



## ARTICLE

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# Functional mechanisms underlying pleiotropic risk alleles at the 19p13.1 breast-ovarian cancer susceptibility locus

Kate Lawrenson *et al.*<sup>#</sup>

A locus at 19p13 is associated with breast cancer (BC) and ovarian cancer (OC) risk. Here we analyse 438 SNPs in this region in 46,451 BC and 15,438 OC cases, 15,252 *BRCA1* mutation carriers and 73,444 controls and identify 13 candidate causal SNPs associated with serous OC ( $P=9.2 \times 10^{-20}$ ), ER-negative BC ( $P=1.1 \times 10^{-13}$ ), *BRCA1*-associated BC ( $P=7.7 \times 10^{-16}$ ) and triple negative BC ( $P\text{-diff}=2 \times 10^{-5}$ ). Genotype-gene expression associations are identified for candidate target genes *ANKLE1* ( $P=2 \times 10^{-3}$ ) and *ABHD8* ( $P<2 \times 10^{-3}$ ). Chromosome conformation capture identifies interactions between four candidate SNPs and *ABHD8*, and luciferase assays indicate six risk alleles increased transactivation of the *ADHD8* promoter. Targeted deletion of a region containing risk SNP rs56069439 in a putative enhancer induces *ANKLE1* downregulation; and mRNA stability assays indicate functional effects for an *ANKLE1* 3'-UTR SNP. Altogether, these data suggest that multiple SNPs at 19p13 regulate *ABHD8* and perhaps *ANKLE1* expression, and indicate common mechanisms underlying breast and ovarian cancer risk.

Correspondence and requests for materials should be addressed to A.C.A. (email: [aca20@medschl.cam.ac.uk](mailto:aca20@medschl.cam.ac.uk)).  
<sup>#</sup>A full list of authors and their affiliations appears at the end of the paper.

Genome-wide association studies (GWAS) have identified more than 100 different genetic susceptibility regions for breast cancer (BC)<sup>1–6</sup> and 20 regions for epithelial ovarian cancer (EOC)<sup>7–13</sup>. A few of these regions, and in some cases the same genetic variants, are associated with risks of both cancers (pleiotropy), suggesting there may be underlying functional mechanisms and biological pathways common to different cancers. The *TERT-CLPTMIL* locus (5p15) is one such example in which the same variants are associated with risks of oestrogen receptor (ER)-negative BC, BC in *BRCA1* mutation carriers and serous invasive OC<sup>10</sup>.

Few studies have comprehensively described the functional mechanisms underlying common variant susceptibility loci<sup>10,14–18</sup>. More than 90% of risk alleles lie in non-protein-coding DNA and there is now unequivocal evidence that susceptibility regions are enriched for risk-associated single-nucleotide polymorphisms (SNPs) intersecting regulatory elements, such as transcriptional enhancers, predicted to control the expression of target genes *in cis*<sup>19–21</sup>. Establishing causality for risk SNPs is very challenging; of the thousands of risk associations identified by GWAS, functional validation of causal variants using genome editing has only been experimentally performed for two SNPs, one for prostate cancer<sup>22</sup> using the CAUSEL pipeline and the other for obesity<sup>23</sup>. Thus, there is a critical need to identify the causal risk SNP(s) and the overlapping regulatory element(s) and the target gene(s) regulated in an allele-specific manner.

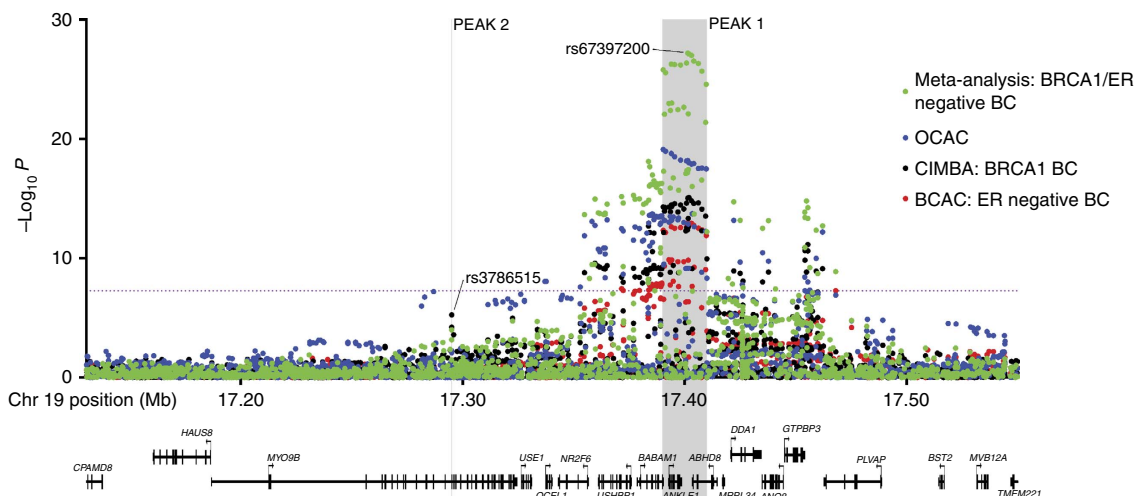
Breast and high-grade serous OC share common genetic and non-genetic risk factors, with mutations in *BRCA1* and *BRCA2* the most significant risk factors for both cancers, suggesting similar biological mechanisms drive breast and OC development. A region on chromosome 19p13.1 has previously been associated with susceptibility to BC and OC in the general population, and to modify the risks of *BRCA1*-related BC and *BRCA2*-related OC<sup>9,24–27</sup>. Initial studies indicated that the association signal was centred around the SNP rs8170 located in the *BRCA1*-interacting gene *BABAM1* (ref. 9), and subsequent studies have refined the subtype specific BC risks associated with these SNPs<sup>24–26,28</sup>.

In the current study, we hypothesized that the same functional mechanism underlies the 19p13.1 risk association in both BC and OC. To evaluate this hypothesis we performed genetic fine mapping in BC and OC patients and in *BRCA1* mutation carriers,

and performed a wide range of functional assays in breast and ovarian tissues and *in vitro* models to identify the likely causal alleles, and target regulatory elements and susceptibility gene(s). Our data indicate that multiple SNPs are involved in the regulation of *ABHD8* and perhaps *ANKLE1* at this locus.

## Results

**Genetic association analyses with breast and OC risks.** A total of 438 SNPs spanning 420 kb at the chromosome 19p13 locus (nucleotides 17,130,000–17,550,000 (NCBI build 37)) were genotyped successfully in the following populations: 46,451 BC cases (of which 7,435 cases had ER-negative tumours) and 42,599 controls from the Breast Cancer Association Consortium (BCAC); 15,438 cases of EOC (of which 9,630 were of serous histology) and 30,845 controls from the Ovarian Cancer Association Consortium (OCAC); and 15,252 *BRCA1* mutation carriers from the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA; 7,797 with BC and 7,455 unaffected; Supplementary Table 1). Genotypes for variants identified through the 1,000 genomes project (minor allele frequency (MAF) > 0.1%) were imputed for all participants of European ancestry. A total of 2,269 genotyped and imputed SNPs were analysed for their associations with ER-negative BC risk in the general population, 2,311 SNPs with BC/OC risk for *BRCA1* mutation carriers, and 2,565 SNPs with risk of serous OC. Results for all SNPs associated with these phenotypes at  $P < 10^{-4}$  are illustrated in Fig. 1 and Supplementary Fig. 1. Two perfectly correlated SNPs rs61494113 and rs67397200 located between the *ANKLE1* and *ABHD8* genes demonstrated the strongest association with BC risk among *BRCA1* mutation carriers ( $\chi^2$ -test  $P = 7.8 \times 10^{-16}$ ) and ER-negative BC in BCAC ( $\chi^2$ -test  $P = 1.3 \times 10^{-13}$ ,  $P$ -meta-analysis =  $7.3 \times 10^{-28}$ ). There was no association for ER-positive BC ( $\chi^2$ -test  $P = 0.21$  for rs61494113). The strongest association with invasive and serous OC was for rs4808075 (correlated with rs61494113 with  $r^2 = 0.99$ ) located in the *BABAM1* gene ( $\chi^2$ -test  $P = 9.2 \times 10^{-20}$ ). We observed no associations with risk of other histological subtypes of invasive OC (Supplementary Table 2). The correlations between the SNP exhibiting the strongest risk association (rs67397200) in the meta-analysis of BC risk for *BRCA1* mutation carriers and ER-negative BC, with the previously reported risk-associated SNPs



**Figure 1 | Regional association plot disease-specific risk associations.** Results for ER negative breast cancer from BCAC, for ovarian cancer from OCAC and for *BRCA1* mutation carriers with breast cancer from CIMBA are shown. Also shown are the results of a meta-analysis for *BRCA1* and general population ER negative breast cancer cases. The grey bars indicate the boundaries of the two association peaks, and the dotted horizontal line indicates the cutoff for genome-wide significance ( $\chi^2$ -test  $P = 5 \times 10^{-8}$ ). Previously identified GWAS SNPs are indicated with italic font. Genes in the region are displayed beneath the association results.

for breast, OC and *BRCA1*-associated BCs can be found in Supplementary Table 3.

All SNPs with an association *P* value < 0.001 with each phenotype were included in forward stepwise Cox regression models for risks of *BRCA1* BC, and logistic regression models for ER-negative BC and serous OC. The most parsimonious models for ER-negative BC and serous OC each included one SNP, rs67397200 for ER-negative BC and rs4808075 for serous OC (referred to as Peak 1). The most parsimonious model in the analysis of BC risk for *BRCA1* mutation carriers included two virtually uncorrelated SNPs (pairwise correlation  $r^2 = 0.018$ ) rs61494113 (*P* value =  $4.4 \times 10^{-16}$  in conditional regression analysis), and rs3786515 (Peak 2, conditional regression *P* value =  $9.6 \times 10^{-5}$ , pairwise correlation  $r^2 = 0.018$ ; Fig. 1). No other SNP was retained in the model at the *P* value threshold of 0.0001.

**Candidate causal variants.** Peak 1 includes SNPs that encompass the *BABAM1*, *ABHD8* and *ANKLE1* gene and are associated with serous OC, ER-negative BC and BC risk for *BRCA1* mutation carriers (Fig. 1 and Supplementary Fig. 1); Peak 2 includes SNPs located in the *MYO9B* gene associated only with BC risk in *BRCA1* mutation carriers. SNPs in Peaks 1 and 2 are virtually uncorrelated.

To identify the strongest candidate causal SNPs, we computed likelihood ratios of each SNP relative to the SNP with the strongest association in each peak for risks of each phenotype.

Due to the similarities in associations between ER-negative BC and *BRCA1*-associated BC in Peak 1, we computed the likelihood ratios on the basis of the meta-analysis results. Table 1 includes the SNPs that cannot be excluded at a likelihood ratio of > 1:100 fold. In Peak 1, all but 12 SNPs can be excluded from being causal for ER-negative BC and *BRCA1*-associated BC. An additional SNP (rs10424198) cannot be excluded from being causal for serous OC. All 13 SNPs were highly correlated ( $r^2 > 0.95$ ) and spanned a region of 19.4 kb. In Peak 2, the likelihood ratios of each SNP were calculated on the basis of the *BRCA1* association analysis conditional on the top SNP rs61494113. All but seven SNPs correlated with rs3786515 ( $r^2 > 0.10$ ) cannot be excluded from being the causal SNP for *BRCA1*-associated BC risk. With the exception of rs3786514 (pairwise  $r^2$  with rs3786515 = 0.87) all other SNPs had  $r^2$  with rs3786515 between 0.13 and 0.20.

**Associations for *BRCA1* and *BRCA2* mutation carriers.** SNPs in Peak 1 were only associated with risk of ER-negative BC for *BRCA1* mutation carriers and provided no evidence of association with ER-positive BC for *BRCA1*. SNPs in Peak 1 were also associated with OC risk for *BRCA1* mutation carriers. SNPs in Peak 2 were also primarily associated with *BRCA1*-related ER-negative BC but there was no evidence of association with OC risk (Supplementary Table 4). SNPs in peak 1 were not associated with overall risk of BC in *BRCA2* carriers (for example, rs67397200 HR for BC = 1.00 (95% confidence interval (CI): 0.93–0.89)); however, SNP rs67397200 showed evidence of

**Table 1 | SNPs associated with risk ovarian cancer, ER-negative breast cancer or breast cancer in *BRCA1* carriers at the 19p13 locus.**

SNP*	Nucleotide position (build 37)	Allele freq.	BRCA1 breast cancer		ER-negative breast cancer		BRCA1/ER-negative breast cancer meta-analysis ( <i>P</i> value)	Serous ovarian cancer	
			HR (95% confidence intervals)	<i>P</i> value	OR (95% confidence intervals)	<i>P</i> value		OR (95% confidence intervals)	<i>P</i> value
<b>Peak 1</b>									
rs4808075 (I)	17390291	0.30	1.19 (1.14–1.24)	$4.77 \times 10^{-15}$	1.16 (1.11–1.21)	$4.42 \times 10^{-13}$	$1.55 \times 10^{-26}$	1.19 (1.14–1.23)	$9.17 \times 10^{-20}$
rs10419397 (I)	17391328	0.30	1.19 (1.14–1.24)	$5.55 \times 10^{-15}$	1.16 (1.11–1.21)	$6.57 \times 10^{-13}$	$2.7 \times 10^{-26}$	1.19 (1.14–1.23)	$1.29 \times 10^{-19}$
rs56069439 (I)	17393925	0.30	1.19 (1.14–1.24)	$3.33 \times 10^{-15}$	1.16 (1.12–1.21)	$2.22 \times 10^{-13}$	$5.26 \times 10^{-27}$	1.19 (1.14–1.23)	$1.94 \times 10^{-19}$
rs4808076 (I)	17395401	0.30	1.19 (1.14–1.24)	$2.55 \times 10^{-15}$	1.16 (1.12–1.21)	$2.9 \times 10^{-13}$	$5.59 \times 10^{-27}$	1.18 (1.14–1.23)	$3.72 \times 10^{-19}$
rs111961716 (I)	17398085	0.30	1.19 (1.14–1.24)	$3.22 \times 10^{-15}$	1.16 (1.12–1.21)	$2.63 \times 10^{-13}$	$6.07 \times 10^{-27}$	1.18 (1.14–1.23)	$6.97 \times 10^{-19}$
rs113299211 (I)	17400765	0.30	1.19 (1.14–1.24)	$2.33 \times 10^{-15}$	1.16 (1.12–1.21)	$2.4 \times 10^{-13}$	$4.22 \times 10^{-27}$	1.18 (1.14–1.23)	$8.13 \times 10^{-19}$
rs67397200 (G)	17401404	0.30	1.19 (1.14–1.24)	$8.88 \times 10^{-16}$	1.16 (1.12–1.21)	$1.10 \times 10^{-13}$	$6.18 \times 10^{-28}$	1.18 (1.14–1.23)	$7.75 \times 10^{-19}$
rs61494113 (G)	17401859	0.30	1.19 (1.14–1.25)	$7.77 \times 10^{-16}$	1.16 (1.12–1.21)	$1.27 \times 10^{-13}$	$7.31 \times 10^{-28}$	1.18 (1.14–1.23)	$1.14 \times 10^{-18}$
rs4808616 (G)	17403033	0.31	1.19 (1.14–1.24)	$1.44 \times 10^{-15}$	1.16 (1.12–1.21)	$1.10 \times 10^{-13}$	$9.37 \times 10^{-28}$	1.18 (1.14–1.23)	$1.51 \times 10^{-18}$
rs55924783 (I)	17404072	0.30	1.19 (1.14–1.24)	$2.44 \times 10^{-15}$	1.16 (1.12–1.21)	$1.61 \times 10^{-13}$	$2.81 \times 10^{-27}$	1.18 (1.14–1.23)	$1.35 \times 10^{-18}$
rs28473003 (I)	17406167	0.30	1.19 (1.14–1.24)	$2.11 \times 10^{-15}$	1.16 (1.12–1.21)	$2.8 \times 10^{-13}$	$4.55 \times 10^{-27}$	1.18 (1.14–1.22)	$3.43 \times 10^{-18}$
rs13343778 (I)	17407695	0.30	1.19 (1.14–1.24)	$7.44 \times 10^{-15}$	1.16 (1.12–1.21)	$3.92 \times 10^{-13}$	$2.06 \times 10^{-26}$	1.18 (1.14–1.22)	$3.18 \times 10^{-18}$
rs10424198 (I)	17409671	0.30	1.18 (1.13–1.24)	$3.13 \times 10^{-14}$	1.16 (1.12–1.20)	$1.18 \times 10^{-12}$	$2.56 \times 10^{-25}$	1.18 (1.14–1.22)	$3.85 \times 10^{-18}$
<b>Peak 2</b>									
rs3786514 (G)	17294954	0.48	1.08 (1.04–1.13)	$5.85 \times 10^{-05}$	1.02 (0.98–1.06)	0.364	$6.52 \times 10^{-04}$	1.05 (1.01–1.08)	$8.01 \times 10^{-03}$
rs3786515 (G)	17295023	0.45	1.10 (1.05–1.14)	$5.42 \times 10^{-06}$	1.02 (0.98–1.06)	0.281	$9.94 \times 10^{-05}$	1.05 (1.01–1.09)	$5.62 \times 10^{-03}$
rs891205 (G)	17354586	0.61	1.09 (1.05–1.13)	$4.16 \times 10^{-05}$	1.05 (1.01–1.09)	0.0164	$5.39 \times 10^{-06}$	1.07 (1.04–1.11)	$1.26 \times 10^{-04}$
rs7247493 (G)	17362941	0.60	1.09 (1.04–1.13)	$5.85 \times 10^{-05}$	1.05 (1.01–1.09)	0.014	$5.73 \times 10^{-06}$	1.07 (1.04–1.11)	$9.68 \times 10^{-05}$
rs7246243 (I)	17363068	0.60	1.09 (1.04–1.13)	$5.28 \times 10^{-05}$	1.05 (1.01–1.09)	0.0149	$5.74 \times 10^{-06}$	1.08 (1.04–1.11)	$3.73 \times 10^{-05}$
rs4464206 (G)	17367585	0.62	1.10 (1.05–1.14)	$7.28 \times 10^{-05}$	1.06 (1.02–1.10)	0.0172	$8.87 \times 10^{-06}$	1.08 (1.04–1.12)	$2.54 \times 10^{-05}$
C19pos17261271 (G)	17400271	0.50	0.92 (0.88–0.96)	$2.41 \times 10^{-05}$	0.96 (0.92–0.99)	0.020	$4.76 \times 10^{-06}$	0.92 (0.89–0.96)	$9.26 \times 10^{-06}$
<b>Peak 2 (conditional <i>P</i> values on top SNP from Peak 1)</b>									
rs3786514	17294954			$1.40 \times 10^{-03}$					
rs3786515	17295023			$9.13 \times 10^{-05}$					
rs891205	17354586			0.0107					
rs7247493	17362941			0.0131					
rs7246243	17363068			0.0122					
rs4464206	17367585			115					
c19_pos17261271	17400271			$6.31 \times 10^{-03}$					

