

# Bioaccessibility of phosphorus sorbed to amorphous oxides: Evaluation of plant supply potential

Maximilian Koch<sup>1</sup>, Stella Gypser<sup>2</sup>, Nina Siebers<sup>1</sup>, Wulf Amelung<sup>3</sup>

<sup>1</sup> Institute for Bio- and Geosciences – IBG-3, Agrosphere, Forschungszentrum Jülich GmbH, 52425 Jülich, Germany

<sup>2</sup> Brandenburg University of Technology Cottbus-Senftenberg, Chair of Soil Protection and Recultivation, 03046 Cottbus, Germany

<sup>3</sup> Institute for Crop Science and Resource Conservation (INRES) – Soil Science and Soil Ecology, University of Bonn, Nussallee 13, 53115 Bonn, Germany



## Introduction and Objectives

In sparsely rooted subsoils secondary minerals, such as iron and aluminum oxides are predominant sorption sites for phosphorus (P) and thus important sinks for P fertilizers. P supply from these oxides can contribute to effective strategies for a sustainable future use of P fertilizers. Here we are going to combine (1) P radio-isotope tracing and imaging techniques, (2) diffuse gradient in thin films technology (DGT), and (3) magnetic resonance imaging (MRI) with the **objective** to assess the P accessibility and use efficiency from stable P reservoirs.

## Bioimaging technology – Radioluminography

Using radioimaging it is possible to locate <sup>33</sup>P labelled P within the plant and roots. The CR 35 Bio is a compact high-speed image plate scanner (Figure 1) to detect a variety of different radio nucleotides (radioluminographic imaging). The Imaging plate (IP) is a flexible, coated radiation sensor plates, which contains fine crystals (< 5 µm) of photo-stimulable barium fluorobromide spiked with bivalent europium as luminescence center (Nakajima, 1993). By comparison with conventional X-ray imaging, referred to as autoradiography, this technique enables shorter exposition times, high resolutions (till 30 µm) and an easy handling due to digital processing.

