PCL filament is higher
 335 than that in the FPLA filament because the extrusion temperature is much lower (60°C), so the
 336 salicylic acid remains more stable during extrusion, but this still represents a significant degree
 337 of degradation.

338

339 TGA data show also signs of drug degradation (**Fig. 3**). The amount of salicylic acid at different
340 temperatures compared with the starting amount in terms of percentage implies the degradation
341 temperature range of the tested substance. The percentage of weight of salicylic acid is greatly
342 reduced at the temperature above 140°C and salicylic acid completely degraded at the
343 temperature about 200°C (**Fig. 3**).
344



345
346 **Fig. 2.** TGA results for Salicylic acid (SA), the extruded filaments (blank) and the drug loaded
347 filaments.

348 The weight loss percentage for the FPLA-salicylic acid filament was higher than non-drug
349 loaded filament by approximately 1.5%, which corresponded to salicylic acid degradation and
350 reduced heat stability of the polymer due to hydrolysis. It was assumed that salicylic acid
351 hydrolysed ester bonds of the polymer, resulting in decreased strength and heat stability of the
352 polymer [23]. However, the stability of the FPLA-salicylic acid filament was acceptable at the
353 printing temperature since the weight loss percentage was not significant.

354

355 The mass of PCL-salicylic acid filament decreased more than the non-drug loaded PCL
356 filaments at temperatures above 200°C. This may be because the incorporation of drug into
357 PCL affects the properties of polymer, reducing heat stability.

358

359 For the 3D printing process, sections of filaments with the diameter closer to the optimum for the
360 FDM printer (1.75mm) were selected. The FPLA-salicylic acid filament was successfully printed
361 as a flat circular patch. The quality of printing was considered good, as the shape was well
362 defined and there were no printing inconsistencies. Pure PCL and the PCL-salicylic acid
363 filaments were also successfully 3D printed as circular patches. As mentioned previously, the
364 NF-salicylic acid filament was not printable due to the lack of flexibility (and so NF-salicylic acid
365 filament was not further evaluated in this study).

366

367 The evaluation of the drug loading from 3D printed devices showed lower values to those of the
368 drug-loaded filaments used for printing, indicative of degradation during the 3DP process. The
369 mean amount of salicylic acid content in the FPLA-salicylic acid printed circle was $0.35 \pm 0.01\%$
370 w/w. This shows that the percentage of salicylic acid in the 3D printed device decreases
371 compared with the initial percentage of drug in the filament because of the printing process,
372 which was performed at 230°C. The mean amount of salicylic acid content in the PCL-salicylic
373 acid printed patch was $1.21 \pm 0.02\%$ w/w, which indicates that some salicylic acid in the filament
374 decomposed while 3D printing at 170°C. The decomposition of the drug while printing, due to
375 the heat involved process, has been noted previously [24].