EOC, epithelial ovarian cancer; ER, oestrogen receptor; freq., frequency; HR, hazards ratio; OR, odds ratio; SNP, single-nucleotide polymorphism. SNPs in Peak 1 and Peak 2 that cannot be excluded at a likelihood ratio of > 1:100 fold relative to the most significant SNP for the meta-analysis and serous EOC (Peak 1) and *BRCA1* association breast cancer for Peak 2. \*Imputed (I) or genotyped (G) SNPs.

association with OC for *BRCA2* mutation carriers (hazards ratio (HR) = 1.18, 95% CI: 1.06–1.36,  $\chi^2$ -test  $P = 0.0056$ ). SNPs in peak 2 did not show any evidence of association with breast or OC risk for *BRCA2* mutation carriers.

**Associations with risk among BC subtypes.** None of the Peak 1 SNPs were associated with risk of ER-positive BC. When analyses were restricted to triple negative BC, the odds ratio (OR) estimates for SNPs in Peak 1 were larger than the corresponding OR estimates for ER-negative disease (Supplementary Table 4). There was no evidence of association with ER-negative and HER2-positive BC risk, with the association restricted only to triple-negative BC (test of difference between triple-negative versus ER-negative/HER2+,  $P$ -diff =  $2.2 \times 10^{-5}$  for SNP rs61494113).

**Analysis in Asian and African ancestry studies.** None of the SNPs in the fine-mapping region were associated with ER-negative BC in samples of Asian ancestry after adjusting for multiple testing ( $P$  values  $\geq 0.0018$ ). However, the risk alleles of the 13 candidate causal SNPs in Peak 1 are uncommon in the Asian population (MAF = 0.0079–0.011); hence, the power to detect an association was limited and, due to the wide CIs for the estimated ORs for these SNPs, we cannot rule out that the minor allele of these SNPs in Asian subjects is associated with similar level of risk as in Europeans. In samples of African ancestry only rs4808616 (MAF = 0.22) showed evidence of association with risk for overall BC or ER-negative disease (OR for BC = 1.19, 95% CI: 1.02–1.39,  $\chi^2$ -test  $P = 0.03$ ; OR for ER-negative BC = 1.59, 95% CI: 1.02–2.49,  $\chi^2$ -test  $P = 0.04$ ).

**Functional characterization of the 19p13.1 region.** Functional characterization focused on the 13 candidate causal SNPs for ER-negative and *BRCA1*-associated BC and serous OC in Peak 1, based on the hypothesis that the functional mechanisms mediated by one or more of these SNPs were the same for these phenotypes.

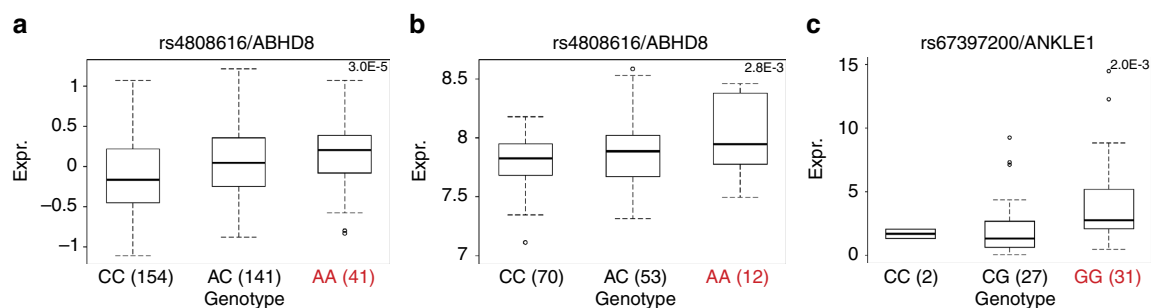
**Genotype-gene expression associations.** We used expression quantitative trait locus (eQTL) analyses to evaluate associations between risk SNPs and the expression of genes in a 1 Mb region spanning rs4808075 in: 135 normal breast tissues<sup>29</sup>, 60 normal ovarian and fallopian tube epithelial cell cultures, 391 ER positive BCs<sup>30</sup>, 59 ER-negative BCs<sup>29</sup> and 340 high-grade serous OCs<sup>30</sup>. We identified significant eQTL associations for *ABHD8* expression (linear regression  $P$  value range  $2 \times 10^{-3}$ – $7 \times 10^{-3}$ ) in normal breast tissues and between rs480816 and *ABHD8*

expression in OCs (linear regression  $P = 3 \times 10^{-5}$ ). In both instances the risk allele was associated with higher *ABHD8* expression (Fig. 2a, Supplementary Data 1 and 2 and Supplementary Table 5). We examined whether risk SNPs were the top eQTL SNPs in this region. rs4808616 was the strongest predictor of *ABHD8* expression in OCs. However, in normal breast tissues the top eQTL SNP for *ABHD8* was rs11666308 (linear regression  $P = 3.3 \times 10^{-4}$ ), a marginally better predictor than rs4808616 (linear regression  $P = 2.8 \times 10^{-3}$ ). The two SNPs were correlated ( $r^2 = 0.79$ ) and regressing out effects of either SNP from the expression levels of *ABHD8* and repeating eQTL analysis abolished the eQTL signal for the other SNP, confirming their statistical inseparability. In addition we found significant associations between rs4808616 and *NXNLI* expression in OCs (linear regression  $P = 4 \times 10^{-3}$ ) and with *ANKLE1* expression ( $P = 0.002$ ) in normal ovarian surface epithelial cells (OSECs). There were no eQTL associations for any other genes in the region.

We also performed allele-specific expression analysis in BC using RNA sequencing data<sup>31</sup> for coding SNPs in *ABHD8* (rs56069439) and *BABAMI* (rs10424198). Both SNPs were correlated with rs4808616 ( $r^2 = 0.91$ ). There was a significant association between rs56069439 and the allelic ratio of *ABHD8* transcripts (F-test  $P = 0.016$ ) with greater expression associated the risk allele (Supplementary Fig. 2; Supplementary Data 3).

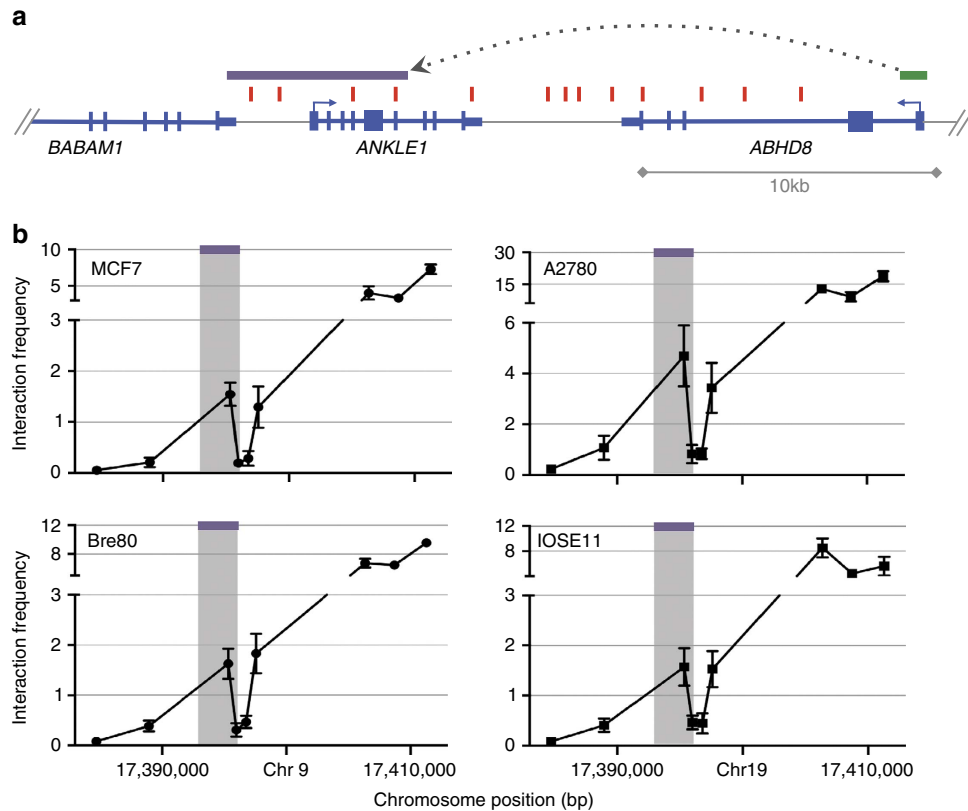
**Chromosome conformation capture.** Chromosome conformation capture (3C) analysis was used to investigate DNA–DNA interactions between *ABHD8* and 5 of 13 candidate causal SNPs in Peak 1. Eight SNPs close to the *ABHD8* promoter were too near to be resolved, and the close proximity of candidate causal SNPs to *ANKLE1* precluded 3C analysis for this gene. The *ABHD8* promoter showed an interaction with a 6.3 kb region  $\sim 20$  kb telomeric to the gene in both normal breast (Bre80) and ovarian (IOSE11) epithelial cells, and in breast (MCF7) and ovarian (A2780) cancer cell lines (Fig. 3). This region spans the *ANKLE1* promoter and includes four candidate causal SNPs: rs4808075, rs10419397, rs56069439 and rs4808076. There was no evidence of interaction for any candidate causal SNP with *BABAMI* (Supplementary Fig. 3).

**Annotation of candidate causal SNPs.** All 13 candidate causal SNPs were located in non-protein coding DNA. We annotated putative functional regulatory elements that coincided with the candidate causal SNPs in normal human mammary epithelial cells (HMECs), and normal fallopian tube and ovarian epithelial cells<sup>19</sup>, and in OC cell lines. Five of the 13 SNPs coincide with



**Figure 2 | Expression quantitative trait locus analyses.** Significant eQTL associations identified between rs4808616 and *ABHD8* expression in (a) ovarian cancer tissues and (b) in normal breast tissues. (c) A significant association was also identified between rs4808616 and *ANKLE1* expression in primary normal ovarian/fallopian tube epithelial cell cultures. The horizontal line indicates the median expression, the limits of the boxes denote the first and third quartiles, and the whiskers represent 1.5 times the interquartile range of the data. Outliers are indicated with circles.





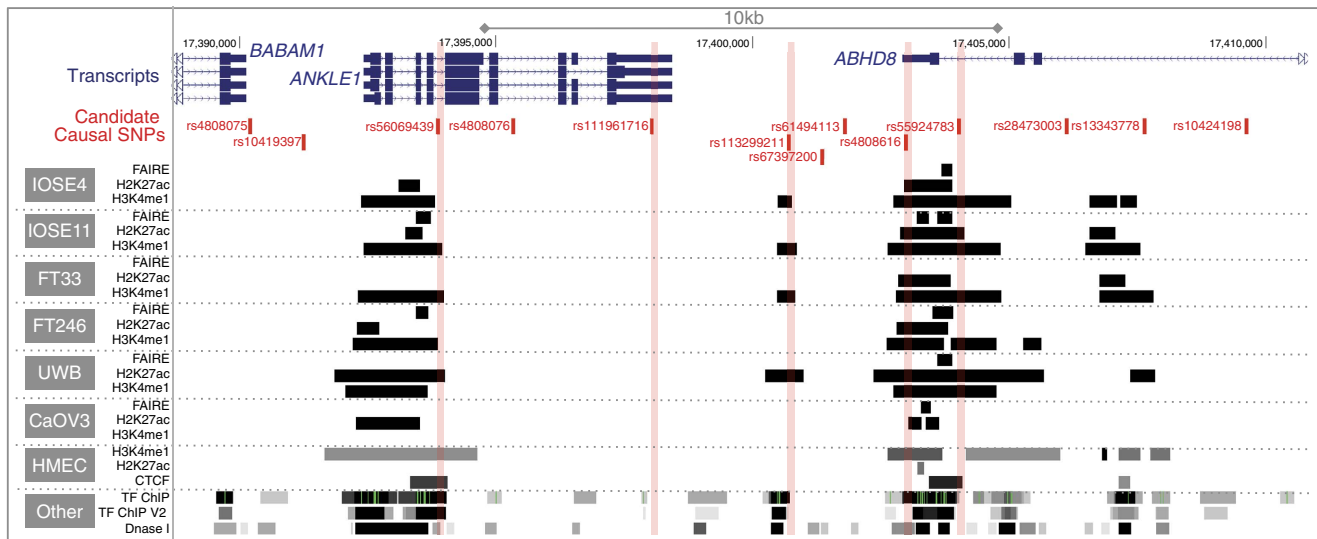
**Figure 3 | Chromosome conformation capture analysis of long-range interactions at the 19p13 region.** 3C interaction profiles in breast and ovarian cell lines. 3C libraries were generated with *Nco*I, with the anchor point set at the *ABHD8* promoter region. **(a)** A physical map of the region interrogated by 3C is shown, with annotated genes shown in blue, the 13 risk-associated SNPs shown in red, the *ABHD8* promoter fragment shown in green and the position of the interacting *Nco*I fragment represented by the purple bar (not to scale). **(b)** Relative interaction frequencies between the *ABHD8* promoter and regions spanning risk associated SNPs in normal breast (Bre80) and ovarian (IOSE11) epithelial cell lines, and in breast (MCF7) and ovarian (A2780) cancer cell lines. A peak of interaction with the *ABHD8* was observed for one region (purple bar) in all four cell lines. There were no interactions detected between the purple region and the *BABAM1* or *USHBP1* promoters. The interacting region contains four candidate causal SNPs (from left to right) rs4808075, rs10419397, rs56069439 and rs4808076. Error bars represent s.d. ( $N=3$ ).

regulatory elements that were reproducible in two biological replicate samples (Fig. 4). Three SNPs were located in epigenetic marks in breast and/or ovarian cells: rs55924783 coincided with insulator marks in HMECs and enhancer marks in ovarian cells; rs113299211 coincided with enhancer marks in ovarian cells and is predicted to alter transcription factor binding sites for ELF1, ELK4 and GABP; and rs56069439 coincided with experimentally derived ChIP-seq footprints (for CTCF, ATF2 and ZNF263), enhancer marks in ovarian cells and both enhancer (H3K4me1) and insulator (CTCF) marks in breast cells. Two SNPs were located in 3'-untranslated regions (UTRs) of protein coding genes: rs111961716 in *ANKLE1* and rs4808616 in *ABHD8*. rs4808616 also coincided with enhancer marks in ovarian and breast cells. Finally, rs10419397 lay within the putative promoter of *ANKLE1*, ~1,200 bp from the transcription start site.

