

Case Study

Design and fabrication of a three-dimensional microfluidic device for blood separation using micro-injection moulding

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Abstract: Micro-manufacturing is a fast developing area due to the increasing demand for components and systems of high-precision and small dimensions. A number of challenges are yet to be overcome before the full potential of such techniques is realised. Examples of such challenges include limitations in component geometry, materials selection and suitability for mass production. Some micro-manufacturing techniques are still at early development stages, whilst other techniques are at higher stage of manufacturing readiness level but require adaptation in part design or manufacturing procedure to overcome such limitations. This paper presents a case

study, where the design of a micro-scale, biomedical device is adapted for functionality and manufacturability by a high-volume micro-fabrication technique. Investigations are described towards a disposable three-dimensional, polymer-based device for the separation of blood-cells and plasma. The importance of attempting a 3-D device design and fabrication route was to take advantage of the high-throughput per unit volume that such systems can in principle allow. The importance of a micro-moulding fabrication route was to allow such blood containing devices to be cheaply manufactured for disposability. Initial device tests showed separation efficiency up to approximately 80% with diluted blood samples. The produced prototype indicated that the process flow was suitable for high-volume fabrication of 3-D microfluidics.

Keywords: microfluidics design, micro-manufacturing, three-dimensional, blood separation, lamination, micro-injection moulding

1. Introduction

This paper presents investigations towards a disposable three-dimensional polymer-based device for the separation of blood-cells and plasma. The paper is structured into three main sections. The first is about the design of the blood separator in terms of functionality and manufacturability, showing how to adapt a micro-scale, complex

design of a biomedical device into a high-volume manufacturing technique. The second section focuses on fabrication procedure and considerations for micro-injection moulding. Finally, results from a prototype device are presented and assessed.

To introduce the topic: four areas of research are briefly summarised here: how blood-plasma can be separated in microfluidic devices, why consider three-dimensional devices, laminable layers as a method of producing such devices, and the production of such layers by micro-injection moulding (μ IM).

1.1 Blood-Plasma Separation by Flow-Focusing

Blood-plasma separation is an important step in several applications related to human health monitoring and disease diagnostics [1]. The presented microfluidic device offers an alternative blood-separation approach to conventional plasma extraction techniques such as centrifugation, blood filtration or CD-like platforms. It depends in operation on several biomechanical separation principles that are combined to produce a separation between plasma and whole blood within micro-channels.

Briefly, the three principles of separation are outlined as follows:

- (a) Laminar flow: When blood flows at relatively low Reynolds number (0.01 to 1) in microchannels of dimension comparable to the cells dimension, red blood cells (RBCs) exhibit a number of flow behaviours that causes the cells to concentrate at the centre of a microfluidic channel, creating a plasma-rich layer adjacent to the channel walls.

(b) Flow focusing: The movement of RBCs in the blood flow is dependent on the magnitude of shear forces exerted on the cells. This relation has led to the assumption that the existence of a constriction in the blood flow will create a zone of high shear stress, which pushes the cells to the middle of the flow, creating a flow-focusing effect as shown in Fig. 1.

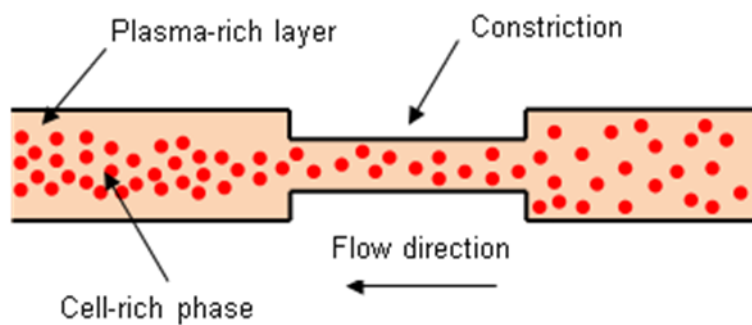


Fig. 1 An illustration of the flow-focusing effect of microfluidic constriction

(c) Bifurcations: RBCs exhibit a specific behaviour in bifurcations. As shown in Fig. 2, at bifurcations, RBCs have the tendency to travel to the high-flow-rate channels, with the largest cross section, whilst plasma tends to move in low-flow-rate channels. One condition for this effect is that the flow rate ratio is at least 2.5:1.

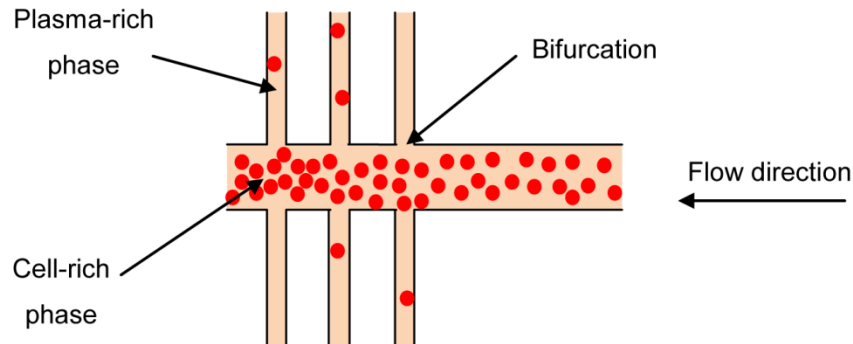


Fig. 2 An illustration of the plasma-separation effect caused by bifurcations

More details about the hemodynamic effects involved in the above three principles can be found in the literature [1-3]

1.2 Three Dimensional Microfluidic Devices

A major design limitation in microfluidics is the geometrical constraint, which limits the majority of designed microfluidic devices to “flat”, so-called 2½-D, structures. These are usually defined as 2-D structures with a finite depth [4]. Due to the increasing demand for more complex microfluidic devices, there is a need for high-volume techniques to manufacture relatively complex, truly three-dimensional microfluidic systems. Such true 3-D structures, also sometimes known as “out-of-plane” or “vertical” architectures [5], offer the advantages of optimising the use of space to integrate more functionality or increase throughput.

