

Structure of the Toll/Interleukin-1 Receptor (TIR) Domain of the B-cell Adaptor That Links Phosphoinositide Metabolism with the Negative Regulation of the Toll-like Receptor (TLR) Signalosome*[§]

Received for publication, October 2, 2016, and in revised form, November 15, 2016. Published, JBC Papers in Press, December 1, 2016, DOI 10.1074/jbc.M116.761528

Samer Halabi¹, Eiki Sekine, Brett Verstak, Nicholas J. Gay², and Martin C. Moncrieffe³

From the Department of Biochemistry, Cambridge University, Cambridge CB2 1GA, United Kingdom

Edited by Alex Tokor

Ligand binding to Toll-like receptors (TLRs) results in dimerization of their cytosolic Toll/interleukin-1 receptor (TIR) domains and recruitment of post-receptor signal transducers into a complex signalosome. TLR activation leads to the production of transcription factors and pro-inflammatory molecules and the activation of phosphoinositide 3-kinases (PI3K) in a process that requires the multimodular B-cell adaptor for phosphoinositide 3-kinase (BCAP). BCAP has a sequence previously proposed as a “cryptic” TIR domain. Here, we present the structure of the N-terminal region of human BCAP and show that it possesses a canonical TIR fold. Dimeric BCAP associates with the TIR domains of TLR2/4 and MAL/TIRAP, suggesting that it is recruited to the TLR signalosome by multitypic TIR-TIR interactions. BCAP also interacts with the p85 subunit of PI3K and phospholipase C γ , enzymes that deplete plasma membrane phosphatidylinositol 4,5-bisphosphate (PIP₂), and these interactions provide a molecular explanation for BCAP-mediated down-regulation of inflammatory signaling.

Phosphoinositide 3-kinases (PI3Ks) are crucial mediators of various cellular processes including growth, survival, proliferation, migration, metabolism, and membrane transport. Dysfunction in the PI3K signaling network is a major driver of oncogenesis (1–4). The well characterized class Ia PI3Ks phosphorylate primarily at the D-3 position of the inositol ring, creating a site that enables the interaction of proteins with the plasma membrane. Class Ia PI3Ks are heterodimers comprised of regulatory (p85, p55, or p50) and catalytic (p110) subunits

(5). The regulatory subunit not only represses the activity of the catalytic subunit in unactivated cells but mediates its activation by interacting directly with phosphorylated tyrosine residues present on activated growth factor receptors or adaptor molecules.

One such adaptor, B-cell adaptor for phosphoinositide 3-kinase (BCAP),⁴ functions in linking the B-cell receptor (BCR) and the co-receptor CD19 to the activation of PI3K via interaction with the SH2 domains on the regulatory p85 subunit (6). Architecturally, BCAP contains well defined ankyrin repeats and a Dof/BANK/BCAP (DBB) domain; the latter was first reported in the *Drosophila* protein Dof, where it is required for FGF-dependent signaling (7, 8), and a homologous domain exists in the B-cell scaffold with ankyrin repeat (BANK) protein (9).

The expression and cellular distribution of BCAP suggest that it is involved in immunological processes in addition to regulation of BCR (10, 11). In mice, BCAP is specifically expressed not only in B-cells but also in macrophages, dendritic cells, and lymphocyte natural killer cells (13). BCAP is mainly localized in the cytosol and the plasma membrane of expressing cells, and there are two splice variants in chicken, humans, and mice (6). Full-length BCAP is designated BCAP_L, and the second largest splice variant is designated BCAP_S.

BCAP_L is recruited into activated complexes of Toll-like receptors (TLRs) and associated signaling adaptor proteins. TLRs are pattern recognition receptors that respond to microbial products such as lipopolysaccharide from Gram-negative bacteria. The TLRs and the signaling adaptors MyD88 and MAL/TIRAP contain TIR domains, and these interact via homo- or heterotypic interactions to form complexes that constitute the post-receptor signalosome. A recent study predicted the existence of a TIR domain in the N-terminal region of BCAP_L that is required for negative regulation of inflammatory signaling by TLRs (14). RNAi depletion of BCAP_L from macro-

* This work was supported by a program grant from the UK Medical Research Council (G1000133) and a Wellcome Investigator award (WT100321/z/12/Z to N. J. G.). The authors declare that they have no conflicts of interest with the contents of this article.

§ Author's Choice—Final version free via Creative Commons CC-BY license.

§ This article contains supplemental Figs. 1–4 and supplemental Tables 1 and 2.

The atomic coordinates and structure factors (code 5for) have been deposited in the Protein Data Bank (<http://www.pdb.org/>).

¹ Supported by the Islamic Development Bank (Jeddah, Kingdom of Saudi Arabia) and the Cambridge Trust (Cambridge, UK).

² To whom correspondence may be addressed: Dept. of Biochemistry, Cambridge University, Sanger Bldg., 80 Tennis Court Rd., Cambridge CB2 1GA, UK. Tel.: 44-1223-333-626; E-mail: njg11@cam.ac.uk.

³ To whom correspondence may be addressed: Dept. of Biochemistry, Cambridge University, Sanger Bldg., 80 Tennis Court Rd., Cambridge CB2 1GA, UK. Tel.: 44-1223-333-626; E-mail: mcm35@cam.ac.uk.

⁴ The abbreviations used are: BCAP, B-cell adaptor for phosphoinositide 3-kinase; TLR, Toll-like receptor; TIR, Toll/interleukin-1 receptor; Btk, Bruton's tyrosine kinase; IP₃, inositol 1,4,5-trisphosphate; PIP₂, phosphatidylinositol 4,5-bisphosphate; PIP₃, phosphatidylinositol 1,4,5-trisphosphate; BCR, B-cell receptor; PLC γ , phospholipase C γ ; I3C, 5-amino-2,4,6-triiodoisophthalic acid monohydrate; AUC, analytical ultracentrifugation; SEC-MALS, size exclusion chromatography with multiangle light-scattering analysis; FL, full-length.

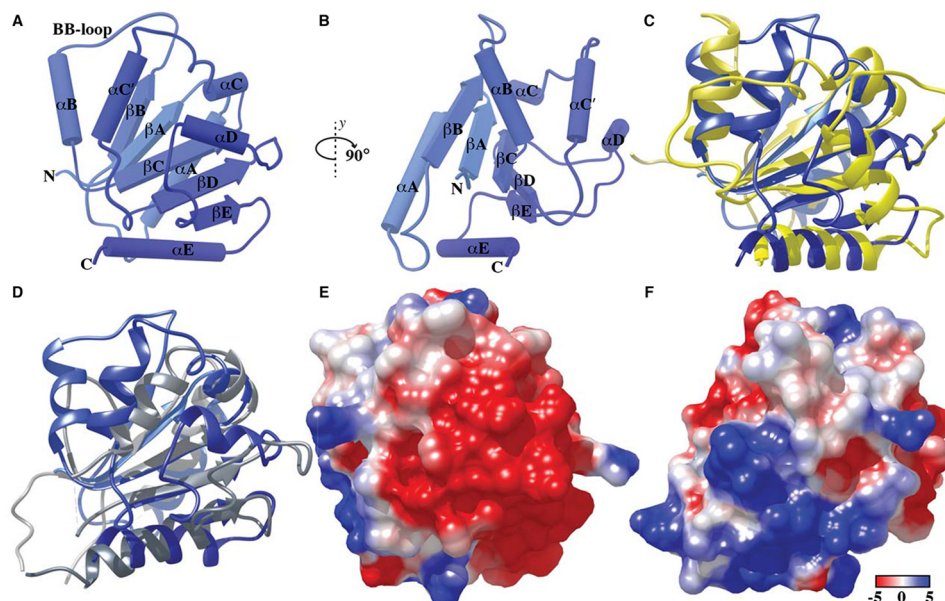


FIGURE 1. **Structure of the TIR domain of human BCAP.** *A* and *B*, the BCAP TIR domain has the typical TIR fold consisting of five parallel β -strands (β A– β E) surrounded by five helices (α A– α E). *C* and *D*, structural superposition of BCAP-TIR and the TIR domain of TLR2 (*C*) and MAL/TIRAP (*D*). Electrostatic surface potential of BCAP-TIR (*E* and *F*) using the orientations shown in *panels A* and *B* is colored from -5 (red) to $+5$ kcal/mol (blue). Electrostatic potentials were calculated using Chimera.

phages enhanced the LPS-induced production of NF- κ B and the inflammatory cytokines IL-6 and IL-10 but not TNF- α (10), which suggests that it negatively regulates inflammation. Similar results were obtained upon stimulation of TLR2/6, TLR3, and TLR7 by their respective ligands (10). In contrast, RNAi-mediated depletion of BCAP_s, a splice variant that lacks the putative TIR domain, is a positive regulator of inflammation (10).

Crosstalk between the TLR and PI3K pathways could be facilitated by the TIR domains on the TLR receptors or any of the well characterized TLR adaptor proteins TIRAP, MyD88, TRIF, TRAM, or SARM. However, the situation is more complex. Tyrosine phosphorylation of the YXXM motif on BCAP, the first of which lies outside of the putative TIR domain, enables interaction with the SH2 domain of p85 (6, 12, 15). Additionally, isolated B-cells from BCAP knockdown mice exhibited decreased calcium mobilization, which was attributed to a decrease in the activation of the enzyme PLC γ 2 (16). PLC γ 2, like p85, has both SH2 and SH3 domains, and its decreased activation may be the result of direct interaction with BCAP (16). The PLC γ 2 signaling pathway has been implicated in TLR4 signaling (17), but the molecular mechanisms and interactions are poorly understood.

Here we present the structure of the N-terminal domain of BCAP, firmly establishing it as a TIR domain. We also show that full-length BCAP is dimeric and that its oligomerization is dependent on its ANK and DBB domains. Furthermore, we show that BCAP TIR associates with the MAL/TIRAP adaptor and the TIR domains of TLRs. BCAP also binds to PLC γ 2, and this interaction does not inhibit its association with the tyrosine kinases Syk and Lyn. The interaction between BCAP and the TLRs, as well as its interaction with PI3K and PLC γ 2, suggests that BCAP plays a central role in crosstalk between the TLR, PI3K, and PLC γ 2 pathways.