**Functional analysis of candidate causal SNPs in UTRs.** We evaluated the effects on mRNA stability of the SNPs located in 3' UTRs of *ANKLE1* (rs111961716) and *ABHD8* (rs4808616, Figs 4 and 5a) in normal primary ovarian epithelial cell lines carrying different SNP genotypes. RNA transcript abundance was measured after blocking mRNA transcription by treating cells with actinomycin D. For rs111961716, *ANKLE1* transcript expression was significantly more stable in cell lines homozygous for the A (risk) allele of rs111961716 compared with

heterozygous cells or cells homozygous for the C allele ( $P=0.006$ , analysis of variance; Fig. 5b). There was no association between *ABHD8* mRNA stability and genotypes of rs4808616 (Fig. 5b).

**Functional analysis of promoter and enhancer SNPs.** Seven of the 13 candidate causal SNPs in Peak 1 resided either in the *ANKLE1* promoter or in putative regulatory elements (PREs-A-C) in breast and ovarian normal and cancer cell lines (Figs 4 and 5a). SNP rs10419397 fell within the *ANKLE1* promoter region, but had no effect on promoter activity (Fig. 5c). PRE-A contained SNP rs56069439, PRE-B contained SNPs rs113299211, rs67397200, rs61494113 and PRE-C contained SNPs rs4808616 and rs55924783. We examined the effect of these PREs, and of the risk alleles of each SNP cloned into luciferase constructs containing the *ABHD8* or *ANKLE1* promoters. Inclusion of the reference allele of PREs A, B and C significantly increased *ABHD8* promoter activity in both OC (A2780) and normal breast (Bre80) cell lines (Fig. 5). Constructs containing the risk alleles further enhanced *ABHD8* promoter activity compared with the reference allele for PREs A, B and C in Bre80 cells ( $P$  values = 0.0027, 0.0308 and 0.0342, respectively, two-way analysis of variance (ANOVA)) and for PREs A, B and C in A2780 cells ( $P$  values = 0.0193, 0.0115 and <0.0001, respectively, two-way ANOVA; Fig. 5d,e). Constructs containing the reference allele of PRE-A showed a silencing effect on the *ANKLE1* promoter in both cell types with the risk allele further silencing the activity of the



**Figure 4 | Epigenetic marks intersecting candidate causal SNPs in the 19p13 susceptibility region and analyses of UTR SNPs.** The thirteen candidate SNPs were aligned with open chromatin and enhancer marks (H3K27ac and H3K4me1) in high-grade serous ovarian cancer cells (UWB1.289 and CaOV3) and ovarian cancer precursor cells (ovarian epithelial cells, IOSE and fallopian epithelial cells, FT). Enhancer and insulator (CTCF) data for human mammary epithelial cells (HMECs) were obtained from ENCODE. Five SNPs coincide with biofeatures in breast and/or ovarian cells (indicated in red).

reference allele in A2780 cells ( $P=0.0049$ , two-way ANOVA). The reference allele of PRE-B had no effect on *ANKLE1* promoter activity, while the risk allele significantly increased activity compared with the reference allele in A2780 cells ( $P=0.0034$ , two-way ANOVA). Constructs containing the reference allele of PRE-C significantly increased *ANKLE1* promoter activity in both ovarian ( $P=0.0004$ , two-way ANOVA) and breast cell lines ( $P=0.0067$ , two-way ANOVA). However the risk allele showed a silencing effect on the reference allele in only Br80 cells ( $P=0.0289$ , two-way ANOVA; Fig. 5d,e).

**Functional effects of rs56069439 deletion.** Collectively, the data above suggested that rs56069439 may regulate the expression of *ANKLE1* and/or *ABHD8*. We used Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR)/Cas9-mediated genome editing to delete a 57 bp region containing the regulatory region that includes rs56069439 in breast (MCF10A) and ovarian (IOSE19) epithelial cells (Fig. 6a). Analysis of multiple clones containing confirmed homozygous deletions (Fig. 6b,c) indicated a significant reduction in *ANKLE1* expression compared with parental cells ( $P=0.025$ , two-tailed paired *T*-test) and a trend towards reduced *ANKLE1* expression in IOSE19 cells ( $P=0.29$ , two-tailed paired *T*-test; Fig. 6d). Expression of *ABHD8* and *BABAM1* was unchanged following deletion of the region containing rs56069439.

**In vitro functional analysis of candidate genes.** We analysed the effects of perturbing *ABHD8*, *ANKLE1* and *BABAM1* expression in *in vitro* models of ‘normal’ breast (MCF10A) and ovarian (IOSE19 (ref. 32)) epithelial cells. For each gene, we overexpressed full length, green fluorescent protein-tagged constructs, because genes at 19p13 were frequently overexpressed in ovarian and BCs<sup>9</sup> and because eQTL analyses indicated that risk alleles were associated with increased expression of *ABHD8* and *ANKLE1*. After confirming gene overexpression (Supplementary Fig. 3a) we evaluated cell growth, migration and invasion, and anchorage-independent growth (Fig. 7 and Supplementary Fig. 3b). Overexpression of *ABHD8* caused a significant reduction in cell migration ( $P=0.007$  in

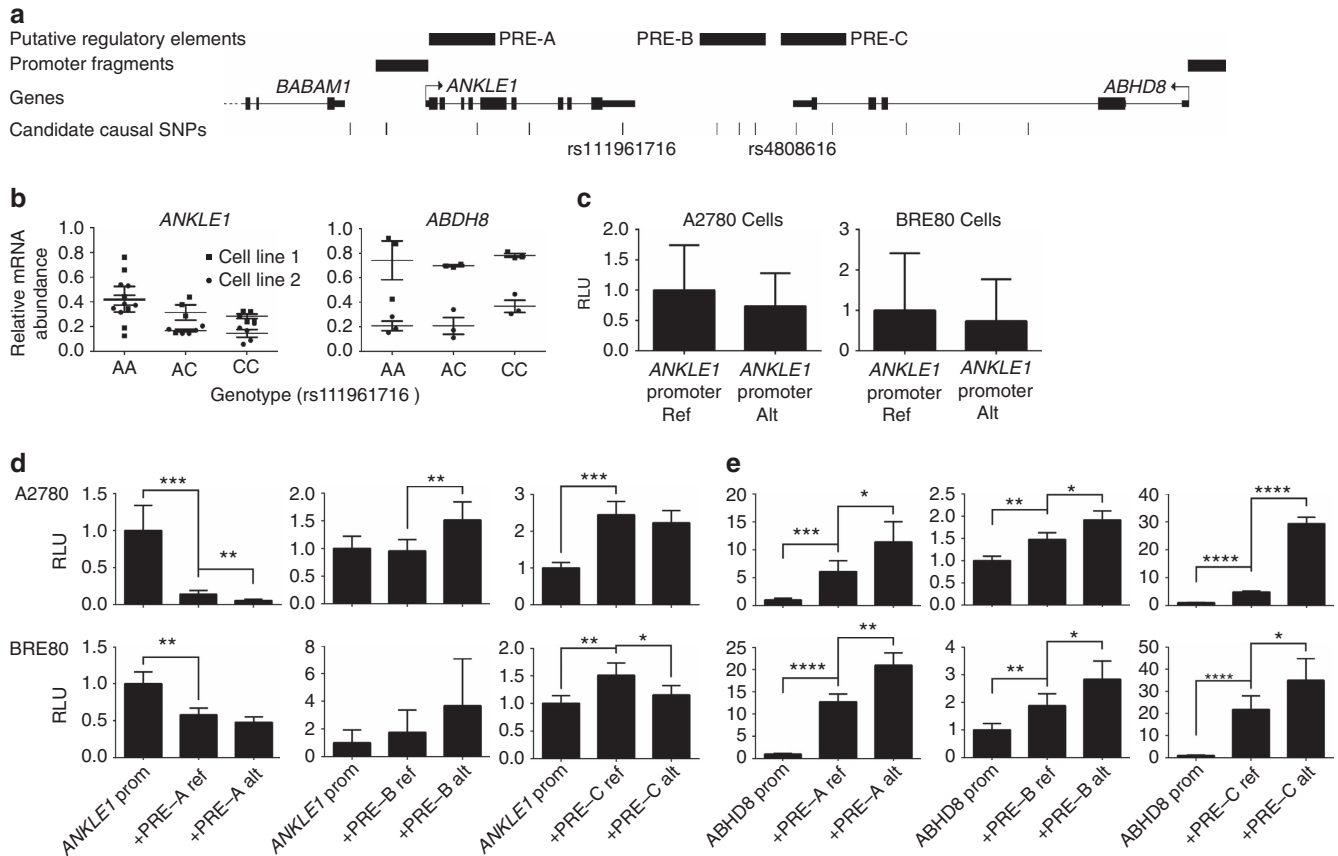
MCF10A;  $P=0.047$  in IOSE19, two-tailed paired *T*-test) and a decrease in invasion ( $P=0.018$  in MCF10A;  $P=0.063$  in IOSE19, two-tailed paired *T*-test; Fig. 7). *BABAM1* and *ANKLE1* overexpression had no effect on these cellular phenotypes for either cell type.

RNA sequencing was used to profile transcriptomic changes caused by overexpression of *ABHD8*, *ANKLE1* and *BABAM1* and pathway analyses performed using Ingenuity Pathway Analysis. We found no indication of significant changes in relevant pathways after overexpressing *BABAM1* in breast or ovarian epithelial cells. Cells overexpressing *ANKLE1* showed a significant enrichment for cancer-associated and cell growth/proliferation pathways in both breast ( $P=3.36 \times 10^{-6}$ ) and ovarian ( $P=2.43 \times 10^{-27}$ ) epithelial cells. Cells overexpressing *ABHD8* were enriched for expression changes in cancer related pathways ( $P<5.52 \times 10^{-8}$ ) and fibrosis pathways ( $P<1.23 \times 10^{-2}$ , all right-tailed Fisher’s exact tests; Supplementary Tables 6-8).

## Discussion

Through fine-scale mapping of the 19p13.1 region we have found evidence of two independent regions of genetic association with BC and/or OC risk among women of European ancestry. The minor alleles of all candidate causal variants in Peak 1 conferred increased risks of ER-negative BC and serous OC and increased risks of both cancers for *BRCA1* mutation carriers. We were able to rule out associations with ER-positive BC and risks for other OC histotypes. There was weaker evidence that SNPs in Peak 2 were independently associated with BC risk among *BRCA1* mutation carriers only. When analyses in BCAC were restricted to triple-negative BC, the strength of association was greater and there was no evidence of association with ER-negative/HER2-positive BC. Thus, our results suggested that these variants are primarily associated with triple-negative BC, the predominant tumour subtype in *BRCA1* mutation carriers<sup>33</sup>. These results are in line with previous findings for the initial SNPs identified through GWAS<sup>26</sup>.

The increased sample size resulting from combining data from BCAC, OCAC and CIMBA for variants in Peak 1 have enabled us to restrict the likely functional variants at 19p13.1 to 13 SNPs.

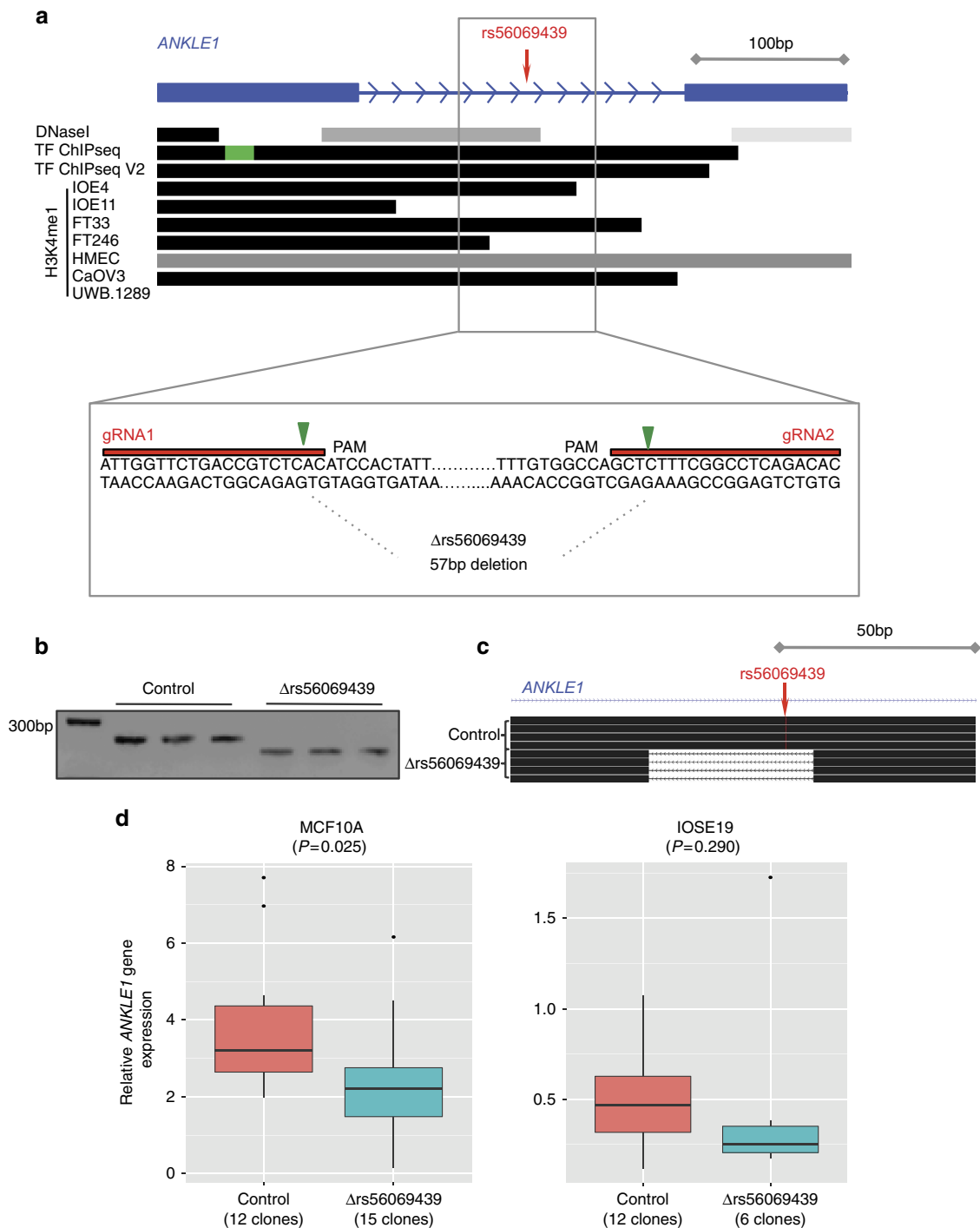


**Figure 5 | Allele specific analysis of susceptibility SNPs.** (a) Location of SNPs in putative regulatory elements (PREs) and 5' untranslated regions. (b) RNA stability assays in primary ovarian epithelial cell lines for risk-associated UTR SNPs in *ABHD8* and *ANKLE1*. Normal ovarian epithelial cell lines carrying different genotypes of the risk SNP rs4808616, located in the 3' UTR of *ABHD8*. Rs4808616 is tightly correlated with rs111961716 ( $R^2 = 0.98$ ) located in the 3' UTR of *ANKLE1*. The risk allele of rs111961716 was associated with decreased mRNA stability of *ANKLE1* compared with the protective allele ( $P = 0.006$ , ANOVA). Different genotypes of rs4808616 are not associated with the stability of the *ABHD8* transcript. (c–e) Luciferase assays to evaluate SNP-dependent promoter and enhancer activity. (c) The *ANKLE1* promoter SNP did not affect *ANKLE1* expression in ovarian cancer cells (A2780) and normal breast cells (Bre80). (d) Allele-specific activity of PRE-A, PRE-B and PRE-C on the *ANKLE1* promoter. (e) Allele-specific activity of PRE-A, PRE-B and PRE-C on *ABHD8* promoter activity. \* $P > 0.05$ , \*\* $P > 0.01$ , \*\*\* $P > 0.001$ , \*\*\*\* $P > 0.0001$ , two-way ANOVA. RLU, relative light units.