Such advantages are evident in, for example, microfluidics, where transforming conventional 2½-D designs into truly 3-D devices could render higher throughput

within a constrained volume [6]. In addition, 3-D structures have potentially more flexibility in terms of interconnection between the micro-component and the surrounding environment. In a true 3-D microfluidic device, for example, interconnection positioning would not be limited to the plane of the microfluidic structure, as the case with conventional 2½-D devices.

Realising these advantages of 3D structures, the blood separator presented in the previous section would be more efficient in blood-plasma separation if it was designed as an out-of-plane 3D structure rather than a flat chip. Section 2 will give more details about how separation efficiency and throughput is improved by such a design-change.

1.3 Laminated Microfluidics

Turning the blood separator design into a truly 3D structure would make it difficult to directly fabricate with a state-of-the-art mass-manufacturing method. One method for overcoming this would be to manufacture sub-components of the device with relatively simple geometries and assemble them in post-processing.

An approach to assembling microfluidic components to produce 3-D structures is that of lamination. In the literature, a number of attempts have been reported in which 3-D microfluidic devices have been fabricated by laminating, for example, paper and tape [7], laser-cut thin-film plastics [8], soft-lithographed elastomers [9] and as multi-material layered microfluidics [10,11]. Common limitations of these approaches include [5]:

- 1) The reported examples implement prototyping techniques that are not particularly intended for high-volume and low cost production.
- 2) Some techniques are not suitable for fabricating layers with high-aspect ratio microstructures.
- 3) Some techniques are limited to specific types of materials.

In light of such limitations, it would be an important step forward to use the capabilities of a high-volume micro-fabrication technique to produce laminated, polymer 3-D microfluidic devices for disposable lab-on-a-chip (LOC) applications. This would combine both the flexibility of laminate microfluidics and the low-cost associated with high-volume production. In this paper, μ IM is assessed as a potential candidate process for such an application due to its advantages, as highlighted in the next section.

1.4 Micro-Injection Moulding of Laminable Layers

Micro-injection moulding (μ IM) is a micro-replication technique that offers mass-production capabilities of polymeric, metallic and ceramic parts at relatively low cost and short-cycle times (a few seconds up to a few minutes). It has the potential for full-automation, accurate replication and dimensional control [12].

In addition to high-volume manufacturing capabilities, μ IM was particularly selected for this application because suitable mould design would enable the production of all

sub-components in a single shot. This would avoid differences in shrinkage between in sub-components. (More details about this are presented in Sections 2.3 and 2.4).

Very few 3-D laminate structures have been produced by μ IM, where a “laminate structure” in this case refers to a microfluidic multiple-layer structure and not simply a microfluidic substrate and lid. In one example, an LOC device was designed for detecting metabolic parameters [13,14], where 3-4 layers of micro-moulded thermoplastics were fabricated and joined by thermal fusion.

In spite of the considerable potential of the technique, little has been mentioned in the literature on how to design a microfluidic laminate structure for manufacturing by micro-injection moulding. Specifically, how to adapt the device design for manufacturing limitations, namely mould making and replication, how to consider design for assembly options, and how to deal with possible contradictions between different design constraints.

2. Experimental

The approach taken to designing a micro-mouldable laminate microfluidic device was to assess the design limitations of each activity in the process flow and then resolve potential conflicts between different design requirements. The approach balanced the functional requirements of the device (dimensions, tolerances, etc.) with the

manufacturability limitations of the process (μIM in this case). Fig. 3 summarises this approach.