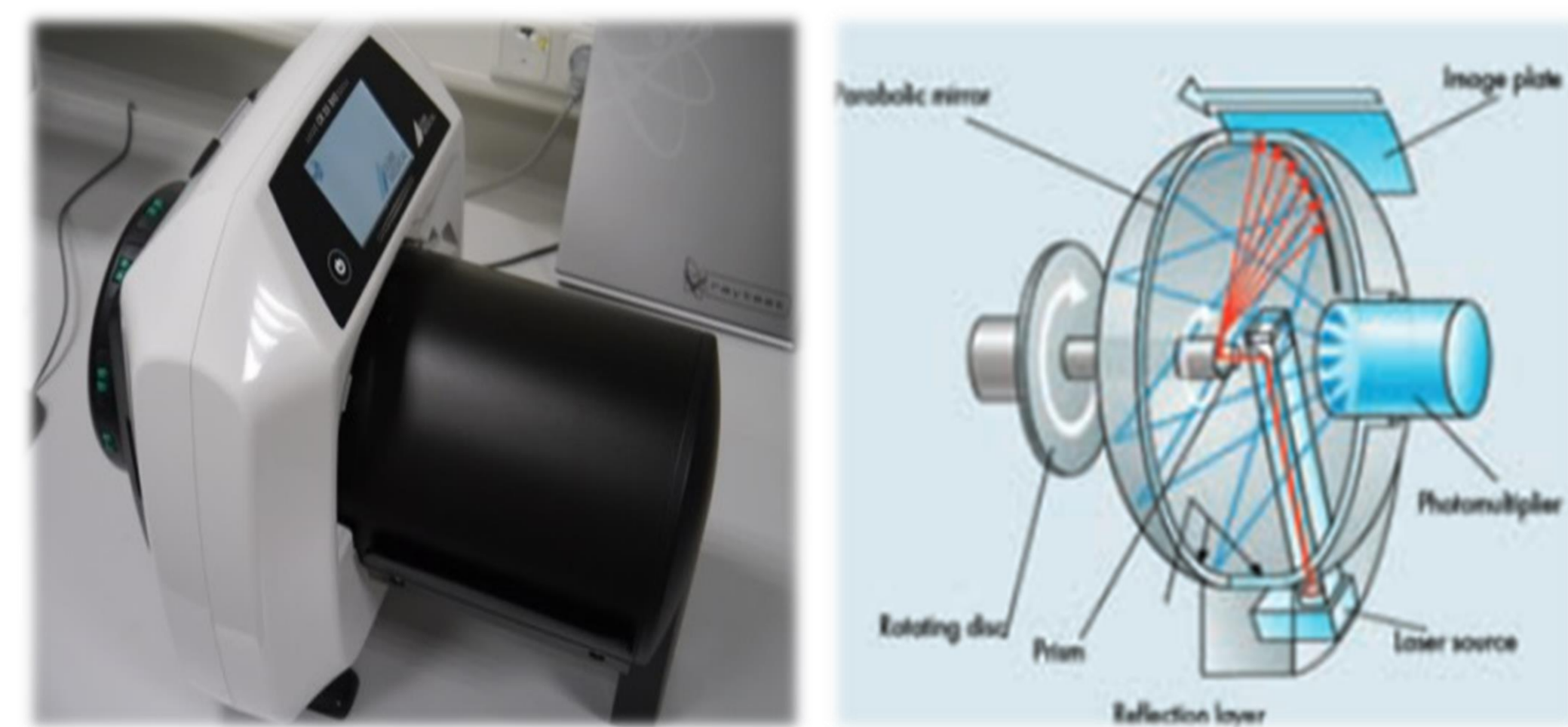


Figure 1: Left: CR 35 Bio compact high-speed image plate scanner (Photo: Koch; 2015). Right: Insights of a scanner unit with activated laser beam (red lines) reading an image plate (blue) (Picture: www.biostep.de).

## Study design and first results

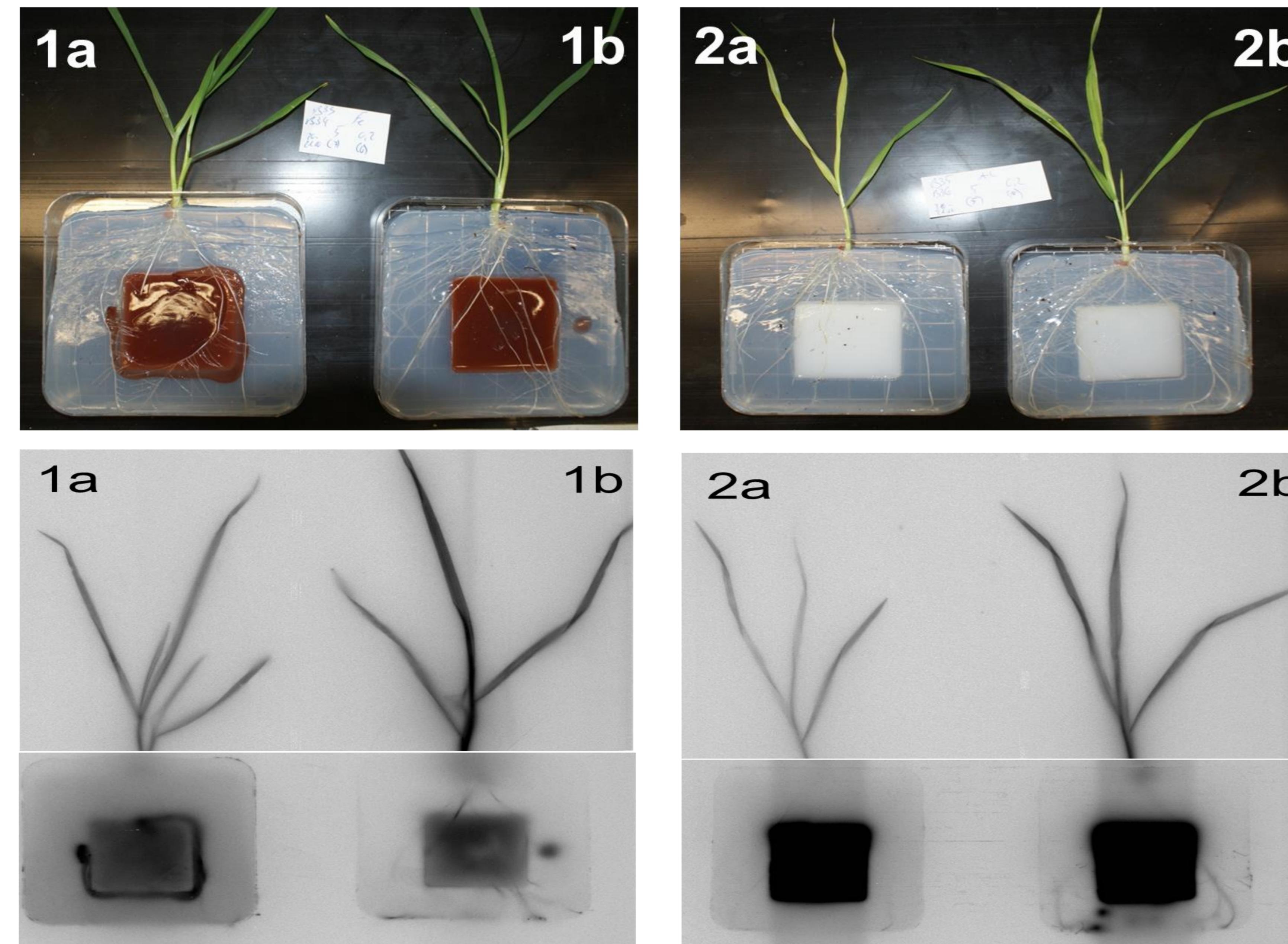


Figure 2: Photo and bioimager image of plant-agar system after 18 d of growth. P sources were provided as (1) P sorbed on to amorphous iron oxide and (2) on to amorphous aluminum oxide, both labelled with <sup>32</sup>P-Orthophosphate mixed in agar gel and implemented in surrounding agar gel with basal nutrition solution containing (a) 5mM and (b) 0,2 mM of KH<sub>2</sub>PO<sub>4</sub>. The experiment was performed in a growth chamber with 16 h day-length at a light intensity of 320 µmol m<sup>-2</sup> s<sup>-1</sup> (PAR). Day-time conditions were 20 °C and 60% relative air humidity, while at night temperature decreased to 12 °C at 80% relative air humidity. IPs was exposed for 6 h and scanned with 100µm sensitivity.

