

Dynamics of a Sex-Linked Deleterious Mutation in Populations Subject to Sex Reversal

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Abstract

The heterogametic sex chromosomes (i.e. mammalian *Y* and avian *W*) do not usually recombine with the homogametic sex chromosomes which is known to lead into rapid degeneration of *Y* and *W* due to accumulation of deleterious mutations. On the other hand, some 96% of amphibian species have homomorphic, i.e. non-degenerate *Y* chromosomes. Nicolas Perrin's fountain-of-youth hypothesis states that this is a result of recombination between *X* and *Y* chromosomes in sex-reversed individuals. In this study, I model the consequences of such recombination for the dynamics of a deleterious mutation occurring in *Y* chromosomes. As expected, even relatively low levels of sex reversal help to purge deleterious mutations. However, the population-dynamic consequences of this depend on the type of selection that operates on the population undergoing sex reversal. Under fecundity selection, sex reversal can be beneficial for some parameter values, whereas under survival selection, it seems to be always harmful.

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Introduction

Some recombination needs to occur between homologous chromosomes to enable proper pairing during meiosis. In heterogametic sex chromosomes (*Y* or *W* for male or female heterogamety, respectively), this recombination occurs only in so-called pseudoautosomal regions. Typically, the pseudoautosomal regions are relatively small, and as a result of this, sex chromosomes missegregate more frequently than autosomes [1]. It is thought that large non-recombining regions have evolved because of the need to maintain linkage between sex-determining loci and other loci which are subject to sexual-antagonistic selection [2]. Presumably as a result of this, the heterogametic sex chromosomes are often 'degenerate', i.e. they have fewer functional genes than the homogametic sex chromosomes [3,4,5].

Charlesworth and Charlesworth [3] provide an excellent review of how the *Y* chromosomes degenerate. To start with, deleterious mutations occurring in the non-recombining regions of *Y* chromosomes are always heterozygous, and selection against them is weaker, accordingly. Secondly, the effective population size of *Y* chromosomes is only one third of that of *X* chromosomes, and one fourth of that of autosomes, and thus drift effects are more important in case of *Y* chromosomes. The consequences of a weakened selection and a reduced effective population size are manifold. Charlesworth and Charlesworth [3] discuss four main effects: Firstly, Muller's ratchet [6] implies that deleterious mutations may reach fixation purely as a result of random drift, turning off one gene after another. In *Y* chromosomes, this fixation process is much faster than in freely recombining chromosomes. Secondly, mildly deleterious mutations may hitchhike into fixation when they are linked with beneficial mutations [7]. Thirdly, the

need to purify the genome of strongly deleterious mutations may drive mildly deleterious ones into fixation, if they are in negative linkage with the more deleterious ones [8]. Finally, Hill-Robertson effects [9] may cause that new, functional variants are unable to rejuvenate the *Y* chromosomes, because they usually occur in different haplotypes, which causes a diversifying selection pressure.

However, not all *Y* chromosomes are equally degenerate. For example, in amphibians, the vast majority of species have homomorphic sex chromosomes, so that the *Y* and *X* chromosomes are indistinguishable on basis of the karyotype [10]. It looks that there has to be some mechanism counter-acting the decay of these *Y* chromosomes [10,11,12]. One explanation could be a high turnover rate: new sex-determining loci may appear on autosomes so rapidly that the *Y* chromosomes do not have time to decay [13]. Another possibility is that *Y* chromosomes recombine more than has been previously thought. In humans, it is likely that duplications and *Y/Y* gene conversions help to prevent the decay of the *Y* chromosome, and this may also be the case of other mammalian species [14,15]. Even recombination between *Y* chromosomes and autosomes has been described [16].

Nicolas Perrin's fountain-of-youth hypothesis [10] states that in certain species, *X/Y* recombination help to maintain the *Y* chromosomes. This concerns recombination outside the pseudoautosomal regions which is thought to occur in sex-reversed females (i.e. *XY* females). This hypothesis rests on two cornerstones: Firstly, *XY* females need to be produced by some mechanism, which we will refer to as 'sex reversal'. Secondly, it has to be assumed that the recombination pattern depends on the phenotypic sex.

Concerning the plausibility of Perrin's hypothesis [10], it is known that sex-reversed individuals (i.e. *XY* females and *XX*

males) occur in considerable quantities in many species of fish, amphibians and reptiles [17,18,19,20,21,22]. In fact, a pure genetic sex determination system where XY individuals are bound to be males, is a special case in the animal kingdom, and no more ‘natural’ than an environmental sex determination system [23]. In addition, the rate of recombination, both in autosomes and sex chromosomes, typically depends on the sex of the individual. This pattern is known as ‘heterochiasmy’ [24]. Specifically, there is a growing body of evidence indicating that the recombination rate depends on the phenotypic (instead of the genotypic) sex [25,26,27,28,29]. Why this is the case, is at present unknown, but differences in the physiological process of male and female meiosis have been offered as an explanation [30]. Secondly, sex-specific differences in gene regulation may play a role [10].

In this study, I provide a mathematical demonstration of Perrin’s fountain-of-youth hypothesis. To this end, I develop analytical theory by writing a dynamical system for allele frequencies in sex chromosomes of populations that are subject to sex reversal. Secondly, I test the predictions of this theory by using individual-based simulations.

