Supplemental information

Regulatory T cells inhibit CD34+ cell differentiation into NK cells by blocking their proliferation

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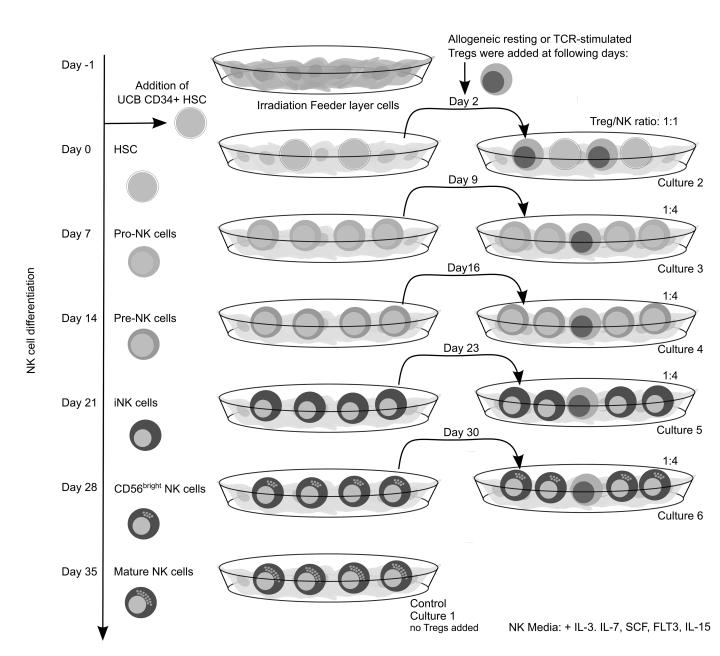


Figure S1: Experimental design used to assess the effect of Tregs on NK cell differentiation *in vitro*. HSC ± resting or activated Tregs were cultured in the presence of irradiated EL08.1D2 feeder layer cells and cytokines for 35 days. Tregs were added to HSC cultures at a 1:4 ratio (Tregs:NK cells), except when added at day 2, where a ratio of 1:1 was used. Activated Tregs were activated with plate bound anti-CD3/soluble anti-CD28 and 1000 IU/mL IL-2 for 24 h and washed before addition to HSC cultures.

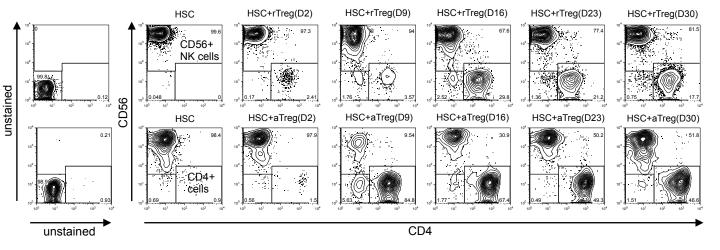


Figure S2: Percentages of NK cells and CD4+ cells in HSC cultures. HSC were cultured ± resting or activated Tregs added at days 2, 9, 16, 23 and 30 of differentiation at a 1:4 ratio (Tregs:HSC). Representative flow cytometric analysis of NK cells (CD56+) and Tregs (CD4+) from all cultures at day 35 of HSC cultures.