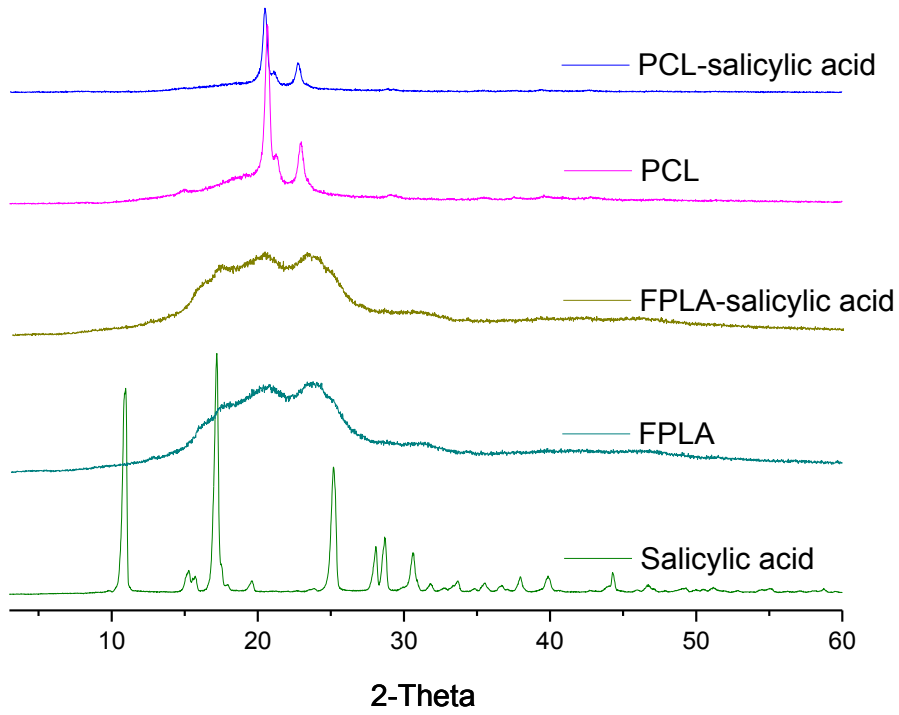
376

377 The XRD data reveals that FPLA is semi-crystalline because crystalline peaks and an
378 amorphous halo are noticeable (**Fig. 4**). However, the XRD data of the FPLA-salicylic acid could
379 not confirm the physical form of salicylic acid in the formulation. There are not peaks similar to
380 those in the XRD pattern of pure salicylic acid powder, which indicates that the drug is either
381 dissolved in the polymers or it is not detected due to the low drug loading percentage.

382

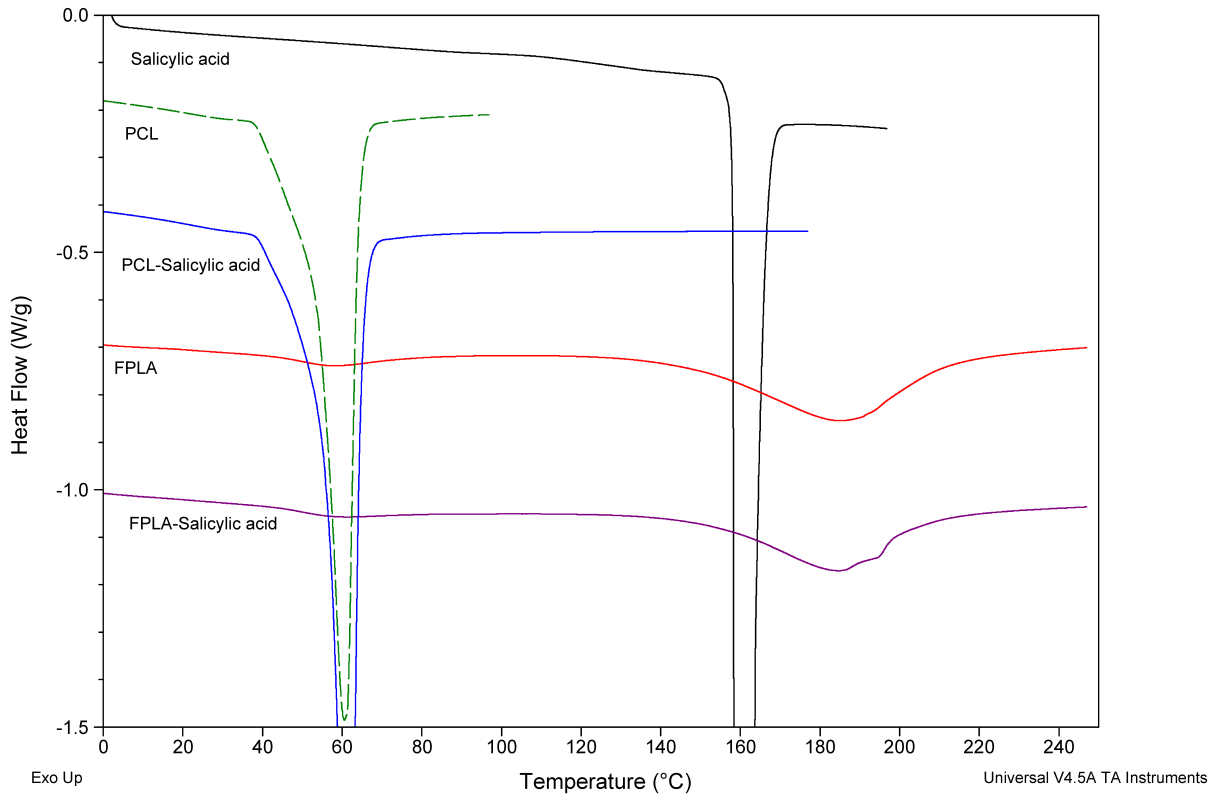
383 PCL is also in a semi-crystalline form as the XRD pattern consists of both crystalline peaks and
384 an amorphous halo (**Fig. 4**). No evidence for salicylic acid as a crystalline phase was seen in
385 the PCL-salicylic acid XRD data.