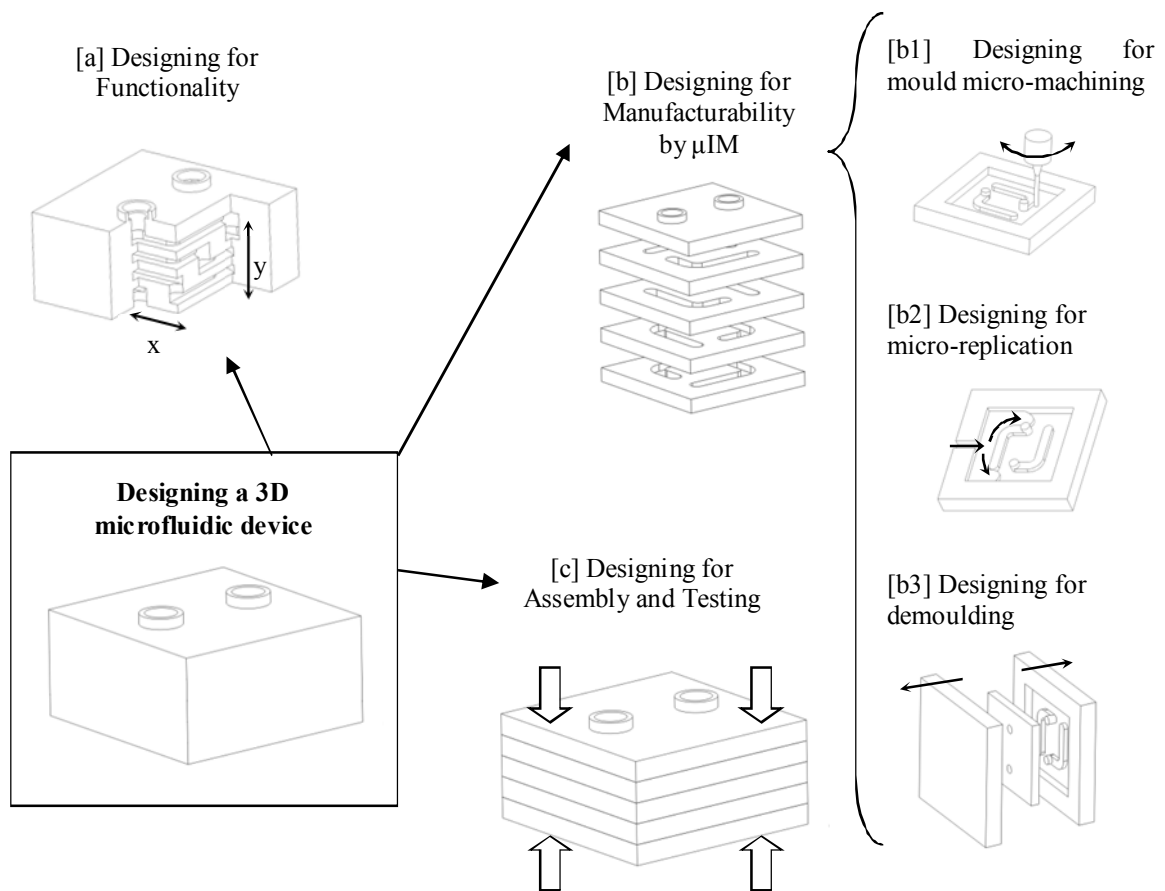


Fig 3. An illustration of the design approach to produce 3-D microfluidic devices by μIM .

Designing for functionality was a first priority in the process; fluid channel sizes, tolerances and orientations were determined (step [a]). The device “layers” were

designed to be micro-mouldable without compromising the functionality of the device (step [b]).

In μ IM as a net-shape micro-fabrication method, part design must involve assessing the three major steps of the process: mould-machining, feature replication and demouldability (denoted in Fig. 3 as steps [b1], [b2] and [b3], respectively). Each of the three steps had its design limitations in the micro-scale manufacturing.

The third major design consideration was joining the layers to produce a leak-proof system (step [c]). This involved not only designing for a specific joining technique but also considerations of part separation and handling, alignment techniques and testing.

The design stages are considered in more detail below.

2.1 Design for Functionality Requirements

The main functional requirements of the device were summarized into the following main groups:

2.1.1 Dimension-Related Requirements. (1) Laminar flow: plasma flow is laminar in microfluidic channels with hydraulic diameter ranging from 10-500 μm [1]. In addition, after the constriction zone, the blood flow should go into a bifurcation zone, where the plasma separates from the main flow path and is then extracted from a separate output. The flow ratio between the main channel and the side paths should be at least 2.5:1. (2) Flow focusing: a constriction zone is required in the blood flow path to induce a high-

shear zone for flow focusing. From the literature, there are no specific guidelines on the dimensions of the constriction relative to the main flow path.

2.1.2 Geometry-Related Requirements. (3) High throughput: A 3-D structure of the microfluidic device would allow for potentially higher throughput of the device, more flexibility in interconnection design and the possibility of future integration into a total analysis cell. Concerning separation efficiency, increasing the number of sequential bifurcation points will increase the separation yield. However, as the number of bifurcation points increases, the flow rate of plasma in the side channels decreases to the extent that no more plasma is extracted from the last bifurcations. A flexible design would be required to change the number of bifurcation points when desired.

2.1.3 Assembly-Related Requirements. (4) Interconnection: The device should be connectable to surrounding environment. (5) Leak-proof system: The microfluidic device should be designed to minimise possible leakage of fluidic samples, especially in bonding/joining positions.

2.1.4 Material/Manufacturing-Related Requirements. (6) Optical transparency: This is required to observe the flow inside the channels during testing stage, but may also be useful for the device in service so that any channel clogging can be observed. (7) Biocompatibility: The device material should be suitable for blood-flow applications. (8) Disposability: The device should be designed such that it is manufacturable by a high-volume technique that allows for low-cost disposable devices.

2.2 Conceptual Design

Fig. 4 presents a design for the blood plasma separator based on the functional requirements outlined in the previous section.

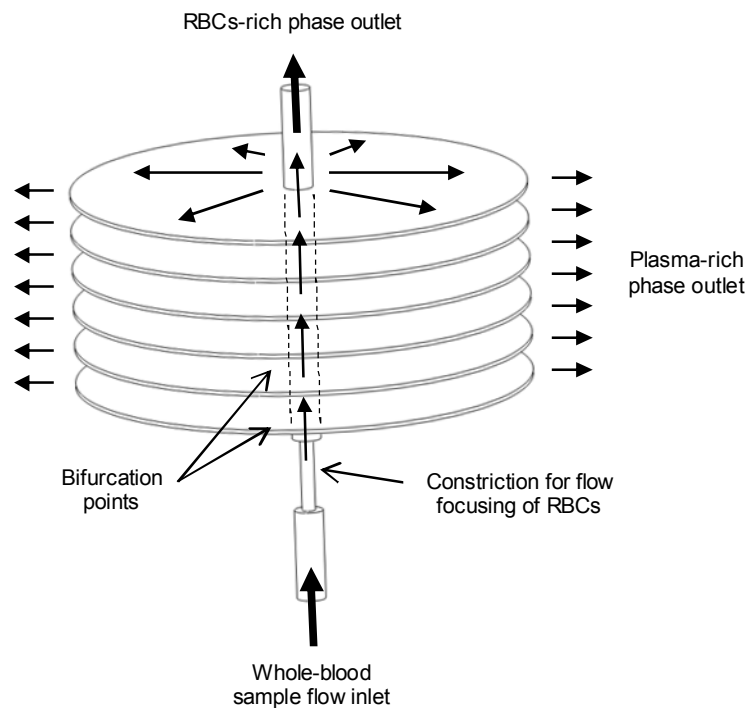


Fig 4. An illustration of the 3-D plasma-separation microfluidic device.

