

Cairo University

Journal of Advanced Research



REVIEW

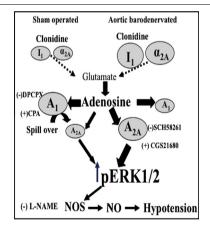
Brain stem adenosine receptors modulate centrally mediated hypotensive responses in conscious rats: A review



Noha N. Nassar^a, Abdel A. Abdel-Rahman^{b,*}

^a Department of Pharmacology and Toxicology, Faculty of Pharmacy, Cairo University, Egypt
^b Department of Pharmacology and Toxicology, Brody School of Medicine, East Carolina University, NC, USA

G R A P H I C A L A B S T R A C T



Abbreviations: A_{2A} , adenosine subtype A_{2A} receptor; A_1 , adenosine subtype A_1 receptor; ABC, avidin biotin complex; ABD rat, aortic barodenervated rat; α_2 AR, alpha 2 adrenergic receptor; α MNE, alpha methyl norepinephrine; ATP, adenosine triphosphate; BP, blood pressure; cAMP, cyclic adenosine monophosphate; CGS21680, 2-[4-[(2-carboxyethyl)phenyl]ethylaminophenyl]ethylamino]-5'-N-ethylcarboxamidoadenosine. Selective A_{2A} receptor agonist; CNS, central nervous system; CPA, N⁶-cyclopentyladenosine. Selective A_1 receptor agonist; DAG, diacylglycerol; DPCPX, 8-cyclopentyl-1,3-dipropylxanthine. Selective A_1 receptor antagonist; I₁, imidazoline subtype 1 receptor; I.C., intracisternal; IP3, Inositol Triphosphate; I.V., intravenous; JNK, C-Jun N-terminal kinase; L-NAME, N⁶⁰-nitro-L-arginine methyl ester hydrochloride. Non-selective nitric oxide synthase inhibitor; NOS, nitric oxide synthase; NO, nitric oxide; NTS, nucleus tractus solitarius; PC-PLC, phosphatidyl choline-selective phospholipase C; PC12 cells, pheochromocytoma cells; PD98059, selective extracellular signal regulated kinase; inhibitor; ERK1/2, extracellular signal regulated kinase; PDE, phosphodiesterase; PKA, protein kinase A; RVLM, rostral ventrolateral medulla; SAPK, stress activated protein kinase; SCH58261, 5-amino-7-(2-phenylethyl)-2-(2-furyl)-pyrazolo[4,3- \in]-1,2,4-triazolo[1,5-c]pyrimidine. Selective adenosine A_{2A} antagonist; SHR, spontaneously hypertensive rat; SND, sympathetic neuronal discharge; SO, sham operated = conscious normotensive rats; 8-SPT, 8-(p-sulfophenyl)-theophylline. Non-selective adenosine receptor blocker; WKY, Wistar Kyoto rat. * Corresponding author.

E-mail address: abdelrahmana@ecu.edu (A.A. Abdel-Rahman).

Peer review under responsibility of Cairo University.



ARTICLE INFO

Article history: Received 19 August 2014 Received in revised form 8 December 2014 Accepted 9 December 2014 Available online 18 December 2014

Keywords: Imidazoline I1-receptor Centrally mediated hypotension Clonidine Central adenosine receptors Conscious rats MAPK-NOS signaling

ABSTRACT

Adenosine is implicated in the modulation of cardiovascular responses either at the peripheral or at central level in experimental animals. However, there are no dedicated reviews on the involvement of adenosine in mediating the hypotensive response of centrally administered clonidine in general and specifically in aortically barodenervated rats (ABD). The conscious ABD rat model exhibits surgically induced baroreflex dysfunction and exaggerated hypotensive response, compared with conscious sham-operated (SO) rats. The current review focuses on, the role of adenosine receptors in blood pressure (BP) regulation and their possible crosstalk with other receptors e.g. imidazoline (I₁) and alpha (α_{2A}) adrenergic receptor (AR). The former receptor is a molecular target for clonidine, whose hypotensive effect is enhanced approx. 3-fold in conscious ABD rats. We also discussed how the balance between the brain stem adenosine A_1 and A_{2A} receptors is regulated by baroreceptors and how such balance influences the centrally mediated hypotensive responses. The use of the ABD rat model yielded insight into the downstream signaling cascades following clonidine-evoked hypotension in a surgical model of baroreflex dysfunction.

© 2014 Production and hosting by Elsevier B.V. on behalf of Cairo University.



Noha Nassar is an Associate Professor of Pharmacology and Toxicology, Faculty of Pharmacy, Cairo University. She has 20 refereed scientific papers in addition to many poster and talks at international conventions. Her research findings have been published in top journals and have received fair citations. Dr. Nassar's research interests focus on signaling mechanisms and mediators implicated in neurodegenerative diseases as well as those involved in neural regulation of circulation.

In addition to her contributions to research, Dr. Nassar has been active as a member of many scientific societies for the past 10 years including The American Society for Pharmacology and Experimental Therapeutics, and Society for Neuroscience and has served as a reviewer for a number of scientific journals.



Dr. Abdel-Rahman is Distinguished Professor of Pharmacology and Vice Chair of the Department of Pharmacology and Toxicology, Brody School of Medicine at East Carolina University, Greenville, NC, USA. He published over 120 refereed scientific papers in addition to 10 education-related articles. His research findings have been published in top journals in his discipline and cited hundreds of times in scientific literature. Dr. Abdel-Rahman research deals with neural control of

circulation and neurobiology of hypertension. Two National Institutes of Health grants fund his research. In the first project his research team investigates the effect of ethanol on neuronal pathways that control blood pressure and cardiac reflexes. The second project deals with the neuroprotective and cardioprotective actions of estrogen and how concurrent alcohol use might compromise these beneficial physiological effects of estrogen. In addition to his contributions to research, Dr. Abdel-Rahman has been active as a member of many scientific societies for the past 30 years and has been named a Fellow of the American Heart Association. Dr. Abdel-Rahman also served as President of the East Carolina University Neuroscience Chapter in addition to his services as editor/associate editor and reviewer for a number of scientific journals. He has also served as a member of review boards (study sections) of the National Institutes of Health and the American Heart Association.

Introduction

The current review focuses on, the role of adenosine receptors in BP regulation and their possible crosstalk with other receptors e.g. imidazoline (I₁) and α_{2A} AR in a rat model of surgicallyinduced baroreflex dysfunction, the ABD rat. Furthermore, the current review delineates the role of the downstream adenosinergic-signaling pathway in mediating the centrally evoked hypotension elicited by clonidine and clonidine such as drugs. The review covers data generated in our laboratory and reported pertinent studies over the past 10 years, which covered the following: (i) imidazoline I₁-receptor and centrally mediated hypotension; (ii) clonidine and aortic barodenervation; (iii) clonidine and SHR rats; (iv) clonidine effects in the RVLM; (v) clonidine effects in the NTS; (vi) central adenosine receptor signaling; and (vii) central MAPK-NOS signaling.