Results

Structure of the N-terminal TIR Domain of Human BCAP—The N-terminal domain of BCAP comprising residues 7–142 was solubly expressed in *Escherichia coli*. The structure was solved at 2.5 Å using data derived from protein crystals in which the lysine residues were dimethylated and single-wavelength anomalous dispersion phasing with 5-amino-2,4,6-triiodo-isophthalic acid monohydrate (18). Data collection, phasing, and refinement statistics are summarized in [supplemental Table 1](#). There was a single molecule in the asymmetric unit, and the structure reveals that the N-terminal region adopts a typical TIR domain fold (Fig. 1*A*). The TIR domain of BCAP is composed of five parallel β -strands (β A– β E) surrounded by five α -helices (α A– α E). The third α -helix has a kink and forms two helices, α C and α C', and the fourth is distorted. The loop regions, with the exception of the biologically important BB-loop, which connects the β B-strand to the α B-helix, were well defined in the electron density, and unlike the structure of the MAL-TIR domain (19), there are no disulfide bonds.

Using the PDBeFOLD (20) and VAST (21) servers, the closest structural homologues were both TLR2-TIR domain mutants (22, 23) (Protein Data Bank (PDB) codes 1FYX and 1O77, respectively), although the sequence identity between the TLR2-TIR domain and BCAP-TIR is only 12%. Other close homologues include the TIR domain of the interleukin-1 receptor, IL-1RAPL (24), as well as MAL/TIRAP (19, 25) and the TIR domain of TLR10 (26). Secondary structural alignment of the BCAP TIR domain to its closest structural homologues is shown in Fig. 1, *C* and *D*. The BB-loop in BCAP is longer and not as rigid as that in TLR2. BCAP is less compact than MAL/TIRAP, which lacks the BB-loop and also contains two disulfide bonds. Strikingly, the orientation of the β B-sheet and the α B-helix of BCAP TIR domain is such that it extends out of the

TIR Domain of the BCAP Adaptor

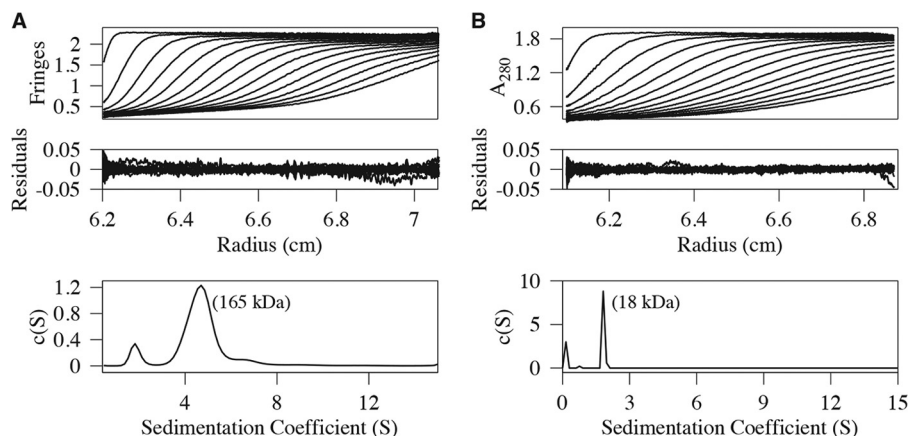


FIGURE 2. AUC analysis of full-length BCAP and TIR domain. *A* and *B*, sedimentation velocity data, residuals, and the sedimentation coefficient distribution, $c(S)$, for full-length BCAP (*A*) and the N-terminal region comprising residues 7–142 (*B*). The recovered molecular masses from a $c(M)$ analysis are also shown. The frictional coefficient was well determined, and as a result, the molecular masses are reliable. Full-length BCAP is dimeric, and the N-terminal domain is monomeric. Every third scan is shown for clarity.

domain core in comparison with that of MAL/TIRAP. With regard to sequence conservation across the structurally homologous TIR domains, there are five highly conserved residues including Ser-17 (αA -loop), Leu-32 (αA), Pro-49 (βB), Leu-101, and Leu-102 (βD) (supplemental Fig. 1). The electrostatic potential of the solvent-accessible surface of BCAP (Fig. 1, *E* and *F*) reveals distinct regions of negative and positive potential in addition to hydrophobic patches, suggesting that the TIR domain of BCAP may interact with diverse TIR domain-containing proteins. For example, the TIR domain of MAL/TIRAP is predominantly hydrophobic but has acidic and small basic patches (19), which could, in principle, interact with acidic or basic surfaces on BCAP-TIR. In contrast, the TIR domain of MyD88 has significant regions of basic residues that may interact with the acidic surface on the TIR domain of BCAP.

Given the necessity of TIR-TIR interactions to signal propagation, several mutations were made to explore possible regions on BCAP that may be involved in its association with MAL/TIRAP. Sequence alignments of BCAP and proteins known to interact with MAL/TIRAP (MyD88, p85, A46, and TRAF6) revealed three conserved residues (Leu-70, Asp-117, and Leu-401). Two of these (Leu-70 and Asp-117) lie within the TIR domain, and all were mutated to alanine. Additionally, the highly conserved βB loop proline residue (Pro-49), which directs adaptor specificity in TLRs (27), was mutated to aspartic acid and alanine, respectively. The co-immunoprecipitation data for these mutants and MAL/TIRAP (supplemental Fig. 3) suggest that Pro-49 is not required for the interaction. Both the D75A and L401A mutations exhibited reduced binding, whereas the D117A mutation abolishes the interaction between BCAP and MAL/TIRAP. Asp-117 is located in a region of acidic surface potential sandwiched between the αD -helix and βE -sheet, and it is likely that this region interacts with a basic patch on MAL/TIRAP. The reduced association observed with the L401A mutant suggests that the determinants of the BCAP-MAL/TIRAP interaction may not solely reside within the TIR domain, and it is likely that there are other regions involved in the BCAP-MAL/TIRAP hetero-association.

Full-length BCAP Is Dimeric, but the TIR Domain Is Monomeric—Analytical ultracentrifugation (AUC) experiments were performed on purified samples of full-length BCAP_L and an N-terminal region spanning residues 7–142, which contains the TIR domain. The sedimentation velocity data, the residuals after fitting, and the $c(S)$ distribution are shown in Fig. 2. The sedimentation coefficient values were 4.70×10^{-13} s and 1.82×10^{-13} s for BCAP_L and the N-terminal domain, respectively. The qualities of the fits were very good with root mean square deviation values of 0.007 (BCAP_L) and 0.005 (N-terminal domain). Additionally, the frictional coefficient was well determined, and this allows estimation of molecular masses from the sedimentation velocity data. Using $c(M)$ analyses, the molecular masses are 165 and 18 kDa for BCAP_L and the TIR domain, respectively. These values are within ~10% of those expected for dimeric BCAP_L and monomeric N-terminal domain on the basis of their amino acid sequence. Weight-averaged molecular masses obtained from an analysis of the SEC-MALS (supplemental Fig. 2, *A* and *C*) data are consistent with those obtained by AUC.

The DBB and ANK Domains Mediate BCAP Oligomerization—The monomeric nature of the TIR domain implies that the determinants of BCAP_L self-association lie outside this region. To clearly define the regions responsible for BCAP_L oligomerization, the splice variant (BCAP_S) and a truncated molecule comprising the first 330 residues of BCAP_L (BCAP₃₃₀), which contains the TIR and DBB domains, were expressed, and their hydrodynamic behavior was studied by sedimentation velocity. Fig. 3 shows an analysis of the sedimentation velocity data for these constructs, and both, like BCAP_L, are dimeric. SEC-MALS data for BCAP₃₃₀ (supplemental Fig. 2*B*) also confirm that it is dimeric. Because BCAP₃₃₀ contains the DBB domain, this confirms that it is, at least in part, responsible for the self-association of BCAP_L. The ankyrin repeats are also known to facilitate oligomerization (28), and together with the DBB domain, they may enhance self-association.

Human BCAP Interacts with MyD88, MAL/TIRAP, TLR2, TLR4, and SARM but Not TRAM—TIR domains associate via homotypic interactions, and the presence of an N-terminal TIR

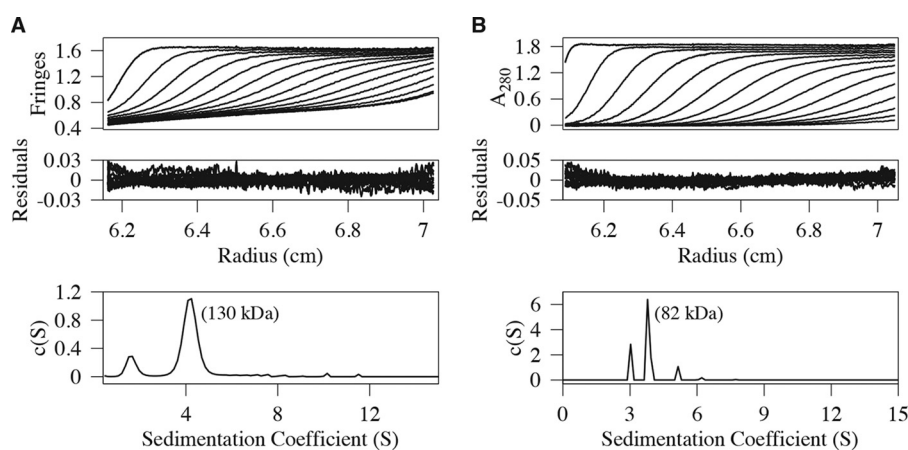


FIGURE 3. **AUC analysis of BCAP splice variant and N-terminal variant.** A and B, sedimentation velocity profiles, residuals, and sedimentation coefficient distribution, $c(S)$, for the splice variant BCAP_s, which lacks the first 180 amino acids and contains the DBB domain and ANK repeats (residues 322–403) (A), and BCAP₃₃₀, which consists of the N-terminal 330 residues and includes the putative TIR and DBB domains (B). Both are dimeric, revealing that the DBB and ANK domains are responsible for BCAP_L self-association. Every third scan is shown for clarity.

domain in BCAP could potentially enable binding with the cytosolic adaptor proteins involved in the TLR pathway. To determine whether BCAP_L interacts with TRAM, SARM, and MyD88, pulldown assays were performed. HEK 293T cells were transiently transfected with Myc-TRAM, FLAG-SARM, Myc-MyD88, and Myc-MyD88-FXXM mutant in the presence of either Myc-BCAP or FLAG-BCAP. BCAP showed no interaction with Myc-TRAM (Fig. 4A) but interacted with FLAG-SARM (Fig. 4B) and also with Myc-MyD88 and the Myc-MyD88-FXXM mutant (Fig. 4C).