The device consists of an inlet channel where the continuous flow of a blood sample is inserted at a controlled flow rate. The diameter of the microfluidic channel was

selected to be 400 μm to ensure continuous laminar conditions as stated in Section 2.1.1.

A constriction is placed after the inlet to induce the flow-focusing effect resulting from high shear conditions. Since no recommended dimensions were available in the literature, the dimensions were selected as 100 μm in diameter and 600 μm in length.

After the flow focusing zone, a number of bifurcations were introduced to separate the plasma from the main flow. Instead of conventional 2½-D bifurcations usually presented in literature, the design was based on 3-D bifurcations, where microfluidic “surfaces” intersect with the main flow path allowing for a relatively larger amount of plasma separation. The thickness of each separation space was selected to be 50 μm .

Since separation yield is directly related to the number of bifurcations, the design was made to accommodate a number of separation layers across the flow path. 20 bifurcations were suggested as a maximum for a 2½-D design to avoid having excess channels that do not extract any plasma [1]. With the available knowledge, the optimum number of bifurcations is a matter of trial and error, so the 3-D structure was designed such that the manufacturing process allows for the flexibility of adding extra separation cavities when needed.

Numerical simulation tests were made to evaluate the effect of the selected dimensions on the separation efficiency of the 3-D device and of possible geometry modifications. The simulations were performed using a developed computational fluid

dynamics (CFD) code in ANSYS CFX5. The simulations demonstrated that separation throughput for the 3D design is much higher than a 2½-D design, because the cross-section of the main channel and the coupled intersection between the main and side channels are largely increased in 3D separator, leading to a much higher throughput and separated flow rates than the 2D separator [15,16].

2.3 Design for Micro-Injection Moulding

2.3.1 Designing for Demouldability. Similar to conventional “macro-scale” injection moulding, μ IM poses a number of limitations on mouldable part geometries. The major limitations are related to the demouldability of the part, i.e. separating the moulded component from the mould at the end of the injection process. Design considerations include [17], for example, the location of the parting line (separation surface between the two mould halves), introduction of draft angles (tapering of vertical surfaces aligned with the opening direction of the mould) and elimination of undercuts (elements that prevent either the core mould-half from being extracted after the component has been formed, or the component from being ejected out of the cavity [18]).

The latter design consideration, that of undercuts, is the main design concern in this case study, since this is what prevents the direct fabrication of three-dimensional microfluidic components by injection moulding. As a result, the design shown in Fig. 4 would not be producible by μ IM as a single part due to geometrical complexity that prevented it from being directly demouldable. It was, therefore, necessary to adapt the

design to the geometry-limitation of the process, and the approach followed in this case was to produce the device as a number of parts that could be assembled in a subsequent assembly process.

The device was designed as a laminate of mouldable layers. Fig. 5 illustrates how the microfluidic device in Fig. 4 was adapted to μ IM by transforming it into a laminate structure. The dotted lines highlight suggested separation levels, which would eventually render a set of demouldable 2½-D layers.

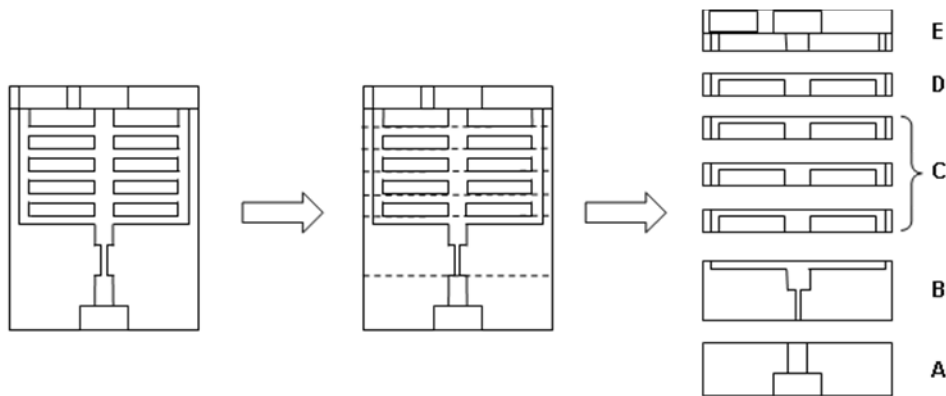


Fig 5. Transferring a 3-D structure into a laminable structure. All individual layers are 2½-D in structure. Extra layers of C correspond to extra separation bifurcations.

The design in Fig. 5 features five layers, denoted from A to E, where each layer performs a specific function or functions. Though not obligatory to the design process, this meant that repeats of functionality would be achievable simply by adding more

layers of a particular type. All the layers are effectively demouldable in the sense that they all feature undercut-free, 2½-D geometries that could be ejected from basic two-half moulds. Layer functionality was as follows:

- Layer A: An input port for tube interconnection and sample delivery.
- Layer B: A multi-functional layer, which contains both a constriction for flow focusing and a separation depression.
- Layer C: A stackable separation surface (bifurcation junction). Theoretically speaking, this layer could have been omitted from the design, since separation is already performed in Layer B. However, it was designed as a repeatable unit to meet the requirement for expandable bifurcations.
- Layer D: A plasma collection channel.
- Layer E: Output port for tube interconnection and extraction of both RBC- and plasma-rich phases.

2.3.2 Designing for Micro-Replication. A major challenge in μ IM is the ability to fill micro-size cavities in the mould. Filling quality is a factor of several interdependent parameters, including mould geometry (e.g. feature sizes, aspect ratios [19]), mould surface properties [20,21], polymer thermal and viscoelastic properties [22] and processing conditions [23-26].

For the blood separator, the critical replication challenge was to accurately create the through-hole micro features located at each of the five layers. Each layer was designed

as thickness of 1 mm to allow for stable material flow inside the cavity during moulding. In addition, the runner and gate system was designed to supply enough volume of filling material inside the mould cavity to avoid premature freezing. Process conditions were also optimised for complete part filling.