The nucleus of the solitary tract (NTS)

The NTS mediates inhibitory actions of baroreceptors on sympathetic discharge and is considered the main site of termination of the baroreceptor afferent fibers via both the aortic depressor nerve and the glossopharyngeal (IX) from the carotid sinus [1-3]. Notably, lesions to the NTS abolish the baroreflex responses [3]. Several reports have shown important roles for activation of NTS glutamate [1,3] as well as adenosine receptors in BP regulation [1,2,4–7].

Rostral and caudal ventrolateral medulla

A large body of evidence supports the view that the RVLM is the major brain stem area that controls sympathetic drive by projecting directly to the spinal cord [1,8–10]. Neuronal activation in the RVLM causes an increase in arterial pressure mediated by an increase in peripheral resistance, cardiac output, and secretion of catecholamines [1]. Electrical and chemical stimulation of the RVLM produces immediate and marked increases in arterial pressure. The direct connection with the

sympathetic preganglionic neurons explains why an alteration in the RVLM neuronal activity dramatically influences sympathetic neuronal discharge (SND) and arterial pressure (AP) [1,8–10]. The RVLM-spinal neuronal connection plays at least two important roles in sympathetic and cardiovascular control. First, RVLM-spinal neurons set the tone for AP by providing a basal SND. This tone generating ability explains why chemical inhibition or lesioning of the RVLM causes a dramatic fall in arterial pressure [1]. Second, a dominant aspect of the RVLM neurons is the control of the baroreflex response. By serving as a major neuroanatomical target for centrally acting antihypertensive agents including clonidine, moxonidine, and rilmenidine [3], the RVLM plays a fundamental role in BP regulation and in the control of BP in treated hypertensives. Similar to the NTS, the RVLM expresses receptors including the adenosine, α_{2A} adrenergic and imidazoline receptors [11,12]. It is not surprising that the RVLM shares with the NTS a similar receptor population since it receives inhibitory projections from the NTS and is involved in mediating baroreceptor efferent response via the sympathetic nervous system [1]. It must also be remembered that the caudal ventrolateral medulla (CVLM) plays important intermediate role between the NTS and RVLM, particularly in regulating the baroreflex function [3]. Unlike the anatomically and functionally (sympathoexcitatory) well defined neurons of the RVLM, the CVLM neurons are more heterogeneous and scattered [3]. However, functional and retrograde studies revealed projections from the NTS to the CVLM, which sends tonic sympathoinhibitory projections to the RVLM [3].

The aortic barodenervated (ABD) rat model

Various genetic models of hypertension, knockout mice, pheochromocytoma (PC12) cells and anesthetized animals have been used extensively to outline the signaling cascades triggered by adenosine, imidazoline (I₁) and α_{2A} adrenergic receptor activation [13–26]. However, little is known about the role of these receptors in BP control or BP responses to centrally acting drugs in conscious rats. Notably, clonidine-evoked hypotension is evident in conscious or anesthetized hypertensive rats [27–29], but only occurs in anesthetized normotensive rats [25,30]. In conscious intact rats, the hypotensive response elicited by clonidine is virtually absent in marked contrast to the case in the conscious aortic barodenervated rats. Following denervation, acute rises in BP, heart rate, and peripheral resistance are apparent in the ABD rat while cardiac index and stroke volume were not altered. Forty-eight hours later, when cardiovascular measurements were conducted in the absence of anesthesia, the reductions in cardiac index and stroke volume were paralleled by a return of the BP of conscious ABD rats to sham-operated levels while the peripheral resistance remained significantly elevated. Compared to sham operated rats, clonidine (30 µg/kg, i.v.) elicited greater decreases in BP in ABD rats via decreases in cardiac index and stroke volume because peripheral resistance did not change [31-33]. However, these studies focused on the role of baroreceptor dysfunction and sympathetic nervous system over-activity as underlying causes for the enhanced response to some centrally acting hypotensive drugs [31,34]. Other reported studies built on these findings to delineate the central pathways and cellular mechanisms implicated in this response in the ABD rat. Specifically, this review focuses on studies that elucidated the role of central adenosine receptor signaling in the conscious ABD rat model and their involvement in centrally mediated hypotension.

Adenosine receptors in the CNS

The high affinity A_1 and the A_{2A} receptors in the brain are tonically activated by extracellular adenosine, which set the basal "purinergic" tone seen in most systems. This notion is supported by the ability of caffeine to antagonize the actions of endogenous adenosine and reversing the tonic inhibition [35]. Four different adenosine receptors have been characterized pharmacologically, structurally and functionally and are denoted A_1 , A_{2A} , A_{2B} and A_3 [35–37].

Role of central adenosine receptors in blood pressure control

The primary neurons that regulate sympathetic outflow located in the NTS and the RVLM, express adenosine receptors [1,2,12,13,23,38-43]. While activation of the A₁ receptor by adenosine, or by the more selective agonist N⁶-cyclopentyl-adenosine, causes a pressor response, adenosine A_{2A} receptor activation by adenosine, or by the more selective agonist, 2-*p*-(2-carboxyethyl) phenylethylamino-5'-N-ethylcarboxamido-adenosine (CGS21680), causes a depressor response [2,44-47].

Adenosine receptor signal transduction mechanisms

The original delineation of adenosine receptors is based on their regulation of cyclic adenosine monophosphate (cAMP) levels. The A_1 and A_3 receptors mediate a reduction in cAMP via $G_{\alpha i/o}$ whereas the A_{2A} receptor mediates elevation in cAMP via $G_{\alpha s}$ [20,48–50]. Notably, the A_{2A} and A_{2B} are also linked to $G_{\alpha\alpha}$ and the activation of PKC [20,21,51]. Contrary to previous views where receptor activation leads to a sequential downstream signaling paradigm, recent evidence suggests that single receptor activation may converge on a multitude of downstream signaling cascades. In line with this concept, adenosine receptor activation results in the phosphorylation of the mitogen-activated protein kinase (MAPK p44/42), also known as pERK1/2, through either PLC-DAG or the PKA pathways [20]. The well-conserved and diverse MAPK family, which covers three main groups, the extracellular signal-regulated protein kinases (ERK), the stress-activated protein kinases (SAPK; p38) and the c-Jun N-terminal kinases (JNK), is involved in cell cycle progression, proliferation and differentiation in all organisms including mammals. Adenosine receptor signaling may enhance or inhibit proliferation of a variety of cell types depending on the adenosine receptor (or combination of adenosine receptor) subtypes and the tissue type. All adenosine receptors activate at least one MAPK. For example, the $G_{\alpha s}$ -coupled adenosine A_{2A} receptor activation enhances ERK1/2 phosphorylation as summarized in Fig. 1.