Methods

Modeling framework

I consider three loci in sex chromosomes, denoted by X/Y , K/k and $+/-$. Locus X/Y determines the genotypic sex which can be environmentally reversed. Locus K/k is subject to sexual-antagonistic selection such that K alleles (k alleles) are subject to a selection differential $0 \leq s \leq 0.5$ in phenotypic males (females), i.e. it is harmful to inherit alleles that are adapted for the other sex. This represents the fitness cost associated with sex reversal. Locus $+/-$ is subject to directional selection so that $-$ alleles are deleterious mutations having a selection differential $0 \leq s_- \leq 0.5$ in both sexes. Mutation occurs by probability μ per locus per generation. Recombination occurs between X/Y and either of the other two loci by probability r per meiosis. (Here it is assumed that K/k and $+/-$ segregate independently of each other on condition of X/Y , but see material S1.)

I assume that gene action is additive within loci, but the fitness consequences are multiplicative, so that the relative fitness of individual i is

$$w_i = (1 - s_{n_-i})(1 - \delta_{fi} s_{n_{Ki}})(1 - \delta_{mi} s_{n_{ki}}) \in [0, 1]. \tag{1}$$

In the individual-based simulations, this is the expected relative number of offspring. Above, n_-i , n_{Ki} and n_{ki} are the numbers of the respective alleles in the sex chromosomes of i , whereas δ_{fi} and δ_{mi} are the indicators of the phenotypic sexes (denoted by f and m).

Biologically, the proportions of phenotypic females in different genetic sexes (denoted by p_{XXf} , p_{XYf} and p_{YYf}) depend on each other [31]. Unless otherwise noted, it is assumed in the rest of this study that $\rho = (1 - p_{XXf}) = p_{XYf}$, i.e. sex reversal is equally likely for both genotypic sexes. According to analysis of Grossen et al. [31], it is likely that $p_{YYf} = 0$ in this case, which is assumed. This makes it possible to specify the amount of sex reversal as a scalar rate ρ . ($\rho = 0$ produces a pure genetic sex-determination system, and $\rho = 0.5$ produces a pure environmental sex-determination system. In many species, values of ρ differing significantly from zero are observed [20,23,32,33,34].)

Initially, all Y chromosomes are Y^{K-} haplotypes, and all X chromosomes are X^{k+} haplotypes, and the genetic sex ratio is even (i.e. 25% of all sex chromosomes are Y chromosomes). Locus K/k has been subject to sexual-antagonistic differentiation, whereas the

fixation of the deleterious mutation represents the degeneration of the Y chromosomes. In this study, I investigate how undergoing sex reversal helps a population to rejuvenate its Y chromosomes, i.e. to purge deleterious mutations. Except for Equation 1, this framework has been directly adopted from Perrin [10].

Dynamics of allele frequencies

I now consider how the allele frequencies evolve in time, given the framework stated above. To this end, I compose a system of difference equations. In this section, I give the equations, whereas discussion on their coefficient matrices is included in the material S1, and the matrices themselves are given in material S3. The key idea is that it is possible to model this biological system by using matrix algebra, because there are a finite number of haplotypes in the modeling framework. However, selection operates on the ‘phenotype’ of the individual which in this case comprises the phenotypic sex (two options, m and f), in addition to the diploid haplotype (64 options, e.g. $X^{k+} Y^{K-}$). Therefore, it is necessary to consider the distribution of these phenotypes in each generation t , denoted by p_t . Two of the phenotypes are always biologically equivalent (e.g. $mX^{k+} Y^{K-}$ and $mY^{K-} X^{k+}$), but an artificial distinction has to be made between them to model the union of gametes at a later stage (Equation 2). Out of p_t , the frequencies of all alleles and the sex ratio can be calculated.

First, selection and recombination change the frequencies of the phenotypes by matrix multiplication:

$$p_S = W p_t,$$

$$p_R = \Gamma p_S.$$

Matrix W has values corresponding to selective pressure, and matrix Γ has values corresponding to probability of recombination. Then, gametes are produced. For clarity, I consider the allele frequencies of eggs and sperm separately.

$$p_f = X_f p_R \quad \text{for eggs, and}$$

$$p_m = X_m p_R \quad \text{for sperm.}$$

Mutation operates on p_f and p_m . Matrix Λ has values corresponding to probability of mutation.

$$p_{fM} = \Lambda p_f$$

$$p_{mM} = \Lambda p_m.$$

Then, union of gametes occurs. Assuming random mating, the distribution of diploid haplotypes is given by the matrix

$$P = p_{fM}' p_{mM} \tag{2}$$

Note that Equation 2 implies non-linearity. If this was not the case, it would be a trivial task to solve the equilibrium allele frequencies

Table 1. Summary of model parameters.

Parameter	Baseline value	Lower value	Upper value	Comments
Per locus rate of mutation μ	10^{-4}	10^{-5}	10^{-3}	Applies on loci $+/-$ and K/k
Probability of recombination r	0.25	0.10	0.50	Upper value implies no molecular linkage
Rate of sexual-antagonistic selection s	0.05	0.02	0.10	Lower value implies no fitness cost for sex reversal
Rate of directional selection s_{-}	0.05	0.02	0.10	Lower value implies a neutral mutation
Initial population size n_0	1,000	200	5,000	Applies only on finite populations
Optimal growth rate r_0	1.10	1.00	1.20	Maximal expected growth rate; used to scale the fitness in finite populations

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explicitly. The frequencies of the phenotypes on generation $t+1$ are obtained by sexing the diploid haplotypes and renormalizing the system:

$$p_{t+1} = \frac{X_{mf}c(P)}{\sum_{i=1}^{128} p_{ti}} \quad (3)$$

As a technical convenience, I use a mapping c that transforms P into a vector, column by column. In the material S2, there is an R code to calculate p_t given an arbitrary initial state p_0 .