2.3.3. Designing for Mould Micro-Machining. The mould was designed to be manufactured as a set of five interchangeable elements that produced the features of the five layers of the device. Five micro-structured aluminium inserts, the carrying steel holder and the mould housing all fit together within an H7 transitional fit. This reconfigurable structure was selected to allow for the flexibility of changing the design and/or re-machining the features when deemed necessary. The time and cost involved in machining moulds, especially with micro-features, makes it a high risk to machine the whole mould as a single piece. This manufacturing path was selected as an alternative approach for prototyping purposes.

When designing the parts, micro-machining limitations were taken into consideration. The main concern was the aspect ratio of the microfeatures, which is limited by the dimensions of the micro-milling cutting tools used. Considering that the overall thickness of each layer was 1 mm, the maximum aspect ratio in all the parts was 5, which was safely below the reported maximum aspect ratio of micro-milling, which is approximately 10 to 15 [27,28].

It should be noted that micro-milling poses other limitations in terms of, for example, surface finish ($R_a \approx 0.3 \mu\text{m}$), feature accuracy (3-10 μm) and burr-generation. In this particular case study, these aspects were not critical to the part design, given the functional dimensions noted in Section 2.1.

2.4 Design for Micro-Assembly

Designing for assembling micro-moulded parts requires the consideration of a number of issues including [29], for example, the use of the runner system as a handling tool, the effect of parting-line marks, draft angles and gate location. In addition, it is advisable to avoid bonding microfluidic laminates directly over voids created by, for example, channels or reservoirs [30].

The blood separator was designed such that the separator layers could be stacked and bonded between flat mating surfaces using a suitable polymer-joining method. Several design elements were incorporated to aid post-moulding assembly:

- All the layers were designed with axially symmetric features, such that no specific axial orientation was required between adjacent layers. This limited the requirements of assembling to alignment rather than both alignment and orientation.

- The layers were all designed in circular shapes of equal diameters. When the parts would be stacked for welding, the outer diameter of the layers would be selected as the alignment edge. This registration approach, also known as “edge-alignment” [5], relies on using a jig to align laminated layers using edges or corners. The jig was designed to

align the parts without interference from side-gate marks that might result from separating the parts from the runner system.

- The outer diameter of the layers was set to 10 mm with the separation features located in the centre of the parts. This allowed for a peripheral area, usable for the joining process, that was away from the microfluidic features, to minimise feature deformation that might compromise the functionality of the device.

It should be noted that when joining laminates, the flatness of the mating surfaces was critical for ensuring leak-proof joints. In the case study presented in this paper, the flatness of the joining surfaces was minimised during the micro-moulding process using statistical quality control [31].

2.5 Micro-Mould Manufacture

The mould inserts shown in Fig. 6 were manufactured by micromilling using a KERN micromilling centre. The surface finish of the micro-features of two of the aluminium mould inserts are shown in the micrographs of Fig. 6. It should be noted that due to its full axial symmetry requirement, Part A was finished with diamond turning for a better surface finish.

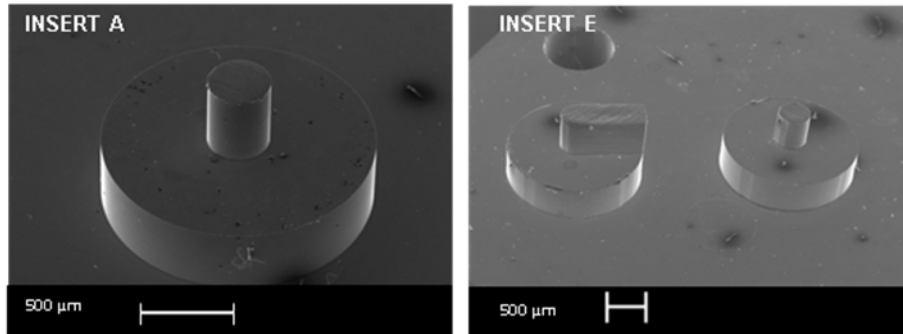


Fig 6. SEM micrographs of the layer A and layer E micro-structured mould inserts.

2.6. Micro-Replication of Component Parts

The polymer layers were moulded with a Battenfeld Microsystems 50 μ IM machine. The material used was Polymethylmethacrylate (PMMA) of the grade Altuglas® VS-UVT. The grade was selected for its ease of flow (MFI = 24 g/10 min), optical transparency (light transmittance 92%) and compatibility with medical applications involving blood. It was also compatible with ultrasonic welding.

A design-of-experiment approach was implemented to optimise processing conditions for complete filling of the features [32-34].

The optimised process conditions were selected as shown in Table 1. The cycle time for the one moulding operation was approximately 5 seconds.

Parameter	T_p [°C]	T_m [°C]	V_i [mm/s]	P_h [bar]	t_c [s]
Value	250	84	200	300	3

Table 1. Processing parameters for a five-part micro-moulded component. From left to right: melt temperature, mould temperature, injection speed, holding pressure and cooling time.

3. Results and Discussion

Fig. 7 shows a photo of the moulded parts, while Fig. 8 shows SEM micrographs of plastic parts A and E.