Reported studies including ours implicated central adenosine receptors in BP modulation in at least some forms of hypertension. Microinjection of adenosine into the nucleus tractus solitarius (NTS) elicited enhanced depressor and reduced pressor responses in the SHR compared to its

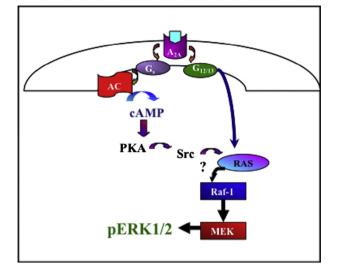


Fig. 1 ERK1/2 activation following stimulation of A_{2A} receptor based on findings obtained from receptor transfected CHO cells, PC12 cells and human endothelial cells. Abbreviations are: cyclic adenosine monophosphate (cAMP); protein kinase A (PKA); Raf-1 is a serine/threonine-specific kinase (Raf-1); G protein (specifically a small GTPase) (RAS). Adopted with modification [20].

respective control, the WKY rat [13]. These findings inferred alteration in the central adenosine receptor signaling as a result of hypertension or due to baroreceptor dysfunction, which is a hallmark of hypertension. As detailed below, similar alterations occur in adenosine receptor function in the aortic barodenervated (ABD) rat, which shares with the SHR a reduced baroreceptor function [31,52,53]. These findings suggest a functional link between baroreceptor function/dysfunction and central adenosine receptor signaling in the ABD rat model.

Imidazoline receptors and centrally acting antihypertensive agents

In clinical or experimental hypertension, central sympatholytics such as clonidine, rilmenidine and moxonidine reduce sympathetic tone and renin release, which ultimately reduces peripheral resistance and BP [54]. These centrally acting medications lower BP primarily by targeting the RVLM neurons in the brain stem to cause inhibition of the activity of bulbospinal sympathoexcitatory presympathetic neurons [11,55]. Additionally, clonidine-like drugs can reduce norepinephrine released by activating peripheral presynaptic α_{2A} adrenergic receptors on axon terminals of postganglionic sympathetic neurons [55].

There has been an ongoing debate regarding the primary target in the medulla oblongata that is mediating the central sympathoinhibitory action of central sympatholytic drugs. Originally, for clonidine-like drugs, it was thought that the primary target was the α_{2A} AR. However, in 1984, Bousquet et al. [56] proposed that activation of the imidazoline I₁ receptor in the RVLM accounts for the central sympathoinhibition caused by clonidine. The fact that direct administration of α -adrenergic receptor agonists with a phenylethylamine structure into the RVLM did not mimic the effects of agonists with imidazoline structure supported the imidazoline receptor hypothesis [56,57]. Further, blockade of the α_{2A} AR in the RVLM did

not reverse the hypotension elicited by local imidazoline I_1 receptor activation [58,59]. On the contrary, the hypotensive action of clonidine analogues was attenuated by microinjections of idazoxan or efaroxan, antagonists with imidazoline structures, into RVLM [11,60-62]. Several imidazoline preferring compounds such as rilmenidine and moxonidine possess preferential binding to the I₁ receptor over the α_{2A} AR compared to clonidine, which is a mixed I_1/α_{2A} AR agonist [11,55,63–65]. However, functional studies in α_{2A} AR knockouts have shown that despite rilmenidine and moxonidine I_1R selectivity, the α_{2A} AR is an important mediator of their hypotensive action [58,66-69]. Other studies have suggested synergy between the α_{2A} AR and the I₁ receptor signaling pathways [14,15]. The imidazoline binding site is a separate entity based on binding and functional studies that demonstrated the ability of selective I1 receptor agonists (LNP509) to lower BP when microinjected into the brain stem of D79N mice [14,68,70]. D79N mice constitute a functional α_{2A} AR knockout model, which has been useful in elucidating the role of α_{2A} AR in several functions including hypotension and sedation [71].

Although it is not known whether the I₁ and α_{2A} AR are operating in parallel or in series, there is evidence that the I_1 receptor downstream signaling is distinct from that of the α_{2A} AR receptor. Several reports have shown that in PC12 cells, which exhibit neuronal phenotype when differentiated, activation of the I₁ receptor involves the phosphatidylcholine-selective phospholipase-C (PC-PLC) and PKC (β_{11} and ζ isoforms) pathway and the increased formation of the second messenger diacylglycerol (DAG). As a consequence of the activation of PKC, ERK1/2 phosphorylation is increased [19,72-74]. These cellular events contribute to I₁ (rilmenidine) mediated hypotension because similar to I₁ receptor blockade (efaroxan), PC-PLC (D609), or pERK1/2 (PD98059) inhibition abrogated the hypotensive response and the corresponding cellular events elicited by the I_1 receptor activation [18,19,22,72,74]. Noteworthy, other neuromodulators in the CNS, including L-glutamate and adenosine, which also enhance ERK1/2 phosphorylation [20,75] might be implicated in I₁ receptor signaling. In support of this notion, L-glutamate release increases following clonidine or rilmenidine administration [17,76–78] and L-glutamate releases adenosine [79,80] (Fig. 2).

Crosstalk between adenosine and imidazoline receptors signaling underlies clonidine-evoked hypotension in conscious ABD rats

Evidence for the involvement of central adenosine receptors in clonidine-evoked hypotension is supported by a number of pharmacological studies. The finding that systemic administration of theophylline virtually abolished the hypotensive effect of clonidine inferred a central interaction of these two drugs because clonidine lowers BP via a central mechanism of action [81], and theophylline gains access to the CNS to block central adenosine receptors [13]. This finding was consolidated by the observation that intracisternal, but not systemic, administration, of the water-soluble adenosine receptor blocker 8-p-sulphophenyl-theophylline (8-SPT) attenuated clonidine-evoked hypotension. The inability of systemic 8-SPT, which blocks peripheral, but not central, adenosine receptors [13,23] to influence clonidine-evoked hypotension [82] bolsters the conclusion that central adenosine receptors are implicated in clonidine-

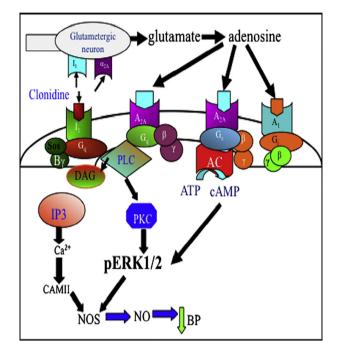


Fig. 2 Schematic overview of a potential I_1 and adenosine receptors crosstalk. Reference is made to the signaling cascades of the adenosine A_1 and A_{2A} receptor subtypes. Adenosine A_{2A} receptor activation with the nonselective agonist adenosine or the more selective agonist CGS21680 leads to enhanced expression of pERK1/2 via a cAMP-dependent or independent mechanism. I_1 activation with its respective agonist, clonidine or rilmenidine, enhances expression of pERK1/2 via a PC-PLC pathway. pERK1/2 activates neuronal nitric oxide synthase (NOS) which causes increased production of NO and decreased sympathetic neuronal activity.

evoked hypotension. Further, central administration of SCH58261, a selective A_{2A} receptor blocker [83,84], virtually abolished the clonidine-evoked hypotension [82]. Together, these findings suggest the dependence of clonidine-evoked hypotension on central adenosine A_{2A} receptor.