Individual-based simulations

The above system of difference equations is essentially a model of an infinite population. Individual-based simulations are needed for three things: to test the predictions of this theory, to investigate drift effects in the allele frequencies of small populations, and to study the population-dynamic consequences of sex reversal. To this end, I consider organisms with discrete, non-overlapping generations. The life history of these organisms is simulated by the following algorithm:

1. In each generation, the phenotypic sex of each individual is randomized. Depending on the respective genotype, an

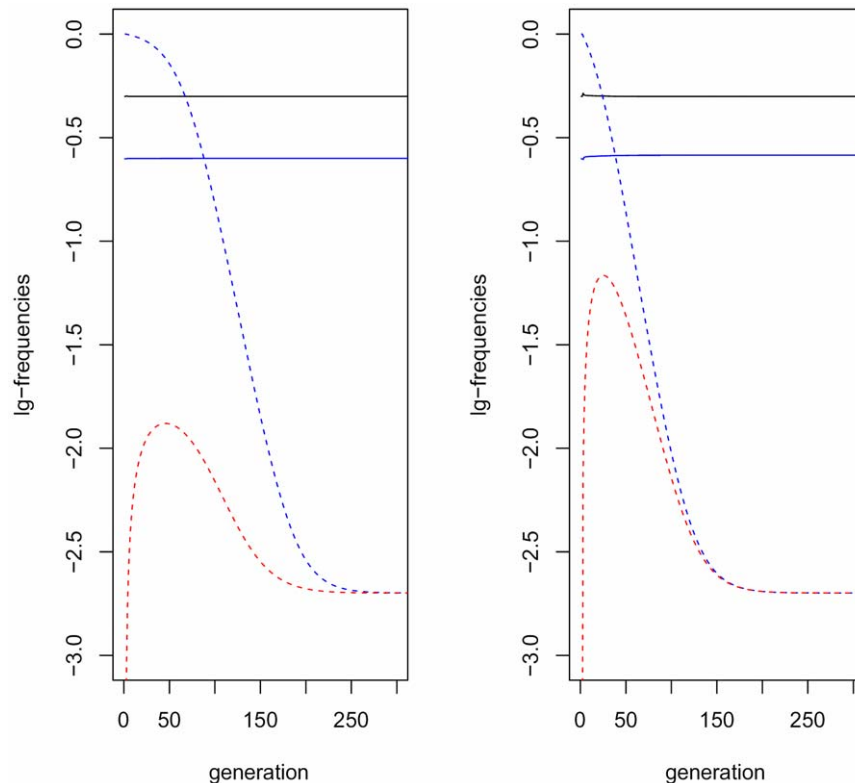


Figure 1. Dynamics of the deleterious mutation. In this figure, dynamics of the deleterious mutation is illustrated by two examples. In both panels, the black lines represent the proportion of phenotypic females, the blue lines the proportion of Y chromosomes out of all sex chromosomes, the dashed blue lines prevalence of the deleterious mutation in Y chromosomes, and the dashed red lines prevalence of the deleterious mutation in X chromosomes. (Note the \log_{10} scale of the y axis.) The rates of sex reversal are 0.01 (panel A) and 0.10 (panel B). The rest of the parameters are as in 'Baseline', Table 1.

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- individual is a phenotypic female by probability p_{XXf} , p_{XYf} or p_{YYf} , i.e. sex reversal happens at this stage.
2. The relative fitness of each individual is calculated by using Equation 1.
 3. Natural selection and breeding occur. Here, I consider two options.
 - a. Survival selection: individual i dies with probability $1 - w_i$ before producing offspring, but each surviving female gets Poisson($2r_0$) offspring. A random mate is assigned for each female from the surviving males. In this case, the expected number of offspring for individual i is $2r_0w_i$.
 - b. Fecundity selection: the total number of offspring produced depends on all w_i values, but no individual dies before getting chance to reproduce. Namely, each phenotypic female i is first assigned a random mate j among the phenotypic males. They get Poisson($2r_0w_iw_j$) offspring, depending on their relative fitness values. In this case, the expected number of offspring for individual i is $2r_0w_i\bar{w}_j$ (\bar{w}_j denoting the average of w_j over the other sex), which is proportional to $2r_0w_i$.
 4. Gametes are produced. In phenotypic females, recombination occurs by probability r between each pair of loci. Mutations occur by probability μ per locus per generation in both sexes. Each offspring gets one random gamete from the sire, and one random gamete from the dam. I assume that K and k alleles may mutate into one other, but $-$ alleles cannot change into $+$ alleles.

5. The reproducing individuals are removed, and the procedure is carried out for a new generation. If population exceeds a carrying capacity $K = 2n_0$ after reproduction, individuals are deleted by random. This is merely a technical convenience to avoid simulating very large populations; natural selection occurs on step 3.

The R codes used for these simulations are given in the online material S2.

Parameter values and sensitivity analysis

Table 1 lists the relevant parameters. Their values are more or less arbitrary, as they are only meant to illustrate the range of factors that affect the dynamics of the $+/-$ locus. A sensitivity analysis is performed by giving each parameter higher and lower values. For ρ , a number of values from 0 to 0.50 are tried in each scenario. Apart from changing the parameter values, a few structural assumptions are tested, namely: the effect of the carrying capacity K , symmetric sex reversal (i.e. assumption that $p_{XXf} = 1 - p_{XYf}$), and the pattern of recombination (i.e. that segregation of $+/-$ and K/k is independent on condition of X/Y). The results of sensitivity analysis are discussed in detail in material S1 and the accompanying Figures S1, S2, S3, S4, S5. Shortly, as is to be expected, selective pressures s and s_- determine the advantage related to sex reversal (see below, 'Individual-based simulations'). Secondly, the advantage related to sex reversal is lesser, if the $+/-$ locus is in a closer linkage with the sex-determining locus, yet it is still observable.