The 13 candidate causal risk SNPs in this region were the same for both BC and OC leading us to hypothesize that the underlying functional mechanisms are the same in both cancers and the overlap between these SNPs and functional elements provided multiple testable hypotheses, necessitating a range of different functional assays to evaluate their possible causality. Multiple assays were performed in breast and ovarian tissues and cell lines to establish if there is true evidence of pleiotropy. The candidate causal SNPs in Peak 1 clustered around two candidate genes, *ANKLE1* and *ABHD8*, neither of which have been previously implicated in BC or OC. Proximal to these SNPs is *BABAM1*, a gene involved in recruiting *BRCA1* to sites of DNA damage<sup>34,35</sup> and therefore a compelling candidate gene at this locus. While gene regulation can be mediated across long genomic distances, the majority of interactions occur over a distance of 1 Mb) or less<sup>36,37</sup>. We, therefore, evaluated all candidate genes within a 1 Mb region centred on the Peak 1 risk SNPs for eQTL associations. We found significant eQTL associations for *ABHD8* in OCs and normal breast tissues, plus allele-specific expression of *ABHD8* in BCs, but no compelling evidence for any other gene at this locus. Nonetheless, the identification of *ABHD8* as the most likely target susceptibility gene must be treated with some caution as it is plausible that more distant *cis*-eQTL or even *trans*-eQTL associations exist for these risk SNPs. Unfortunately, the limited power of eQTL analysis based on the current sample

size precluded us from performing genome-wide eQTL analysis to address these hypotheses.

The weight of our functional data, in particular the eQTL associations, indicates that *ABHD8* is a target of functional SNPs at this locus, and therefore a novel breast and OC susceptibility gene. 3C identified an interaction between a region containing four candidate causal SNPs and the *ABHD8* promoter in both breast and OC and normal epithelial cell lines. The luciferase assays of three PREs (including one encompassing rs56069439 in the interacting region) consistently showed that they acted as enhancers, and furthermore the risk-associated alleles of rs56069439, rs113299211, rs67397200, rs61494113, rs4808616 and rs55924783 (within PREs A-C) further increase *ABHD8* promoter activity in both breast and ovarian cells. These results were consistent with our eQTL studies and support the hypothesis that increased *ABHD8* expression is associated with an increased cancer risk. *ABHD8* is a poorly studied lipase<sup>38</sup>. The Achilles heel project identified *ABHD8* as a lineage-specific cancer cell vulnerability in OC cell lines<sup>39</sup> and a recent study identified *ABHD8* as a potential OC susceptibility gene though its participation in a homeobox transcription factor-centred gene network associated with serous OC risk<sup>40</sup>. Overexpression of *ABHD8* led to significant reductions in the invasive and migratory potential of breast and ovarian cells and enriched for genes involved in cellular movement (IOSE19) and mTOR



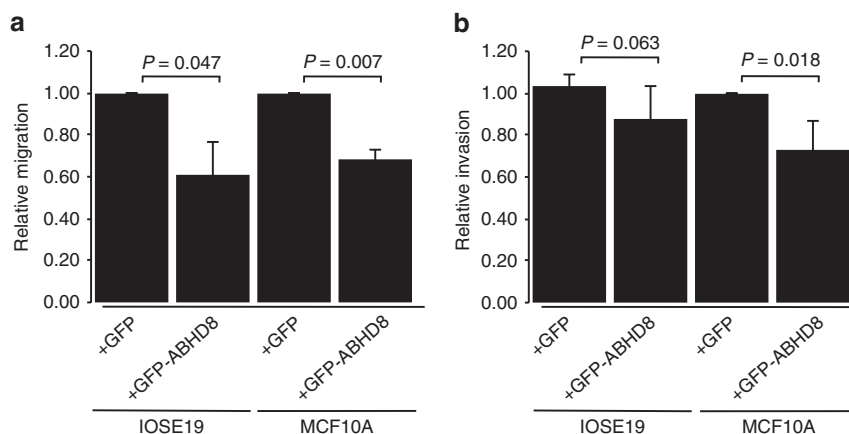
**Figure 6 | Effects of deletion of the putative enhancer containing the rs56069439 risk SNP in breast and ovarian epithelial cells. (a)** Illustration of the 57 bp region in an intron of *ANKLE1* containing rs56069439; H3K4me1 marks overlapped rs56069439 in ovarian, fallopian and breast cells. Location of the two guide RNAs (gRNAs) used to create the stable  $\Delta$ rs56069439 deletion by CRISPR/Cas9 genome editing, cutting sites are indicated with the green arrow. PAM, protospacer adjacent motif. **(b)** PCR analysis of targeted region in representative MCF10A (breast) epithelial cell clones. Control clones were transfected with the vector backbone only. **(c)** Verification of deletions by Sanger sequencing, and alignment to the genome using BLAT. **(d)** Gene expression analysis using TaqMan probes showing downregulation of *ANKLE1* was associated with deletion of a region containing rs56069439.

signalling (MCF10A), consistent with the observed changes in invasion and migration. The direction of the effect was opposite to what we might expect from the eQTL data, which might reflect different functions of *ABHD8* in different contexts, similar to the observations for another BC susceptibility gene, *TOX3* (ref. 41). For example, under specific microenvironmental cues or in a tumour cell (rather than the normal cells used in these

experiments) increased *ABHD8* may promote rather than inhibit migration and invasion.

Nonetheless, we cannot unequivocally exclude other genes as the targets of candidate causal variants at this locus, in particular *ANKLE1*. The close proximity of the candidate causal SNPs to the *ANKLE1* gene precluded 3C analysis; but in the luciferase assays, these same PREs and SNPs had variable, context-dependent





**Figure 7 | Phenotypic effects of overexpressing full length *ABHD8GFP* fusion transcript in normal breast and ovarian epithelial cells. (a)** *ABHD8* overexpression induced a significant decrease in migration in both breast (MCF10A) and ovarian (IOSE19) cells; **(b)** *ABHD8* overexpression induced a significant decrease in invasion in breast epithelial cells and a similar trend of decreased invasion in ovarian epithelial cells.

effects on *ANKLE1* promoter activity. This raises the possibility that the SNPs were cooperatively acting to alter *ANKLE1* expression although it was difficult to predict the overall direction of their effects from this assay. We were able to rule out the SNP rs10419397 in the promoter of *ANKLE1* as a likely causal variant. The SNP rs111961716 in the 3'-UTR of *ANKLE1* was associated with allele-specific *ANKLE1* mRNA stability; but stable overexpression of *ANKLE1* had no influence on the phenotype of normal breast and ovarian epithelial cells even though pathway after overexpression of *ANKLE1* found a significant enrichment for cancer and cell death/proliferation associated pathways in both breast and ovarian epithelial cells. More recently, *ANKLE1* has been implicated in DNA damage responses, while other, better-characterized endonucleases (for example, ERCC1) are involved in nucleotide excision repair, which are important for the repair of bulky adducts<sup>42</sup>.

This study has highlighted the challenges in establishing causality for both candidate causal SNPs at common variant susceptibility loci and the susceptibility genes targets. The multitude of functional assays that can be used to test allele specific functional activity rarely provide unequivocal evidence of one SNP over another. Genome editing, which allows the creation of isogenic experimental models carrying the different alleles of candidate causal SNP, is emerging as a single assay approach that can evaluate the function of common variants. However, until now the technical challenges of genome editing have restricted its application to two non-coding risk SNPs identified by GWAS at susceptibility loci for prostate cancer and obesity, respectively<sup>22,23</sup>. It was beyond the scope of the current study to utilize genome editing to test all 13 candidate causal SNPs in Peak 1 at 19p13 in BC and OC and normal cell line models. Instead, we used CRISPR-Cas9 genome editing to evaluate the effects of a putative enhancer containing most plausible functional SNP (rs56069439) identified from 3C analysis and mapping of putative regulatory elements. This revealed strong functional evidence for a breast/ovarian epithelial cell enhancer, within an intron of *ANKLE1*. When this enhancer containing rs56069439 was deleted *ANKLE1* expression was significantly reduced, without any reduction in *BABAM1* or *ABHD8* expression. Further experiments using homology-directed repair will be required to determine if there is allele-specific activity of the rs56069439 SNP in regulating *ANKLE1* expression, and to determine whether shadow enhancers are employed to maintain *ABHD8* expression<sup>43</sup>.

In conclusion, we have performed detailed functional analysis of SNPs and candidate target genes at the 19p13 locus in breast

and ovarian normal and cancer cells. *ABHD8* is the most likely target gene although we cannot rule out a role for *ANKLE1* in the development of breast and OC or the possibility that both genes, acting independently or in synergy may be functional targets of candidate causal SNPs. Using a combination of genetic fine mapping, and a spectrum of *in silico* and functional assays, seven of thirteen showed evidence of functionality.

These data suggest that the underlying functional mechanism(s) at the 19p13 locus may be mediated by many SNPs rather than by a single causal allele. This hypothesis is supported by studies showing tissue-specific enrichment of correlated risk-associated SNPs at susceptibility loci within regulatory biofeatures, including enhancers and transcription factor binding sites<sup>19,20</sup>. Such enrichments would not be detected if a single causal SNP at a locus was driving disease development. Taken together these data suggest that common molecular mechanisms are likely to underlie this pleiotropic risk locus.

## Methods

**Study populations.** All specimens used in this study were collected with informed consent and under the approval of local Institutional Review Boards. We used epidemiological and genotype data from studies participating in the BCAC<sup>44</sup>, the OCAC<sup>12</sup> and the CIMBA<sup>45</sup> that have been genotyped using the iCOGS array that included ~200,000 SNPs.

**BC association consortium.** Data were available from 52 BC case-control studies, 41 studies of European ancestry, 9 studies of Asian ancestry and 2 studies of African-American ancestry. Details of all studies, the genotyping process and the quality control process have been described elsewhere<sup>6,44</sup>, standard sample and genotyping QC criteria were applied. After the quality control process, data on 46,451 cases and 42,599 controls of European ancestry, 6,269 cases and 6,624 controls of Asian ancestry and 1,117 cases and 932 controls of African-American ancestry were available for analysis. Data on the BC ER status were available for 34,509 cases of European ancestry, 7,435 (22%) of whom had ER-negative tumours.

**OC association consortium.** Data were available from 41 case-control studies of EOC from OCAC that were genotyped using the iCOGS array<sup>12</sup>. In addition to the OCAC iCOGS data, genotype data were available for stage 1 of three population-based OC genome-wide association studies. The final data set comprised genotype data for 11,069 cases and 21,722 controls from COGS ('OCAC-iCOGS'), 2,165 cases and 2,564 controls from a GWAS from North America ('US GWAS')<sup>46</sup>, 1,762 cases and 6,118 controls from a UK-based GWAS ('UK GWAS')<sup>7</sup>, and 441 cases and 441 controls from the Mayo Clinic. All subjects included in this analysis provided written informed consent as well as data and blood samples under ethically approved protocols. Overall, 43 studies from 11 countries provided data on 15,437 women diagnosed with invasive EOC, 9,627 of whom were diagnosed with serous EOC and 30,845 controls from the general population.

**Consortium of investigators of modifiers of BRCA1/2.** Data on *BRCA1* mutation carriers were obtained through CIMBA. Eligibility in CIMBA is restricted to females 18 years or older with pathogenic mutations in *BRCA1* or *BRCA2*. The majority of the participants were sampled through cancer genetics clinics<sup>47</sup>, including some related participants. Fifty-one studies from 25 countries contributed data on *BRCA1* mutation carriers who were genotyped using the iCOGS array<sup>45</sup>. After quality control of the phenotypes and genotypes, data were available on 15,252 *BRCA1* mutation carriers of whom 7,455 had been diagnosed with BC, 2,639 with ER-negative BC and 1,724 with OC, all of European ancestry. Analyses in *BRCA1* mutation carriers focused on assessing associations with BC risk, following the evidence from the original GWAS in *BRCA1* mutation carriers<sup>48</sup>.

URLs: 1000 Genomes Project, <http://www.1000genomes.org/>; BCAC, <http://ccge.medschl.cam.ac.uk/consortia/bcac/index.html>; CIMBA, <http://ccge.medschl.cam.ac.uk/consortia/cimba/index.html>; COGS, <http://www.cogseu.org/>; iCOGS, <http://ccge.medschl.cam.ac.uk/research/consortia/icogs/>; SNAP <https://www.broadinstitute.org/mpg/snap/>; TCGA, <https://tcga-data.nci.nih.gov/>; CGHub, <https://cghub.ucsc.edu/>

**iCOGS SNP selection for fine mapping and imputation.** The fine mapping region was defined as Chromosome 19 positions: 17,130,000–17,550,000 (NCBI build 37). To identify the set of variants potentially responsible for the original GWAS reports, we considered all variants with minor allele frequencies of  $>0.02$  from the 1,000 Genomes Project (March 2010 version) and selected all SNPs correlated ( $r^2 > 0.1$ ) with either of the two SNPs that had been identified through the *BRCA1* and EOC GWAS studies (rs8170 and rs2363956)<sup>12,45</sup>, plus an additional set of SNPs that tagged all remaining SNPs in the region with  $r^2 > 0.9$ . A total of 438 SNPs that were included on iCOGS in the 19p13 region passed QC and were available for the analyses. Data on these SNPs were used to impute the genotypes of all known variants from the 1,000 genomes project (V3, April 2012 release49) using the IMPUTE (version 2) software. After excluding SNPs with MAF  $< 0.001$  and SNPs with imputation  $r^2$  accuracy score of  $\leq 0.3$ , there were 2,269 imputed SNPs in BCAC, 2,565 in OCAC and 2,311 in *BRCA1* mutation.

**BCAC and OCAC association analysis and logistic regression.** To evaluate the association of each SNP with breast and EOC risk in BCAC and OCAC we used a Wald test statistic based on logistic regression, by estimating the per-allele OR and its s.e. Analyses restricted to specific tumour subtypes (ER-negative BC or high-grade serous EOC) were assessed separately using all available controls. All analyses were adjusted for principal components, described in more detail elsewhere<sup>12,44</sup>. Conditional logistic regression was used to assess the evidence that there are multiple independent association signals in the region, by evaluating the associations of genetic variants in the region while adjusting for the SNP with the smallest *P* value. We considered only SNPs with *P* values of association of  $< 10^{-3}$  and MAF  $> 0.1\%$  and the most parsimonious model was identified using step-wise forward logistic regression and a threshold of  $P < 10^{-4}$  for retaining SNPs in the model.

**CIMBA retrospective cohort analysis.** All associations between genotypes and BC risk in *BRCA1* mutation carriers were evaluated using a 1 *df* per allele trend-test (*P*-trend), based on modelling the retrospective likelihood of the observed genotypes conditional on BC phenotypes<sup>49</sup>. To allow for the non-independence among related individuals, an adjusted test statistic was used which took into account the correlation in genotypes<sup>48</sup>. Per allele HR estimates were obtained by maximizing the retrospective likelihood. All analyses were stratified by country of residence. To identify the most parsimonious model that includes multiple SNPs, forward-selection Cox-regression analysis was performed, using the same *P* value thresholds as in the BCAC and OCAC analysis. This approach provides valid tests of association, although the parameter estimates can be biased<sup>49,50</sup>. Parameter estimates for the most parsimonious model were obtained using the retrospective likelihood approach.

**Meta-analysis.** It is well established that the majority of BCs in *BRCA1* mutation carriers are ER-negative<sup>51,52</sup>. To increase the statistical power for identifying the most likely causal variants, we also performed a meta-analysis of the associations of BC risk for *BRCA1* mutation carriers and ER-negative BC in the general population (in BCAC) for both genotyped and imputed SNPs. We used an inverse variance approach assuming mixed effects, by combining the logarithm of the per-allele HR for the association with BC risk for *BRCA1* mutation carriers and the logarithm of the OR estimate for the association with ER-negative BC in BCAC.

**eQTL and allele-specific expression analyses.** Germline genotype data were obtained from the Affymetrix SNP 6.0 (METABRIC) and Illumina 1M-Duo (TCGA HGSOC). No SNPs from Peak 1 and 2 were present on the Affymetrix platform so these genotypes were imputed into the 1000 Genomes European reference panel (March 2012, version 3) using IMPUTE version 2 (ref. 53). All analyses were restricted to patients of  $>90\%$  European ancestry as per LAMP estimates<sup>54</sup> and SNPs with info score  $> 0.3$ . For METABRIC, gene expression data consisted of probe-level measurements from the Illumina HT-12 v3 microarray

platform for a total of 135 samples obtained from normal breast tissue adjacent to tumour and 59 samples obtained from ER-negative breast tumours were analysed. For TCGA HGSOC, gene expression data consisted of measurements from the Agilent 244 K microarray for 340 HGSOC tumours downloaded from the cBioportal. Only genes and probes  $< 1$  Mb from the top Peak 1 SNP were analysed. Tumour gene expression data was first adjusted for copy number (TCGA and METBRIC, Affymetrix SNP 6.0 calls) and methylation (TCGA only, Illumina 27 K beta values) using the method of Li *et al.*<sup>51</sup>. Expression QTL analysis was conducted by linear regression with genotypes as predictors, as implemented in the R package Matrix eQTL<sup>55</sup>.

