Identification of host cell proteins involved in *Shigella flexneri* pathogenesis

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Abstract

Shigella flexneri is the etiological agent of bacillary dysentery (shigellosis). It is transmitted via the faecal-oral route and is a significant human pathogen due to the high morbidity among children <5 years in developing countries. The key pathogenic features of Shigella include cell death induction in myeloid immune cells and circumventing cell death in colonic epithelial cells, the site of bacterial infection. Shigella also interact with host proteins to initiate de novo actin synthesis to facilitate its intra- and intercellular spread to disseminate in the host.

In this thesis, the role of three host proteins: myosin IIA, dynamin II, and dynamin-related protein 1 (Drp1) during *Shigella* cell-to-cell spreading was examined. The myosin IIA specific kinase, myosin like chain kinase (MLCK), was previously shown to be important for *Shigella* plaque formation. Myosin IIA and MLCK have also been implicated in septin caging of non-motile *Shigella* which are targeted for degradation. Chemical inhibition and siRNA knockdown of myosin IIA reduced *Shigella* plaque formation. Curiously HeLa cells infected with *Shigella* mutants defective in cell-to-cell spreading have significantly reduced myosin IIA levels when quantified by immunofluorescence microscopy.

Dynamin II and Drp1 are members of the dynamin superfamily. Both proteins have self-assembly driven GTPase activation. Dynamin II is important for clathrin-mediated endocytosis and pinches the budding clathrin-coated vesicle, and Drp1 is essential for mitochondrial fission. It was hypothesized that *Shigella* protrusion formation into adjacent host cells resembles endocytic and exocytic processes, and components of these processes may facilitate *Shigella* dissemination. When dynamin II GTPase was inhibited with dynasore and dynamin II was knocked down with siRNA, *Shigella* cell-to-cell spreading was significantly reduced. The *in vivo* efficacy of dynasore was tested in a murine Sereny model. No significant reduction in inflammation was observed but mice were protected against weight loss during infection. Further experimentation suggested dynasore protected mice against cytotoxic effects from the three secretion system (TTSS) effectors expressed by *Shigella* during infection.

Drp1 was investigated in this thesis as dynasore also inhibits the GTPase of this mitochondrial fission protein. Mitochondrial fission is important in maintaining mitochondrial

dynamics and also in events downstream of intrinsic apoptosis and programmed necrosis pathways activation. Loss of mitochondrial function in *Shigella*-induced epithelial cell death has been reported previously. Hence the role of Drp1 in *Shigella* plaque formation and HeLa death was examined with the Drp1-specific inhibitor, Mdivi-1, and siRNA knockdown. HeLa cell death was significantly reduced; suggesting loss of mitochondrial function observed previously may now be attributed to Drp1 and subsequent Drp1-mediated mitochondrial fission. The impairment in *Shigella* cell-to-cell spreading in the absence of Drp1 suggests maintaining an intact mitochondrial network is essential for *Shigella* lateral spread since loss of Drp1 function would result in excessive mitochondrial fusion, leading to formation of net-like or perinuclear structures.

The outcomes of this thesis highlight the importance of host proteins during different stages of *Shigella* infection. By improving our understanding on the host and bacteria interaction, future work on novel approaches to prevent *Shigella* dissemination can be developed.

Declaration

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Publications

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Abbreviations

~ approximately

aa amino acids

ABM actin-based motility

ADP adenosine diphosphate

AJ(s) adherens junction(s)

APC(s) apical junctional complex(es)

ATP adenosine triphosphate

ATPase adenosine triphosphatase

BDM 2,3-butanedione monoxime

BSE bundle signalling element

CFU colony forming units

D0/D1/D2/D3 day 0/day 1/day 2/day 3

DCCR DharmaFECT Cell Culture Reagent

DLP(s) dynamin-like protein(s)

DLP1 dynamin-like protein 1 (alternate name for DNM1L/Drp1)

DNM1L dynamin-1-like protein (alternate name for DLP1/Drp1)

DNM2 dynamin II gene

DMSO dimethyl sulfoxide

Drp1 dynamin-related protein 1 (alternate name for DLP1/DNM1L)

GAPDH glyceraldehyde 3-phosphate dehydrogenase

GED GTPase effector domain guanosine triphosphate

GTPase(s) guanosine triphosphatase(s)

h hour(s)

IF immunofluorescence

IP intraperitoneal

kDa kilodaltons

LDH lactate dehydrogenase

Lo low myosin IIA protein levels

LPS lipopolysaccharide

MEFs mouse embryonic fibroblasts

Mdivi-1 <u>m</u>itochondrial <u>div</u>ision <u>i</u>nhibitor-<u>1</u>

MYH9 myosin, heavy chain 9, non-muscle gene

(myosin IIA heavy chain gene)

min minute(s)

MLCK myosin light chain kinase moi multiplicity of infection

MOMP mitochondrial outer membrane permeabilisation

myosin IIA / B / C non-muscle myosin IIA / B / C

N-WASP Neural Wiskott-Aldrich syndrome protein

NF-κB nuclear factor-κB

NMP *N*-methyl-2-pyrrolidone

NPF(s) nucleation promoting factor(s)

OM outer membrane

PEG / PEG300 polyethylene glycol 300

PGN peptidoglycan

PH pleckstrin homology

PMN(s) polymorphonuclear cell(s)

PRD(s) proline-rich domain(s)

PtK2 Potorous tridactylis kidney epithelial (cells)

R-LPS rough LPS

ROS reactive oxygen species

S-LPS smooth LPS

siRNA small interfering RNA

STS staurosporine

t time

TJ(s) tight junction(s)

TTSS type three secretion system

VP virulence plasmid

VP⁻ / VP- virulence plasmid-cured

WT wild type