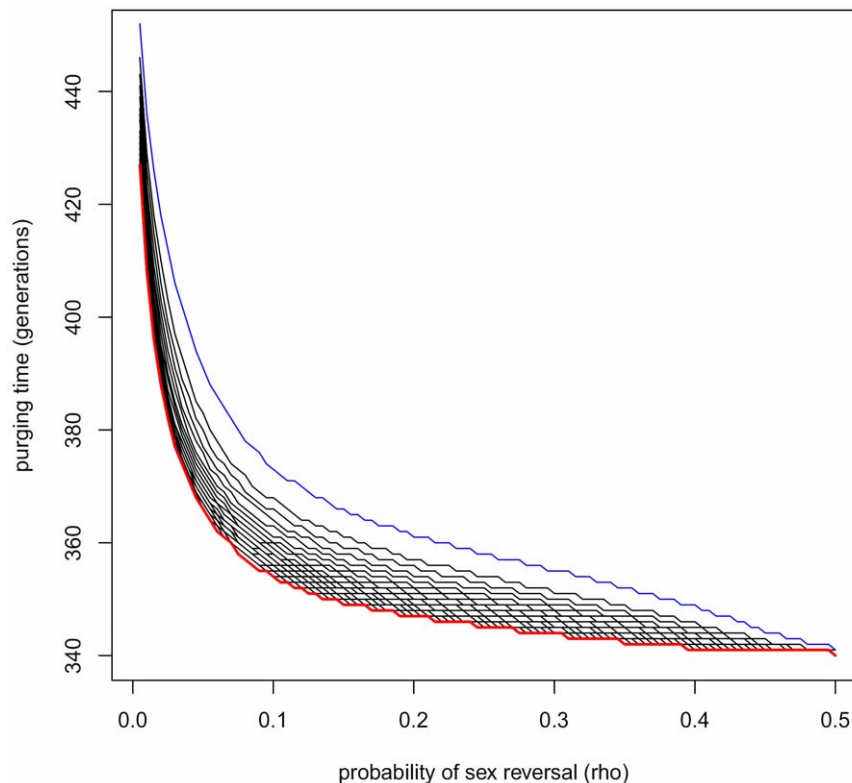


Figure 2. Purging time. This figure illustrates the joint effect of the probabilities of sex reversal (ρ) and recombination (r). The probability of sex reversal can be read from the x axis, whereas each curve corresponds to a specific probability of recombination from 0.1 (the blue line) to 0.5 (the red line) with even spaces. Other parameter values are as in the baseline scenario of Table 1. On the y axis, the 'purging time' is the number of generations that is required for the allele frequencies to reach the mutation-selection balance by 10^{-8} . doi:10.1371/journal.pone.0025362.g002

Results

Dynamics of allele frequencies

The system of equations laid out above is nonlinear, because the union of gametes (Eq. 2) follows a mass-action principle. Because of this, it is possible that multiple equilibria exist. I investigated that possibility by initializing the system with random initial values (i.e. initial frequencies p_0), but only one equilibrium was observed for each set of parameter values, having properties discussed below.

Initially, sex reversal creates a burst of polymorphism, rapidly generating new phenotypes (such as $fX^{k+}Y^{k-}$) and haplotypes (such as X^{k-}). Eventually, frequencies of some phenotypes (i.e. those with the deleterious mutation) decline, as natural selection wipes them out while the system slowly approaches the equilibrium. However, all phenotypes are maintained in populations of sufficient size, because mutation and recombination always produce them.

Figure 1 illustrates the dynamics of a few key statistics, namely the sex ratio, proportion of Y chromosomes, and prevalence of the deleterious mutation in X and Y chromosomes. As can be seen from Figure 1, the deleterious mutation is purged so that the prevalence in X chromosomes first catches up with that of Y chromosomes, and then both prevalences start to approach their equilibrium value. The end result of this process is the usual mutation-selection balance, i.e. roughly m/s_+ , and it does not depend on ρ . After an initial disturbance (barely visible in Fig. 1B),

proportion of Y chromosomes and the sex ratio remain constant. This was to be expected on basis of earlier research: using difference equations, Grossen et al. [18] have shown that this is the equilibrium state for $\rho < 0.5$, which they interpret as sex-ratio selection. However, this concerns the sex ratio after birth, given by p_t , not that of adult life stage which is given by p_S ; as long as there are more — in Y chromosomes, there are less breeding males than females in p_S .

Higher values of ρ always imply that the mutation can be purged more rapidly. Using the parameter values in Table 1 and $\rho = 0.01 - 0.50$, it takes 167 to 452 generations for the deleterious mutation to reach the mutation-selection balance by 10^{-8} accuracy, except of the case of neutral mutation ($s_+ = 0$) where more than 3,000 generations are required to break the linkage between — and Y . Most of the advantage is produced by relatively low values of ρ (see Fig. 2). As a rule of thumb, it could be said that $\rho \approx 0.1$ purges mutations almost as effectively as a pure environmental sex-determination system ($\rho = 0.5$).

Finally, linkage disequilibrium is a necessary condition for the fountain-of-youth hypothesis to work in this modeling framework. If the system is initialized at Hardy-Weinberg equilibrium, or by even phenotypic frequencies, sex reversal does not affect the dynamics of $+/-$ locus in any way. Because no epistasis has been assumed, recombination only helps to break the linkage disequilibrium between — and Y . If this disequilibrium does not exist initially, recombination is not relevant, and hence sex reversal is not relevant.