Although Bousquet et al. [85] classified clonidine as ligand at the imidazoline-binding site, clonidine is still considered a mixed I_1/α_{2A} AR agonist [27,85]. Therefore, it was difficult to ascertain the type of receptor, I_1 or α_{2A} , whose activation triggers central adenosine signaling. Findings from our laboratory indicate that the central hypotensive response elicited by selective activation of the central I₁ (rilmenidine) or α_{2A} (α -MNE) receptor was attenuated by central adenosine receptor blockade [82]. It is imperative to note that although α -MNE is considered a "pure" α_{2A} receptor agonist [65], the selective I₁ agonist rilmenidine also exhibits α_{2A} agonist activity [27,55]. Together, these findings raise the interesting possibility that α_{2A} receptor activation might also trigger central adenosine receptor signaling [82]. However, an alternative explanation is that I₁ activation by rilmenidine might depend on a downstream α_{2A} AR activation as proposed by Head [66]. Collectively, these findings suggest that the adenosinergic system plays a critical role in mediating centrally mediated hypotension. However, the use of non-selective adenosine receptor blockers (theophylline or 8-SPT) in these earlier studies precluded ascertaining the adenosine receptor subtype implicated in the mediation of clonidine-evoked hypotension. Building on the A_{2A} receptor as a viable candidate because its activation within the brain stem leads to hypotension [4], data from our laboratory confirmed A_{2A} involvement because the selective A_{2A} receptor antagonist SCH58261 virtually abolished clonidine-evoked hypotension in conscious ABD rats [82].

Reciprocal roles for central \mathbf{A}_1 and \mathbf{A}_{2A} in blood pressure regulation

A number of studies including ours demonstrated functionally opposite roles for central A_{2A} and A_1 adenosine receptors in BP regulation because they mediate depressor, and pressor responses, respectively [2,4,6,47]. These findings lead to the postulate that concomitant activation of the adenosine A_1 receptor might counterbalance (mask) the adenosine A_{2A} dependent hypotensive action of clonidine, as discussed above.

Our laboratory showed that upregulations of α_{2A} AR and I1 receptors were paralleled with similar A2A receptor upregulation in the same brain stem areas of the ABD rat model [29]. The latter confirms and extends earlier findings, which demonstrated the upregulation of α_2 and I_1 receptors in the same animal model [32,86]. It might be argued that aortic barodenervation caused nonspecific upregulation of adenosine A_{2A} as well as the $\alpha_{2A}\;AR$ and I_1 receptors because they followed the same pattern in the investigated brain stem nuclei. However, such parallel upregulation might be physiologically relevant because: (i) the A_{2A} receptor, the α_{2A} and the I₁ receptors in the NTS and RVLM are spatially associated, (ii) all three receptors mediate hypotension [2,55,65,86], and (iii) their shared signaling pathways make it highly likely that these receptors physiologically interact [18-20]. These findings are consistent with a key role for central adenosine A2A in clonidine evoked hypotension in conscious ABD rats [82].

Overexpressed a denosine $\mathbf{A}_{2\mathbf{A}}$ receptor in brain stem is functionally relevant

Immunohistochemical evidence demonstrated approximately twofold increase in the number of A_{2A} receptors in the NTS and RVLM of ABD, compared to SO, rats [87]. These findings were functionally relevant because the selective adenosine A_{2A} agonist CGS21680 elicited significantly greater dose-dependent hypotensive responses in the ABD, compared to SO, rats [29]. Notably, particularly in the NTS and RVLM, the A_{2A} receptor activation produces sympathoinhibition and hypotension [2,12,47], which are shared by clonidine and similar drugs [18,31,82,86]. Together, these findings establish a link between the anatomical and functional upregulation of brain stem adenosine A_{2A} receptor in the ABD rat [31,32]. Equally important, these findings might explain, at least partly, the enhanced hypotensive response elicited by clonidine in ABD rats [31,82] and its dependence on central adenosine A_{2A} receptor signaling [82].

ERK1/2-NOS activation underlies centrally mediated hypotension

As discussed earlier, ERK1/2 phosphorylation constitutes important signaling event in clonidine-evoked hypotension. Noteworthy, pERK1/2 involvement in I_1 receptor-evoked hypotension has been based on two findings: (i) pERK1/2 expression in the RVLM is enhanced in association with centrally mediated hypotension elicited by rilmenidine, but not by α -methylnorepinephrine [18] and (ii) the ERK1/2 phosphorylation inhibitor PD98059 significantly attenuated rilmenidineevoked hypotension [18]. By the same token, the exaggerated hypotensive response elicited by central A_{2A} receptor activation with i.c. CGS21680 in ABD rats might involve enhancement of ERK1/2 phosphorylation [87]. Further, central A2A receptor blockade, which virtually abolished clonidine-evoked hypotension [82], abrogated the associated increase in brain stem ERK1/2 phosphorylation (pERK1/2). The latter findings suggest the involvement of the A2A receptor signaling in the centrally evoked hypotensive response elicited by clonidine and other I1R agonists. It was reasoned that NOS activation (phosphorylation) is triggered by pERK1/2 based on an established signaling pathway in cultured cells [88,89], and because NOS-derived NO causes sympathoinhibition and hypotension [90]. This intriguing possibility is supported: (i) by pharmacologic inhibition of ERK1/2 phosphorylation attenuated clonidine-evoked hypotension and **ERK1/2** and NOS phosphorylation in the RVLM and (ii) while L-NAME abrogated clonidine-evoked hypotension without affecting the enhanced ERK1/2 phosphorylation in the RVLM [87]. These findings are consistent with a role for pERK1/2 as an upstream activator of NOS [87,91] and bolster the conclusion that pERK1/2 plays a pivotal role in centrally-mediated hypotension via downstream NOS activation (enhanced NO production). Further, these reported findings rule out the possibility that ERK1/2 phosphorylation was consequence of clonidineevoked hypotension in the ABD model system. Together, these findings delineate the molecular events in the brain stem triggered by central adenosine A2A receptor activation and suggest a biological relevance for the pERK1/2-NOS pathway in-vivo. By contrast, we showed that the latter signaling pathway contributes to the central CB₁R-mediated pressor response [92] via GABA dependent mechanisms. Future studies are needed to address this controversy because the adenosine A_2 receptors are expressed on GABAergic neurons of the medulla oblongata of the developing rat brain.

Why clonidine fails to lower BP in conscious normotensive rats?