Sixty early passage primary normal OSECs and fallopian tube epithelial cells were collected and cultured as previously described<sup>27,56</sup>. Briefly, OSECs were harvested from ovaries using a sterile cytobrush and cultured in Medium 199 and MCDB105, mixed in a 1:1 ratio and supplemented with 15% fetal bovine serum (FBS, Hyclone), 10 ng ml<sup>-1</sup> epidermal growth factor, 0.5 mg ml<sup>-1</sup> hydrocortisone, 5 mg ml<sup>-1</sup> insulin (all Sigma, St Louis, MO, USA) and 34 mg protein per ml bovine pituitary extract (Life Technologies). Fresh fallopian specimens were subjected to 48–72 h Pronase (Roche) and DNase I digests to release the epithelial cells. Epithelial cells were pelleted and cultured on collagen in DMEM/F12 supplemented with 10% FBS (Seradigm). RNA was isolated from cell cultures harvested at  $\sim 80\%$  confluency using the QIAgen miRNAeasy kit with on-column DNase 1 digestion. 500 ng of RNA was reverse transcribed using SuperScript III First-Strand Synthesis System (Invitrogen). The cDNA was diluted to 10 ng  $\mu$ l<sup>-1</sup> and 12.5 ng was used in target specific amplification before real-time PCR using TaqMan PreAmp Master Mix Kit (Applied Biosystems) following Fluidigm's Specific Target Amplification Protocol. 1.25  $\mu$ l of the 25  $\mu$ l pre-amplified cDNA was added to each chip. Each sample was run in triplicate and each experiment included no template controls and no template controls from the cDNA reactions. 96.96 Dynamic Array Integrated Fluidic Circuits (Fluidigm) were loaded with 96 pre-amplified cDNA samples and 96 TaqMan gene expression probes (Applied Biosystems) using the BioMark HD System (Fluidigm). Expression levels for each gene were normalized to the average expression of control genes (*GAPDH* and *ACTB*). Relative expression levels were calculated using the  $\Delta\Delta$ Ct method. Correlations between genotype and gene expression were calculated in R 2.14.1. Genotype specific gene expression was compared using the Jonckheere-Terpstra test. Genes with significant eQTL results were validated by individual Taqman (Applied Biosystems, Warrington UK) reactions run on ABI 7900HT Sequence Detection System equipment and analysed with SDS software according to the manufacturer's instructions. Normal cell line DNAs were analysed on iCOGS arrays to obtain genotype information. We analysed all protein-coding genes within a 1 Mb region of the risk association. The method for allele specific expression analysis has been described previously<sup>31</sup>.

**Breast and ovarian normal and cancer cell lines.** Breast and OC cell lines MCF7 (ER+, breast; ATCC #HTB-22) and A2780 (ER+, ovarian; kindly provided by Thomas Hamilton, NCI, Maryland) were grown in RPMI medium with 10% FBS and antibiotics. The normal breast epithelial cell lines Bre-80 (kindly provided by Roger Reddel, CMRI, Sydney) and MCF10A (ATCC #CRL-10317) were grown in DMEM/F12 medium with 5% horse serum, 10 mg ml<sup>-1</sup> insulin, 0.5 mg ml<sup>-1</sup> hydrocortisone, 20 ng ml<sup>-1</sup> epidermal growth factor, 100 ng ml<sup>-1</sup> cholera toxin and antibiotics. The phenotypically normal TERT immortalized ovarian epithelial cell lines IOSE11 and IOSE19 (ref. 32) were grown in NOSE-CM. All cell lines were maintained under standard conditions, were routinely tested for *Mycoplasma* and were profiled with short tandem repeats to confirm their identity.

**Functional annotation of risk SNPs.** FAIRE-seq and ChIP-seq for H3K27ac and H3K4me1 marks in normal ovarian (IOSE4, IOSE11) and fallopian epithelial cell lines (FT33, FT246) and OC cell lines (CaOV3, UWB1.289) were generated in-house using standard protocols and have been previously described<sup>19,27</sup>. Epigenetic marks in HMECs were downloaded from ENCODE (genome.ucsc.edu).

**Chromosome conformation capture.** 3C libraries were generated using *NcoI* as described previously<sup>14</sup>. To quantify interactions by real-time quantitative PCR (qPCR) was performed using primers listed in Supplementary Table 9. All qPCRs were performed on a RotorGene 6,000 using MyTaq HS DNA polymerase with the addition of 5 mM of Syto9, annealing temperature of 66 °C and extension of 30 s. Each experiment was performed three times in duplicate. The BAC clone (CTD-2278110) covering the 19p13 region was used to normalize for PCR efficiency and a by reference region within *GAPDH* used to calculate relative interaction frequencies. All qPCR products were resolved on 2% agarose gels, gel purified and sequenced to verify the 3C product.

**RNA stability assays.** For each genotype (two homozygotes and the heterozygote) two early passage primary normal ovarian epithelial cell lines were incubated with actinomycin D for 20 h. RNA was extracted using the QIAgen RNeasy extraction kit and reverse transcribed using MMLV RT enzyme and random hexamers (Promega). Quantitative PCR was performed using TaqMan gene expression probes for *ABHD8* (Hs00225984\_m1) and *ANKLE1* (Hs01094673\_g1). Signal for each gene of interest was normalized to signal for *ACTB* (Hs01060665\_g1) and

*GAPDH* (Hs02758991\_g1) and relative gene expression calculated using the  $\Delta\Delta Ct$  method, relative to untreated cells. 18s rRNA (Hs99999901\_s1) and *MYC* (Hs00153408\_m1) mRNA levels were included as internal controls.

**Promoter and allele specific enhancer assays.** A 1119 bp fragment containing the *ABHD8* promoter was cloned into the pGL3 basic luciferase reporter. Reference and risk associated *ANKLE1* promoter fragments were synthesized by GenScript and cloned into pGL3 basic. We generated PCR fragments corresponding to PRE A and PRE B and had PRE C haplotype fragments synthesized by GenScript and these were also sub-cloned into *ABHD8* and *ANKLE1* promoter constructs. PCR primers are listed in Supplementary Table 10. Bre80 and A2780 cells were transiently transfected with equimolar amounts of luciferase reporter constructs using *Renilla* luciferase as an internal control reporter. Luciferase was measured 24 h after transfection using Dual-Glo Luciferase (Promega). To correct for any differences in transfection efficiency or cell lysate preparation, *Firefly* luciferase activity was normalized to *Renilla* luciferase, and the activity of each construct was measured relative to the promoter alone construct, which had a defined activity of 1. Association was assessed by log transforming the data and performing two-way ANOVA, followed by Dunnett's multiple comparisons test; for ease of interpretation, values were back transformed to the original scale for the graphs.

**Genome editing.** Guide RNAs targeting the region flanking rs56069439 (5'-GTGAGACGGTCAGAACCAAT-3' and 5'-GTGTCTGAGGCCGAAAGAGC-3') were designed using the CRISPR design tool from the Zhang lab ([www.crispr.mit.edu](http://www.crispr.mit.edu))<sup>57</sup>. The gRNAs were cloned into the lentiCRISPR (Addgene Plasmid 49535) vector by using the *BsmBI* restriction enzyme site and lentiviral supernatants made by cotransfection of HEK293T cells. IOSE19 and MCF10A cells were transduced with viral supernatants and infected cells selected using 400 ng ml<sup>-1</sup> and 500 ng ml<sup>-1</sup> puromycin (Sigma Aldrich) respectively. Selected cells were sorted into single cells using flow cytometry and expanded *in vitro*. Screening for clones containing the deletion was performed using the following primers: Forwards: 5'-CCCTGACATC CAGGGTCTTC-3' and Reverse: 5'-AGTCCAGCGTCTCATCGGTA-3'. For sequence verification of the deletion the following primers were used: Forwards: 5'-TCTGGACACAGTCCCTGACA-3' and Reverse: 5'-CAGCGTCTCATCGGT AGGTC-3'. RNA was isolated from positive clones using the Zymo Quick-RNA kit and reverse transcribed using Superscript III (Life Technologies). Real time gene expression analysis was performed using TaqMan probes, as described above.

**In vitro analysis of candidate genes.** The three candidate genes were overexpressed as green fluorescent protein fusion proteins. The *BABAM1* overexpression construct was a kind gift from Dr S Elledge<sup>58</sup>. *ANKLE1* and *ABHD8* constructs were purchased from Genecopoeia. Virus was made in-house by cotransfection of HEK293Ts and used to transduce MCF10A and IOSE19 cells. Positive cells were selected using 400 ng ml<sup>-1</sup> (for IOSE19 cells) or 500 ng ml<sup>-1</sup> (for MCF10A cells) puromycin. Anchorage dependent and independent growth assays were performed as previously described<sup>32,59</sup>. For invasion and migration assays Millipore luminescent transwell assays (24 well plate format) were used, following the manufacturer's protocol.

**Data availability.** The relevant SNP genotype data underpinning these analyses can be accessed by applying to the OCAC, BCAC and CIMBA consortia (see URLs). EQTL data are available in supplementary information. All other data are available on request.

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### Author contributions

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Kate Lawrenson<sup>1,\*</sup>, Siddhartha Kar<sup>2,\*</sup>, Karen McCue<sup>3</sup>, Karoline Kuchenbaecker<sup>4</sup>, Kyriaki Michailidou<sup>4</sup>, Jonathan Tyrer<sup>2</sup>, Jonathan Beesley<sup>3</sup>, Susan J. Ramus<sup>1</sup>, Qiyuan Li<sup>5,6</sup>, Melissa K. Delgado<sup>1</sup>, Janet M. Lee<sup>1</sup>, Kristiina Aittomäki<sup>7</sup>, Irene L. Andrulis<sup>8,9</sup>, Hoda Anton-Culver<sup>10</sup>, Volker Arndt<sup>11</sup>, Banu K. Arun<sup>12</sup>, Brita Arver<sup>13</sup>, Elisa V. Bandera<sup>14</sup>, Monica Barile<sup>15</sup>, Rosa B. Barkardottir<sup>16</sup>, Daniel Barrowdale<sup>4</sup>, Matthias W. Beckmann<sup>17</sup>, Javier Benitez<sup>18,19</sup>, Andrew Berchuck<sup>20</sup>, Maria Bisogna<sup>21</sup>, Line Borge<sup>22,23</sup>, Carl Blomqvist<sup>24</sup>, William Blot<sup>25,26</sup>, Natalia Bogdanova<sup>27</sup>, Anders Bojesen<sup>28</sup>, Stig E. Bojesen<sup>29,30,31</sup>, Manjeet K. Bolla<sup>4</sup>, Bernardo Bonanni<sup>15</sup>, Anne-Lise Børresen-Dale<sup>32,33</sup>, Hiltrud Brauch<sup>34,35,36</sup>, Paul Brennan<sup>37</sup>, Hermann Brenner<sup>11,36,38</sup>, Fiona Bruinsma<sup>39</sup>, Joan Brunet<sup>40</sup>, Shaik Ahmad Buhari<sup>41</sup>, Barbara Burwinkel<sup>42,43</sup>, Ralf Butzow<sup>44,45</sup>, Sandra S. Buys<sup>46</sup>, Qiuyin Cai<sup>25</sup>, Trinidad Caldes<sup>47</sup>, Ian Campbell<sup>48</sup>, Rikki Canniotto<sup>49</sup>, Jenny Chang-Claude<sup>50,51</sup>, Jocelyne Chiquette<sup>52</sup>, Ji-Yeob Choi<sup>53,54</sup>, Kathleen B.M. Claes<sup>55</sup>, GEMO Study Collaborators<sup>‡</sup>, Linda S. Cook<sup>56</sup>, Angela Cox<sup>57</sup>, Daniel W. Cramer<sup>58,59</sup>, Simon S. Cross<sup>60</sup>, Cezary Cybulski<sup>61</sup>, Kamila Czene<sup>62</sup>, Mary B. Daly<sup>63</sup>, Francesca Damiola<sup>64</sup>, Agnieszka Dansonka-Mieszkowska<sup>65</sup>, Hatf Darabi<sup>62</sup>, Joe Dennis<sup>4</sup>, Peter Devilee<sup>66,67</sup>, Orland Diez<sup>68</sup>, Jennifer A. Doherty<sup>69</sup>, Susan M. Domchek<sup>70</sup>, Cecilia M. Dorfling<sup>71</sup>, Thilo Dörk<sup>27</sup>, Martine Dumont<sup>72</sup>, Hans Ehrencrona<sup>73,74</sup>, Bent Ejlersen<sup>75</sup>, Steve Ellis<sup>4</sup>, EMBRACE<sup>‡</sup>, Christoph Engel<sup>76</sup>, Eunjung Lee<sup>1</sup>, D. Gareth Evans<sup>77</sup>, Peter A. Fasching<sup>17,78</sup>, Lidia Feliubadalo<sup>79</sup>, Jonine Figueroa<sup>80</sup>, Dieter Flesch-Janys<sup>81,82</sup>, Olivia Fletcher<sup>83,84</sup>, Henrik Flyger<sup>85</sup>, Lenka Foretova<sup>86</sup>, Florentia Fostira<sup>87</sup>, William D. Foulkes<sup>88</sup>,



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Massuger<sup>150</sup>, Keitaro Matsuo<sup>151</sup>, Sylvie Mazoyer<sup>64</sup>, Lesley McGuffog<sup>4</sup>, Catriona McLean<sup>152</sup>, Iain McNeish<sup>153</sup>, Alfons Meindl<sup>154</sup>, Usha Menon<sup>95</sup>, Arjen R. Mensenkamp<sup>155</sup>, Roger L. Milne<sup>39,96</sup>, Marco Montagna<sup>156</sup>, Kirsten B. Moysich<sup>157</sup>, Kenneth Muir<sup>144,158</sup>, Anna Marie Mulligan<sup>159,160</sup>, Katherine L. Nathanson<sup>70</sup>, Roberta B. Ness<sup>161</sup>, Susan L. Neuhausen<sup>162</sup>, Heli Nevanlinna<sup>44</sup>, Silje Nord<sup>33</sup>, Robert L. Nussbaum<sup>163</sup>, Kunle Odunsi<sup>164</sup>, Kenneth Offit<sup>165</sup>, Edith Olah<sup>166</sup>, Olufunmilayo I. Olopade<sup>167</sup>, Janet E. Olson<sup>102</sup>, Curtis Olswold<sup>102</sup>, David O'Malley<sup>168</sup>, Irene Orlow<sup>169</sup>, Nick Orr<sup>83</sup>, Ana Osorio<sup>170,171</sup>, Sue Kyung Park<sup>54,129,172</sup>, Celeste L. Pearce<sup>1</sup>, Tanja Pejovic<sup>173,174</sup>, Paolo Peterlongo<sup>175</sup>, Georg Pfeiler<sup>176</sup>, Catherine M. Phelan<sup>177</sup>, Elizabeth M. Poole<sup>178,179</sup>, Katri Pylkäs<sup>180,181</sup>, Paolo Radice<sup>182</sup>, Johanna Rantala<sup>183</sup>, Muhammad Usman Rashid<sup>109,184</sup>, Gad Rennert<sup>185</sup>, Valerie Rhenius<sup>2</sup>, Kerstin Rhiem<sup>186</sup>, Harvey A. Risch<sup>187</sup>, Gus Rodriguez<sup>188</sup>, Mary Anne Rossing<sup>189,190</sup>, Anja Rudolph<sup>50</sup>, Helga B. Salvesen<sup>22,23</sup>, Suleeporn Sangrajrang<sup>191</sup>, Elinor J. Sawyer<sup>192</sup>, Joellen M. Schildkraut<sup>193,194</sup>, Marjanka K. Schmidt<sup>195</sup>, Rita K. Schmutzler<sup>196,197,198,199</sup>, Thomas A. Sellers<sup>177</sup>, Caroline Seynaeve<sup>122</sup>, Mitul Shah<sup>2</sup>, Chen-Yang Shen<sup>200,201</sup>, Xiao-Ou Shu<sup>25</sup>, Weiva Sieh<sup>202</sup>, Christian F. Singer<sup>176</sup>, Olga M. Sinilnikova<sup>203,204</sup>, Susan Slager<sup>102</sup>, Honglin Song<sup>2</sup>, Penny Soucy<sup>72</sup>, Melissa C. Southey<sup>205</sup>, Marie Stenmark-Askmal<sup>74,206</sup>, Dominique Stoppa-Lyonnet<sup>207,208</sup>, Christian Sutter<sup>209</sup>, Anthony Swerdlow<sup>84,94</sup>, Sandrine Tchatchou<sup>8</sup>, Manuel R. Teixeira<sup>210,211</sup>, Soo H. Teo<sup>113,114</sup>, Kathryn L. Terry<sup>58,59</sup>, Mary Beth Terry<sup>212</sup>, Mads Thomassen<sup>213</sup>, Maria Grazia Tibiletti<sup>214</sup>, Laima Tihomirova<sup>215</sup>, Silvia Tognazzo<sup>216</sup>, Amanda Ewart Toland<sup>217</sup>, Ian Tomlinson<sup>218</sup>, Diana Torres<sup>109,219</sup>, Thérèse Truong<sup>107,108</sup>, Chiu-chen Tseng<sup>1</sup>, Nadine Tung<sup>220</sup>, Shelley S. Tworoger<sup>178,179</sup>, Celine Vachon<sup>102</sup>, Ans M.W. van den Ouweland<sup>221</sup>, Helena C. van Doorn<sup>222</sup>, Elizabeth J. van Rensburg<sup>71</sup>, Laura J. Van't Veer<sup>195</sup>, Adriaan Vanderstichele<sup>223</sup>, Ignace Vergote<sup>223</sup>, Joseph Vijai<sup>165</sup>, Qin Wang<sup>4</sup>, Shan Wang-Gohrke<sup>224</sup>, Jeffrey N. Weitzel<sup>117</sup>, Nicolas Wentzensen<sup>225</sup>, Alice S. Whittemore<sup>202</sup>, Hans Wildiers<sup>226</sup>, Robert Winqvist<sup>180,181</sup>, Anna H. Wu<sup>1</sup>, Drakoulis Yannoukakos<sup>227</sup>, Sook-Yee Yoon<sup>228,229</sup>, Jyh-Cherng Yu<sup>230</sup>, Wei Zheng<sup>25</sup>, Ying Zheng<sup>231</sup>, Kum Kum Khanna<sup>3</sup>, Jacques Simard<sup>72</sup>, Alvaro N. Monteiro<sup>232</sup>, Juliet D. French<sup>3</sup>, Fergus J. Couch<sup>102,233</sup>, Matthew L. Freedman<sup>6</sup>, Douglas F. Easton<sup>2,4</sup>, Alison M. Dunning<sup>2</sup>, Paul D. Pharoah<sup>2</sup>, Stacey L. Edwards<sup>3</sup>, Georgia Chenevix-Trench<sup>3,\*\*</sup>, Antonis C. Antoniou<sup>4,\*\*</sup> & Simon A. Gayther<sup>1\*\*,†</sup>