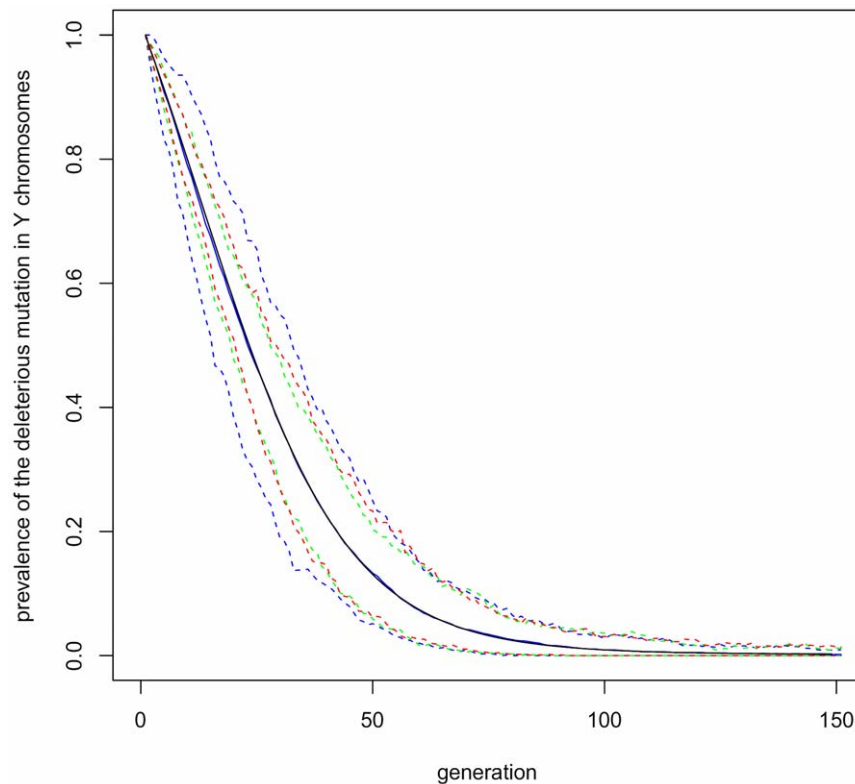


Figure 3. Drift effects under survival selection. Drift effects are studied in populations undergoing survival selection (Option a. in the main matter), using prevalence of the deleterious mutation in Y chromosomes as a summary statistic. The black line represents the theoretical value of this statistic, calculated by applying Equation 3 iteratively. The colored lines illustrate the distribution of this statistic in small populations ($n_0 = 200$ blue, $n_0 = 1,000$ green, and $n_0 = 5,000$ red). The dashed lines indicate the 0.05 and 0.95 quantiles of these distributions. The solid blue line indicates the sample mean for $n_0 = 200$. (For $n_0 = 1,000$ and $n_0 = 5,000$, the sample mean has been omitted, because it is so near to the expectation.) For each n_0 , 150 Monte Carlo replicates have been generated. The rate of sex reversal is $\rho = 0.1$, and the other parameter values are as in 'Baseline', Table 1. doi:10.1371/journal.pone.0025362.g003

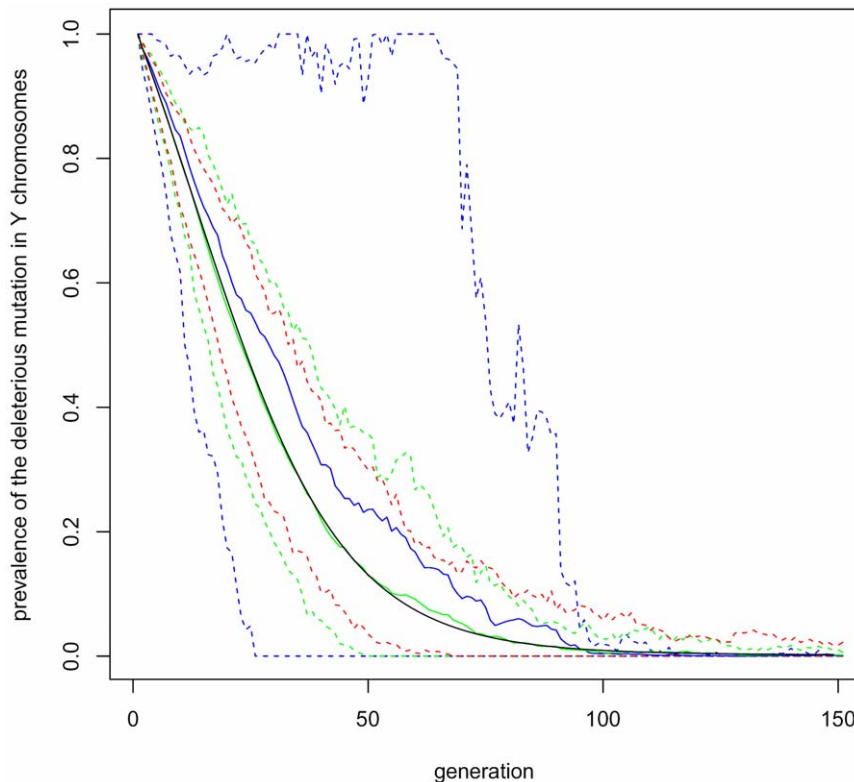


Figure 4. Drift effects under fecundity selection. This figure illustrates drift effects in populations undergoing fecundity selection (Option b. in the main matter), presenting the prevalence of the deleterious mutation in *Y* chromosomes. The black line represents the theoretical value of this prevalence, calculated by applying Equation 3 iteratively. The colored lines illustrate the distribution of this statistic in small populations ($n_0 = 200$ blue, $n_0 = 1,000$ green, and $n_0 = 5,000$ red). The dashed lines indicate the 0.05 and 0.95 quantiles of these distributions. The solid colored lines indicate the sample mean for $n_0 = 200$ and $n_0 = 1,000$. (This has been omitted for red, i.e. $n_0 = 5,000$, because it is so near to the expectation.) For each n_0 , 150 Monte Carlo replicates have been generated. The rate of sex reversal is $\rho = 0.1$, and the other parameter values are as in 'Baseline', Table 1. doi:10.1371/journal.pone.0025362.g004