Many reported findings, including ours showed that clonidine does not lower BP [31,34,93,94] or influence ERK1/2 phosphorylation in the NTS and RVLM [87] in conscious normotensive rats. By contrast, as discussed above, clonidine enhances pERK1/2 expression and lowers BP in conscious ABD rats via adenosine A_{2A} receptor dependent mechanisms. These findings set forth the postulate that concomitant adenosine A₁ receptor activation serves a negative (counterbalancing) role against adenosine A2A receptor signaling triggered by clonidine in conscious normotensive rats. In support of this hypothesis are the findings that clonidine significantly reduced BP and increased brain stem pERK1/2 expression following central adenosine A1 receptor blockade (DPCPX) in conscious normotensive rats [29]. Interestingly, these molecular and BP responses were similar to those elicited by clonidine in ABD rats [31,82]. Collectively, these findings support a dampening role for central adenosine A1 receptor against clonidineevoked hypotension and advance our knowledge in this area

of research because central adenosine A_1 receptor blockade (i) unmasked clonidine-evoked hypotension and the enhanced phosphorylation of brain stem pERK1/2 in conscious normotensive rats and (ii) had no effect on the neurochemical (pERK1/2) or the hypotensive response elicited by clonidine in ABD rats. These findings are consistent with opposite roles for central A_1 (pressor) and A_{2A} (depressor) receptor activation [2,6] and further support a pivotal role for brain stem pERK1/2 in the hypotensive action of clonidine and similar drugs [18].

Finally, it is imperative to comment on the differential expression of the adenosine A_1 receptor in the NTS and RVLM of SO and ABD rats. We demonstrated an inverse relationship between the level of adenosine A_1 receptor expression and the BP response to clonidine [29] in marked contrast to a direct relationship between A_{2A} receptor expression in the same brain nuclei and the hypotensive effect of clonidine in ABD rats [29,87]. It is likely, therefore, that the balance between the A_1 and the A_{2A} adenosine receptor populations

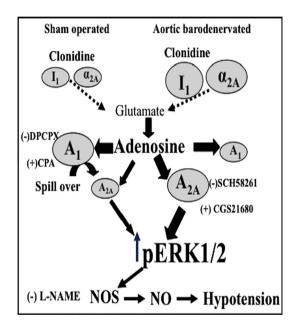


Fig. 3 Conceptual overview of the major findings discussed in this review. Upregulation of A_{2A} (large circle) and the molecular targets for clonidine $(I_1/\alpha_{2A},\mbox{ large circles})$ are more evident in ABD rats (right hand side) compared to sham-operated, SO, rats (left hand side, small circles). Note the downregulation of A₁ (small circle) in ABD compared to SO rats (large circle) in the NTS and RVLM. Direct (CGS21680) or indirect (clonidine) central A2A activation enhances pERK1/2 expression, which subsequently phosphorylates NOS (increased NO) and ultimately reduces BP. Blockade of central A2A receptor (SCH58261) or inhibition of NOS (L-NAME) abrogated clonidine-evoked hypotension, but only the former abrogated clonidine-evoked elevation in pERK1/2 expression. Intracisternal A1 receptor blockade (DPCPX) (large circle) unravels clonidine-evoked hypotension and enhances pERK1/2 expression in conscious normotensive rats. Central A1 receptor is downregulated in the NTS and RVLM (small circle) in ABD compared to SO rats (large circle), which is paralleled by an attenuated pressor response to adenosine A₁ receptor activation (CPA) in ABD, compared to SO, rats.

in the brain stem determines the magnitude of the BP response elicited by clonidine and perhaps other centrally acting drugs. Tipping the balance toward adenosine A2A dominance might explain the enhanced clonidine-evoked hypotension in conscious ABD rats [82,87] and SHRs [13]. It is also important to discuss the role of the NTS adenosine A₁ receptor in BP regulation and how it might be impacted by anesthesia. In general, anesthesia dampens the NTS A1-mediated pressor response because Machado and de Paula [95] showed that intra-NTS adenosine produced pressor response via activation of the local A₁ receptor in conscious rats. These findings explain, at least partly, why systemic or intracisternal clonidine lowers BP in anesthetized, but not in conscious rats. Consistent with this knowledge, as discussed above, suppression of adenosine A_1 (and concomitant upregulation of A_2) receptors in the brain stem occurs in the ABD rat and clonidine lowers BP in this animal model in the conscious state [82]. Nonetheless, the NTS neurons are heterogeneous because our reported studies showed that under the same experimental condition (anesthetized rats), microinjection of adenosine into the rostral and caudal NTS produced pressor and depressor responses, respectively [13]. Whether the adenosine A_1/A_2 ratios are different in these two subareas of the NTS remains to be elucidated.

Conclusions

The reviewed pharmacological and molecular findings support a differential role of adenosine A2A and A1 receptors in mediating and opposing clonidine-evoked hypotension, respectively. This review also provides a brief account on the role of pERK1/2-NOS-NO activation in brain stem nuclei as a molecular mechanism for the centrally mediated hypotension elicited by direct and indirect activation of the central A2A receptor by CGS21680 and clonidine, respectively. Further, the reviewed findings support the conclusion that pERK1/2is a mediator and not a result of the hypotension elicited by direct or indirect A_{2A} receptor activation. This is the first review that discussed the novel mechanism that central A₁ receptor signaling masks clonidine-evoked hypotension in conscious normotensive rats (summarized in Fig. 3). Since clonidine is clinically used for the management of hypertension, possible drug interactions with the adenosine agonists and antagonists that cross the blood brain barrier might have clinical implications.

Conflict of interest

The authors have declared no conflict of interest.

Compliance with Ethics Requirements

This article does not contain any studies with human or animal subjects.

Acknowledgment

This study was supported in part by National Institutes of Health Grant 2R01 AA07839.