The nature of the interaction between BCAP and TIRAP was also probed by a yeast two-hybrid assay utilizing a construct consisting of the first 434 amino acids of BCAP (BCAP-TDA). This construct includes the dimerization site, two of the four YXXM motifs present in the full-length protein, and the TIR, DBB, and ANK domains. BCAP-TDA was tested against full-length constructs of MAL/TIRAP and its mutants D96N and S180L, the TIR domains of TLR2 and TLR4, and also the regulatory p85 subunit of PI3K. In these experiments, all proteins tested interacted with BCAP-TDA (see supplemental Fig. 4). Interestingly, the disease-associated MAL/TIRAP variants S180L (29) and D96N (30) also bind BCAP-TDA, strongly suggesting a different binding mode to that proposed for MyD88 (19).

BCAP Associates with PLC γ 2, Lyn, and Syk—BCAP_L is involved in the BCR-mediated activation of PLC γ 2 and PI3K (6). The interaction between BCAP_L and PI3K is direct and is known to involve at least one of two SH2 domains on the regulatory p85 subunit of PI3K. The p85 subunit also has an SH3 domain that is separated from the first N-terminal SH2 domain by a Rho-GAP domain (5, 31), and it is not known whether this domain is involved in the interaction with BCAP. Like p85, PLC γ 2 contains two SH2 domains and an SH3 domain, and given the similarities in domain structure, we hypothesized that BCAP could interact with PLC γ 2 in a manner analogous to its interaction with the p85 subunit. Additionally, there is ambiguity regarding the identity of the kinase that activates PLC γ 2, with both Syk and Btk having been implicated (17, 32).

To address these questions, pulldown assays using BCAP_L and human PLC γ 2 (PLC γ 2-V5) were carried out in the pres-

ence or absence of either human YFP-tagged Syk or GFP-tagged Lyn. The results presented in Fig. 5A show that FLAG-BCAP_L associates with PLC γ 2-V5 in the presence and absence of either YFP-Syk or GFP-Lyn. Moreover, the addition of Syk or Lyn did not increase the apparent binding of BCAP_L to PLC γ 2, suggesting that the interaction between BCAP and PLC γ 2 may be promoted by endogenous tyrosine kinases in these experiments. However, it is apparent from the presence of a higher molecular mass band of BCAP that it is phosphorylated by the transfected Lyn kinase. BCAP was also able to pull down both Lyn and Syk in the absence and presence of PLC γ 2 (Fig. 5A). Consequently, BCAP is phosphorylated by Lyn and forms a complex containing either Syk or Lyn kinases and PLC γ 2 irrespective of its activation by kinase.

We also investigated the ability of PLC γ 2 to interact with the TIR domain-containing proteins SARM, MAL/TIRAP, and MyD88. Both MyD88 and MAL/TIRAP were able to interact with PLC γ 2. In contrast, SARM appears not to associate directly with PLC γ 2 (Fig. 5B).

Discussion

Information flow through the TLR pathway is initiated by ligand binding to leucine-rich repeats on the receptor ectodomain followed by receptor dimerization. Subsequently, homotypic associations of cytoplasmic receptor TIR domains and those of various adaptor molecules including MyD88, TRIF, MAL/TIRAP, TRAM, and SARM ensue, setting in motion a cascade of molecular interactions that result in many cellular responses including the production of pro-inflammatory molecules (33). Until recently, the known cytoplasmic TIR-containing adaptors were thought to be involved in direct stimulus-driven TLR signaling only. However, the observation that an adaptor molecule, BCAP, which is involved in BCR signaling, a pathway seemingly unrelated to TLRs, was a negative regulator of inflammation prompted a search for “missing” TIR domains in the human genome and the postulate that BCAP possesses a hidden N-terminal TIR domain (14).

Here we show conclusively that BCAP has an N-terminal TIR domain that structurally bears closest resemblance to the homologous domain in TLR2. The isolated TIR domain of BCAP

TIR Domain of the BCAP Adaptor

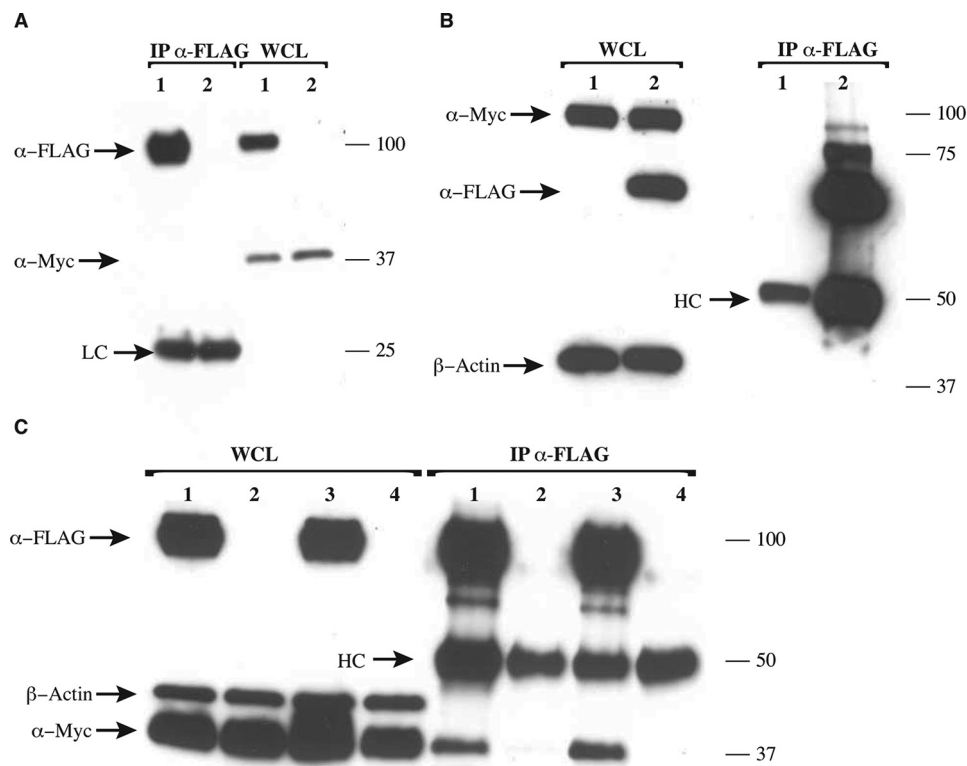


FIGURE 4. Interaction of BCAP and other TIR domain-containing proteins. HEK 293T cells were overexpressed with Myc-TRAM, FLAG-BCAP, Myc-BCAP, FLAG-SARM, Myc-MyD88, and a Myc-MyD88-FXXM mutant as indicated. Cells were lysed, and immunoprecipitation (IP) was performed using anti-FLAG antibody. Precipitates were split and assayed for the presence of FLAG- or Myc-tagged proteins. *A*, BCAP shows no interaction with TRAM (lane 1, Myc-TRAM + FLAG-BCAP-FL; lane 2, Myc-TRAM). *WCL*, whole cell lysate. *B* and *C*, BCAP interacts with SARM (lane 1, Myc-BCAP-FL; lane 2, Myc-BCAP-FL and FLAG-SARM) (*B*) and with both MyD88 and the FXXM mutant of MyD88 (lane 1, Myc-MyD88-WT and FLAG-BCAP-FL; lane 2, Myc-MyD88-WT; lane 3, Myc-MyD88-FXXM and Myc-BCAP-FL; lane 2, Myc-MyD88-FXXM) (*C*). *HC* and *LC* are the heavy and light chains of the mouse anti-FLAG antibody, respectively.

is monomeric both in the crystal and in solution, a feature shared with most non-plant TIR domains (34), suggesting that vertebrate TIR domain-containing proteins require other domains to promote self-oligomerization: leucine-rich repeats (LRRs) for the TLR, a death domain for MyD88, and the DBB domain and likely the ANK repeats for BCAP. The BCAP_L paralogue, BANK1, which regulates CpG-induced IL-6 production (35), also has a TIR domain as the corresponding domains have high sequence homology (40%).

A strong interaction with MAL/TIRAP, TLR4, and TLR2 is also detected in yeast two-hybrid assays. This implies that the dimeric BCAP TIR domain makes multiple interactions with the TLR post-receptor complex. MAL/TIRAP is of particular interest because this adaptor localizes to the plasma membrane by binding to PIP2, an association that is required for signaling (36).

Together our findings provide evidence that BCAP is part of a membrane-bound network that down-regulates inflammatory signaling by the TLRs. Thus, activation of PI3K and PLC γ 2 results in the conversion of PIP2 to PIP3 and inositol 1,4,5-triphosphate (IP3), respectively, leading to depletion of MAL/TIRAP from the membrane (Fig. 6*B*) and negative regulation of MAL/TIRAP-dependent TLR signaling.

The association between BCAP and PLC γ 2 appears to be more stable than the transient interaction observed between BANK1 and PLC γ 2 (37) and may reflect cooperative effects in complex assembly. In contrast, the association between human MAL/TIRAP or MyD88 and PLC γ 2 appears to be less stable

than that of BCAP with PLC γ 2, which likely reflects the central role played by BCAP in modulating all three pathways. Our finding that BCAP interacts directly with PLC γ 2 is consistent with published reports linking BCAP to PLC γ 2-dependent calcium release (16). Calcium release initiated by the cleavage of PIP2 by PLC γ 2 releases diacylglycerol and the potent second messenger IP3, which subsequently interacts with IP3 receptors in the endoplasmic reticulum (38). The resulting calcium flux activates both TLR signaling (17, 39–41) and the trafficking of TLR4 to endosomal compartments (17).