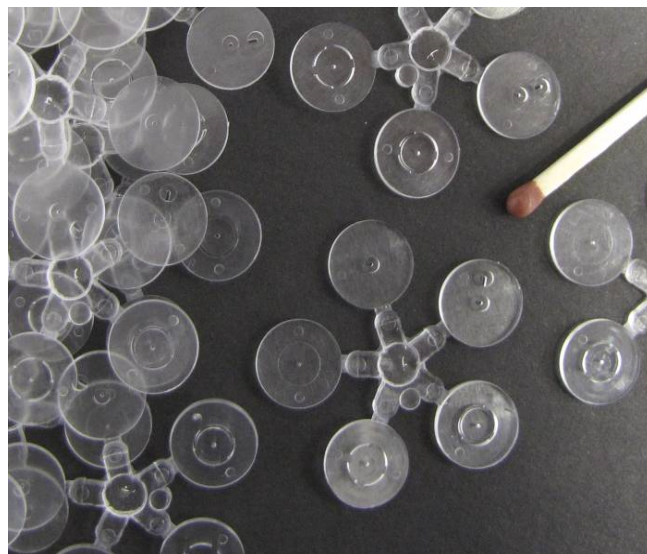


Fig 7. Micro-injection moulded plastic components.

Defibrinated horse blood was used to test the functionality of the device. In order to achieve higher separation efficiency, several separation layers (C-type layers) were added to the tested device.

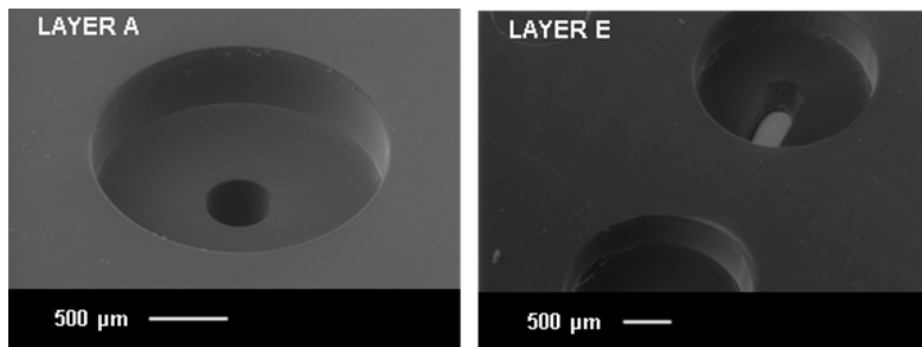


Fig 8. SEM micrographs of PMMA micro-moulded components: layers A and E.

A preliminary assembly was created using a commercial PMMA-compatible adhesive using a pre-designed fixture system. The layers were aligned based on their external edges, and the assembled prototypes were inspected under the microscope, where observed misalignments were in the order of 10 μm. Some deformations related to the flatness of the mating surfaces of the layers were observed. They were found to be due to the ejection forces of the ejector pins. These deformations were detected and minimised using a design-of-experiment approach, where further details are available in the literature [31].

The assembled device was then immersed in a resin to enable cross-sectioning and polishing for optical inspection. Fig. 9 presents an image of an assembled device, with the fitted tubes, while being proof-tested with water.

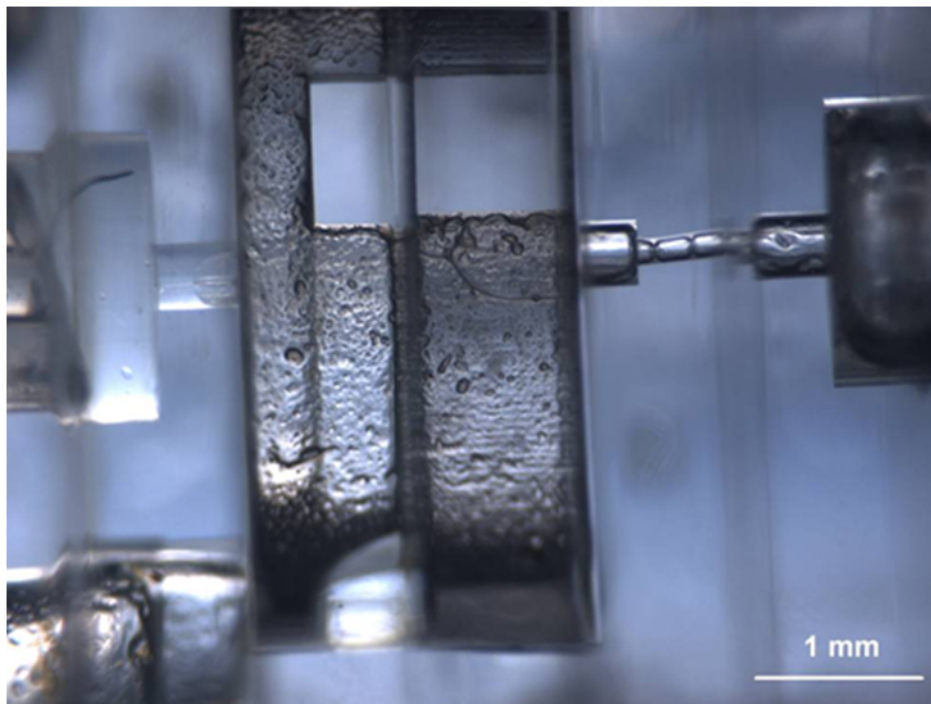


Fig 9. An assembled device being tested with water. The five layers, A to E, run from left to right across the optical micrograph, with the flow-focus constriction in Layer B. An inlet tube is inserted into the device, centre left

Fig. 10 shows a recording of the test, taken using a digital video camera.

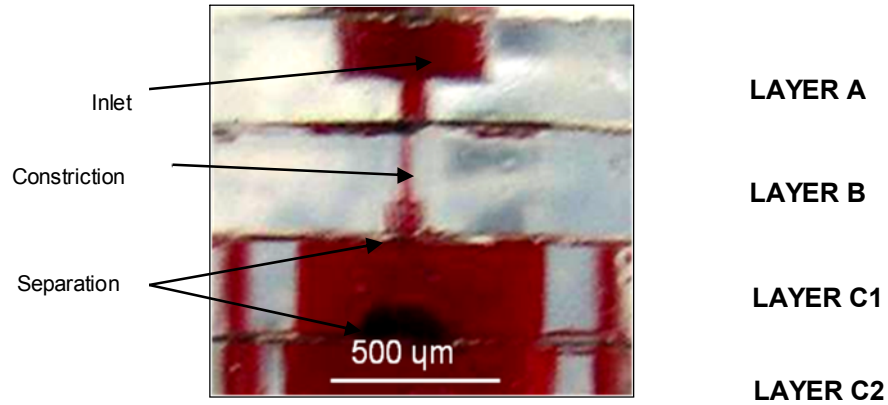


Fig 10. Blood flow through an assembled separator.

