Defining and Characterising the anti-HCV Actions of the Interferon-Induced Transmembrane proteins

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Abstract

Hepatitis C virus (HCV) is a significant human pathogen of the liver that in the majority of infected individuals causes a chronic infection of the liver. This may over time culminate in the development of severe liver disease such as cirrhosis and hepatocellular carcinoma. Prior to 2012, the only treatment option available for HCV infection was combination therapy with pegylated interferon- α (IFN- α) and ribavirin. However, the recent development and addition of direct acting antivirals (DAAs) into treatment regimes has significantly improved sustained virological response rates. While the ultimate aim is for IFN- α free therapy, IFN- α is often required in combination with the DAAs to reduce the development of viral resistance and due to cost. It is clear that IFN (either exogenous or endogenous) can induce an antiviral state in HCV infected cells; however, the exact mechanisms that underpin this action remain unclear.

The interferon-induced transmembrane (IFITM) family of proteins - IFITM1, IFITM2 and IFITM3 has recently been identified as important host effector molecules of the type I IFN response against a broad range of RNA viruses. During the course of this PhD study, a number of investigations identified the IFITM proteins to be potent antiviral effectors against HCV; however, the mechanism(s) for this antiviral activity remains contradictory. In this thesis, we demonstrate that IFITM1, IFITM2 and IFITM3 play an integral role in the IFN response against HCV and act specifically to inhibit early and late stages of HCV entry to inhibit infection. We reveal that IFITM1 localises to the cell surface in hepatocytes and interacts with the host entry factor CD81 to limit HCV entry. Furthermore, the N-terminus, in particular amino acids 21-28, of

IFITM1 plays an important role in this anti-HCV activity, while the C-terminus is found to be important for localisation to the cell surface.

We also established that in hepatocytes, IFITM2 and IFITM3 localise to the late and early endosomes respectively, as well as the lysosome, indicating that IFITM2 and IFITM3 follow the established paradigm of targeting the late entry stages of HCV infection. Furthermore, we have demonstrated that S-palmitoylation of all three IFITM proteins is essential for both anti-HCV activity and cellular localisation, while the conserved tyrosine residue in the N-terminus of IFITM2 and IFITM3 plays a significant role in protein localisation. However, this tyrosine was found to be dispensable for anti-HCV activity, with mutation of the tyrosine resulting in an IFITM1-like phenotype with the retention of anti-HCV activity and co-localisation of IFITM2 and IFITM3 with CD81.

In conclusion, we propose that the IFITM proteins act in a coordinated manner to restrict HCV infection by targeting the endocytosed HCV virion for lysosomal degradation and demonstrate that the actions of the IFITM proteins are indeed virus and cell-type specific. We believe we have significantly added to our understanding of the interplay between HCV and the host innate immune response and that in the long term these findings will aid in the generation of novel and targeted anti-HCV therapeutics for patients chronically infected with HCV.

Declaration

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Sumudu Kumari Narayana

October 9th, 2015

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Publications Arising from this PhD

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Awards Received During PhD

| 2012 | School of Molecular and Biomedical Science School Symposium Poster Prize – 2^{nd} Place - $\$200$ |
|------|---|
| 2012 | Nomination by the Golden Key International Honour Society to attend the 2013 International Scholar Laureate Program (ISLP) Delegation on Medicine |
| 2012 | Australian Centre for Hepatitis Virology Travel Award for HCV Research – \$2500 National award - one awarded per year for travel to international HCV meeting. |
| 2011 | Dawes Top-Up Scholarship, Royal Adelaide Hospital Research Fund Grant Round – funded from 2012 to 2014 |

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K.J. Helbig, J.M. Carr, S.K. Narayana, E.M. McCartney, N.S. Eyre and M.R. Beard. The ISG viperin has novel anti-hepatitis C virus activity through interaction with a

HCV pro-host cell factor. Keystone Symposia 2012 - Innate Immunity, Colorado, USA, 2012. (poster presentation)

McCartney E.M., Helbig, K.J., Eyre, N.S, **Narayana, S.K**. and Beard, M.R. The role of STAT3 in the life cycle of HCV. 18th International Symposium on Hepatitis C Virus and Related Viruses, Seattle, USA, 2011. (poster presentation)

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Narayana S.K., Eyre, N.S., Van der Hoek, K., Helbig, K.J., and Beard, M.R. The interferon-induced transmembrane (IFITM) proteins 1 and 3 have anti-HCV activity *in vitro*. Australian Centre for Hepatitis Virology workshop, Maroochydore, Australia, 2011. (oral presentation)

Beard M.R., **Narayana**, **S.K.**, Eyre, N.S., Yip, E.Y., Lemon, S.M. and Helbig, K.J. The ISGs viperin and the IFITM family have novel anti-Hepatitis C Virus activity. Infection and Immunity Conference, Lorne, Australia, 2011. (oral presentation)

Narayana S.K., Eyre N.S., Van der Hoek, K., Helbig K.J., and Beard, M.R. Identification of novel interferon stimulated genes (ISGs) that control HCV *in vitro*. Australian Centre for Hepatitis Virology workshop, Yarra Valley, Australia, 2010. (oral presentation)