It is also likely that BCAP is the “missing link,” coupling LPS activation of TLR4 to endocytosis and the TRAM/TRIF pathway that induces interferon- β . Zanoni *et al.* (42) have shown that the extrinsic membrane pattern recognition receptor (PRR) CD14 is required for the LPS-induced internalization of the TLR4 post-receptor complex and that this function of CD14 is dependent upon the activation of both Syk and PLC γ 2. A further correlation involves the potential role of MAL/TIRAP as a promiscuous lipid-binding protein (43). Although this sorting adaptor had only been implicated in signaling from the cell surface TLR2 and TLR4, Bonham *et al.* (43) present evidence that MAL/TIRAP is required for resident endosomal TLRs such as TLR9, at least in response to natural viral infection rather than synthetic ligands such as phosphorothioate CpG-containing oligonucleotides. Endosomal membranes lack PIP2, but MAL/TIRAP may bind to other phospholipids that are abundant in this compartment such as phosphatidylinositol 3-phosphate (PI(3)P). As noted, BCAP_L also negatively regulates

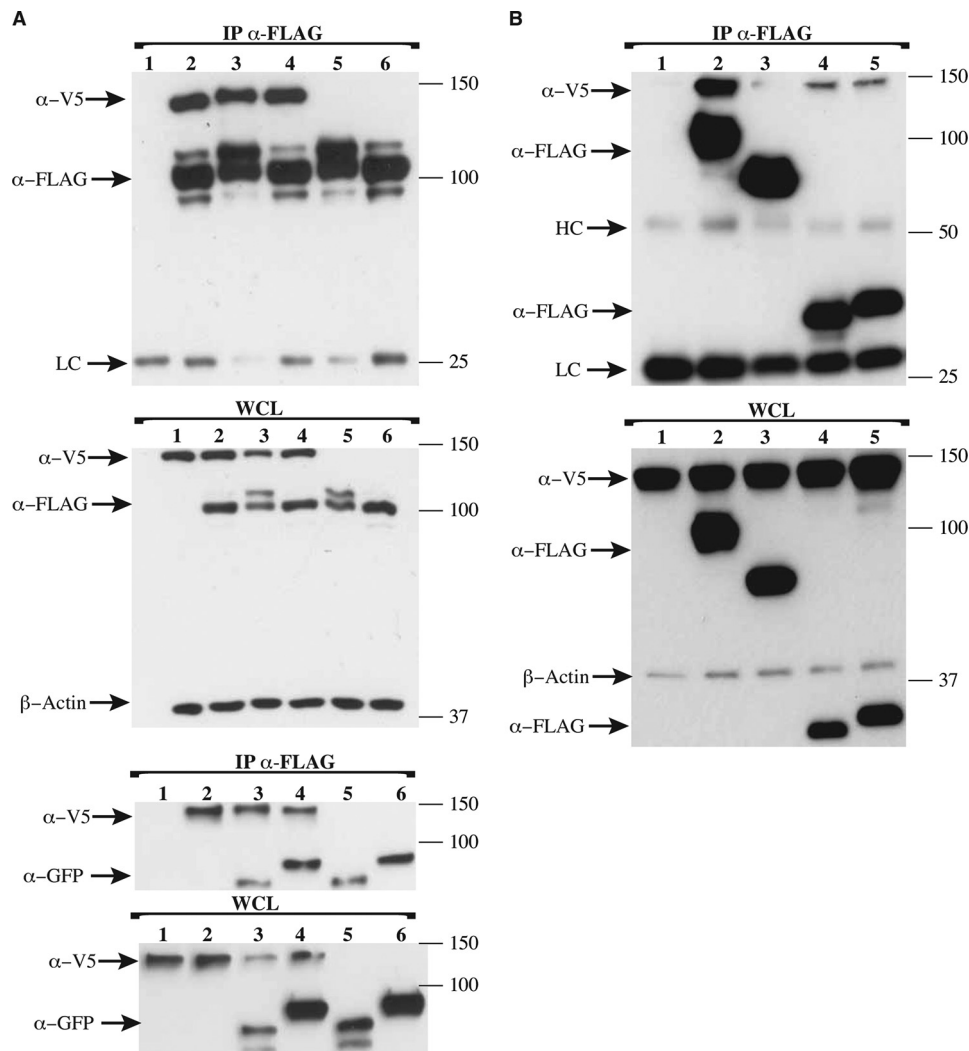


FIGURE 5. BCAP associates with phospholipase C γ 2 and the kinases Lyn and Syk. HEK 293T cells were overexpressed with PLC γ 2-V5, FLAG-BCAP, Lyn-GFP, YFP-Syk, and TIRAP-FLAG. Cells were lysed, and immunoprecipitation was performed using anti-FLAG antibody. Precipitates were split and assayed for the presence of V5-, FLAG-, GFP-, and YFP-tagged proteins as shown. *A*, the interaction between BCAP and PLC γ 2 is independent of the tyrosine kinases Syk and Lyn; BCAP associates with Syk or Lyn both in the presence and in the absence of PLC γ 2 (*lane 1*, PLC γ 2-V5; *lane 2*, PLC γ 2-V5 and FLAG-BCAP-FL; *lane 3*, PLC γ 2-V5, FLAG-BCAP-FL, and GFP-Lyn; *lane 4*, PLC γ 2-V5, FLAG-BCAP-FL, and YFP-Syk; *lane 5*, FLAG-BCAP-FL and GFP-Lyn; *lane 6*, FLAG-BCAP-FL and YFP-Syk). *B*, both MAL/TIRAP and MyD88 show reduced association with PLC γ 2 relative to BCAP, whereas SARM does not interact with PLC γ 2 (*lane 1*, PLC γ 2-V5; *lane 2*, PLC γ 2-V5 and FLAG-BCAP-FL; *lane 3*, PLC γ 2-V5 and FLAG-SARM; *lane 4*, PLC γ 2-V5 and TIRAP-FLAG; *lane 5*, PLC γ 2-V5 and MyD88-FLAG). The expression of β -actin was used to assess the relative expression levels of the transformed plasmids. HC and LC are the heavy and light chains of the mouse anti-FLAG antibody, respectively.

endosomal TLRs, perhaps by modulating the levels of these alternative phospholipids in endocytic vesicles. The precise pathways that mediate this crosstalk remain to be elucidated.

There are other scenarios in which PLC γ 2 activity is linked to TLR pathway outcomes. PLC γ 2 is crucial in the calcium signaling response of TLR2 and TLR4 stimulation by their respective ligands and is also required for the peptidoglycan-dependent production of TNF α and IL-6 (40). Additionally, the calcium flux produced by PLC γ 2 is crucial for TRIF-dependent activation of IRF-3 but not the LPS-induced activation of TNF α (17). The activation of PLC γ 2 in BCR signaling in murine splenic B-cells is the result of phosphorylation on Tyr-753, Tyr-759, and Tyr-1217 by Btk, but in Ramos cells, phosphorylation of Tyr-1217 is Btk-independent, implying the presence of Btk-dependent and Btk-independent pathways (44). Indeed, its phosphorylation by Syk is implicated in the production of TNF α following stimulation of TLR9 by CpG. BCAP, however, is

involved in the production of IL-6 and IL-10 but not TNF α upon TLR stimulation (10). Thus, our finding that the association of BCAP and PLC γ 2 is not enhanced by co-expression with Syk is consistent with those observations.

As summarized in Fig. 6, this study partially uncovers a complex membrane-associated network that regulates inflammation by establishing a negative feedback loop between TLR activation and the metabolism of inositol phospholipids. Future studies will elucidate more fully the interactions in this network and the molecular basis of the multitypic interactions mediated by the BCAP TIR domain.

Experimental Procedures

Protein Expression and Purification for Biophysical Analysis—The N-terminal domain of human BCAP comprising residues 7–142 was cloned into pMCSG7 (DNASU) and expressed as an N-terminal hexahistidine-tagged fusion protein in *E. coli*