Diluted samples (1:20) of horse blood were pumped into the device at a rate of 0.35 ml/min. Separation was assessed for a device example, which contained 3 separation layers (1 B-layer and 2 C-layers). Separation efficiency was characterised by plasma selectivity (σ) defined as:

$$\sigma = 100 \left(1 - \frac{C_{plasma}}{C_{input}} \right) \% \quad (1)$$

In Equation 1, C_{plasma} and C_{input} are the concentrations of RBC in the sample collected at the plasma outlet and in the whole blood sample, respectively. A plasma selectivity of 100% implies that the plasma is completely free of red blood cells. The selectivity measured for this particular experiment was 79.9%.

It should be noted that there are a number of limitations that need to be addressed with regard to the presented design. One limitation is the complexity of the design. Increasing the complexity of the geometry results in increased processing steps, such as alignment and joining. This potentially increases the fabrication cost and time, in addition to increasing risk of leakage. Automation of the process is key to achieve controlled process of manufacturing and joining. Another limitation is the sample dilution, which was selected deliberately high for testing purposes. Using the device for separating concentrated blood samples requires better control of feature dimensions and surface properties. Mould making should be done with a manufacturing technique that offers better control over dimensions and surface properties, such as micro-EDM.

4. Conclusions

This paper described a case study of adapting high-volume micro-fabrication technique to realise a complex three-dimensional component. This was illustrated through designing and fabricating a prototype disposable three-dimensional polymer-based device for the separation of blood-cells and plasma. The design goal of a three-dimensional structure was in order to investigate the potential for three-dimensional maximisation of throughput and of separation efficiency.

The paper indicated the design stages required in such an approach: design for functionality, design for manufacturability, design for assembly and testing. It showed how the micro-moulding of individual micro-parts with different functionality, followed by their lamination could be used to achieve the goal of a 3-D blood separation device.

The designed structure was replicated by μ IM of PMMA, as five different laminable layers. The required micro-features, were successfully replicated. Layers were successfully edge-registered and assembled.

Separation tests of diluted animal blood samples in manually assembled devices showed successful separation of red blood cells and plasma to a 79.9% efficiency.

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References

1. Kersaudy-Kerhoas, M.; Dhariwal, R.; Desmulliez, M.P.Y.; Jovet, L. Hydrodynamic blood plasma separation in microfluidic channels. *Microfluidics and Nanofluidics* 2010, 8, 105-114.

2. Fung, Y.C. *Biomechanics: Mechanical Properties of Living Tissues*, 2nd ed. New York: Springer-Verlag; 1993.
3. Faivre, M.; Abkarian, M.; Bickraj, K.; Stone, H.A. Geometrical focusing of cells in a microfluidic device: an approach to separate blood plasma. *Biorheology* 2006, 43, 147-159.
4. Alting, L.; Kimura, F.; Hansen, H.N.; Bissacco, G. Micro engineering. *CIRP Annals – Manufacturing Technology*, 2003, 52, 635-658.
5. Paul, B.K.; Peterson, R.B. Microlamination for microtechnology-based energy, chemical, and biological systems. In *ASME Advanced Energy Systems Division Proceedings of the ASME International Mechanical Engineering Congress and Exposition*, Nashville, TN, USA, 14-19 November 1999, pp. 45-52.
6. Topham, D.; Harrison, D. The conceptual design of products benefiting from integrated micro-features. 2nd *Electronics Systemintegration Technology Conference (ESTC)*, Greenwich, 1-4 September 2008. DOI: 10.1109/ESTC.2008.4684544
7. Martinez, A.W.; Phillips, S.T.; Whitesides, G.M. Three-dimensional microfluidic devices fabricated in layered paper and tape. *Proceedings of the National Academy of Sciences of the United States of America* 2008, 105, 19606-11.

8. Weigl, B.H.; Bardell, R.; Schulte, T.; Battrell, F.; Hayenga, J. Design and rapid prototyping of thin-film laminate-based microfluidic devices. *Biomedical Microdevices* 2001, 3, 267-274.
9. Unger, M.A.; Chou, H-P.; Thorsen, T.; Scherer, A.; Quake, S.R. Monolithic microfabricated valves and pumps by multilayer soft lithography. *Science* 2000, 288, 113-116.
10. Yang, J.M.; Bell, J.; Huang, Y.; Tirado, M.; Thomas, D.; Forster, A.H.; et al. An integrated, stacked microlaboratory for biological agent detection with DNA and immunoassays. *Biosensors and Bioelectronics* 2002, 17, 605-618.
11. King Jr., C.R.; Sekar, D.; Bakir, M.S.; Dang, B.; Pikarsky, J.; Meindl, J.D. 3D stacking of chips with electrical and microfluidic I/O interconnects. 58th Electronic Components and Technology Conference (ECTC), Lake Buena Vista, FL; 27 - 30 May 2008. DOI: 10.1109/ECTC.2008.4549941
12. Attia, U.M.; Marson, S.; Alcock, J.R. Micro-injection moulding of polymer microfluidic devices. *Microfluidics and Nanofluidics* 2009, 7, 2009, 1-28.
13. Ahn, C.H.; Choi, J-W.; Beaucage, G.; Nevin, J.H.; Lee, J.B.; Puntambekar, A.; Lee, J.Y. Disposable smart lab on a chip for point-of-care clinical diagnostics. *Proceedings of the IEEE* 2004, 92, 154-173.
14. Choi, J.-W.; Puntambekar, A.; Hong, C.-C.; Gao, C.; Zhu, X.; Trichur, R.; Han, J.; Chilukuru, S.; Dutta, M.; Murugesan, S.; Kim, S.; Sohn, Y.-S.; Nevin, J.H.;