Individual-based simulations

The drift effects are illustrated by Figures 3 and 4, using the prevalence of the deleterious mutation in the *Y* chromosomes as a summary statistic. Regardless of the drift effects, even populations of the modest initial size $n_0 = 200$ behave in great consistency with the theory developed above in 'Dynamics of allele frequencies'. However, the drift effects are more visible for any initial population size if fecundity selection is assumed. Under survival selection, the fate of individual's genetic material depends on its own fitness, only. Under fecundity selection, the expected number of offspring depends also on the fitness of a random mate. This creates a hitch-hike type opportunity for the deleterious mutations. In some simulation runs, it was observed that the random drift drove — back to fixation in the *Y* chromosomes under fecundity selection, which was never observed under survival selection.

Under fecundity selection, sex reversal has a non-uniform effect on the expected population size. This is illustrated by Figure 5. If the — allele is neutral ($s_- = 0$), sex reversal always decreases the expected population size, because it only breaks down the co-adaptation of the *X/Y* and *K/k* loci. For $s_- > 0$, there is a trade-off between this effect and the need to purify deleterious mutations. This is shown in Figure 4B by using 'Baseline' parameter values. In this case, an intermediate value of ρ is optimal, whereas pure genetic ($\rho = 0$) and pure environmental ($\rho = 0.5$) sex-determination systems decrease the expected population size. However, it should be added that due to demographic stochasticity, even populations with a nearly optimal ρ will sometimes drift into extinction.

It should be stressed that the advantage related to sex reversal concerns only fecundity selection (Option b.) and a suitable range of parameter values. I was unable to find parameter values for which sex reversal would have a significant positive effect on the expected population size under survival selection (Option a.). This is understandable by recalling the mechanism of survival selection: in Option a., the number of offspring depends only on the number of surviving females, as long as there is at least one male to breed with. In this case, it is presumably optimal to keep the — and *K* alleles out of the *X* chromosomes, even if it means that the deleterious mutation cannot be purged.

Discussion

I have used Perrin's framework [10] to study his fountain-of-youth hypothesis. According to theoretical considerations, environmental sex reversal helps to purge deleterious mutations out of the *Y* chromosomes. This was also observed in individual-based simulations for a wide range of parameters. The fitness consequences of this effect depend on the type of natural selection used in the simulation model: For populations undergoing pure survival selection, sex reversal seems to be always harmful, but for populations undergoing fecundity selection, intermediate rates of sex reversal could be optimal. Doubtless, other selection mechanisms than these two could be implemented, but the key seems to be that in survival selection, the number of males is not important, whereas in fecundity selection, male and female fitness

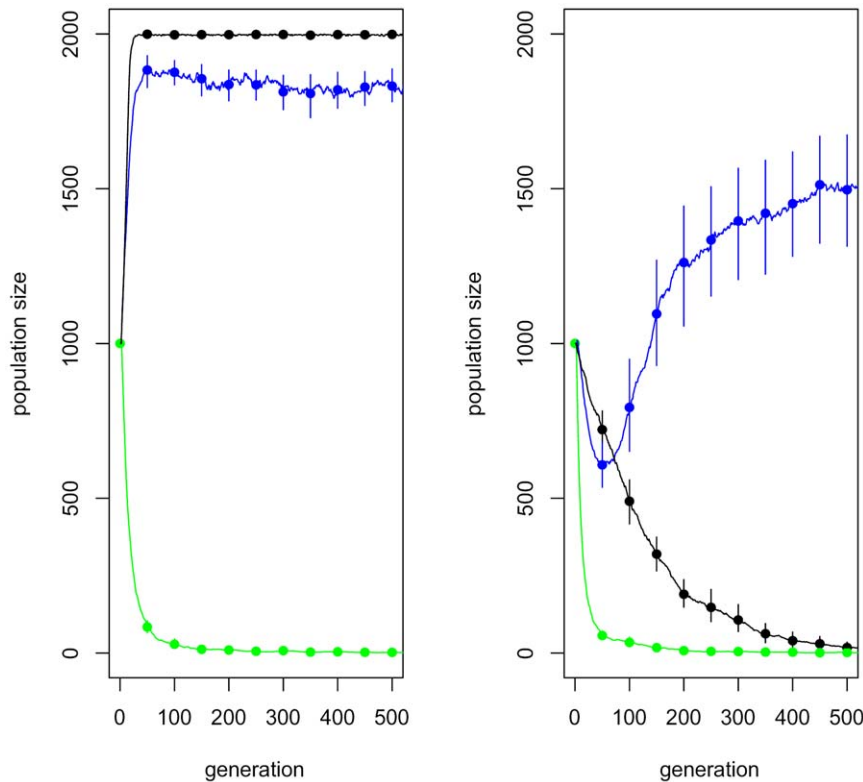


Figure 5. How sex reversal affects the population size. In panel A, the $-$ allele is neutral. In panel B, the $-$ allele is mildly deleterious ($s_- = 0.05$), with other parameters as in ‘Baseline’, Table 1. Fecundity selection (Option b.) is assumed. The dots are estimates of the expected population size, whereas the error bars represent their uncertainty due to sampling error (95% confidence intervals). The black dots are calculated for a population with no sex reversal ($\rho = 0$), the blue dots for a moderate rate of sex reversal ($\rho = 0.1$), and the green dots for a pure environmental sex-determination system ($\rho = 0.5$). A sample of 150 populations has been used in each case. doi:10.1371/journal.pone.0025362.g005

are equally important. In short, it is only desirable to rejuvenate the Y chromosomes, if the males matter.