References

- Sun MK. Central neural organization and control of sympathetic nervous system in mammals. Prog Neurobiol 1995;47(3):157–233.
- [2] Scislo TJ, O'Leary DS. Purinergic mechanisms of the nucleus of the solitary tract and neural cardiovascular control. Neurol Res 2005;27(2):182–94.
- [3] Colombari E, Sato MA, Cravo SL, Bergamaschi CT, Campos Jr RR, Lopes OU. Role of the medulla oblongata in hypertension. Hypertension 2001;38(3 Pt 2):549–54.
- [4] Scislo TJ, Kitchen AM, Augustyniak RA, O'Leary DS. Differential patterns of sympathetic responses to selective stimulation of nucleus tractus solitarius purinergic receptor subtypes. Clin Exp Pharmacol Physiol 2001;28(1–2):120–4.
- [5] Scislo TJ, O'Leary D S. Vasopressin V1 receptors contribute to hemodynamic and sympathoinhibitory responses evoked by stimulation of adenosine A2a receptors in the NTS. Am J Physiol Heart Circ Physiol 2005.
- [6] Scislo TJ, O'Leary DS. Mechanisms mediating regional sympathoactivatory responses to stimulation of NTS A(1) adenosine receptors. Am J Physiol Heart Circ Physiol 2002;283(4):H1588–99.
- [7] Scislo TJ, Tan N, O'Leary DS. Differential role of nitric oxide in regional sympathetic responses to stimulation of NTS A2a adenosine receptors. Am J Physiol Heart Circ Physiol 2005;288(2):H638–49.
- [8] Guyenet PG. The sympathetic control of blood pressure. Nat Rev Neurosci 2006;7(5):335–46.
- [9] Kumagai H, Oshima N, Matsuura T, Iigaya K, Imai M, Onimaru H, et al. Importance of rostral ventrolateral medulla neurons in determining efferent sympathetic nerve activity and blood pressure. Hypertens Res 2012;35(2):132–41.
- [10] Braga VA, Colombari E, Jovita MG. Angiotensin II-derived reactive oxygen species underpinning the processing of the cardiovascular reflexes in the medulla oblongata. Neurosci Bull 2011;27(4):269–74.
- [11] Head GA, Mayorov DN. Imidazoline receptors, novel agents and therapeutic potential. Cardiovasc Hematol Agents Med Chem 2006;4(1):17–32.
- [12] Thomas T, St Lambert JH, Dashwood MR, Spyer KM. Localization and action of adenosine A2a receptors in regions of the brainstem important in cardiovascular control. Neuroscience 2000;95(2):513–8.
- [13] Abdel-Rahman AA, Tao S. Differential alteration of neuronal and cardiovascular responses to adenosine microinjected into the nucleus tractus solitarius of spontaneously hypertensive rats. Hypertension 1996;27(4):939–48.
- [14] Bruban V, Estato V, Schann S, Ehrhardt JD, Monassier L, Renard P, et al. Evidence for synergy between alpha(2)adrenergic and nonadrenergic mechanisms in central blood pressure regulation. Circulation 2002;105(9):1116–21.
- [15] Bruban V, Feldman J, Greney H, Dontenwill M, Schann S, Jarry C, et al. Respective contributions of alpha-adrenergic and non-adrenergic mechanisms in the hypotensive effect of imidazoline-like drugs. Br J Pharmacol 2001;133(2):261–6.
- [16] Arslan G, Kull B, Fredholm BB. Signaling via A2A adenosine receptor in four PC12 cell clones. Naunyn Schmiedebergs Arch Pharmacol 1999;359(1):28–32.
- [17] Zhang J, Abdel-Rahman AA. The hypotensive action of rilmenidine is dependent on functional N-methyl-D-aspartate receptor in the rostral ventrolateral medulla of conscious spontaneously hypertensive rats. J Pharmacol Exp Ther 2002;303(1):204–10.
- [18] Zhang J, Abdel-Rahman AA. Mitogen-activated protein kinase phosphorylation in the rostral ventrolateral medulla plays a key

role in imidazoline (i1)-receptor-mediated hypotension. J Pharmacol Exp Ther 2005;314(3):945–52.

- [19] Zhang J, El-Mas MM, Abdel-Rahman AA. Imidazoline I(1) receptor-induced activation of phosphatidylcholine-specific phospholipase C elicits mitogen-activated protein kinase phosphorylation in PC12 cells. Eur J Pharmacol 2001;415(2– 3):117–25.
- [20] Schulte G, Fredholm BB. Signalling from adenosine receptors to mitogen-activated protein kinases. Cell Signal 2003;15(9):813–27.
- [21] Schulte G, Fredholm BB. The G(s)-coupled adenosine A(2B) receptor recruits divergent pathways to regulate ERK1/2 and p38. Exp Cell Res 2003;290(1):168–76.
- [22] Separovic D, Kester M, Haxhiu MA, Ernsberger P. Activation of phosphatidylcholine-selective phospholipase C by Ilimidazoline receptors in PC12 cells and rostral ventrolateral medulla. Brain Res 1997;749(2):335–9.
- [23] Tao S, Abdel-Rahman AA. Neuronal and cardiovascular responses to adenosine microinjection into the nucleus tractus solitarius. Brain Res Bull 1993;32(4):407–17.
- [24] Yamamoto J. Cardiovascular response to acute stress in spontaneously hypertensive rats. Hypertension 1987;10(5):550.
- [25] Yang J, Wang WZ, Shen FM, Su DF. Cardiovascular effects of agmatine within the rostral ventrolateral medulla are similar to those of clonidine in anesthetized rats. Exp Brain Res 2005;160(4):467–72.
- [26] Barac YD, Bar-Am O, Liani E, Amit T, Frolov L, Ovcharenko E, et al. II imidazoline receptor: novel potential cytoprotective target of TVP1022, the S-enantiomer of rasagiline. PLoS One 2012;7(11):e47890.
- [27] Estato V, Araujo CV, Bousquet P, Tibirica E. Effects of centrally acting antihypertensive drugs on the microcirculation of spontaneously hypertensive rats. Braz J Med Biol Res 2004;37(10):1541–9.
- [28] Jastrzebski Z, Czyzewska-Szafran H, Gozlinska B, Remiszewska M, Mazurek AP. Clonidine hypotension in spontaneously hypertensive rats (SHR) depends on the functional state of GABAergic and glutamatergic systems. Neurosci Lett 1995;184(2):94–6.
- [29] Nassar N, Abdel-Rahman AA. Brainstem adenosine A1 receptor signaling masks phosphorylated extracellular signalregulated kinase 1/2-dependent hypotensive action of clonidine in conscious normotensive rats. J Pharmacol Exp Ther 2009;328(1):83–9.
- [30] El-Mas MM, Tao S, Carroll RG, Abdel-Rahman AA. Ethanolclonidine hemodynamic interaction in normotensive rats is modified by anesthesia. Alcohol 1994;11(4):307–14.
- [31] Abdel-Rahman AA. Aortic baroreceptors exert a tonically active restraining influence on centrally mediated depressor responses. J Cardiovasc Pharmacol 1992;19(2):233–45.
- [32] El-Mas MM, Abdel-Rahman AA. Aortic barodenervation upregulates alpha2-adrenoceptors in the nucleus tractus solitarius and rostral ventrolateral medulla: an autoradiographic study. Neuroscience 1997;79(2):581–90.
- [33] Medvedev OS, Kunduzova OR, Murashev AN, Medvedeva NA. Influence of sino-aortic barodenervation on the cardiovascular effects of imidazoline-like drugs. J Auton Nerv Syst 1998;72(2–3):205–9.
- [34] Bonham AC, Trapani AJ, Portis LR, Brody MJ. Studies on the mechanism of the central antihypertensive effect of guanabenz and clonidine. J Hypertens Suppl 1984;2(3):S543–6.
- [35] Dunwiddie TV, Masino SA. The role and regulation of adenosine in the central nervous system. Annu Rev Neurosci 2001;24:31–55.
- [36] Fredholm BB, AP IJ, Jacobson KA, Klotz KN, Linden J. International union of pharmacology. XXV. Nomenclature and classification of adenosine receptors. Pharmacol Rev 2001;53(4): 527–52.