TIR Domain of the BCAP Adaptor

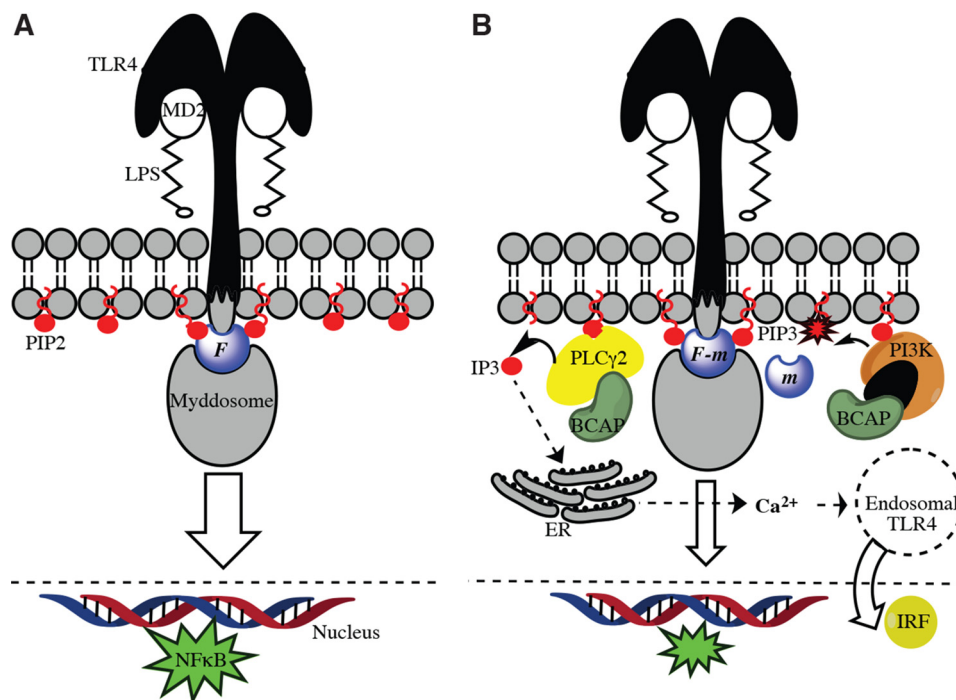


FIGURE 6. Negative regulation of TLR4 signaling by BCAP and PLC γ 2. *A*, MD2/LPS binding to the TLR4 ectodomain enables the recruitment of a cytosolic signalosome comprising the PIP₂-linked adaptor MAL/TIRAP and the Myddosome (MyD88 and interleukin-1 receptor-associated kinase (IRAK) kinases), which interact with the receptor TIR domains. The flux, *F*, through this pathway stimulates the production of NF κ B. *B*, PLC γ 2-linked BCAP cleaves PIP₂ to DAG and IP₃, which deprives MAL/TIRAP of its membrane anchor. Additionally, PI3K further depletes PIP₂ by converting it to PIP₃. MAL/TIRAP is still capable of interacting with PIP₃, but it is inactive presumably because it can no longer recruit the Myddosome. The net result is a reduction in flux, (*F* – *m*), and thus negative regulation, of the MD2/LPS-induced stimulation of TLR4. IP₃ produced by PLC γ 2 cleavage of PIP₂ stimulates calcium release from the endoplasmic reticulum (ER), which in turn drives the internalization of TLR4 into endosomal compartments, leading to the production of interferon regulatory factors (IRFs).

BL21DE3 cells (Novagen). After nickel-nitrilotriacetic acid purification in 20 mM Tris, 20 mM NaCl, pH 7.5, samples were treated with tobacco etch virus (TEV) protease to remove the hexahistidine tag, and further purification was performed using S200 (GE Healthcare) size exclusion chromatography in the same buffer supplemented with 5 mM DTT. BCAP_L (1–805), BCAP_S, and BCAP₃₃₀ were cloned into the same vector with a Strep-tag at the C terminus. Proteins were purified using a 5-ml StrepTrap column (GE Healthcare), and proteins were eluted using 2.5 mM desthiobiotin (Sigma) dissolved in the same buffer with final purification on S200 size exclusion as described above.

Analytical Centrifugation and SEC-MALS—Sedimentation velocity measurements were performed using a Beckman XL-A ultracentrifuge equipped with absorbance and interference optical systems. Sample concentrations ranged from 0.3 to 1 mg/ml. Data were acquired every 300 s at rotor speeds of 30,000–40,000 rpm using an An60Ti rotor (Beckman Coulter). Buffer density and viscosity were estimated using SEDNTERP, and the experimental data were analyzed using SEDFIT (45). SEC-MALS data were obtained using an injection volume of 50 μ l and a Superdex 200 Increase 10/300 column (GE Healthcare). Multiangle light scattering and differential refractometry data were acquired using DAWN 8⁺ and Optilab T-rEX instruments, respectively (Wyatt Technology), and data analysis was performed using the ASTRA (v6.1) software package from the manufacturer. Protein concentrations were 4, 2, and 0.8 mg/ml for the TIR, BCAP₃₃₀, and BCAP_L, respectively.

Protein Production for Crystallization—Following nickel-nitrilotriacetic acid purification of the N-terminal domain of

BCAP, samples were treated with tobacco etch virus protease to remove the hexahistidine tag and then buffer-exchanged using a Hi-Prep 26/10 desalting column (GE Healthcare) into 50 mM HEPES, 100 mM NaCl, 2 mM DTT, pH 7.5. Following lysine dimethylation (46), samples were purified by size exclusion chromatography into 20 mM TRIS, 20 mM NaCl, 5 mM DTT, pH 7.5. Crystals were obtained using the sitting-drop vapor diffusion method at 20 °C and protein concentrations of 5–7 mg/ml. The well solution contained 100 mM MES, pH 6.5, 2–10% PEG 400, 1.5–2.4 M (NH₄)₂SO₄, and 1 mM 5-amino-2,4,6-triiodo-isophthalic acid monohydrate (I3C) (18).

Crystallographic Data Collection and Structure Determination—Diffraction data were collected on the IO3 beamline (Diamond Light Source, Didcot, Oxfordshire, UK). Data reduction and scaling were performed using XDS (47) and Aimless (48). Initial phases using the anomalous signal from I3C were obtained using SHELXD (49), and a partial model was obtained using Buccaneer (50). Further rounds of manual model building and refinement were performed using Coot (51), O (52), and PHENIX (53), respectively.

Antibodies—The antibodies used were mouse α -Myc (Santa Cruz Biotechnology), mouse monoclonal α -FLAG M2 (Sigma-Aldrich), rabbit α -FLAG (Sigma-Aldrich), and α -V5-HRP conjugate (Invitrogen).

Yeast Two-hybrid Assays—The bait plasmid for yeast two-hybrid assay was generated by inserting the first 434 amino acids of human BCAP (Source Bioscience) in-frame into the pGBKT7 vector (Clontech) to be expressed as a GAL4-DBD fusion protein. The prey plasmids were generated by inserting

either the full-length proteins or different protein mutations and truncations in-frame into the pGADT7 vector (Clontech) to be expressed as GAL4-AD fusion proteins. The TIR domain of TLR2 contained residues 636–784, whereas the TIR domain of TLR4 comprised residues 671–820. Protein-protein interactions were assayed by transforming the bait plasmid into the yeast strain AH109 (Clontech) and the prey plasmids into the yeast strain Y187 (Clontech) using standard lithium acetate transformation (54) with the Matchmaker yeast two-hybrid system (Clontech). The Leu⁺ and Trp⁺ transformants were selected on SD/–Leu and SD/–Trp plates, respectively. For the interaction analysis, equally dense suspensions of each transformant were prepared in 0.5× yeast extract, peptone, dextrose and adenine broth, mixed, and allowed to mate for 21 h at 30 °C with shaking at 50 rpm. The respective pair of transformants (bait and prey) was then plated onto SD/–Leu/–Trp plates for assessing mating efficiency and on SD/–Leu/–Trp/–His/–Ade+Xα-Gal+ 5 mM 3-amino-1,2,4-triazole (3-AT) plates for assessing the interaction. Negative controls were the empty vectors of pGBKT7 and pGADT7 treated similarly.

Pulldown Assays—Human BCAP and splice variant was cloned into either p3XFLAG-CMV-10 or pCMV/Myc vectors. Protein constructs were transfected into HEK 293T cells (ATCC, Manassas, VA) (70–80% confluence) using JetPEI (Polyplus-transfection SA) according to the manufacturer's recommendations. 3 μg of plasmid DNA was transfected per well in a 6-well plate. When required, pcDNA3.1 was included to ensure that the total DNA transfected was 3 μg. The transfection mixture was incubated for 24 h, after which cells were washed once in 1× PBS and lysed using 300 μl of a solution containing 20 mM Tris, 150 mM NaCl, 1 mM EDTA, 0.5% Nonidet P-40, pH 8.0, which was supplemented with 1% Triton X-100, 0.5% *n*-octylglucoside, 1 mM PMSF, and 1× protease inhibitor cocktail (Calbiochem). After incubation on ice for 10 min, the mixture was transferred to an Eppendorf tube, and lysis was continued for an additional 30 min at 4 °C with agitation, after which the mixture was centrifuged at 16,000 × *g* for 10 min at 4 °C, and the supernatant was collected for analysis by Western blotting or for immunoprecipitation with anti-FLAG M2 beads (EZview Red anti-FLAG M2 affinity gel, Sigma). 250 μl of the extracted protein sample was incubated with anti-FLAG beads for 2 h at 4 °C with constant rotation, after which non-specifically bound proteins were removed by repeated centrifugation at 1000 × *g* and resuspension of the beads with 500 μl of lysis buffer. After the final wash, excess solution was aspirated, and 30 μl of 1× SDS loading buffer was added to the beads and heated at 100 °C for 10 min. Finally, the protein samples from whole cell lysate and immunoprecipitation were analyzed by Western blotting using the indicated antibodies.

Author Contributions—S. H. produced proteins, conducted Y2H and pulldown experiments and assisted with the AUC and crystallographic analysis and writing. E. S. and B. V. supplied the data for supplemental Fig. 3. M. C. M. wrote most of the paper and performed AUC analysis and X-ray structure determination and SEC-MALS. N. J. G. initially conceived the project and helped with writing the paper.

Acknowledgments—Yfp-Syk was a gift from Dr. Peter Jordan (Instituto Nacional de Saúde, Portugal). Dr. Michael Carty (Trinity College Dublin, Ireland) provided the pEF-Bos FLAG-tagged human SARM, and PLCγ2-V5 was a gift from Dr. Marta Alarcón Riquelme (University of Granada, Spain). Our thanks go to Dr. Joseph Mahman for help with the SEC-MALS experiments, Dr. Owen R. Davies for initial help with the yeast two-hybrid experiments, and Drs. Fernando Bazan (Bio-Techne Corp. and the Department of Pharmacology, University of Minnesota) and Chandrashekar Pasare (Department of Immunology, University of Texas-Southwestern Medical Center) for helpful discussions.