The concept of sex reversal deserves to be discussed in some length. As far as this study is concerned, it comprises both environmental sex determination and sex changes before reproductive age. Bull [23] distinguishes between these two, so that in environmental sex determination, sex is permanently fixed as a result of environmental conditions, e.g. temperature, during ontogeny. Ah-King and Nylin [21] have proposed that phenotypic sex could be seen as a reaction norm. In that context, sex reversal can occur in all species, but some species are extremely insensitive to environmental conditions (genetic sex-determination system), whereas some other species are infinitely sensitive to them (environmental sex-determination system). Pure environmental and pure genetic sex-determination systems can be seen as opposite ends of a spectrum, with many species falling somewhere in between [21,31,35]. The intermediate species could still be said to have sex chromosomes, whereas pure environmental sex determination implies that no such chromosomes exist. Such is the case of many crocodylian species [22].

From the biological side, viability of the population does not imply that a trait such as sex reversal is an evolutionary advantage for the individual. This relates to the discussion about group selection [36]. In our context, turning sex reversal on in an individual-based simulation does not imply that it would evolve in nature out of the need to rejuvenate Y chromosomes. The next step would be to endogenize the evolution of the sex-determination system. This could be done by including a fourth locus in the

present framework, one that determines liability for sex reversal, and studying its allele frequencies. Perrin also hints at this [10].

Finally, this study is insufficient to judge the truth of Perrin’s hypothesis. It is almost self-evident that sex reversal helps to purify deleterious mutations out of Y chromosomes, if recombination of sex chromosomes occurs only in phenotypic females. I have only verified the quantitative consequences of this notion. The evolutionary consequences are more complicated, as they depend on the type of selection that the species undergoes. Judging the importance of fountain-of-youth hypothesis (or more appropriately, ‘fountain-of-youth effects’), calls for studying sex reversals and recombination rates, but also the selective pressures acting in the wild.

Supporting Information

Material S1 Further details of the model. In this section, further details of the model are given. Firstly, the coefficient matrices of ‘Dynamics of allele frequencies’ are discussed. Secondly, results of the sensitivity analysis are discussed. (DOC)

Material S2 The R programs. In this .zip file, the R programs used for this study are given. Further information is available in a readme file within the .zip file. (ZIP)

Material S3 The coefficient matrices. In this Excel file, the coefficient matrices from ‘Dynamics of allele frequencies’ are given in their full extent, in a symbolic form. (XLS)

Figure S1 The effect of uneven sex reversal. The black lines represent the proportion of phenotypic females, the blue lines the proportion of *Y* chromosomes out of all sex chromosomes, the dashed blue lines prevalence of the deleterious mutation in *Y* chromosomes, and the dashed red lines prevalence of the deleterious mutation in *X* chromosomes. (Note the \log_{10} scale of the y axis.) In panel A, it is assumed that $p_{XXf} = 1$, $p_{XYf} = 0.1$, $p_{YYf} = 0.05$, i.e. the female phenotype is strongly favored so that even some of the *YY* individuals are female. In panel B, the male phenotype is favored so that $p_{XXf} = 0.7$, $p_{XYf} = 0.1$, $p_{YYf} = 0$. (TIF)

Figure S2 Less favorable sex reversal under fecundity selection. This figure illustrates the sensitivity of the results towards the rates of directional selection (s_{-}) and recombination (r). In panel A, $s_{-} = 0.02$. In panel B, $r = 0.10$. The other parameters are as in 'Baseline', Table 1. Fecundity selection has been assumed. The black data are for population with no sex reversal, the green data for populations with environmental sex determination, and the blue data for $\rho = 0.10$. The dots and the continuous lines represent the sample mean, and the error bars represent 95% confidence intervals. Both of these figures should look like Figure 5B, if the results were not sensitive to (s_{-}, r). (TIF)

Figure S3 The effect of recombination pattern. In Figure 1, it is implicitly assumed that the chromosomal organization is $(+/-, X/Y, K/k)$, i.e. that loci $+/-$ and K/k are conditionally independent. In this figure, that assumption is relaxed by modifying the recombination matrix Γ . In panel A, the chromosomal organization is $(X/Y, +/-, K/k)$, and in panel B, it is $(X/Y, K/k, +/-)$. In both panels, the black lines represent the proportion of phenotypic females, the blue lines the proportion of *Y* chromosomes out of all sex chromosomes, the dashed blue

lines prevalence of the deleterious mutation in *Y* chromosomes, and the dashed red lines prevalence of the deleterious mutation in *X* chromosomes. (TIF)

Figure S4 The recombination pattern matters for phenotypes. In this figure, the effect of chromosomal organization is illustrated by investigating the dynamics of phenotypic frequencies. In panel A, the chromosomal organization is $(X/Y, +/-, K/k)$, and in panel B, it is $(X/Y, K/k, +/-)$. The blue lines are the frequencies of the 64 male phenotypes, and the red lines are the frequencies of the 64 female phenotypes. The parameter values are as in the 'Baseline' scenario, Table 1. (TIF)

Figure S5 The recombination pattern and population dynamics. This figure illustrates the sensitivity of the population-dynamic consequences of sex reversal towards the chromosomal organization. In panel A, the chromosomal organization is $(X/Y, +/-, K/k)$, and in panel B, it is $(X/Y, K/k, +/-)$. Fecundity selection has been assumed, and the parameter values are as in 'Baseline', Table 1. If the chromosomal organization did not matter, both of these figures should look like Figure 5B. (TIF)

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Author Contributions

Conceived and designed the experiments: MK. Performed the experiments: MK. Analyzed the data: MK. Contributed reagents/materials/analysis tools: MK. Wrote the paper: MK.