386



387

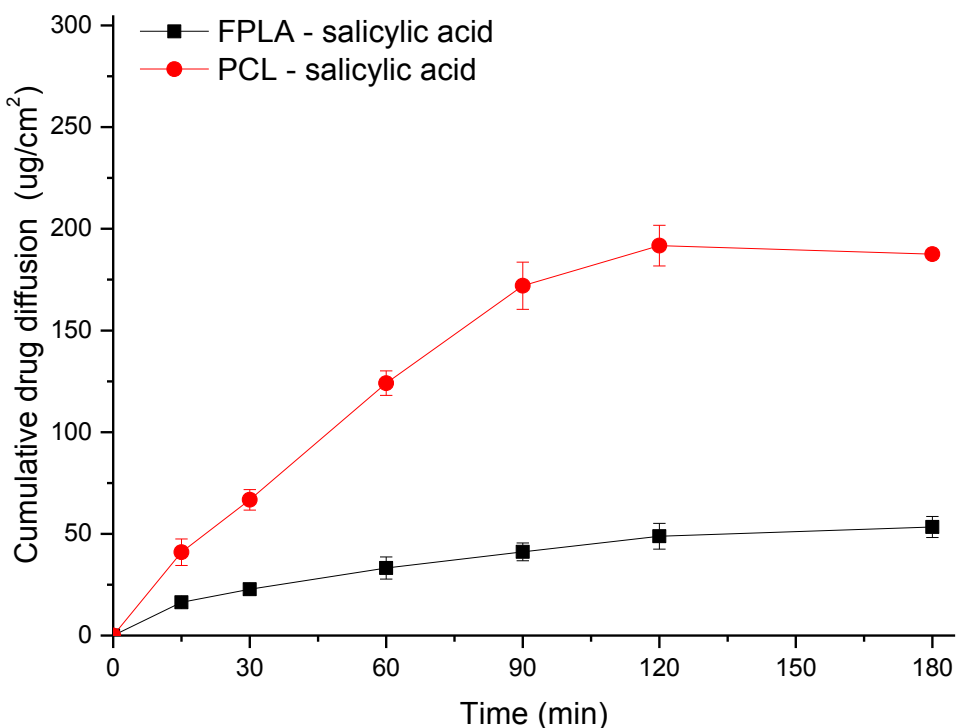
388 **Fig. 4.** X-ray powder diffraction patterns for salicylic acid and various FDM printed discs



389

390 **Fig. 5.** DSC thermal traces for salicylic acid and various FDM printed discs

391
392 The DSC thermograms for salicylic acid show a sharp endothermic peak (melting point, T_m) at
393 160°C (**Fig. 5**). This implies that salicylic acid powder was in the crystalline form. The
394 comparable DSC profiles of FPLA and FPLA-salicylic acid discs are shown in **Fig. 5**. A salicylic
395 acid melt is not seen in the FPLA-salicylic acid disc. Although the T_m of FPLA patch (187°C) is
396 slightly higher compared with the two endothermic peaks at 183°C and 202°C that became a
397 combined peak with T_m of 185°C of FPLA-SA, the overall properties of FPLA are not significantly
398 affected by the presence of SA. For the PCL, PCL-salicylic acid disc does not show the melting
399 peak of the SA, and PCL printed disk shows also similar T_m (60°C) to the T_m of the PCL-salicylic
400 acid disc (61°C).



401
402
403 **Fig. 6.** Cumulative amounts of salicylic acid permeated from the FDM 3D printed devices.
404
405 Drug permeation from the patches occurred slowly (**Fig. 6**), as expected for a topical drug
406 delivery device (even though the synthetic membrane used is considered as a high flux material
407 [25]).

408

409 The PCL-salicylic acid printed sample showed a higher rate of drug diffusion than the FPLA-
410 salicylic acid. The release profiles show that the cumulative percentage of drug diffused from
411 the printed FPLA-salicylic acid patch was 16 and 22 $\mu\text{g}/\text{cm}^2$ at 15 and 60 min, respectively. For
412 the 3D printed PCL-salicylic acid sample the values were higher; 40 and 66 $\mu\text{g}/\text{cm}^2$ at 15 and 60
413 min, respectively. In use, such a mask could be worn on numerous occasions since the
414 maximum diffusion reached only 191 $\mu\text{g}/\text{cm}^2$.