References

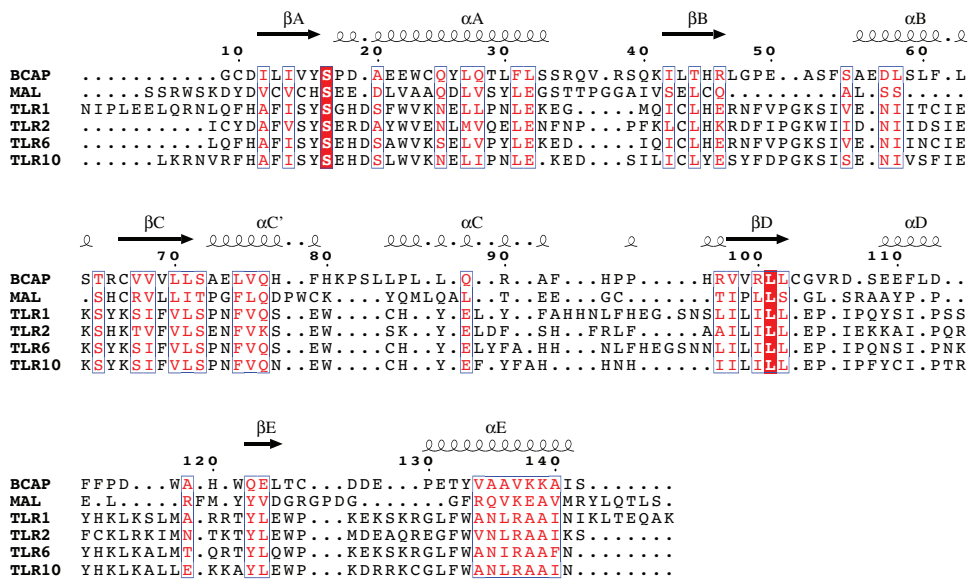
- Rameh, L. E., and Cantley, L. C. (1999) The role of phosphoinositide 3-kinase lipid products in cell function. *J. Biol. Chem.* **274**, 8347–8350
- Leevers, S. J., Vanhaesebroeck, B., and Waterfield, M. D. (1999) Signalling through phosphoinositide 3-kinases: the lipids take centre stage. *Curr. Opin. Cell Biol.* **11**, 219–225
- Cantley, L. C. (2002) The phosphoinositide 3-kinase pathway. *Science* **296**, 1655–1657
- Fan, Q. W., and Weiss, W. A. (2010) Targeting the RTK-PI3K-mTOR axis in malignant glioma: overcoming resistance. *Curr. Top. Microbiol. Immunol.* **347**, 279–296
- Okkenhaug, K. (2013) Signaling by the phosphoinositide 3-kinase family in immune cells. *Annu. Rev. Immunol.* **31**, 675–704
- Okada, T., Maeda, A., Iwamatsu, A., Gotoh, K., and Kurosaki, T. (2000) BCAP: the tyrosine kinase substrate that connects B cell receptor to phosphoinositide 3-kinase activation. *Immunity* **13**, 817–827
- Vincent, S., Wilson, R., Coelho, C., Affolter, M., and Leptin, M. (1998) The *Drosophila* protein Dof is specifically required for FGF signaling. *Mol. Cell* **2**, 515–525
- Battersby, A., Csiszár, A., Leptin, M., and Wilson, R. (2003) Isolation of proteins that interact with the signal transduction molecule Dof and identification of a functional domain conserved between Dof and vertebrate BCAP. *J. Mol. Biol.* **329**, 479–493
- Yokoyama, K., Su Ih, I. H., Tezuka, T., Yasuda, T., Mikoshiba, K., Tarakhovskiy, A., and Yamamoto, T. (2002) BANK regulates BCR-induced calcium mobilization by promoting tyrosine phosphorylation of IP₃ receptor. *EMBO J.* **21**, 83–92
- Matsumura, T., Oyama, M., Kozuka-Hata, H., Ishikawa, K., Inoue, T., Muta, T., Semba, K., and Inoue J.-i. (2010) Identification of BCAP-L as a negative regulator of the TLR signaling-induced production of IL-6 and IL-10 in macrophages by tyrosine phosphoproteomics. *Biochem. Biophys. Res. Commun.* **400**, 265–270
- Song, S., Chew, C., Dale, B. M., Trauma, D., Peacock, J., Yamazaki, T., Clynes, R., Kurosaki, T., and Greenberg, S. (2011) A requirement for the p85 PI3K adapter protein BCAP in the protection of macrophages from apoptosis induced by endoplasmic reticulum stress. *J. Immunol.* **187**, 619–625
- Ni, M., MacFarlane, A. W., 4th, Toft, M., Lowell, C. A., Campbell, K. S., and Hamerman, J. A. (2012) B-cell adaptor for PI3K (BCAP) negatively regulates Toll-like receptor signaling through activation of PI3K. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 267–272
- MacFarlane, A. W., 4th, Yamazaki, T., Fang, M., Sigal, L. J., Kurosaki, T., and Campbell, K. S. (2008) Enhanced NK-cell development and function in BCAP-deficient mice. *Blood* **112**, 131–140
- Troutman, T. D., Hu, W., Fulencheck, S., Yamazaki, T., Kurosaki, T., Bazan, J. F., and Pasare, C. (2012) Role for B-cell adapter for PI3K (BCAP) as a signaling adapter linking Toll-like receptors (TLRs) to serine/threonine kinases PI3K/Akt. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 273–278
- Inabe, K., and Kurosaki, T. (2002) Tyrosine phosphorylation of B-cell adaptor for phosphoinositide 3-kinase is required for Akt activation in response to CD19 engagement. *Blood* **99**, 584–589

16. Yamazaki, T., Takeda, K., Gotoh, K., Takeshima, H., Akira, S., and Kurosaki, T. (2002) Essential immunoregulatory role for BCAP in B cell development and function. *J. Exp. Med.* **195**, 535–545
17. Chiang, C.-Y., Veckman, V., Limmer, K., and David, M. (2012) Phospholipase C γ 2 and intracellular calcium are required for lipopolysaccharide-induced Toll-like receptor 4 (TLR4) endocytosis and interferon regulatory factor 3 (IRF3) activation. *J. Biol. Chem.* **287**, 3704–3709
18. Beck, T., Krasauskas, A., Gruene, T., and Sheldrick, G. M. (2008) A magic triangle for experimental phasing of macromolecules. *Acta Crystallogr. D Biol. Crystallogr.* **64**, 1179–1182
19. Valkov, E., Stamp, A., Dimaio, F., Baker, D., Verstak, B., Roversi, P., Kellie, S., Sweet, M. J., Mansell, A., Gay, N. J., Martin, J. L., and Kobe, B. (2011) Crystal structure of Toll-like receptor adaptor MAL/TIRAP reveals the molecular basis for signal transduction and disease protection. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 14879–14884
20. Krissinel, E., and Henrick, K. (2004) Secondary structure matching (SSM), a new tool for fast protein structure alignment in three dimensions. *Acta Crystallogr. D Biol. Crystallogr.* **60**, 2256–2268
21. Gibrat, J. F., Madej, T., and Bryant, S. H. (1996) Surprising similarities in structure comparison. *Curr. Opin. Struct. Biol.* **6**, 377–385
22. Xu, Y., Tao, X., Shen, B., Horng, T., Medzhitov, R., Manley, J. L., and Tong, L. (2000) Structural basis for signal transduction by the Toll/interleukin-1 receptor domains. *Nature* **408**, 111–115
23. Tao, X., Xu, Y., Zheng, Y., Beg, A. A., and Tong, L. (2002) An extensively associated dimer in the structure of the C713S mutant of the TIR domain of human TLR2. *Biochem. Biophys. Res. Commun.* **299**, 216–221
24. Khan, J. A., Brint, E. K., O'Neill, L. A., and Tong, L. (2004) Crystal structure of the Toll/interleukin-1 receptor domain of human IL-1RAPL. *J. Biol. Chem.* **279**, 31664–31670
25. Snyder, G. A., Deredge, D., Waldhuber, A., Fresquez, T., Wilkins, D. Z., Smith, P. T., Durr, S., Cirl, C., Jiang, J., Jennings, W., Luchetti, T., Snyder, N., Sundberg, E. J., Wintrose, P., Miethke, T., and Xiao, T. S. (2014) Crystal structures of the Toll/Interleukin-1 receptor (TIR) domains from the *Brucella* protein TcpB and host adaptor TIRAP reveal mechanisms of molecular mimicry. *J. Biol. Chem.* **289**, 669–679
26. Nyman, T., Stenmark, P., Flodin, S., Johansson, I., Hammarström, M., and Nordlund, P. (2008) The crystal structure of the human toll-like receptor 10 cytoplasmic domain reveals a putative signaling dimer. *J. Biol. Chem.* **283**, 11861–11865
27. Verstak, B., Arnot, C. J., and Gay, N. J. (2013) An alanine-to-proline mutation in the BB-loop of TLR3 Toll/IL-1R domain switches signalling adaptor specificity from TRIF to MyD88. *J. Immunol.* **191**, 6101–6109
28. Kohl, A., Binz, H. K., Forrer, P., Stumpp, M. T., Plückthun, A., and Grütter, M. G. (2003) Designed to be stable: Crystal structure of a consensus ankyrin repeat protein. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 1700–1705
29. Khor, C. C., Chapman, S. J., Vannberg, F. O., Dunne, A., Murphy, C., Ling, E. Y., Frodsham, A. J., Walley, A. J., Kyrielleis, O., Khan, A., Aucan, C., Segal, S., Moore, C. E., Knox, K., Campbell, S. J., *et al.* (2007) A Mal functional variant is associated with protection against invasive pneumococcal disease, bacteremia, malaria and tuberculosis. *Nat. Genet.* **39**, 523–528
30. Zhang, Y. X., Xue, Y., Liu, J. Y., Zhao, M. Y., Li, F. J., Zhou, J. M., Wang, H. J., and Li, J. C. (2011) Association of TIRAP (MAL) gene polymorphisms with susceptibility to tuberculosis in a Chinese population. *Genet. Mol. Res.* **10**, 7–15
31. Musacchio, A., Cantley, L. C., and Harrison, S. C. (1996) Crystal structure of the breakpoint cluster region-homology domain from phosphoinositide 3-kinase p85 α subunit. *Proc. Natl. Acad. Sci. U.S.A.* **93**, 14373–14378
32. Kurosaki, T., and Tsukada, S. (2000) BLNK: connecting Syk and Btk to calcium signals. *Immunity* **12**, 1–5
33. Gay, N. J., Symmons, M. F., Gangloff, M., and Bryant, C. E. (2014) Assembly and localization of Toll-like receptor signalling complexes. *Nat. Rev. Immunol.* **14**, 546–558
34. Bernoux, M., Ve, T., Williams, S., Warren, C., Hatters, D., Valkov, E., Zhang, X., Ellis, J. G., Kobe, B., and Dodds, P. N. (2011) Structural and functional analysis of a plant resistance protein TIR domain reveals interfaces for self-association, signaling, and autoregulation. *Cell Host Microbe* **9**, 200–211
35. Wu, Y. Y., Kumar, R., Haque, M. S., Castillejo-López, C., and Alarcón-Riquelme, M. E. (2013) BANK1 controls CpG-induced IL-6 secretion via a p38 and MNK1/2/eIF4E translation initiation pathway. *J. Immunol.* **191**, 6110–6116
36. Kagan, J. C., and Medzhitov, R. (2006) Phosphoinositide-mediated adaptor recruitment controls Toll-like receptor signaling. *Cell* **125**, 943–955
37. Bernal-Quirós, M., Wu, Y. Y., Alarcón-Riquelme, M. E., and Castillejo-López, C. (2013) BANK1 and BLK act through phospholipase C γ 2 in B-cell signaling. *PLoS ONE* **8**, e59842
38. Dal Porto, J. M., Gauld, S. B., Merrell, K. T., Mills, D., Pugh-Bernard, A. E., and Cambier, J. (2004) B cell antigen receptor signaling 101. *Mol. Immunol.* **41**, 599–613
39. Zhou, X., Yang, W., and Li, J. (2006) Ca²⁺- and protein kinase C-dependent signaling pathway for nuclear factor- κ B activation, inducible nitric oxide synthase expression, and tumor necrosis factor- α production in lipopolysaccharide stimulated rat peritoneal macrophages. *J. Biol. Chem.* **281**, 31337–31347
40. Aki, D., Minoda, Y., Yoshida, H., Watanabe, S., Yoshida, R., Takaesu, G., Chinen, T., Inaba, T., Hikida, M., Kurosaki, T., Saeki, K., and Yoshimura, A. (2008) Peptidoglycan and lipopolysaccharide activate PLC γ 2, leading to enhanced cytokine production in macrophages and dendritic cells. *Genes Cells* **13**, 199–208
41. Arsenault, R. J., Kogut, M. H., and He, H. (2013) Combined CpG and poly I:C stimulation of monocytes results in unique signaling activation not observed with the individual ligands. *Cell. Signal.* **25**, 2246–2254
42. Zaroni, I., Ostuni, R., Marek, L. R., Barresi, S., Barbalat, R., Barton, G. M., Granucci, F., and Kagan, J. C. (2011) CD14 controls the LPS-induced endocytosis of Toll-like receptor 4. *Cell* **147**, 868–880
43. Bonham, K. S., Orzalli, M. H., Hayashi, K., Wolf, A. I., Glanemann, C., Weninger, W., Iwasaki, A., Knipe, D. M., and Kagan, J. C. (2014) A promiscuous lipid-binding protein diversifies the subcellular sites of toll-like receptor signal transduction. *Cell* **156**, 705–716
44. Kim, Y. J., Sekiya, F., Poulin, B., Bae, Y. S., and Rhee, S. G. (2004) Mechanism of B-cell receptor-induced phosphorylation and activation of phospholipase C γ 2. *Mol. Cell Biol.* **24**, 9986–9999
45. Schuck, P. (2000) Size-distribution analysis of macromolecules by sedimentation velocity ultracentrifugation and Lamm equation modeling. *Biophys. J.* **78**, 1606–1619
46. Walter, T. S., Meier, C., Assenberg, R., Au, K.-F., Ren, J., Verma, A., Nettleship, J. E., Owens, R. J., Stuart, D. I., and Grimes, J. M. (2006) Lysine methylation as a routine rescue strategy for protein crystallization. *Structure* **14**, 1617–1622
47. Kabsch, W. (2010) XDS. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 125–132
48. Evans, P. R., and Murshudov, G. N. (2013) How good are my data and what is the resolution? *Acta Crystallogr. D Biol. Crystallogr.* **69**, 1204–1214
49. Schneider, T. R., and Sheldrick, G. M. (2002) Substructure solution with SHELXD. *Acta Crystallogr. D Biol. Crystallogr.* **58**, 1772–1779
50. Cowtan, K. (2006) The Buccaneer software for automated model building. 1. Tracing protein chains. *Acta Crystallogr. D Biol. Crystallogr.* **62**, 1002–1011
51. Emsley, P., Lohkamp, B., Scott, W. G., and Cowtan, K. (2010) Features and development of Coot. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 486–501
52. Jones, T. A., Zou, J. Y., Cowan, S. W., and Kjeldgaard, M. (1991) Improved methods for building protein models in electron density maps and the location of errors in these models. *Acta Crystallogr. A* **47**, 110–119
53. Adams, P. D., Afonine, P. V., Bunkóczi, G., Chen, V. B., Davis, I. W., Echols, N., Headd, J. J., Hung, L.-W., Kapral, G. J., Grosse-Kunstleve, R. W., McCoy, A. J., Moriarty, N. W., Oeffner, R., Read, R. J., Richardson, D. C., *et al.* (2010) PHENIX: A comprehensive Python-based system for macromolecular structure solution. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 213–221
54. Gietz, R. D., and Schiestl, R. H. (2007) High-efficiency yeast transformation using the LiAc/SS carrier DNA/PEG method. *Nat. Protoc.* **2**, 31–34