<sup>1</sup> Department of Preventive Medicine, Keck School of Medicine, University of Southern California Norris Comprehensive Cancer Center, Los Angeles, California 90033, USA. <sup>2</sup> Department of Oncology, Centre for Cancer Genetic Epidemiology, University of Cambridge, Cambridge CB1 8RN, UK. <sup>3</sup> QIMR Berghofer Medical Research Institute, Brisbane, Queensland 4029, Australia. <sup>4</sup> Department of Public Health and Primary Care, Centre for Cancer Genetic Epidemiology, University of Cambridge, Cambridge CB1 8RN, UK. <sup>5</sup> Medical College, Xiamen University, Xiamen 361102, China. <sup>6</sup> Department of Medical Oncology, The Center for Functional Cancer Epigenetics, Dana-Farber Cancer Institute, Boston, Massachusetts 02215, USA. <sup>7</sup> Department of Clinical Genetics, Helsinki University Hospital, University of Helsinki, Helsinki 00029 HUS, Finland. <sup>8</sup> Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, Ontario Canada, M5G 1X5. <sup>9</sup> Department of Molecular Genetics, University of Toronto, Toronto, Ontario Canada, M5S 1A8. <sup>10</sup> Department of Epidemiology, Genetic Epidemiology Research Institute, School of Medicine, University of California Irvine, Irvine, California 92697, USA. <sup>11</sup> Division of Clinical Epidemiology and Aging Research, German Cancer Research Center (DKFZ), Heidelberg, 69120, Germany. <sup>12</sup> University of Texas MD Anderson Cancer Center, Houston, Texas 77030, USA. <sup>13</sup> Department of Oncology, Karolinska University Hospital, Stockholm 171 77, Sweden. <sup>14</sup> Cancer Prevention and Control, Rutgers Cancer Institute of New Jersey, New Brunswick, New Jersey 08903, USA. <sup>15</sup> Division of Cancer Prevention and Genetics, Istituto Europeo di Oncologia, Milan 20141, Italy. <sup>16</sup> Department of Pathology, Landspítali University Hospital and BMC (Biomedical Centre), Faculty of Medicine, University of Iceland, Reykjavik 600169-2039, Iceland. <sup>17</sup> University Hospital Erlangen, Department of Gynecology and Obstetrics, Friedrich-Alexander-University Erlangen-Nuremberg, Comprehensive Cancer Center Erlangen-EMN, Erlangen 91054, Germany. <sup>18</sup> Human Cancer Genetics Program, Spanish National Cancer Research Centre, Madrid E-28029, Spain. <sup>19</sup> Centro de Investigación en Red de Enfermedades Raras, Valencia 28029, Spain. <sup>20</sup> Department of Obstetrics and Gynecology, Duke University Medical Center, Durham, North Carolina 27710, USA. <sup>21</sup> Gynecology Service, Department of Surgery, Memorial Sloan-Kettering Cancer Center, New York 10065, USA. <sup>22</sup> Department of Gynecology and Obstetrics, Haukeland University Hospital, 5021 Bergen, Norway. <sup>23</sup> Centre for Cancer Biomarkers, Department of Clinical Science, University of Bergen, N-5020 Bergen, Norway. <sup>24</sup> Department of Oncology, Helsinki University Hospital, University of Helsinki, Helsinki FIN-00029, Finland. <sup>25</sup> Division of Epidemiology, Department of Medicine, Vanderbilt-Ingram Cancer Center, Vanderbilt University School of Medicine, Nashville, Tennessee 37203, USA. <sup>26</sup> International Epidemiology Institute, Rockville, Maryland 20850, USA. <sup>27</sup> Gynaecology Research Unit, Hannover Medical School, Hannover D-30625, Germany. <sup>28</sup> Department of Clinical Genetics, Vejle Hospital, Vejle 7100, Denmark. <sup>29</sup> Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen 2200, Denmark. <sup>30</sup> Department of Clinical Biochemistry, Herlev Hospital, Copenhagen University Hospital, Herlev 2730, Denmark. <sup>31</sup> Copenhagen General Population Study, Herlev Hospital, Copenhagen University Hospital, Herlev 2730, Denmark. <sup>32</sup> Department of Genetics, Institute for Cancer Research, Oslo University Hospital Radiumhospitalet, Oslo N-0310, Norway. <sup>33</sup> K.G. Jebsen Center for Breast Cancer Research, Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo N-0310, Norway. <sup>34</sup> Dr Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart D-70376, Germany. <sup>35</sup> University of Tübingen, Tübingen 72074, Germany. <sup>36</sup> German Cancer Consortium (DKTK), German Cancer Research Center (DKFZ), Heidelberg 69120, Germany. <sup>37</sup> International Agency for Research on Cancer, Lyon 69008, France. <sup>38</sup> Division of Preventive Oncology, German Cancer Research Center (DKFZ), Heidelberg 69121, Germany. <sup>39</sup> Cancer Epidemiology Centre, Cancer Council Victoria, Melbourne, Victoria 3004, Australia. <sup>40</sup> Genetic Counseling Unit, Hereditary Cancer Program, IDIBGI (Institut d'Investigació Biomèdica de Girona), Catalan Institute of Oncology, Girona 08908, Spain. <sup>41</sup> Department of Surgery, National University Health System, Singapore 119077, Singapore. <sup>42</sup> Molecular Epidemiology Group, German Cancer Research Center (DKFZ), Heidelberg 69120, Germany. <sup>43</sup> Department of Obstetrics and Gynecology, University of Heidelberg, Heidelberg 69120, Germany. <sup>44</sup> Department of Obstetrics and Gynecology, University of Helsinki and Helsinki University Central Hospital, Helsinki 00029 HUS, Finland. <sup>45</sup> Department of Pathology, Helsinki University Central Hospital, Helsinki 00029, Finland. <sup>46</sup> Department of Medicine, Huntsman Cancer Institute, University of Utah School of Medicine, Salt Lake City, Utah 84112, USA. <sup>47</sup> Molecular Oncology Laboratory, Hospital Clínico San Carlos, IdISSC (El Instituto de Investigación Sanitaria del Hospital Clínico San Carlos), Madrid 28040, Spain. <sup>48</sup> Cancer Genetics Laboratory, Peter MacCallum Cancer Centre, Melbourne, Victoria 3002, Australia. <sup>49</sup> Cancer Pathology & Prevention, Division of Cancer Prevention and Population Sciences, Roswell Park Cancer Institute, Elm and Carlton Streets, Buffalo 14263, New York, USA. <sup>50</sup> Division of Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg 69121, Germany. <sup>51</sup> University Cancer Center Hamburg (UCC), University Medical Center Hamburg-Eppendorf, Hamburg 20246, Germany. <sup>52</sup> Unité de recherche en santé des populations, Centre des maladies du sein Deschênes-Fabia, Centre de recherche FRSQ du Centre hospitalier affilié universitaire de Québec, Québec City, Québec Canada, G1J 1Z4. <sup>53</sup> Cancer Research Institute, Seoul National University, Seoul 08826, Korea. <sup>54</sup> Department of Biomedical Sciences, Seoul National University College of Medicine, Seoul 03080, Korea. <sup>55</sup> Center for Medical Genetics, Ghent University, Ghent 9000, Belgium. <sup>56</sup> Division of Epidemiology and Biostatistics, Department of Internal Medicine, University of New Mexico, Albuquerque, New Mexico 87131, USA. <sup>57</sup> Sheffield Cancer Research, Department of Oncology, University of Sheffield, Sheffield S10 2TN, UK. <sup>58</sup> Harvard HT Chan School of Public Health, Boston, Massachusetts 02115, USA. <sup>59</sup> Obstetrics and Gynecology Epidemiology Center, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts 02115, USA. <sup>60</sup> Academic Unit of Pathology, Department of Neuroscience, University of Sheffield, Sheffield S10 2TN, UK. <sup>61</sup> Department of Genetics and Pathology, Pomeranian Medical University, Szczecin 70-115, Poland. <sup>62</sup> Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm SE-171 77, Sweden. <sup>63</sup> Department of Clinical Genetics, Fox Chase Cancer Center, Philadelphia, Pennsylvania 19111, USA. <sup>64</sup> INSERM U1052, CNRS UMR5286, Université Lyon, Centre de Recherche en Cancérologie de Lyon, Lyon 69373, France. <sup>65</sup> Department of Pathology and Laboratory Diagnostics the Maria Skłodowska Curie Memorial Cancer Center and Institute of Oncology, Warsaw 44-101, Poland. <sup>66</sup> Department of Pathology, Leiden University Medical Center, Leiden 2333, The Netherlands. <sup>67</sup> Department of Human Genetics, Leiden University Medical Center, Leiden 2333, The Netherlands. <sup>68</sup> Oncogenetics Group, University Hospital Vall d'Hebron, Vall d'Hebron Institute of Oncology (VHIO) and Universitat Autònoma de Barcelona, Barcelona 08035, Spain. <sup>69</sup> Department of Community and Family Medicine, Section of Biostatistics & Epidemiology, The Geisel School of Medicine at Dartmouth, Lebanon, New Hampshire 03755, USA. <sup>70</sup> Department of Medicine, Abramson Cancer Center, Perelman School of Medicine, University of Pennsylvania, Philadelphia, Pennsylvania 19104, USA. <sup>71</sup> Department of Genetics, University of Pretoria, Pretoria 0083, South Africa. <sup>72</sup> Genomics Center, Centre Hospitalier Universitaire de Québec Research Center, Laval University, Québec City, Québec Canada, G1V 4G2. <sup>73</sup> Department of Immunology, Genetics and Pathology, Uppsala University, Uppsala 751 05, Sweden. <sup>74</sup> Department of Clinical Genetics, Lund University Hospital, Lund 221 00, Sweden. <sup>75</sup> Department of Oncology, Rigshospitalet, Copenhagen University Hospital, 2100 Copenhagen, Denmark. <sup>76</sup> Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, 04107 Leipzig, Germany. <sup>77</sup> Genomic Medicine, Manchester Academic Health Sciences Centre, Institute of Human Development, Manchester University, Central Manchester University Hospitals NHS Foundation Trust, Manchester M13 9PL, UK. <sup>78</sup> University of California at Los Angeles, David Geffen School of Medicine, Department of Medicine, Division of Hematology and Oncology, Los Angeles California 90095, USA. <sup>79</sup> Molecular Diagnostic Unit, Hereditary Cancer Program, IDIBELL (Bellvitge Biomedical Research Institute), Catalan Institute of Oncology, Barcelona 08908, Spain. <sup>80</sup> Division of Cancer Epidemiology and Genetics, National Cancer Institute, Rockville, Maryland 20892, USA. <sup>81</sup> Institute for Medical Biometrics and Epidemiology, University Medical Center Hamburg-Eppendorf, Hamburg 20246, Germany. <sup>82</sup> Department of Cancer Epidemiology, Clinical Cancer Registry, University Medical Center Hamburg-Eppendorf, Hamburg 20246, Germany. <sup>83</sup> Breakthrough Breast Cancer Research Centre, The Institute of Cancer Research, London SW3 6JB, UK. <sup>84</sup> Division of Breast Cancer Research, The Institute of Cancer Research, London SW7 3RP, UK. <sup>85</sup> Department of Breast Surgery, Herlev Hospital, Copenhagen University Hospital, 2730 Herlev, Denmark. <sup>86</sup> Masaryk Memorial Cancer Institute and Medical Faculty MU, 625 00 Brno, Czech Republic. <sup>87</sup> Molecular Diagnostics Laboratory, (INRASTES) Institute of Nuclear and Radiological Sciences and Technology, National Centre for Scientific Research 'Demokritos', Aghia Paraskevi Attikis, Athens 153 10, Greece. <sup>88</sup> Program in Cancer Genetics, Departments of Human Genetics and Oncology, McGill University, Montreal, Quebec H2W 1S6, Canada. <sup>89</sup> Biostatistics and Informatics Shared Resource, University of Kansas Medical