415

416 Regarding the printing of intricate devices/masks, the FPLA-salicylic acid printed nose was
417 successfully obtained with the same printing settings as for the FPLA-salicylic acid circular
418 patch. The printed nose was flexible and the shape was clearly defined, although some parts of
419 the nose had small gaps between layers due to the inconsistency of the printing process.

420

421 The nose mask could not be printed using the PCL-salicylic acid filament. At the printing
422 temperature used to print the patch from the PCL-salicylic acid filament (170°C) the layer cooled
423 down slowly and failed to solidify to form stable layers in the curved regions of the nose. Lower
424 temperatures resulted in blockage of the nozzle of the printer. The use of lower printing speed
425 or extra fans to cool it down faster may allow the fabrication of structures more complex than a
426 flat patch.

427

428 **SLA printing**

429

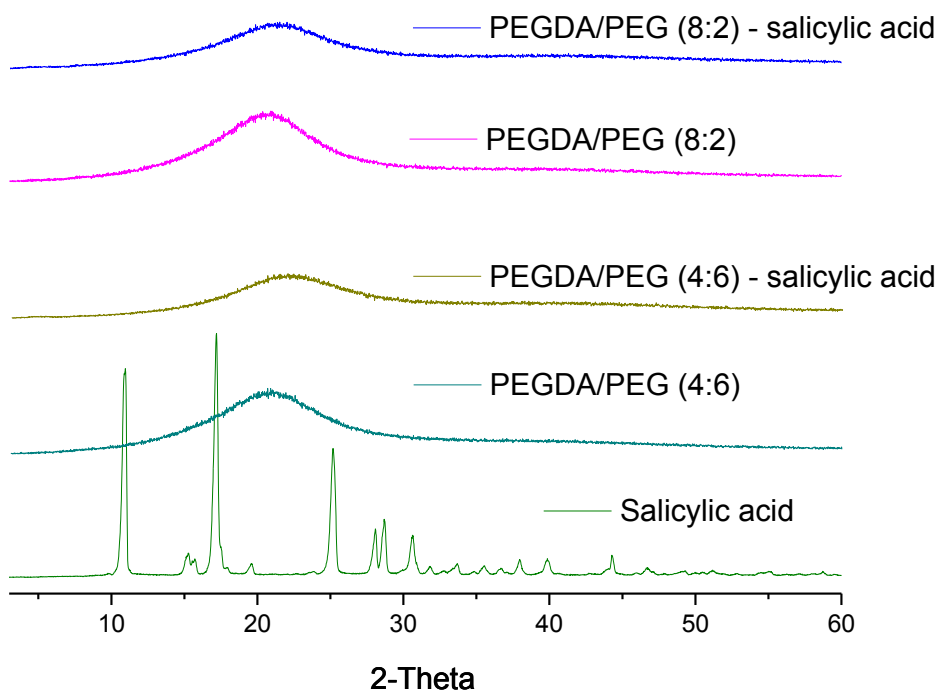
430 It was possible to fabricate patches incorporating drugs by SLA printing. The composition of the
431 formulations included the photocrosslinkable polymer PEGDA and PEG that was added as a
432 filler. PEG chains are interspersed with the PEGDA chains, which reduces the degree of
433 crosslinking between PEGDA chains. The PEGDA/PEG-salicylic acid patches were smooth and
434 slightly flexible; those with higher amounts of PEG: PEGDA/PEG (4:6)-salicylic acid were most
435 flexible.

436

437 The salicylic acid content in the devices obtained by SLA was $1.95 \pm 0.04\%$ w/w for the
438 PEGDA/PEG (4:6)-salicylic acid and $1.96 \pm 0.03\%$ w/w for the PEGDA/PEG (8:2)-salicylic acid,
439 both higher than that in the devices prepared by FDM 3DP and very close to the expected
440 value. Since fabrication of the mask by SLA printing is not based on heat, unlike FDM 3DP,,
441 salicylic acid is not thermally degraded and remains in the mask at a higher concentration.

