Species differential regulation of COX2 can be described by an NFkB-dependent logic AND gate

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Supplementary Material

TCTGAGCAGCGAGCACGTCAGACTGCGCCCCAGTGGGGAGAGGTGA<mark>GGGGATTCCCT</mark>TAGTT AGGACCTTAGATCCCGGGAGGGGAAGCTGTGACACTCTTGAGCTTTTAGGCCCCCACTGGAT GCGCGACTGGGAGGAAACCGGAGACCCCAAAGAGCGCCAGACTAGGCGCAGACTCAGCGAAC CACAGGGCGCCTGGAGGGATGGAGAGGGCGGTGCAGCTCTCTTGGCACCACCTGGGGGCAGCC AAGGGCAGCTTCCCGGCTTCCTTCGTCTCTCATTTGCGTGGGTAAAAGCCTGCCGCTGCGGT TCTT

Supplementary Figure S1. Sequence of the mouse COX2 promoter used to create pGlucmCOX2. The NFκB response element is highlighted in yellow. Supplementary Figure S2. Sequence of the human COX2 promoter used to create pGluc-COX2. The NF κ B response elements are highlighted in yellow.



Supplementary Figure S3. Analysis of protein expression and transcriptional activity as a function of TNFa concentration. (**A**, **B**) The data on COX2 protein expression from mouse (A) and human (B) cells (from Figures 1C and D) were fitted with the Hill equation. (**C**, **D**) The data on transcriptional activity from mouse (C) and human (D) COX2 promoter (from Figures 2C and D) were fitted with the Hill equation. Hill coefficients are given in the respective graphs.



Supplementary Figure S4. The AND gate is functional at higher concentration of TNF α . (A) Transcriptional activity of hCOX2 wild-type promoter or mutants in response to 10ng/ml of TNF α (shown as fold activation over unstimulated; n = 4). (B) Truth table showing the relationship between promoter activity (1 = active due to NF κ B, 0 = basal) and presence (1) or absence (0) of NRE for wild-type and mutants hCOX2 promoter. Significant differences (p<0.05) are denoted by *.

CTAGCCTCAATGACGACCTAAGCTGCACTTTTCCCCCTAGTTGTGTCTTGCCATGCTAAAGG ACGTCACATTGCACAATCTTAATAAGGTTTCCAATCAGCCCCACCCGCTCTGGCCCCACCCT CACCCTCCAACAAAGATTTATCAAATGT<mark>GGGATTTTCCC</mark>ATGAGTCTCAATATTAGAGTCTC AACCCCCAATAAATATAGGACTGGAGATGTCTGAGGCTCATTCTGCCCTCGAGCCCACCGGG AACGAAAGAGAAG

Supplementary Figure S5. Sequence of the human IL6 promoter used for creating the pGluc-

IL6. The NF κ B response element is highlighted in yellow.

CTAGCCTCAATGACGACCTAAGCTGCACTTTTCCCCCTAGTTGTGTCTTGCCATGCTAAAGG ACGTCACATTGCACAATCTTAATAAGGTTTCCAATCAGCCCCACCGGCTCTGGCCCCACCCT CACCCCCAACAAAGATTTATCAAATGT<mark>GGGATTTTCCC</mark>ATGAGTCTCAATATTAGAGTCTC AACCCCCAATAAATATAGGACTGGAGATGTCTGAGGCTCATTCTGCCCTCGAGCCCACCGGG AACGAAAGAGAAG TCTGCCCTCGAGCCCACCGGGAACGAAAGAGAAGGAATTC CTAGCCCACAGCTGCACTTTTCCCCCTAGTTGTGTCTTGCCATGCTAAAGGACGTCACAT TGCACAATCTTAATAAGGTTTCCAATCAGCCCCACCGGCTCTGGCCCCACCCTCAACGCTCAA ACAAAGATTTATCAAATGT<mark>GGGATTTTCCC</mark>ATGAGTCTCAATATTAGAGTCTCAACCCCCAA TAAATATAGGACTGGAGATGTCTGAGGCTCATTCTGCCCTCGAGCCCACCCGGGAACGAAAGA GAAG

Supplementary Figure S6. Sequence used for creating the pGluc-2xIL6. The NF κ B response elements are highlighted in yellow. The green highlighted sequence refers to the original pGluc vector.



Supplementary Figure S7. Ensemble simulation of the 2-sites AND- vs OR-gate model. (A) Simulations of the OR-gate model for 1000 random parameter sets randomly drawn from the ranges [0.0001, 0.01] and [0.001, 0.1] for association and dissociation kinetic rates, respectively. Parameter units are given in Table 1. (B) Mean (solid) and +/- one standard deviation (dashed) curves calculated from the dose-response curves in panel (A). (C, D) Similar simulations as in (A, B) for the AND-gate model.