Center, Kansas City, Kansas 66160, USA. <sup>90</sup> Susanne Levy Gertner Oncogenetics Unit, Sheba Medical Center, Tel-Hashomer 52621, Israel. <sup>91</sup> Section of Genetic Oncology, Department of Laboratory Medicine, University and University Hospital of Pisa, Pisa 56126, Italy. <sup>92</sup> UCLA Schools of Medicine and Public Health, Division of Cancer Prevention & Control Research, Jonsson Comprehensive Cancer Center, Los Angeles, California 90024, USA. <sup>93</sup> Cancer Risk and Prevention Clinic, Dana-Farber Cancer Institute, Boston, Massachusetts 02215, USA. <sup>94</sup> Division of Genetics and Epidemiology, The Institute of Cancer Research, London SW7 3RP, UK. <sup>95</sup> Women's Cancer, UCL EGA Institute for Women's Health, London WC1E 6AU, UK. <sup>96</sup> Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global Health, The University of Melbourne, Melbourne, Victoria 3010, Australia. <sup>97</sup> Cancer Research UK Clinical Trials Unit, The Beatson West of Scotland Cancer Centre, Glasgow G12 0YN, UK. <sup>98</sup> Department of Pathology and Laboratory Medicine, University of Kansas Medical Center, Kansas City, Kansas 66160, USA. <sup>99</sup> Division of Clinical Epidemiology, Royal Victoria Hospital, McGill University, Montreal, Québec H3A 1A1, Canada. <sup>100</sup> Department of Medicine, McGill University, Montreal, Québec H3A 1A1, Canada. <sup>101</sup> Department of Dermatology, Huntsman Cancer Institute, University of Utah School of Medicine, Salt Lake City, Utah 84132, USA. <sup>102</sup> Department of Health Sciences Research, Mayo Clinic, Rochester, Minnesota 55902, USA. <sup>103</sup> Cancer Prevention and Control, Samuel Oschin Comprehensive Cancer Institute, Cedars-Sinai Medical Center, Los Angeles, California 90048, USA. <sup>104</sup> Community and Population Health Research Institute, Department of Biomedical Sciences, Cedars-Sinai Medical Center, Los Angeles, California 90048, USA. <sup>105</sup> Clinical Genetics Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Rockville, Maryland 20892, USA. <sup>106</sup> Department of Genetics and Pathology, Pomeranian Medical University, 70-204 Szczecin, Poland. <sup>107</sup> Environmental Epidemiology of Cancer, Center for Research in Epidemiology and Population Health, INSERM, 94805 Villejuif, France. <sup>108</sup> University Paris-Sud, 91405 Villejuif, France. <sup>109</sup> Molecular Genetics of Breast Cancer, German Cancer Research Center (DKFZ), Heidelberg 69120, Germany. <sup>110</sup> Center for Genomic Medicine, Rigshospitalet, Copenhagen University Hospital, 2100 Copenhagen, Denmark. <sup>111</sup> Department of Oncology, Department of Public Health and Primary Care, University of Cambridge, Strangeways Research Laboratory, Cambridge CB1 8RN, UK. <sup>112</sup> Saw Swee Hock School of Public Health, National University of Singapore Singapore 119077, Singapore. <sup>113</sup> Breast Cancer Research Unit, Cancer Research Institute, University Malaya Medical Centre, 50603 Kuala Lumpur, Malaysia. <sup>114</sup> Cancer Research Initiatives Foundation, Subang Jaya, 47500 Selangor, Malaysia. <sup>115</sup> Department of Gynecology and Gynecologic Oncology, Kliniken Essen-Mitte, 45136 Essen, Germany. <sup>116</sup> Department of Gynecology and Gynecologic Oncology, Dr Horst Schmidt Kliniken Wiesbaden, 65199 Wiesbaden, Germany. <sup>117</sup> Clinical Cancer Genetics, for the City of Hope Clinical Cancer Genetics Community Research Network, Duarte California 91010, USA. <sup>118</sup> Molecular Unit, Department of Pathology, Herlev Hospital, University of Copenhagen, 2730 Copenhagen, Denmark. <sup>119</sup> Department of Virus, Lifestyle and Genes, Danish Cancer Society Research Center, DK-2100 Copenhagen, Denmark. <sup>120</sup> Department of Gynecology, Rigshospitalet, University of Copenhagen, 2100 Copenhagen, Denmark. <sup>121</sup> Family Cancer Clinic, Netherlands Cancer Institute, 1006 Amsterdam, The Netherlands. <sup>122</sup> Department of Medical Oncology, Family Cancer Clinic, Erasmus MC Cancer Institute, 3015 Rotterdam, The Netherlands. <sup>123</sup> Center for Medical Genetics, NorthShore University Health System, Evanston, Illinois 60201, USA. <sup>124</sup> N.N. Petrov Institute of Oncology, St Petersburg 197758, Russia. <sup>125</sup> Lombardi Comprehensive Cancer Center, Georgetown University, Washington District of Columbia 20057, USA. <sup>126</sup> Division of Epidemiology and Prevention, Aichi Cancer Center Research Institute, Aichi 464-8681, Japan. <sup>127</sup> State Research Institute Centre for Innovative Medicine, LT-01102 Vilnius, Lithuania. <sup>128</sup> Department of Epidemiology, Cancer Prevention Institute of California, Fremont, California 94538, USA. <sup>129</sup> Department of Preventive Medicine, Seoul National University College of Medicine, Seoul 08826, Korea. <sup>130</sup> Centre for Epidemiology and Biostatistics, University of Melbourne, Melbourne, Victoria 3010, Australia. <sup>131</sup> Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute, Cedars-Sinai Medical Center, Los Angeles, California 90048, USA. <sup>132</sup> Radboud University Medical Centre, Radboud Institute for Health Sciences, 6500 Nijmegen, The Netherlands. <sup>133</sup> Prosserman Centre for Health Research, Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, Ontario M5G 1X5, Canada. <sup>134</sup> Division of Epidemiology, Dalla Lana School of Public Health, University of Toronto, Toronto, Ontario M5T 3M7, Canada. <sup>135</sup> Imaging Center, Department of Clinical Pathology, Kuopio University Hospital, 70210 Kuopio, Finland. <sup>136</sup> Cancer Center, Kuopio University Hospital, 70210 Kuopio, Finland. <sup>137</sup> Institute of Clinical Medicine, Pathology and Forensic Medicine, University of Eastern Finland, 70210 Kuopio, Finland. <sup>138</sup> Department of Clinical Molecular Biology, Oslo University Hospital, University of Oslo, 1478 Oslo, Norway. <sup>139</sup> The Hong Kong Hereditary Breast Cancer Family Registry, Cancer Genetics Center, Hong Kong Sanatorium and Hospital, Hong Kong, China. <sup>140</sup> Department of Surgery, The University of Hong Kong, Hong Kong, China. <sup>141</sup> Vesalius Research Center, VIB, 3000 Leuven, Belgium. <sup>142</sup> Laboratory for Translational Genetics, Department of Oncology, University of Leuven, 3000 Leuven, Belgium. <sup>143</sup> Department of Molecular Medicine and Surgery, Karolinska Institutet, SE-171 77 Stockholm, Sweden. <sup>144</sup> Division of Health Sciences, Warwick Medical School, Warwick University, Coventry CV4 7AL, UK. <sup>145</sup> Department of Gynecologic Oncology, The University of Texas MD Anderson Cancer Center, Houston, Texas 77030, USA. <sup>146</sup> Unit of Medical Genetics, Department of Preventive and Predictive Medicine, Fondazione IRCCS (Istituto Di Ricovero e Cura a Carattere Scientifico) Istituto Nazionale Tumori (INT), 20133 Milan, Italy. <sup>147</sup> University of Hawaii Cancer Center, Honolulu, Hawaii 96813, USA. <sup>148</sup> Department of Oncology - Pathology, Karolinska Institutet, SE-171 77 Stockholm, Sweden. <sup>149</sup> National Center for Tumour Diseases, University of Heidelberg, 69117 Heidelberg, Germany. <sup>150</sup> Department of Gynaecology, Radboud University Medical Centre, 6500 Nijmegen, The Netherlands. <sup>151</sup> Department of Preventive Medicine, Kyushu University Faculty of Medical Sciences, Fukuoka 812-8582, Japan. <sup>152</sup> Anatomical Pathology, The Alfred Hospital, Melbourne, Victoria 3004, Australia. <sup>153</sup> Institute of Cancer Sciences, University of Glasgow, Wolfson Wohl Cancer Research Centre, Beatson Institute for Cancer Research, Glasgow G61 1BD, UK. <sup>154</sup> Division of Gynaecology and Obstetrics, Technische Universität München, 81675 Munich, Germany. <sup>155</sup> Department of Human Genetics, Radboud University Medical Centre, 6500 Nijmegen, The Netherlands. <sup>156</sup> Immunology and Molecular Oncology Unit, Instituto Oncologico Veneto IOV, IRCCS, 35128 Padua, Italy. <sup>157</sup> Department of Cancer Prevention and Control, Roswell Park Cancer Institute, Buffalo, New York 14263, USA. <sup>158</sup> Institute of Population Health, University of Manchester, Manchester M13 9PL, UK. <sup>159</sup> Laboratory Medicine Program, University Health Network, Toronto, Ontario M5G 1L7, Canada. <sup>160</sup> Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, Ontario M5G 1L7, Canada. <sup>161</sup> The University of Texas School of Public Health, Houston, Texas 77030, USA. <sup>162</sup> Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte, California 91010, USA. <sup>163</sup> Department of Medicine and Genetics, University of California, San Francisco, California 94143, USA. <sup>164</sup> Department of Gynecological Oncology, Roswell Park Cancer Institute, Buffalo, New York 14263, USA. <sup>165</sup> Department of Medicine, Memorial Sloan-Kettering Cancer Center, New York 10065, USA. <sup>166</sup> Department of Molecular Genetics, National Institute of Oncology, 1122 Budapest, Hungary. <sup>167</sup> Center for Clinical Cancer Genetics and Global Health, University of Chicago Medical Center, Chicago, Illinois 60637, USA. <sup>168</sup> The Ohio State University and the James Cancer Center, Columbus, Ohio 43210, USA. <sup>169</sup> Department of Epidemiology and Biostatistics, Memorial Sloan Kettering Cancer Center, New York 10017, USA. <sup>170</sup> Human Genetics Group, Human Cancer Genetics Program, Spanish National Cancer Centre (CNIO), 28019 Madrid, Spain. <sup>171</sup> Biomedical Network on Rare Diseases (CIBERER), 28029 Madrid, Spain. <sup>172</sup> Department of Surgery, Seoul National University College of Medicine, Seoul, 03080 Korea. <sup>173</sup> Department of Obstetrics and Gynecology, Oregon Health and Science University, Portland, Oregon 97239, USA. <sup>174</sup> Knight Cancer Institute, Oregon Health and Science University, Portland, Oregon 97239, USA. <sup>175</sup> IFOM, The FIRC (Italian Foundation for Cancer Research) Institute of Molecular Oncology, 16 20139 Milan, Italy. <sup>176</sup> Department of Obstetrics and Gynecology, Comprehensive Cancer Center, Medical University of Vienna, 1090 Vienna, Austria. <sup>177</sup> Department of Cancer Epidemiology, Moffitt Cancer Center, Tampa, Florida 33606, USA. <sup>178</sup> Channing Division of Network Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts 02115, USA. <sup>179</sup> Department of Epidemiology, Harvard TH Chan School of Public Health, Boston, Massachusetts 02115, USA. <sup>180</sup> Laboratory of Cancer Genetics and Tumour Biology, Northern Finland Laboratory Centre NordLab, FI-90014 Oulu, Finland. <sup>181</sup> Laboratory of Cancer Genetics and Tumour Biology, Department of Clinical Chemistry and Biocenter Oulu, University of Oulu, FI-90014 Oulu, Finland. <sup>182</sup> Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Preventive and Predictive Medicine, Fondazione IRCCS (Istituto Di Ricovero e Cura a Carattere Scientifico) Istituto Nazionale dei Tumori (INT),

20133 Milan, Italy. <sup>183</sup> Department of Clinical Genetics, Karolinska University Hospital, 171 76 Stockholm, Sweden. <sup>184</sup> Department of Basic Sciences, Shaikat Khanum Memorial Cancer Hospital and Research Centre (SKMCH & RC), Lahore 54000, Pakistan. <sup>185</sup> Clalit National Israeli Cancer Control Center and Department of Community Medicine and Epidemiology, Carmel Medical Center and B. Rappaport Faculty of Medicine, Haifa, 34362, Israel. <sup>186</sup> Centre of Familial Breast and Ovarian Cancer, Department of Gynaecology and Obstetrics and Centre for Integrated Oncology (CIO), Center for Molecular Medicine Cologne (CMMC), University Hospital of Cologne, 50931 Cologne, Germany. <sup>187</sup> Department of Chronic Disease Epidemiology, Yale School of Public Health, New Haven, Connecticut 06510, USA. <sup>188</sup> Division of Gynecologic Oncology, NorthShore University HealthSystem, Evanston, Illinois 60201, USA. <sup>189</sup> Program in Epidemiology, Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington 98109, USA. <sup>190</sup> Department of Epidemiology, University of Washington, Seattle, Washington 98109, USA. <sup>191</sup> National Cancer Institute, Bangkok 10400, Thailand. <sup>192</sup> Research Oncology, Guy's Hospital, King's College London, London SE1 9RT, UK. <sup>193</sup> Department of Community and Family Medicine, Duke University Medical Center, Durham, North Carolina 27710, USA. <sup>194</sup> Cancer Control and Population Sciences, Duke Cancer Institute, Durham, North Carolina 27710, USA. <sup>195</sup> Netherlands Cancer Institute, Antoni van Leeuwenhoek Hospital, 1066 CX Amsterdam, The Netherlands. <sup>196</sup> Division of Molecular Gyneco-Oncology, Department of Gynaecology and Obstetrics, University Hospital of Cologne, 50676 Cologne, Germany. <sup>197</sup> Center for Integrated Oncology, University Hospital of Cologne, 50676 Cologne, Germany. <sup>198</sup> Center for Molecular Medicine, University Hospital of Cologne, 50676 Cologne, Germany. <sup>199</sup> Center of Familial Breast and Ovarian Cancer, University Hospital of Cologne, 50676 Cologne, Germany. <sup>200</sup> Taiwan Biobank, Institute of Biomedical Sciences, Academia Sinica, Taipei 115, Taiwan. <sup>201</sup> School of Public Health, China Medical University, Taichung 404, Taiwan. <sup>202</sup> Department of Health Research and Policy - Epidemiology, Stanford University School of Medicine, Stanford California 94305, USA. <sup>203</sup> Unité Mixte de Génétique Constitutionnelle des Cancers Fréquents, Hospices Civils de Lyon - Centre Léon Bérard, Lyon 69008, France. <sup>204</sup> INSERM U1052, CNRS UMR5286, Université Lyon 1, Centre de Recherche en Cancérologie de Lyon, Lyon 69003, France. <sup>205</sup> Department of Pathology, University of Melbourne, Parkville, Victoria 3010, Australia. <sup>206</sup> Division of Clinical Genetics, Department of Clinical and Experimental Medicine, Linköping University, 581 83 Linköping, Sweden. <sup>207</sup> Institut Curie, Department of Tumour Biology, Paris, France; Institut Curie, INSERM U830, 75248 Paris, France. <sup>208</sup> Université Paris Descartes, Sorbonne Paris Cité, 75270 Paris, France. <sup>209</sup> Institute of Human Genetics, Department of Human Genetics, University Hospital Heidelberg, 69120 Heidelberg, Germany. <sup>210</sup> Department of Genetics, Portuguese Oncology Institute, Porto 4200-072, Portugal. <sup>211</sup> Biomedical Sciences Institute (ICBAS), Porto University, Porto 4099-002, Portugal. <sup>212</sup> Department of Epidemiology, Mailman School of Public Health, Columbia University, New York 10027, USA. <sup>213</sup> Department of Clinical Genetics, Odense University Hospital, 5000 Odense C, Denmark. <sup>214</sup> UO Anatomia Patologica, Ospedale di Circolo-Università dell'Insubria, 21100 Varese, Italy. <sup>215</sup> Latvian Biomedical Research and Study Centre, Riga LV-1067, Latvia. <sup>216</sup> Immunology and Molecular Oncology Unit, Istituto Oncologico Veneto IOV - IRCCS (Istituto Di Ricovero e Cura a Carattere Scientifico), 64 - 35128 Padua, Italy. <sup>217</sup> Department of Molecular Virology, Immunology and Medical Genetics, The Ohio State University, Columbus, Ohio 43210, USA. <sup>218</sup> Wellcome Trust Centre for Human Genetics and Oxford Biomedical Research Centre, University of Oxford, Oxford OX3 7BN, UK. <sup>219</sup> Institute of Human Genetics, Pontificia Universidad Javeriana, Cra. 7 #40-62 Bogota, Colombia. <sup>220</sup> Department of Medical Oncology, Beth Israel Deaconess Medical Center, Boston, Massachusetts 02215, USA. <sup>221</sup> Department of Clinical Genetics, Erasmus University Medical Center, 3015 CE Rotterdam, The Netherlands. <sup>222</sup> Department of Gynecology, Family Cancer Clinic, Erasmus MC Cancer Institute, 3015 CE Rotterdam, The Netherlands. <sup>223</sup> Division of Gynecological Oncology, Department of Oncology, University Hospitals Leuven, B-3000 Leuven, Belgium. <sup>224</sup> University Hospital Ulm, 89069 Ulm, Germany. <sup>225</sup> Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda Maryland 20892, USA. <sup>226</sup> Multidisciplinary Breast Center, Department of General Medical Oncology, University Hospitals Leuven, B-3000 Leuven, Belgium. <sup>227</sup> Molecular Diagnostics Laboratory, IRRP, National Centre for Scientific Research 'Demokritos', Athens 153 10, Greece. <sup>228</sup> Cancer Research Initiatives Foundation, Sime Darby Medical Centre, 47500 Subang Jaya, Malaysia. <sup>229</sup> University Malaya Cancer Research Institute, Faculty of Medicine, University Malaya Medical Centre, University Malaya, 59100 Kuala Lumpur, Malaysia. <sup>230</sup> Department of Surgery, Tri-Service General Hospital, National Defense Medical Center, Taipei, 114 Taiwan. <sup>231</sup> Shanghai Center for Disease Control and Prevention, Shanghai, China. <sup>232</sup> Cancer Epidemiology Program, Division of Population Sciences, H. Lee Moffitt Cancer Center & Research Institute, Tampa, Florida 33612, USA. <sup>233</sup> Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota 55905, USA. \* These authors contributed equally to this work. \*\* These authors jointly supervised this work. † Present addresses: Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute, Cedars-Sinai Medical Center, Los Angeles, CA, USA (K.L.); The Center for Bioinformatics and Functional Genomics, Cedars-Sinai Medical Center, Los Angeles, CA, USA (S.A.G.). ‡ A full list of consortia members are listed below.

## Australian Ovarian Cancer Study Group

David Bowtell<sup>206,234,235,236,237</sup>, Anna deFazio<sup>238</sup> & Penny Webb<sup>3</sup>

<sup>234</sup> Peter MacCallum Cancer Centre, East Melbourne, Victoria 3002, Australia. <sup>235</sup> Sir Peter MacCallum Cancer Centre Department of Oncology, University of Melbourne, Parkville, Victoria 3052, Australia. <sup>236</sup> Ovarian Cancer Action Research Centre, Department of Surgery and Cancer, Imperial College London, London W12 0HS, UK. <sup>237</sup> Department of Biochemistry and Molecular Biology, University of Melbourne, Parkville, Victoria 3052, Australia. <sup>238</sup> Department of Gynaecological Oncology, Westmead Institute for Cancer Research, Westmead Hospital Westmead, New South Wales 2145, Australia.