Materials Providers

Abcam Cambridge, UK

Ambion Texas, USA

Amersham Pharmacia Biotech Birminghamshire, UK

Amrad Biotech Boronia, VIC, Australia

Anogen Ontario, Canada

Applied Biosystems Warrington, UK

Becton Dickson Labware New Jersey, USA

Biomol New Jersey, USA

BioRad Laboratories California, USA

Cell Signaling Massachusetts, USA

Chemicon International Massachusetts, USA

Cohu California, USA

DAKO California, USA

Dynatech Virginia, USA

GeneWorks Adelaide, SA, Australia

Invitrogen California, USA

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Mol Bio Laboratories California, USA

Molecular Probes Oregon, USA

Nalge Nunc International Illinois, USA

Nikkon Sydney, NSW, Australia

New England Biolabs Massachusetts, USA

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Olympus New York, USA

Panomics Santa Clara, USA

Perkin Elmer Massachusetts, USA

Promega Wisconsin, USA

QIAgen Hilden, Germany

Roche Indiana, USA

Rockland Pennsylvania, USA

Schering-Plough New Jersey, USA

Schleicher and Schuell Dassel, Germany

Sigma Missouri, USA

SPSS Inc Illinois, USA

Stratagene California, USA

UVP Inc California, USA

Vector Laboratories California, USA

Vision Systems Mount Waverley, VIC, Australia

Abbreviations Used

A adenosine

aa amino acids

bp base pairs

BSA bovine serum albumin

BVDV bovine viral diarrhoea virus

C cytosine

° C degrees Celsius

cDNA complimentary deoxyribosenucleic acid

CLDN1 claudin-1

CHC chronic hepatitis C

CIL cytoplasmic intracellular loop

CMV cytomegalovirus

dATP deoxyadenosine-5'-triphosphate

dCTP deoxycytosine-5'-tripshosphate

DEPC diethyl pyrocarbonate

DAA direct acting antiviral

dGTP deoxyguanosine-5'-triphosphate

dH₂O deionised water

DMEM Dulbecco's Modified Eagle Medium with HEPES

DNA deoxyribonucleic acid

dNTP deoxyribonucleotide triphosphate

dTTP deoxythymidine-5'-triphosphate

DENV Dengue virus

EBoV Ebola virus

EDTA ethylene diamine tetra acetic acid

ER endoplasmic reticulum

FCS foetal calf serum

FITC fluorescein isothiocyanate

FFU focus forming unit

g grams

G guanosine

GAPDH glyceraldehyde-3-phosphate deydrogenase

GAS gamma-activated sequence

GFP green fluorescent protein

HCC hepatocellular carcinoma

HBV hepatitis B virus

HCV hepatitis C virus

hr hour(s)

HRP horse radish peroxidase

HIV human immunodeficiency virus°°°°

HPV human pappilomavirus

IAV influenza virus

IDU infecting drug use

IFITM interferon-induced transmembrane protein

IFN-α interferon alpha

IFN-β interferon beta

IFN-γ interferon gamma

IFN-λ interferon lambda

IRES internal ribosome entry site

ISRE interferon stimulated response element

JAK janus kinase

kb kilobase

kDa kilo Dalton

L-Agar LB + agar

LD lipid droplet

LB Luria Bertani broth

LDL low density lipoproteins

Luc luciferase

μg micrograms

μl microlitres

μM micromolar

mA milliamps

mg milligrams

ml millilitres

mM millimolar

MCS Multiple Cloning Site

MEM Minimum Essential Medium

min minute(s)

mRNA messenger RNA

MW molecular weight

MLV murine leukemia virus

ng nanograms

nM nanomolar

N/A not applicable

nt nucleotide

NTD N-terminal domain

NF-κB nuclear factor-kappa-light chain enhancer

OCLN occludin

ORF open reading frame

PAGE polyacrylamide gel electrophoresis

PBS phosphate buffered saline; 0.15M NaCl, 6M K₂HPO₄, 2mM

 KH_2PO_4 (pH 7)

PAMP pathogen associated membrane patterns

PCR polymerase chain reaction

PHH primary human hepatocyte

PRR pathogen recognition receptors

PTM post-translational modification

pg picograms

pmol picomolar

RNA ribonucleic acid

rpm revolutions per minute

RFP red fluorescent protein

RdRp RNA-dependent-RNA polymerase

RC replication complex

RIG-I retinoic acid inducible gene I

RT room temperature

RT-PCR reverse transcriptase polymerase chain reaction

sd standard deviation

SDS sodium dodecyl sulfate

sec second(s)

SEM standard error of mean

ss single stranded

siRNA small interefering RNA

SR-BI scavenger receptor class B member I

STAT signal transducer and activator of transcription

SVR sustained virological response

T thymidine

TAE 0.04M Tris (pH 8), 0.04M Acetic Acid, 1mM EDTA

TEMED N,N,N',N'-tetramethylethyethylenediamine

Tris 3,3,5,5-tetramethylbenzidine

TLR Toll-like receptor

TYK2 tyrosine kinase 2

U units

UTR untranslated region

V volts

VSV vesicular stomatis virus

w/v weight per volume

WNV West Nile Virus