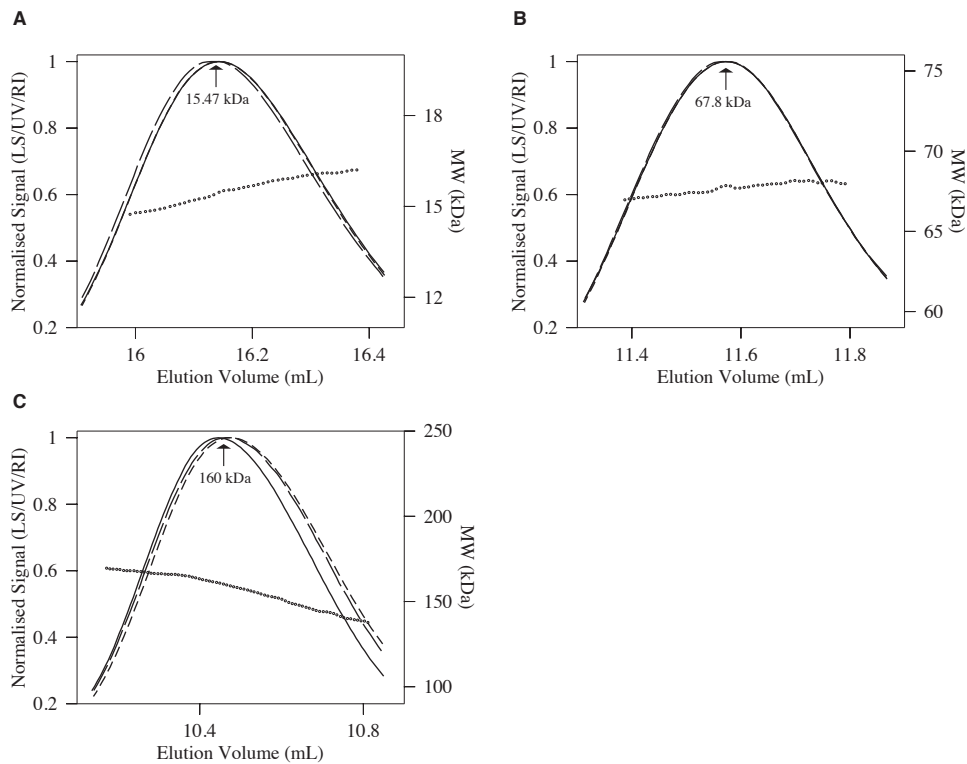
SUPPLEMENTARY MATERIALS

Supplementary Table S1: Crystallographic Statistics for the N-terminal TIR domain of human BCAP

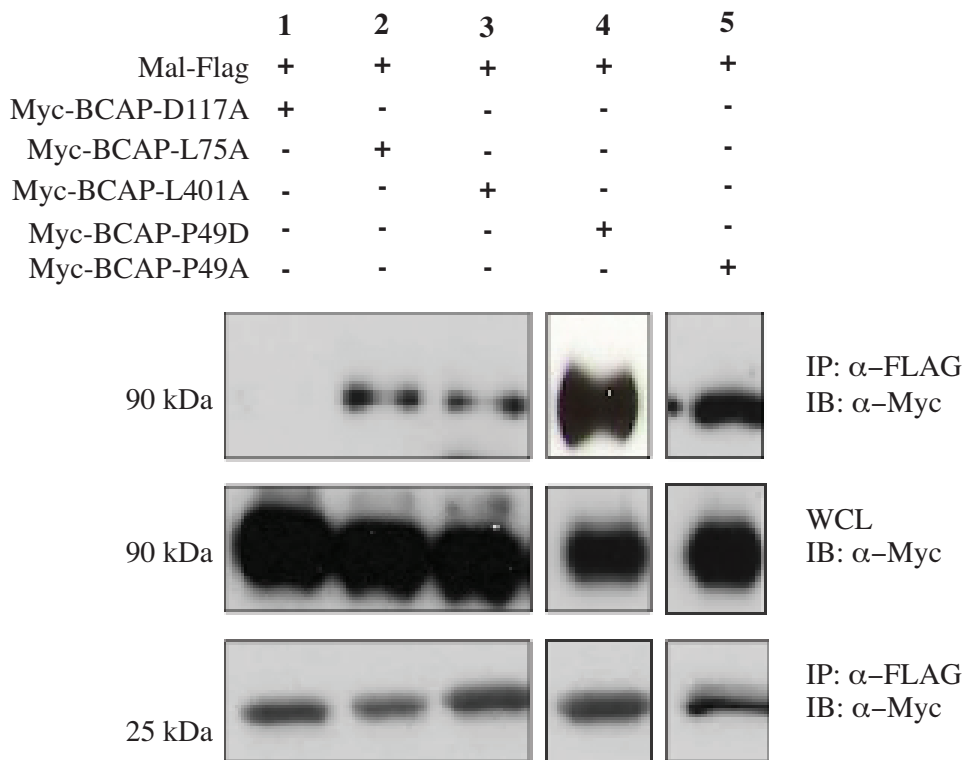
BCAP-TIR (I3C)	
Data Collection	
Space group	P62 2 2
Cell dimensions	
<i>a</i> = <i>b</i> , <i>c</i> (Å)	135.04, 42.91
Resolution (Å)	44.2–2.5
Total reflections	99543 (9785)
Unique reflections	15066 (1489)
<i>I</i> / σ <i>I</i>	24.8(7.2)
Completeness (%)	99(98)
Redundancy	6.6(6.6)
Wilson B-factor	53.84
R-merge	0.047 (0.24)
R-meas	0.051 (0.26)
CC1/2	0.99 (0.98)
CC*	1.0 (1.0)
Phaser-EP FOM	0.35
Refinement	
No. reflections	15066 (1489)
<i>R</i> _{work} / <i>R</i> _{free}	0.19/0.24
No. atoms	
Protein	1088
Ligands	32
Avg. <i>B</i> -factor	59.7
R.m.s. deviations	
Bond lengths (Å)	0.012
Bond angles (Degrees)	1.60
Validation	
Ramachandran favoured/allowed (%)	96/100
Rotamer Outliers (%)	7.8
Clashscore	7.7