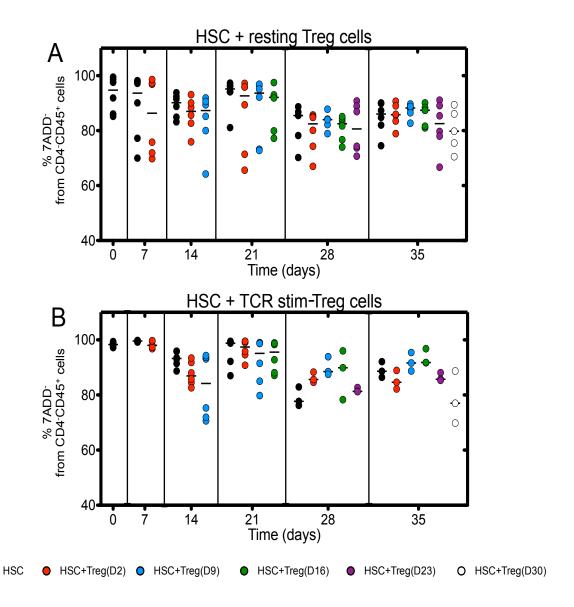


Figure S3: Viability of CD45⁺ cells in HSC cultures in the presence or absence of resting or TCR-stimulated CB Treg cells. Viability of CD45⁺ was assessed by flow cytometry using CD4, CD45 and 7-AAD. HSC were cultured in the presence or absence of allogeneic resting CB Treg cells (A) or TCR-stimulated CB Treg cells (B). Treg cells were added at key time points of differentiation. The lines represent medians. n=6-8.

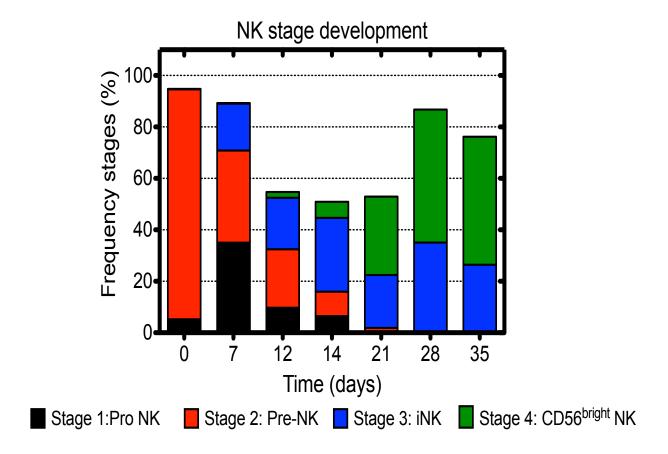


Figure S4: Frequency of cells in each NK cell differentiation stage in HSC cultures. HSC were cultured for 35 days and frequencies of Stage 1-4 NK cells were assessed based on CD4, CD34, CD117 and CD94 expression at days 0, 7, 14, 21, 28 and 35. The values represent medians. n=11

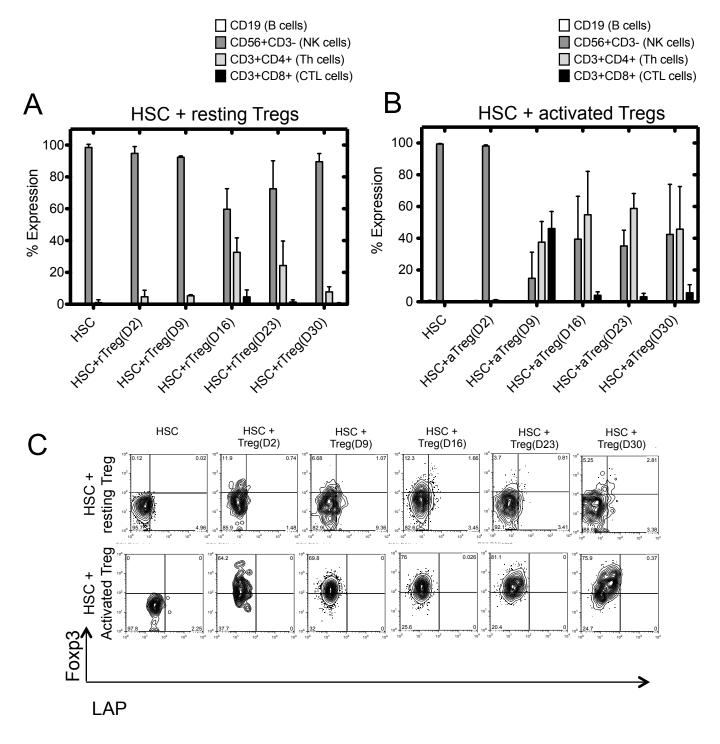


Figure S5: Phenotypic analysis of lymphocyte populations after HSC cultures. Flow cytometry analysis of CD4, CD8, CD19 and CD56 in HSC cultures with resting or activated Tregs (**A-B**). Expression of Foxp3 and LAP on CD4 T cell populations. HSC were used as negative control of expression (**C**).