## Outlook

### (1) Plant - Soil System - P Uptake

<sup>33</sup>P sorbed oxide uptake studies allow to evaluate P utilization potential from P associated with soil minerals in a natural environment.



Figure 3 : Design of soil filled rhizotron for analysing Fe-P/ Al-P accessibility in subsoil by wheat plants

### (2) Oxid-P desorption isothermes - DGT

To unambiguously analyse exchange - and desorption potentials in soils compared to plant acquisition strategies of Fe-P and Al-P oxides.

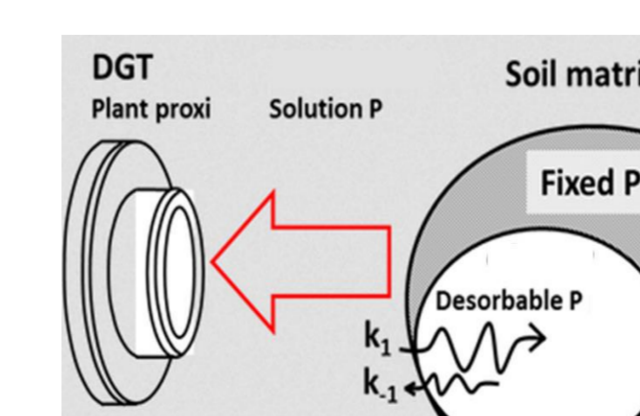


Figure 4 : Functional design of the DGT method. Illustrating the trapping of soil desorbable P in the soil solution (modified Picture: soilforum.com)

### (3) Root morphology - MRI

MRI experiments (Figure 5) for analysing root morphology and to obtain a structural model of plant root systems. This allows to evaluate P acquisition strategies by plant roots at the soil profile scale.

Figure 5 : MRI picture of a Lupine root (Oswald et al., 2015)



In a plant-agar system <sup>32</sup>P labelled Fe-P and Al-P oxides are applied in agar gel (Figure 2; brown and white boxes in the square petri dishes). Wheat plants were grown in the agar for 18 d, afterwards <sup>32</sup>P plant up take was quantified via liquid scintillation counting (LSC).

The **hypothesis** of this study was to analyse (1) whether the plants take up more <sup>32</sup>P in the low P concentration agar medium (0,2 mM) than in the high P medium (5mM)? (2) what is the P supply potential of Fe-P and Al-P oxides? Is the plant able to adapt strategies for P supply to different soil minerals?

Table 1: Dry weights (DW) and <sup>32</sup>P proportions of selected plant compartments and agar nutrition medium after 18 d of growth. <sup>32</sup>P recovery was between 96% and 100%. Significant at \* P < 0.05. n = 3

P concentration	Iron oxide		Aluminium oxide		Control
	5mM (1a)	0,2 mM (1b)	5mM (2a)	0,2 mM (2b)	0,6 mM
Shoot DW [mg]	178±33	183±28	120±6	146±15	176±39
Root DW [mg]	42±20	37±3	36±3	41±2	45±9
Shoot <sup>32</sup> P (%)	1.1±0.3	<b>3.9±0.7*</b>	0.1±0.0	0.3±0.0	-
Root <sup>32</sup> P (%)	1.2±1.0	1.9±0.0	1.1±0.2	1.8±0.3	-
Agar <sup>32</sup> P (%)	98±1.1	94±0.7	99±0.4	98±0.2	-

The results illustrated that it is possible to quantify P plant acquisition from dominant soil P sources via radio-isotopic labelling. Plants grown in 0,2 mM P agar medium showed higher specific <sup>32</sup>P activities. Further, P sorbed to iron oxide was significantly better accessible than from aluminum oxide due to adapted acquisition strategies of the plant to utilize the Fe-P as available P source.

## References

Nakajima, E. (1993). "CRC Handbook of Chromatography: Analysis of Lipids," CRC Press, London, Tokyo.  
 Oswald, S. E., Tötze, C., Haber-Pohlmeier, S., Pohlmeier, A., Kaestner, A. P., and Lehmann, E. (2015). Combining Neutron and Magnetic Resonance Imaging to Study the Interaction of Plant Roots and Soil. *Physics Procedia* 69, 237-243.

## Acknowledgment

The authors gratefully thank for assistance and help Martina Krause. The authors would like to acknowledge the BMBF for funding this work within the InnoSoilPhos project group.