442

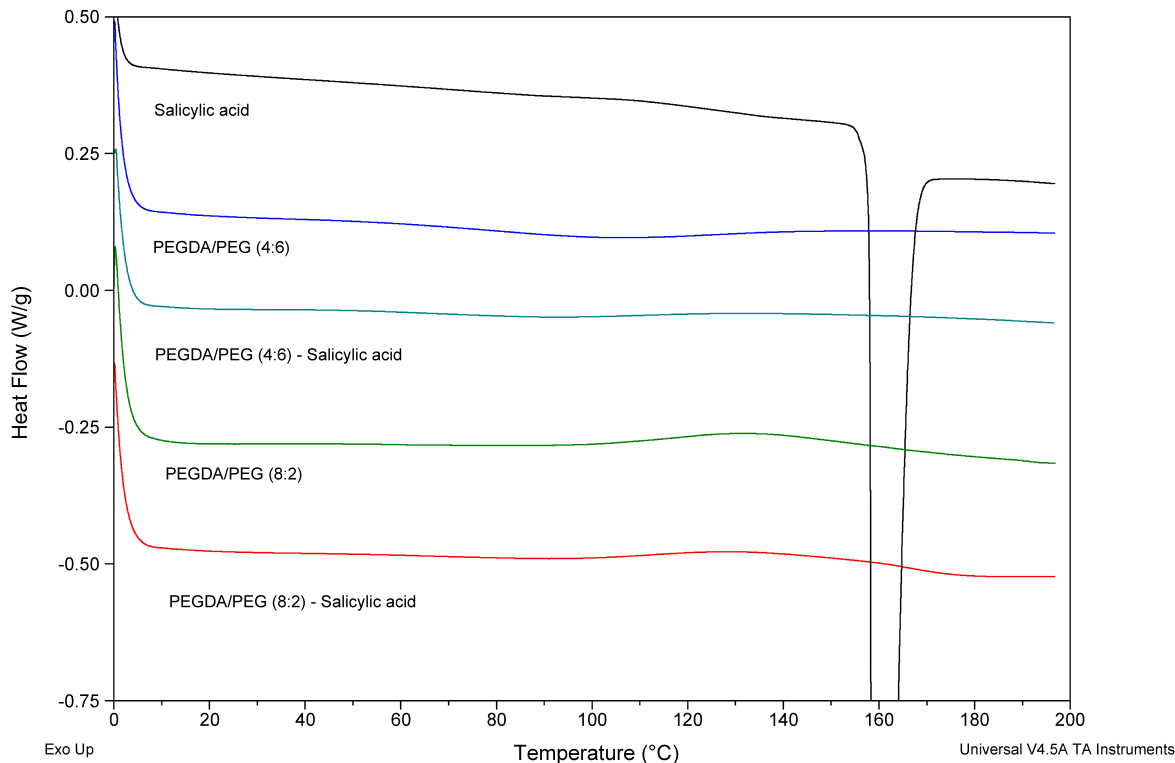
443 XRPD results suggest that salicylic acid is present in the amorphous phase within the patches
444 as no peaks appeared in the patterns of these formulations (**Fig. 7**). The drug is completely
445 dissolved in the photopolymer solution while printing and according to these results there is no
446 crystallization of the drug during the photopolymerization process. DSC data confirm that extent
447 since no drug endotherm peak is observed (Fig. 8)



448

449

450 **Fig. 7.** X-ray powder diffraction patterns for salicylic acid and various SLA 3DP polymer discs.