## GEMO Study Collaborators

Marie-Agnès Collonge-Rame<sup>239</sup>, Alexandre Damette<sup>239</sup>, Emmanuelle Barouk-Simonet<sup>240</sup>, Françoise Bonnet<sup>240</sup>, Virginie Bubien<sup>240</sup>, Nicolas Sevenet<sup>240</sup>, Michel Longy<sup>240</sup>, Pascaline Berthet<sup>241</sup>, Dominique Vaur<sup>241</sup>, Laurent Castera<sup>241</sup>, Sandra Fert Ferrer<sup>242</sup>, Yves-Jean Bignon<sup>243</sup>, Nancy Uhrhammer<sup>243</sup>, Fanny Coron<sup>244</sup>, Laurence Faivre<sup>244</sup>, Amandine Baurand<sup>244</sup>, Caroline Jacquot<sup>244</sup>, Geoffrey Bertolone<sup>244</sup>, Sarab Lizard<sup>244</sup>, Dominique Leroux<sup>245</sup>, Hélène Dreyfus<sup>245</sup>, Christine Rebuschung<sup>245</sup>, Magalie Peysselon<sup>245</sup>, Jean-Philippe Peyrat<sup>246</sup>, Joëlle Fournier<sup>246</sup>, Françoise Révillion<sup>246</sup>, Claude Adenis<sup>246</sup>, Laurence Vénat-Bouvet<sup>247</sup>, Mélanie Léone<sup>248</sup>, Nadia Boutry-Kryza<sup>248</sup>, Alain Calender<sup>248</sup>, Sophie Giraud<sup>248</sup>, Carole Verny-Pierre<sup>249</sup>, Christine Lasset<sup>250</sup>, Valérie Bonadona<sup>250</sup>, Laure Barjhoux<sup>251</sup>, Hagay Sobol<sup>252</sup>, Violaine Bourdon<sup>252</sup>, Tetsuro Noguchi<sup>252</sup>, Audrey Remenieras<sup>252</sup>, Isabelle Coupier<sup>253</sup>, Pascal Pujol<sup>253</sup>, Johanna Sokolowska<sup>254</sup>, Myriam Bronner<sup>254</sup>, Capucine Delnatte<sup>255</sup>, Stéphane Bézieau<sup>255</sup>, Véronique Mari<sup>256</sup>, Marion Gauthier-Villars<sup>257</sup>, Bruno Buecher<sup>257</sup>, Etienne

Rouleau<sup>257</sup>, Lisa Golmard<sup>257</sup>, Virginie Moncoutier<sup>257</sup>, Muriel Belotti<sup>257</sup>, Antoine de Pauw<sup>257</sup>, Camille Elan<sup>257</sup>, Emmanuelle Fourme<sup>257</sup>, Anne-Marie Birot<sup>257</sup>, Claire Saule<sup>257</sup>, Maïté Laurent<sup>257</sup>, Claude Houdayer<sup>257,258</sup>, Fabienne Lesueur<sup>259</sup>, Noura Mebirouk<sup>259</sup>, Florence Coulet<sup>260</sup>, Chrystelle Colas<sup>260</sup>, Florent Soubrier<sup>260</sup>, Mathilde Warcoïn<sup>260</sup>, Fabienne Prieur<sup>261</sup>, Marine Lebrun<sup>261</sup>, Caroline Kientz<sup>261</sup>, Danièle Muller<sup>262</sup>, Jean-Pierre Fricker<sup>262</sup>, Christine Toulas<sup>263</sup>, Rosine Guimbaud<sup>263</sup>, Laurence Gladieff<sup>263</sup>, Viviane Feillel<sup>263</sup>, Isabelle Mortemousque<sup>264</sup>, Brigitte Bressac-de-Paillerets<sup>265</sup>, Olivier Caron<sup>265</sup> & Marine Guillaud-Bataille<sup>265</sup>

<sup>239</sup> Service de Génétique, CHU de Besançon, 25030 Besançon, France. <sup>240</sup> Oncogénétique, Institut Bergonié, 229 cours de l'Argonne, 33076 Bordeaux, France. <sup>241</sup> Centre François Baclesse, 3 avenue Général Harris, 14000 Caen, France. <sup>242</sup> Laboratoire de Génétique Chromosomique, Hôtel Dieu Centre Hospitalier, BP 1125 Chambéry, France. <sup>243</sup> Centre Jean Perrin, BP 392 Clermont-Ferrand cedex, France. <sup>244</sup> Centre de Lutte Contre le Cancer Georges François Leclerc, 1 rue Professeur Marion, BP 77 980 Dijon Cedex, France. <sup>245</sup> Département de Génétique, CHU de Grenoble, BP 217 Grenoble Cedex 9, France. <sup>246</sup> Centre Oscar Lambret, 3 rue Frédéric Combemale, 59020 Lille cedex BP307, France. <sup>247</sup> Department of Medical Oncology, CHU Dupuytren, 87042 Limoges, France. <sup>248</sup> Service de Génétique Moléculaire et Clinique, Hôpital Edouard Herriot, 5 place d'Arsonval, 69437 Lyon cedex 03, France. <sup>249</sup> Centre Léon Bérard, 28 rue Laënnec, 69437 Lyon, France. <sup>250</sup> Unité de Prévention et d'Epidémiologie Génétique, Centre Léon Bérard, 28 rue Laënnec, 69437 Lyon, France. <sup>251</sup> Biopathologie, Centre Léon Bérard, 28 rue Laënnec, 69437 Lyon, France. <sup>252</sup> Département Oncologie Génétique, Prévention et Dépistage, Institut Paoli-Calmettes, 232 boulevard Sainte-Marguerite, 13009 Marseille, France. <sup>253</sup> Unité d'Oncogénétique, CHU Arnaud de Villeneuve, 34295 Montpellier Cedex 5, France. <sup>254</sup> Laboratoire de génétique médicale, Nancy Université, Centre Hospitalier Régional et Universitaire, Rue du Morvan, 54511 cedex 1, Vandoeuvre-les-Nancy, France. <sup>255</sup> Service d'Oncogénétique, Centre René Gauducheau, Boulevard Jacques Monod, 44805 Nantes Saint Herblain Cedex, France. <sup>256</sup> Centre Antoine Lacassagne, 33 Avenue de Valombrose, 06100 Nice, France. <sup>257</sup> Service de Génétique, Institut Curie, 26, rue d'Ulm, 75248 Paris Cedex 05, France. <sup>258</sup> Inserm U830, Université INSERM U830, centre de recherche de l'Institut Curie, 75013 Paris, France. <sup>259</sup> Inserm U900, Institut Curie, Mines ParisTech, PSL University, 26 rue d'Ulm, 75248 Paris Cedex 05, France. <sup>260</sup> Département de Génétique, Groupe Hospitalier Pitié-Salpêtrière, 47-83 boulevard de l'Hôpital, 75013 Paris, France. <sup>261</sup> Service de Génétique Clinique Chromosomique et Moléculaire, Hôpital Nord, CHU Saint Etienne, St Etienne 42055 Cedex 2, France. <sup>262</sup> Unité d'Oncogénétique, Centre Paul Strauss, 3 rue de la Porte de l'Hôpital, BP30042, Strasbourg, France. <sup>263</sup> Oncogénétique, Institut Claudius Regaud, 1 avenue Irène Joliot-Curie, 31059 Toulouse cedex 9, France. <sup>264</sup> Hôpital Bretonneau - CHU de Tours, 2 boulevard Tonnelé, 37004 Tours cedex, France. <sup>265</sup> Service de Génétique, Institut Gustave Roussy, 39, rue Camille Desmoulins, 94805 Villejuif Cedex, France

## EMBRACE

Helen Gregory<sup>266</sup>, Zosia Miedzybrodzka<sup>266</sup>, Patrick J. Morrison<sup>267</sup>, Alan Donaldson<sup>268</sup>, Mark T. Rogers<sup>269</sup>, M. John Kennedy<sup>270,271</sup>, Mary E. Porteous<sup>272</sup>, Angela Brady<sup>273</sup>, Julian Barwell<sup>274</sup>, Claire Foo<sup>275</sup>, Fiona Lalloo<sup>276</sup>, Lucy E. Side<sup>277</sup>, Jacqueline Eason<sup>278</sup>, Alex Henderson<sup>279</sup>, Lisa Walker<sup>280</sup>, Jackie Cook<sup>281</sup>, Katie Snape<sup>282</sup>, Alex Murray<sup>283</sup> & Emma McCann<sup>284</sup>

<sup>266</sup> North of Scotland Regional Genetics Service, NHS Grampian & University of Aberdeen, Foresterhill, Aberdeen AB24 3AA, UK. <sup>267</sup> Northern Ireland Regional Genetics Centre, Belfast Health and Social Care Trust, and Department of Medical Genetics, Queens University Belfast, Belfast BT9 7BL, UK. <sup>268</sup> Clinical Genetics Department, St Michael's Hospital, Bristol BS2 8EG, UK. <sup>269</sup> All Wales Medical Genetics Services, University Hospital of Wales, Cardiff CF14 4XW, UK. <sup>270</sup> Academic Unit of Clinical and Molecular Oncology, Trinity College Dublin, Dublin 2, Ireland. <sup>271</sup> St James's Hospital, Dublin 8, Ireland. <sup>272</sup> South East of Scotland Regional Genetics Service, Western General Hospital, Edinburgh EH4 2XU, UK. <sup>273</sup> North West Thames Regional Genetics Service, Kennedy-Galton Centre, Harrow HA1 3UJ, UK. <sup>274</sup> Leicestershire Clinical Genetics Service, University Hospitals of Leicester NHS Trust, Leicester LE1 5WW, UK. <sup>275</sup> Department of Clinical Genetics, Alder Hey Hospital, Eaton Road, Liverpool L12 2AP, UK. <sup>276</sup> Genetic Medicine, Manchester Academic Health Sciences Centre, Central Manchester University Hospitals NHS Foundation Trust, Manchester M13 9WL, UK. <sup>277</sup> North East Thames Regional Genetics Service, Great Ormond Street Hospital for Children NHS Trust, London WC1N 3JH, UK. <sup>278</sup> Nottingham Clinical Genetics Service, Nottingham University Hospitals NHS Trust, Nottingham NG5 1PB, UK. <sup>279</sup> Institute of Genetic Medicine, Centre for Life, Newcastle Upon Tyne Hospitals NHS Trust, Newcastle upon Tyne NE7 7DN, UK. <sup>280</sup> Oxford Regional Genetics Service, Churchill Hospital, Oxford OX3 7LE, UK. <sup>281</sup> Sheffield Clinical Genetics Service, Sheffield Children's Hospital, Sheffield S10 2TH, UK. <sup>282</sup> South West Thames Regional Genetics Service, St. Georges Hospital, Cranmer Terrace, Tooting, London SW17 0RE, UK. <sup>283</sup> All Wales Medical Genetics Services, Singleton Hospital, Swansea SA2 8QA, UK. <sup>284</sup> All Wales Medical Genetics Service, Glan Clwyd Hospital, Rhyl LL18 5UJ, UK.

## The Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON)

M.A. Rookus<sup>285</sup>, F.E. van Leeuwen<sup>285</sup>, L.E. van der Kolk<sup>286</sup>, M.K. Schmidt<sup>287</sup>, N.S. Russell<sup>288</sup>, J.L. de Lange<sup>285</sup>, R. Wijnands<sup>285</sup>, J.M. Collée<sup>289</sup>, M.J. Hooning<sup>290</sup>, C. Seynaeve<sup>290</sup>, C.H.M. van Deurzen<sup>291</sup>, I.M. Obdeijn<sup>292</sup>, C.J. van Asperen<sup>293</sup>, R.A.E.M. Tollenaar<sup>294</sup>, T.C.T.E.F. van Cronenburg<sup>293</sup>, C.M. Kets<sup>295</sup>, M.G.E.M. Ausems<sup>296</sup>, C.C. van der Pol<sup>297</sup>, T.A.M. van Os<sup>298</sup>, Q. Waisfisz<sup>299</sup>, H.E.J. Meijers-Heijboer<sup>299</sup>, E.B. Gómez-García<sup>300</sup>, J.C. Oosterwijk<sup>301</sup>, M.J. Mourits<sup>302</sup>, G.H. de Bock<sup>303</sup>, H.F. Vasen<sup>304</sup>, S. Siesling<sup>305</sup>, J. Verloop<sup>305</sup> & L.I.H. Overbeek<sup>306</sup>

<sup>285</sup> Department of Epidemiology, Netherlands Cancer Institute, PO Box 90203, 1006 BE Amsterdam, The Netherlands. <sup>286</sup> Family Cancer Clinic, Netherlands Cancer Institute, PO Box 90203, 1006 BE Amsterdam, The Netherlands. <sup>287</sup> Division of Psychosocial Research and Epidemiology, Netherlands Cancer Institute, PO Box 90203, 1006 BE Amsterdam, The Netherlands. <sup>288</sup> Department of Radiotherapy, Netherlands Cancer Institute, PO Box 90203, 1006 BE Amsterdam, The Netherlands. <sup>289</sup> Department of Clinical Genetics, Family Cancer Clinic, Erasmus University Medical Center, PO Box 2040, 3000 CA Amsterdam, The Netherlands. <sup>290</sup> Department of Medical Oncology, Family Cancer Clinic, Erasmus MC Cancer Institute, PO Box 5201, 3008 AE Rotterdam, The Netherlands. <sup>291</sup> Department of Pathology, Family Cancer Clinic, Erasmus University Medical Center, PO Box 2040, 3000 CA Amsterdam, The Netherlands. <sup>292</sup> Department of Radiology, Family Cancer Clinic, Erasmus University Medical Center, PO Box 2040, 3000 CA Amsterdam, The Netherlands. <sup>293</sup> Department of Clinical Genetics, Leiden University Medical Center, PO Box 9600, 2300 RC Leiden, The Netherlands. <sup>294</sup> Department of Surgery, Leiden University Medical Center, PO Box 9600, 2300 RC Leiden, The Netherlands. <sup>295</sup> Department of Human Genetics, Radboud University Medical Center, PO Box 9101, 6500 HB Nijmegen, The Netherlands. <sup>296</sup> Department of Medical Genetics, University Medical Center Utrecht, PO Box 85090, 3508 AB Utrecht, The Netherlands. <sup>297</sup> Department of Oncological and Endocrine Surgery, University Medical Center Utrecht, PO Box 85090, 3508 AB Utrecht, The Netherlands. <sup>298</sup> Department of Clinical Genetics, Academic Medical Center, PO Box 22700, 1100 DE Amsterdam, The Netherlands. <sup>299</sup> Department of Clinical Genetics, VU University Medical Center, PO Box 7057, 1007 MB Amsterdam, The Netherlands. <sup>300</sup> Department of Clinical Genetics and GROW, School for Oncology and Developmental Biology, Maastricht University Medical Center, PO Box 5800, 6202 AZ Maastricht, The Netherlands. <sup>301</sup> Department of Genetics, University Medical Center Groningen, PO Box 30.001, 9700 RB Groningen, The Netherlands. <sup>302</sup> Department of Gynaecology, University Medical Center Groningen, PO Box 30.001, 9700 RB Groningen, The Netherlands. <sup>303</sup> Department of Epidemiology, University Medical Center Groningen, PO Box 30.001, 9700 RB Groningen, The Netherlands. <sup>304</sup> The Netherlands Foundation for Detection of Hereditary Tumours, University Medical Center, Poortgebouw Zuid, 2333 AA Leiden, The Netherlands. <sup>305</sup> The Netherlands Comprehensive Cancer Organization (IKNL), Location Amsterdam, IJsbanaanpad 9-11, 1076 CV Amsterdam, The Netherlands. <sup>306</sup> The nationwide network and registry of histo- and cytopathology in the Netherlands (PALGA), Randhoeve 225A, 3995 GA Houten, The Netherlands.

## KConFab

Stephen Fox<sup>307</sup>, Judy Kirk<sup>308</sup>, Geoff Lindeman<sup>309</sup> & Melanie Price<sup>310</sup>

<sup>307</sup> Pathology Department, Peter MacCallum Cancer Centre, Melbourne, Victoria 3002, Australia. <sup>308</sup> Familial Cancer Service, Department of Medicine, Westmead Hospital, Westmead, New South Wales 2145, Australia. <sup>309</sup> Breast Cancer Laboratory, Walter and Eliza Hall Institute, PO Royal Melbourne Hospital, Parkville, Victoria 3050, Australia. <sup>310</sup> Medical Psychology, University of Sydney, Sydney, New South Wales 2006 Australia.