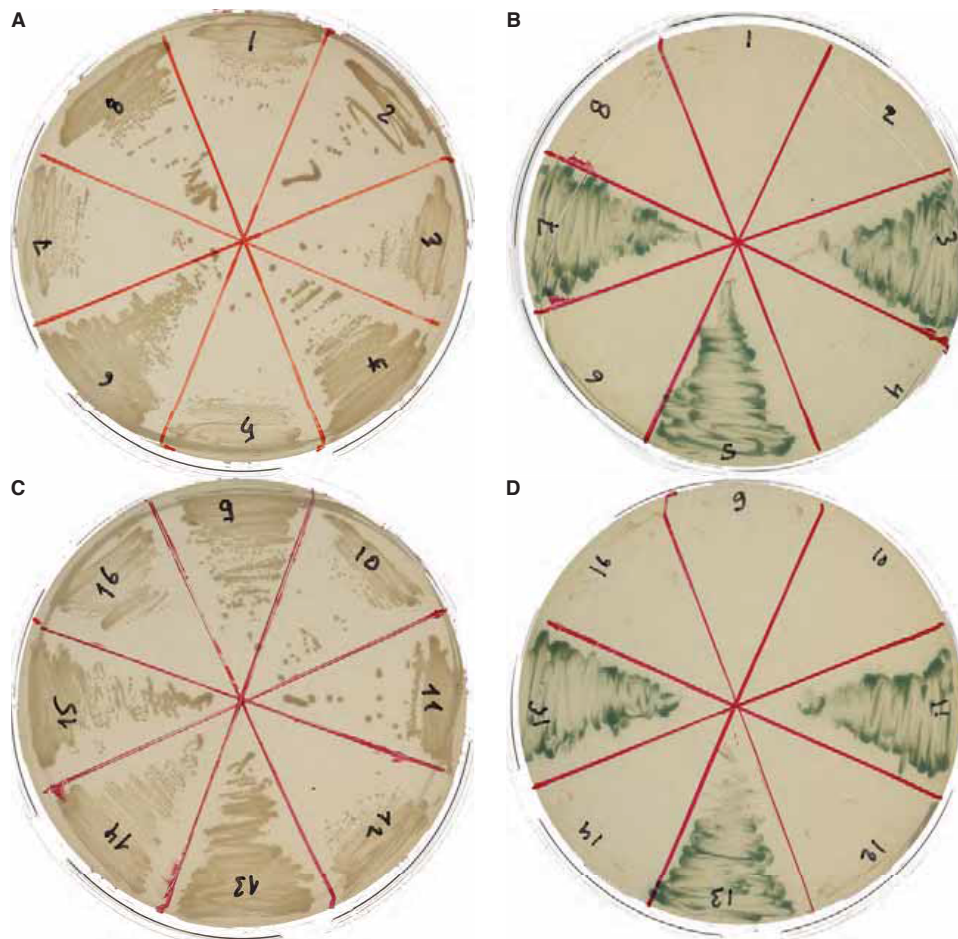
Supplementary Fig. S1: Structural alignment of the BCAP-TIR and the TIR-domains of MAL (2y92), TLR1 (1fyv), TLR2 (1o77), TLR6 (4om7) and TLR10 (2j67). The secondary structural elements shown are those of the BCAP-TIR domain (5for).



Supplementary Fig. S2: Size exclusion chromatography in combination with multi-angle light scattering (SEC-MALS) and differential refractometry data for the N-terminal TIR domain of BCAP (A), BCAP₃₃₀ (B) and BCAP_L (C). Plots show the normalised UV (280 nm) light scattering, refractive index and weight-averaged molecular mass (MW) variation across the peak in addition to the MW at the peak. The molecular weights obtained by SEC-MALS are in complete agreement with those obtained by sedimentation velocity.)



Supplementary Fig. S3: The highly conserved proline residue (P49) in the β B-loop of BCAP is not required for Mal/TIRAP interaction. HEK293T cells were transfected with FLAG-Mal, Myc-Mal and Myc-BCAP as described. Both β B loop proline mutations (P49A/P49D) allowed the BCAP-Mal/TIRAP interaction (lanes 4–5) to occur. Mutations in the α C' helix (L75A) and outside the TIR domain (L401A) both exhibit reduced interaction with Mal/TIRAP (lanes 2–3) while the D117A (lane 1) mutation which lies in the region connecting the α D helix and β D strand completely abolishes the BCAP and Mal/TIRAP interaction.



	pGADT7	pGADT7[TIRAP]	pGADT7[D96N]	pGADT7[S180L]
pGBKT7[BCAP-TDA]	1	3	5	7
pGBKT7	2	4	6	8
	pGADT7	pGADT7[TLR2]	pGADT7[TLR4]	pGADT7[TIRAP]
pGBKT7[BCAP-TDA]	9	11	13	15
pGBKT7	10	12	14	16

Supplementary Table S2: Plasmids and primers used for protein expression in bacteria, yeast and human cells. The restriction sites used, when relevant are shown.

Protein	Oligonucleotide Sequence (5' → 3')	Vector
BCAP-TIR(F)	TACTTCCAATCCAATGCCCCAGAGGATGCGACATCCTCATC	pMCSG7
BCAP-TIR(R)	TTATCCACTTCCAATGTTAGGAAATGGCTTTTTTTCACAGCTGCCAC	
BCAP ₅ (F)	TACTTCCAATCCAATGCCATGGTGGTGCAGCCGGACCG	pMCSG7
BCAP _{L/S} (R)	TTATCCACTTCCAATGTTACTTCTCGAACTGGGGGTGACTC CAGCGTCCTCTGGGTGG	pMCSG7
BCAP _{L/330} (F)	TACTTCCAATCCAATGCCATGGCAGCCTCAGGG	pMCSG7
BCAP ₃₃₀ (R)	TTATCCACTTCCAATGTTACTTTTTCAAACCTGCGGATGCGACCAG GCACTGGCATTGTCATCATATCTTCTTCTTCCAGCTGGTTG	
FLAG-BCAP _L (F)	ATCAAGCTTATGGCAGCCTCAGGGGTGCC (HindIII)	pCMV10
FLAG-BCAP _L (R)	ATCGGATCCTCAGCGTCCTCTGGGTGGAACAGG (BamHI)	
Myc-BCAP _L (F)	Same as FLAG-BCAP _L (F) (HindIII)	pCMV-Myc
Myc-BCAP _L (R)	ATCGGTACCGCGTCCTCTGGGTGGAACAGG (KpnI)	
Myc-BCAP-SV (F)	ATCAAGCTTATGGTGGTGCAGCCGGACCG (HindIII)	pCMV-Myc
Myc-BCAP-SV (R)	ATCGGTACCGCGTCCTCTGGGTGGAACAGG (KpnI)	
BCAP-TDA (F)	ATCCATATGATGGCAGCCTCAGGGGTGCC (NdeI)	pGBKT7 AD
BCAP-TDA (R)	ATCGGATCCTCACGAGCATTTTCATAAGCAGGTCTGTGG (BamHI)	
MAL (F)	ATCCATATGATGGCATCATCGACCTCCCTCCC (NdeI)	pGBKT7 AD
MAL (R)	ATCGGATCCTCAAAGTAGATCAGATACTGTAGCTGAATCCCG (BamHI)	
TLR2-TIR (F)	ATCCATATGAGCAGGAACATCTGCTATGATGCATTTGTTTC (NdeI)	pGBKT7 AD
TLR2-TIR (R)	ATCGGATCCTCAGGACTTTATCGCAGCTCTCAGATTTACC (BamHI)	
TLR4-TIR (F)	ATCCATATGGAAAACATCTATGATGCCTTTGTTATCTACTCAAG (NdeI)	pGBKT7 AD
TLR4-TIR (R)	TCGGATCCTCATGATTTACCATCCAGCAGGGCTTTTCTG (BamHI)	

Structure of the Toll/Interleukin-1 Receptor (TIR) Domain of the B-cell Adaptor That Links Phosphoinositide Metabolism with the Negative Regulation of the Toll-like Receptor (TLR) Signalosome

Samer Halabi, Eiki Sekine, Brett Verstak, Nicholas J. Gay and Martin C. Moncrieffe

J. Biol. Chem. 2017, 292:652-660.

doi: 10.1074/jbc.M116.761528 originally published online December 1, 2016

Access the most updated version of this article at doi: [10.1074/jbc.M116.761528](https://doi.org/10.1074/jbc.M116.761528)

Alerts:

- [When this article is cited](#)
- [When a correction for this article is posted](#)

[Click here](#) to choose from all of JBC's e-mail alerts

Supplemental material:

<http://www.jbc.org/content/suppl/2016/12/01/M116.761528.DC1>

This article cites 54 references, 20 of which can be accessed free at <http://www.jbc.org/content/292/2/652.full.html#ref-list-1>