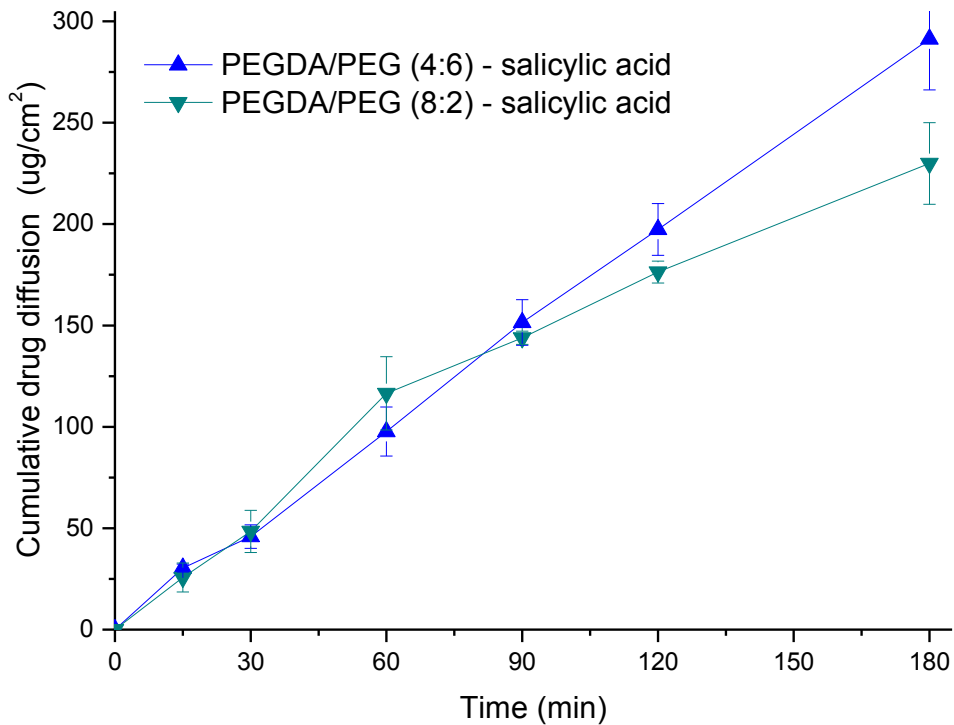
451

452 **Fig. 8.** DSC thermal traces for salicylic acid and various SLA 3DP polymer discs .

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454

455 Drug diffusion from the two 3D printed devices manufactured by SLA was higher than that
 456 obtained from devices printed by FDM 3DP after 3h (**Fig. 9**). The diffusion profiles show that the
 457 cumulative percentage of drug diffused from the printed PEGDA/PEG patch with the higher
 458 amount of PEG is similar (25 and 48 $\mu\text{g}/\text{cm}^2$ at 15min and 60 min, respectively) to that with
 459 lower amount of PEG (30 and 45 $\mu\text{g}/\text{cm}^2$ at 15min and 60min) during the first 2 h. After this time
 460 diffusion is faster from patches with higher amount of PEG. A possible explanation is that the
 461 PEG gets dissolved in the dissolution media, forming pores that let the media have improved
 462 access to more internal regions of the devices easier and faster than in less porous devices.
 463 The effect of the PEGDA/PEG ratio was previously described for oral tablets prepared by SLA
 464 printing [15].



465

466

467 **Fig. 9.** Cumulative amounts of salicylic acid permeated from the devices 3D printed by SLA.

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469 For the fabrication of the nose-shaped device containing salicylic acid, SLA 3DP provided
 470 higher resolution than the FDM approach (**Fig. 10**). The mask is as flexible as the mask
 471 obtained from FDM 3DP with the flexible polymer FPLA-salicylic acid.



472

473 **Fig. 10.** Nose-shaped device fabricated by SLA 3D printing PEGDA/PEG (4:6)-salicylic acid.

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Conclusions

3D printing technologies show potential in the development of personalized anti-acne drug loaded masks/patches. In FDM 3DP, the use of HME produced filaments of FPLA-salicylic acid and PCL-salicylic acid with uniform diameter and suitable for 3D printing, whereas the NF-salicylic acid filament was too brittle for 3D printing. Drug loading was higher in 3D printed objects obtained with PCL-salicylic acid filaments than with FPLA-salicylic acid filaments due to the greater degradation of the drug at higher temperatures (extrusion and 3D printing). However, the resolution and printing characteristics of the objects were better with FPLA-salicylic acid, it was not possible to print nose-shaped mask with the PCL-salicylic acid filament.

Drug diffusion tests conducted in Franz cells revealed that FPLA-salicylic acid and PCL-salicylic acid printed samples diffused only 53 and 187 $\mu\text{g}/\text{cm}^2$ respectively within 3 h.

SLA printing involves a one-step process that leads to 3D printed devices with higher resolution than the obtained with the FPLA-salicylic acid filaments and with higher drug loading than the PCL-salicylic acid filament (1.9% w/w), with no drug degradation. The results of drug diffusion tests conducted under the same conditions revealed that the total drug diffused is also faster than with the FDM approaches, 291 $\mu\text{g}/\text{cm}^2$ within 3 hour for PEGDA/PED (4:6)-salicylic acid and 229 $\mu\text{g}/\text{cm}^2$ for the PEGDA/PED (8:2)-salicylic acid.

Therefore it can be concluded that SLA printing is a more convenient 3D printing technology to manufacture anti-acne devices with salicylic acid. The 3D printed masks may be considered as promising formulations that can be developed further to provide higher efficacy in acne treatment. The dose of drug may be adjusted (reduced) to personalize the device by incorporating a specific dose of drug into the polymer that will then be printed as a patch for each patient, maybe ameliorating the dose-dependent side effects of the treatment. In other pathologies, the combination of 3D scanning and 3D printing, have the potential to offer solutions to produce personalised drug loaded devices, adapted in shape and size to individual patients.

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