- [37] Poulsen SA, Quinn RJ. Adenosine receptors: new opportunities for future drugs. Bioorg Med Chem 1998;6(6):619–41.
- [38] Chen S, He RR. Effect of intracarotid injection of adenosine on the activity of RVLM neurons in barodenervated rats. Sheng Li Xue Bao 1998;50(6):629–35.
- [39] Thomas T, Spyer KM. The role of adenosine receptors in the rostral ventrolateral medulla in the cardiovascular response to defence area stimulation in the rat. Exp Physiol 1996;81(1): 67–77.
- [40] Scislo TJ, O'Leary DS. Dkifferential control of renal vs. adrenal sympathetic nerve activity by NTS A2a and P2x purinoceptors. Am J Physiol 1998;275(6 Pt 2):H2130-9.
- [41] Scislo TJ, O'Leary DS. Activation of A2a adenosine receptors in the nucleus tractus solitarius inhibits renal but not lumbar sympathetic nerve activity. J Auton Nerv Syst 1998;68(3): 145–52.
- [42] Spyer KM, Lambert JH, Thomas T. Central nervous system control of cardiovascular function: neural mechanisms and novel modulators. Clin Exp Pharmacol Physiol 1997;24(9– 10):743–7.
- [43] St Lambert JH, Dashwood MR, Spyer KM. Role of brainstem adenosine A1 receptors in the cardiovascular response to hypothalamic defence area stimulation in the anaesthetized rat. Br J Pharmacol 1996;117(2):277–82.
- [44] Barraco RA, el-Ridi MR, Ergene E, Phillis JW. Adenosine receptor subtypes in the brainstem mediate distinct cardiovascular response patterns. Brain Res Bull 1991;26(1): 59–84.
- [45] Barraco RA, Janusz CJ, Polasek PM, Parizon M, Roberts PA. Cardiovascular effects of microinjection of adenosine into the nucleus tractus solitarius. Brain Res Bull 1988;20(1):129–32.
- [46] Barraco RA, Phillis JW. Subtypes of adenosine receptors in the brainstem mediate opposite blood pressure responses. Neuropharmacology 1991;30(4):403–7.
- [47] Scislo TJ, O'Leary DS. Adenosine receptors located in the NTS contribute to renal sympathoinhibition during hypotensive phase of severe hemorrhage in anesthetized rats. Am J Physiol Heart Circ Physiol 2006.
- [48] Freissmuth M, Selzer E, Schutz W. Interactions of purified bovine brain A1-adenosine receptors with G-proteins. Reciprocal modulation of agonist and antagonist binding. Biochem J 1991;275(Pt 3):651–6.
- [49] Akbar M, Okajima F, Tomura H, Shimegi S, Kondo Y. A single species of A1 adenosine receptor expressed in Chinese hamster ovary cells not only inhibits cAMP accumulation but also stimulates phospholipase C and arachidonate release. Mol Pharmacol 1994;45(5):1036–42.
- [50] Jockers R, Linder ME, Hohenegger M, Nanoff C, Bertin B, Strosberg AD, et al. Species difference in the G protein selectivity of the human and bovine A1-adenosine receptor. J Biol Chem 1994;269(51):32077–84.
- [51] Fresco P, Diniz C, Queiroz G, Goncalves J. Release inhibitory receptors activation favours the A2A-adenosine receptormediated facilitation of noradrenaline release in isolated rat tail artery. Br J Pharmacol 2002;136(2):230–6.
- [52] El-Mas MM, Abdel-Rahman AA. Role of aortic baroreceptors in ethanol-induced impairment of baroreflex control of heart rate in conscious rats. J Pharmacol Exp Ther 1992;262(1): 157–65.
- [53] Takeda K, Hayashi J, Itoh H, Hirata M, Nakata T, Oguro M, et al. Transection of aortic depressor nerve fails to raise blood pressure in spontaneously hypertensive rats. Cardiovasc Res 1989;23(7):573–6.
- [54] Dubar M, Pillion G. II agents: a new approach to the treatment of hypertension. Ann NY Acad Sci 1995;763:642–58.
- [55] Szabo B, Bock C, Nordheim U, Niederhoffer N. Mechanism of the sympathoinhibition produced by the clonidine-like drugs

rilmenidine and moxonidine. Ann NY Acad Sci 1999;881: 253-64.

- [56] Bousquet P, Feldman J, Schwartz J. Central cardiovascular effects of alpha adrenergic drugs: differences between catecholamines and imidazolines. J Pharmacol Exp Ther 1984;230(1):232–6.
- [57] Ernsberger P, Giuliano R, Willette RN, Reis DJ. Role of imidazole receptors in the vasodepressor response to clonidine analogs in the rostral ventrolateral medulla. J Pharmacol Exp Ther 1990;253(1):408–18.
- [58] Reis DJ, Piletz JE. The imidazoline receptor in control of blood pressure by clonidine and allied drugs. Am J Physiol 1997;273(5 Pt 2):R1569–71.
- [59] Bousquet P. Identification and characterization of I1 imidazoline receptors: their role in blood pressure regulation. Am J Hypertens 2000;13(6 Pt 2):84S–8S.
- [60] Chan CK, Head GA. Relative importance of central imidazoline receptors for the antihypertensive effects of moxonidine and rilmenidine. J Hypertens 1996;14(7):855–64.
- [61] Haxhiu MA, Dreshaj I, Schafer SG, Ernsberger P. Selective antihypertensive action of moxonidine is mediated mainly by Ilimidazoline receptors in the rostral ventrolateral medulla. J Cardiovasc Pharmacol 1994;24(Suppl 1):S1–8.
- [62] Feldman J, Tibirica E, Bricca G, Dontenwill M, Belcourt A, Bousquet P. Evidence for the involvement of imidazoline receptors in the central hypotensive effect of rilmenidine in the rabbit. Br J Pharmacol 1990;100(3):600–4.
- [63] Ernsberger P, Haxhiu MA. The I1-imidazoline-binding site is a functional receptor mediating vasodepression via the ventral medulla. Am J Physiol 1997;273(5 Pt 2):R1572–9.
- [64] Tolentino-Silva FP, Haxhiu MA, Waldbaum S, Dreshaj IA, Ernsberger P. Alpha(2)-adrenergic receptors are not required for central anti-hypertensive action of moxonidine in mice. Brain Res 2000;862(1–2):26–35.
- [65] Szabo B. Imidazoline antihypertensive drugs: a critical review on their mechanism of action. Pharmacol Ther 2002;93(1):1–35.
- [66] Head GA. Central imidazoline- and alpha 2-receptors involved in the cardiovascular actions of centrally acting antihypertensive agents. Ann NY Acad Sci 1999;881:279–86.
- [67] Bousquet P, Bruban V, Schann S, Greney H, Ehrhardt JD, Dontenwill M, et al. Participation of imidazoline receptors and alpha(2-)-adrenoceptors in the central hypotensive effects of imidazoline-like drugs. Ann NY Acad Sci 1999; 881:272–8.
- [68] Guyenet PG. Is the hypotensive effect of clonidine and related drugs due to imidazoline binding sites? Am J Physiol 1997;273(5 Pt 2):R1580–4.
- [69] Tan CM, Wilson MH, MacMillan LB, Kobilka BK, Limbird LE. Heterozygous alpha 2A-adrenergic receptor mice unveil unique therapeutic benefits of partial agonists. Proc Natl Acad Sci USA 2002;99(19):12471–6.
- [70] Ernsberger P, Friedman JE, Koletsky RJ. The I1-imidazoline receptor: from binding site to therapeutic target in cardiovascular disease. J Hypertens Suppl 1997;15(1):S9–S23.
- [71] Kable JW, Murrin LC, Bylund DB. In vivo gene modification elucidates subtype-specific functions of alpha(2)-adrenergic receptors. J Pharmacol Exp Ther 2000;293(1):1–7.
- [72] Ernsberger P. The I1-imidazoline receptor and its cellular signaling pathways. Ann NY Acad Sci 1999;881:35–53.
- [73] Separovic D, Kester M, Ernsberger P. Coupling of Ilimidazoline receptors to diacylglyceride accumulation in PC12 rat pheochromocytoma cells. Mol Pharmacol 1996;49(4):668–75.
- [74] Edwards L, Ernsberger P. The I(1)-imidazoline receptor in PC12 pheochromocytoma cells reverses NGF-induced ERK activation and induces MKP-2 phosphatase. Brain Res 2003;980(1):71–9.
- [75] Choe ES, Chung KT, Mao L, Wang JQ. Amphetamine increases phosphorylation of extracellular signal-regulated kinase and