- Beaucage, G.; Lee, J.-B.; Lee, J.Y.; Bissell, M.G.; Ahn, C.H. A disposable plastic biochip cartridge with on-chip power sources for blood analysis. Proceedings of the IEEE Micro Electro Mechanical Systems (MEMS), Kyoto, 19-23 January 2003, pp. 447-450.
15. Xue, X.; Marson, S.; Patel, M.K.; Attia, U.M.; Bailey, C.; O'Neill, W.; Topham, D.; Desmulliez, M.P.Y. Biofluid behaviour in 3D microchannel systems: Numerical analysis and design development of 3D microchannel biochip separators. Proceedings of the 60th Electronic Components and Technology Conference (ECTC 2010), 1-4 June 2010, art. no. 5490827, pp. 1021-1030. DOI: 10.1109/ECTC.2010.5490827
16. Xue, X.; Marson, S.; Patel, M.K.; Bailey, C.; O'Neill, W.; Topham, D.; Kay, R.; Desmulliez, M.P.Y. Progress towards the design and numerical analysis of a 3D microchannel biochip separator. *International Journal for Numerical Methods in Biomedical Engineering* 2011, 27, 1771-1792.
17. Osswald, T.A.; Turng, L-S.; Gramann, P.J.; Eds. *Injection Molding Handbook*. Hanser/Gardner Publications, Cincinnati, OH, 2001.
18. Rosen, D.W.; Dixon, J.R.; Poli, C.; Dong, X. Features and algorithms for tooling cost evaluation in injection molding and die casting. In Proceedings of the ASME international computers in engineering conference and exposition, New York, NY, USA, 2-6 August 1992.

19. Sha, B.; Dimov, S.; Griffiths, C.; Packianather, M.S. Micro-injection moulding: factors affecting the achievable aspect ratios. *International Journal of Advanced Manufacturing Technology* 2007, 33, 147-156.
20. Zhang, H.L.; Ong, N.S.; Lam, Y.C. Mold surface roughness effects on cavity filling of polymer melt in micro injection molding. *International Journal of Advanced Manufacturing Technology*, 2008, 37,1105-1112.
21. Griffiths, C.; Dimov, S.; Brousseau, E.; Hoyle, R. The effects of tool surface quality in micro-injection moulding. *Journal of Materials Processing Technology*, 2007, 189, 418–427.
22. Yao, D.; Kim, B. Scaling issues in miniaturization of injection molded parts. *Journal of Manufacturing Science and Engineering, Transactions of the ASME* 2004, 126, 733-739.
23. Zhao, J.; Mayes, R.; Chen, G.; Xie, H.; Chan, P. Effects of process parameters on the micro molding process. *Polymer Engineering and Science* 2003, 43, 1542-1554.
24. Aufiero, R. The effect of process conditions on part quality in microinjection molding. In: *Proceedings of the annual technical conference (ANTEC 2005)*, Boston, MA, 1–5 May 2005. Newtown, CT: Society of Plastics Engineers.
25. Pirskanen, J.; Immonen, J.; Kalima, V.; Pietarinen, J.; Siitonen, S.; Kuittinen, M. et al. Replication of sub-micrometre features using microsystems technology. *Plastics, Rubber and Composites* 2005, 34, 222-226.

26. Sha, B.; Dimov, S.; Griffiths, C.; Packianather, M. Investigation of micro-injection moulding: factors affecting the replication quality. *Journal of Materials Processing Technology* 2006, 183, 284–296.
27. Bissacco, G.; Hansen, H.; Tang, T.; Fugl, J. Precision manufacturing methods of inserts for injection molding of microfluidic systems. *ASPE spring topical meeting on precision micro/nano scale polymer based component and device fabrication*, April 2005.
28. Uriarte, L.; Herrero, A.; Ivanov, A.; et al. Comparison between microfabrication technologies for metal tooling. *Proc IMechE Part C: J Mechanical Engineering Science* 2006; 220: 1665-1676.
29. Bibber, D.M. Micro and ultra precision assemblies - part 1. *Commercial Micro Manufacturing* 2010, 3, 50-53.
30. Garst, S.; Schuenemann, M.; Solomon, M.; Atkin, M.; Harvey, E. Fabrication of multilayered microfluidic 3D polymer packages. *Proceedings of the 55th Electronic Components and Technology Conference (ECTC)*, Lake Buena Vista, FL, 31 May-4 June 2005
31. Marson, S.; Attia, U.M.; Lucchetta, G.; Wilson, A.; Alcock, J.R.; Allen, D.M. Flatness optimization of micro-injection moulded parts: The case of a PMMA microfluidic component. *Journal of Micromechanics and Microengineering* 2011, 21, article no. 115024

32. Attia, U.M.; Alcock, J.R. An evaluation of process-parameter and part-geometry effects on the quality of filling in micro-injection moulding. *Microsystem Technologies* 2009, 15, 1861-1872.
33. Attia, U.M.; Alcock, J.R. Evaluating and controlling process variability in micro-injection moulding. *International Journal of Advanced Manufacturing Technology* 2010, 52,183-194.
34. Attia, U.M.; Alcock, J.R. Optimising process conditions for multiple quality criteria in micro-injection moulding. *International Journal of Advanced Manufacturing Technology* 2010, 50, 533-542.