Table S1. Primer sequences used for real time PCR. Conc. indicates concentration.

		Conc
Primer	Sequence	(nM)
BCL11B	F: 5'-CTCTCACCCACGAAAGGCAT-3' R: 5'-GCACGCAGAGGTGAAGTGAT-3'	300
E4BP4	F: 5'-CCAAGGGCCCCATCCATTC-3' R: 5'-GATGCCAGTGCTCCGATTTG-3'	300
FOXP3	F: 5'-CACCTGGCTGGGAAAATGG-3' R: 5'-GGAGCCCTTGTCGGATGAT-3'	900
GATA-3	F: 5'-AGCACAGAAGGCAGGGAGTGT-3' R: 5'-TTCGCTTGGGCTTAATGAGGGGC-3'	300
HELIOS	F: 5'-ACACCTCAGGACCCATTCTG-3' R: 5'-TCCATGCTGACATTCTGGAG-3'	600
ID2	F: 5'-CGGATATCAGCATCCTGTCC-3' R: 5'-TCATGAACACCGCTTATTCAG-3'	300
PU.1	F: 5'-TGTTACAGGCGTGCAAAATGGAAGG-3' R: 5'-CTCGTGCGTTTGGCGTTGGTATAGA-3'	300
RORC	F: 5'-AGTCGGAAGGCAAGATCAGA-3' R: 5'-CAAGAGAGGTTCTGGGCAAG-3'	300
Т-ВЕТ	F: 5'-GGATGCGCCAGGAAGTTTCA-3' R: 5'-CTCTGGCTCTCCGTCGTTCA-3'	300
TOX	F: 5'-TATGTGCCAGCCAGCCAGTCCTA-3' R: 5'-TGGTCTGGGAGGGAAGGAGGAGTAA-3'	300

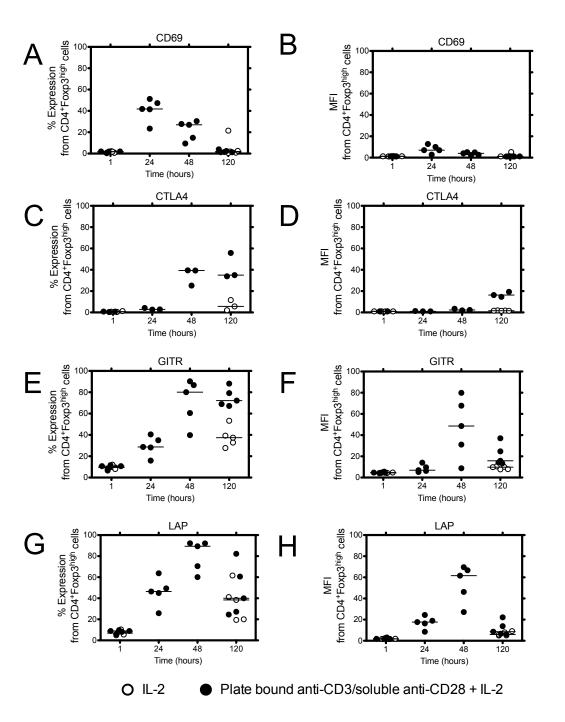


Figure S6: Phenotypic analysis of Tregs after activation. Flow cytometric analysis of Tregs during stimulation with plate bound anti-CD3/soluble anti-CD28 and 1000 IU/mL IL-2. Frequency of expression and MFI of CD69 (**A-B**), CTLA4 (**C-D**), GITR (**E-F**) and LAP (**G-H**) were assessed on gated CD4+Foxp3high Tregs. n = 3-5.