transcription factors in the rat striatum via group I metabotropic glutamate receptors. Neuropsychopharmacology 2002;27(4):565–75.

- [76] Tingley 3rd FD, Arneric SP. Evidence for clonidine presynaptically modulating amino acid release in the rostral ventral medulla: role in hypertension. Brain Res 1990;537(1-2):175-81.
- [77] Wang WZ, Yuan WJ, Su DF. Blockade of N-methyl-D-aspartate receptors within the rostral ventrolateral medulla antagonizes clonidine-induced cardiovascular effects. Auton Neurosci 2003;109(1–2):21–8.
- [78] Wang WZ, Yuan WJ, Tang CS, Su DF. Electrophysiological evidences for the contribution of NMDA receptors to the inhibition of clonidine on the RVLM presympathetic neurons. Brain Res 2004;1023(1):163–6.
- [79] Iliff JJ, D'Ambrosio R, Ngai AC, Winn HR. Adenosine receptors mediate glutamate-evoked arteriolar dilation in the rat cerebral cortex. Am J Physiol Heart Circ Physiol 2003;284(5):H1631–7.
- [80] Paes-de-Carvalho R, Dias BV, Martins RA, Pereira MR, Portugal CC, Lanfredi C. Activation of glutamate receptors promotes a calcium-dependent and transporter-mediated release of purines in cultured avian retinal cells: possible involvement of calcium/calmodulin-dependent protein kinase II. Neurochem Int 2005;46(6):441–51.
- [81] van Zwieten PA. Renewed interest in centrally acting antihypertensive drugs. Cardiovasc J S Afr 2000;11(4):225–9.
- [82] Nassar N, Abdel-Rahman AA. Central adenosine signaling plays a key role in centrally mediated hypotension in conscious aortic barodenervated rats. J Pharmacol Exp Ther 2006;318(1):255–61.
- [83] Beukers MW, den Dulk H, van Tilburg EW, Brouwer J, Ijzerman AP. Why are A(2B) receptors low-affinity adenosine receptors? Mutation of Asn273 to Tyr increases affinity of human A(2B) receptor for 2-(1-Hexynyl)adenosine. Mol Pharmacol 2000;58(6):1349–56.
- [84] Ongini E, Dionisotti S, Gessi S, Irenius E, Fredholm BB. Comparison of CGS 15943, ZM 241385 and SCH 58261 as antagonists at human adenosine receptors. Naunyn Schmiedebergs Arch Pharmacol 1999;359(1):7–10.
- [85] Bousquet P, Greney H, Bruban V, Schann S, Ehrhardt JD, Monassier L, et al. I(1) imidazoline receptors involved in cardiovascular regulation: where are we and where are we going? Ann NY Acad Sci 2003;1009:228–33.
- [86] El-Mas MM, Abdel-Rahman AA. Upregulation of imidazoline receptors in the medulla oblongata accounts for the enhanced hypotensive effect of clonidine in aortic barodenervated rats. Brain Res 1995;691(1–2):195–204.
- [87] Nassar N, Abdel-Rahman AA. Brainstem phosphorylated extracellular signal-regulated kinase 1/2-nitric-oxide synthase signaling mediates the adenosine A2A-dependent hypotensive action of clonidine in conscious aortic barodenervated rats. J Pharmacol Exp Ther 2008;324(1):79–85.
- [88] Lee SA, Park JK, Kang EK, Bae HR, Bae KW, Park HT. Calmodulin-dependent activation of p38 and p42/44 mitogenactivated protein kinases contributes to c-fos expression by calcium in PC12 cells: modulation by nitric oxide. Brain Res Mol Brain Res 2000;75(1):16–24.
- [89] Wyatt AW, Steinert JR, Wheeler-Jones CP, Morgan AJ, Sugden D, Pearson JD, et al. Early activation of the p42/p44MAPK pathway mediates adenosine-induced nitric oxide production in human endothelial cells: a novel calcium-insensitive mechanism. Faseb J 2002;16(12):1584–94.
- [90] Chan SH, Wang LL, Wang SH, Chan JY. Differential cardiovascular responses to blockade of nNOS or iNOS in rostral ventrolateral medulla of the rat. Br J Pharmacol 2001;133(4):606–14.

- [91] Shen SC, Wu MS, Lin HY, Yang LY, Chen YH, Chen YC. Reactive oxygen species-dependent nitric oxide production in reciprocal interactions of glioma and microglial cells. J Cell Physiol 2014.
- [92] Ibrahim BM, Abdel-Rahman AA. Enhancement of rostral ventrolateral medulla neuronal nitric-oxide synthase-nitricoxide signaling mediates the central cannabinoid receptor 1evoked pressor response in conscious rats. J Pharmacol Exp Ther 2012;341(3):579–86.
- [93] El-Mas MM, Carroll RG, Abdel-Rahman AA. Blood pressure normalization in carotid barodenervated rats: role

of cardiac output. Can J Physiol Pharmacol 1993;71(10-11):783-90.

- [94] El-Mas MM, Carroll RG, Abdel-Rahman AA. Centrally mediated reduction in cardiac output elicits the enhanced hypotensive effect of clonidine in conscious aortic barodenervated rats. J Cardiovasc Pharmacol 1994;24(2): 184–93.
- [95] de Paula PM, Machado BH. Antagonism of adenosine A(1) receptors in the NTS does not affect the chemoreflex in awake rats. Am J Physiol Regul Integr Comp Physiol 2001;281(6): R2072-8.