References

- Keeney S, Kauppi L, Barchi M, Baudat F, Romanienko PJ, et al. (2011) Distinct Properties of the XY Pseudoautosomal Region Crucial for Male Meiosis. *Science* 331: 916–920.
- Rice WR (1996) Evolution of the Y sex chromosome in animals. *Bioscience* 46: 331–343.
- Charlesworth B, Charlesworth D (2000) The degeneration of Y chromosomes. *Philos Trans R Soc Lond B Biol Sci* 355: 1563–1572.
- Steinemann M, Steinemann S, Lottspeich F (1993) How Y chromosomes become genetically inert. *Proc Natl Acad Sci U S A* 90: 5737–5741.
- Steinemann S, Steinemann M (2005) Y chromosomes: born to be destroyed. *Bioessays* 27: 1076–1083.
- Muller HJ (1964) The Relation of Recombination to Mutational Advance. *Mutation Research* 1: 2–9.
- Rice WR (1987) Genetic hitchhiking and the evolution of reduced genetic activity of the Y sex chromosome. *Genetics* 116: 161–167.
- Charlesworth B (1994) The Effect of Background Selection against Deleterious Mutations on Weakly Selected, Linked Variants. *Genetical Research* 63: 213–227.
- Hill WG, Robertson A (1966) Effect of Linkage on Limits to Artificial Selection. *Genetical Research* 8: 269–294.
- Perrin N (2009) Sex Reversal: A Fountain of Youth for Sex Chromosomes? *Evolution* 63: 3043–3049.
- Schartl M (2004) Sex chromosome evolution in non-mammalian vertebrates. *Current Opinion in Genetics & Development* 14: 634–641.
- Bachtrog D (2006) A dynamic view of sex chromosome evolution. *Current Opinion in Genetics & Development* 16: 578–585.
- Engelstadter J (2008) Muller's ratchet and the degeneration of Y chromosomes: a simulation study. *Genetics* 180: 957–967.
- Marais GAB, Campos PRA, Gordo I (2010) Can Intra-Y Gene Conversion Oppose the Degeneration of the Human Y Chromosome? A Simulation Study. *Genome Biology and Evolution* 2: 347–357.
- Charlesworth B (2003) The organization and evolution of the human Y chromosome. *Genome Biology* 4: -.
- Switonski M, Szczerbal I, Krumrych W, Nowacka-Wosuzuk J (2010) Y-autosome reciprocal translocation in a bull. *Chromosome Research* 18: 720–721.
- Wedekind C (2010) Searching for sex-reversals to explain population demography and the evolution of sex chromosomes. *Molecular Ecology* 19: 1760–1762.
- Alho JS, Matsuba C, Merila J (2010) Sex reversal and primary sex ratios in the common frog (*Rana temporaria*). *Molecular Ecology* 19: 1763–1773.
- Stelkens RB, Wedekind C (2010) Environmental sex reversal, Trojan sex genes, and sex ratio adjustment: conditions and population consequences. *Molecular Ecology* 19: 627–646.
- Ospina-Alvarez N, Piferrer F (2008) Temperature-Dependent Sex Determination in Fish Revisited: Prevalence, a Single Sex Ratio Response Pattern, and Possible Effects of Climate Change. *Plos One* 3: -.
- Ah-King M, Nylin S (2010) Sex in an Evolutionary Perspective: Just Another Reaction Norm. *Evolutionary Biology* 37: 234–246.
- Janzen FJ, Phillips PC (2006) Exploring the evolution of environmental sex determination, especially in reptiles. *Journal of Evolutionary Biology* 19: 1775–1784.
- Bull JJ (1983) Evolution of sex determining mechanisms. Menlo Park, USA: The Benjamin/Cummings Publishing Company.
- Burt A, Bell G, Harvey PH (1991) Sex-Differences in Recombination. *Journal of Evolutionary Biology* 4: 259–277.
- Lynn A, Schrupp S, Cherry J, Hassold T, Hunt P (2005) Sex, not genotype, determines recombination levels in mice. *Am J Hum Genet* 77: 670–675.
- Matsuba C, Alho JS, Merila J (2010) Recombination Rate between Sex Chromosomes Depends on Phenotypic Sex in the Common Frog. *Evolution* 64: 3634–3637.
- Kondo M, Nagao E, Mitani H, Shima A (2001) Differences in recombination frequencies during female and male meioses of the sex chromosomes of the medaka, *Oryzias latipes*. *Genetical Research* 78: 23–30.
- Wallace H, Wallace BMN, Badawy GMI (1997) Lampbrush chromosomes and chiasmata of sex-reversed crested newts. *Chromosoma* 106: 526–533.
- Merila J, Matsuba C (2009) Isolation and characterization of 145 polymorphic microsatellite loci for the common frog (*Rana temporaria*). *Molecular Ecology Resources* 9: 555–562.
- Tease C, Hulten MA (2004) Inter-sex variation in synaptonemal complex lengths largely determine the different recombination rates in male and female germ cells. *Cytogenetic and Genome Research* 107: 208–215.
- Grossen C, Neuenschwander S, Perrin N (2011) Temperature-Dependent Turnovers in Sex-Determination Mechanisms: A Quantitative Model. *Evolution* 65: 64–78.

32. Matsuba C, Miura I, Merila J (2008) Disentangling genetic vs. environmental causes of sex determination in the common frog, *Rana temporaria*. *BMC Genetics* 9: -.
33. Shine R, Elpick MJ, Donnellan S (2002) Co-occurrence of multiple, supposedly incompatible models of sex determination in a lizard population. *Ecological Letters* 5: 486–489.
34. Aida T (1936) On the inheritance of color in a fresh-water fish *Aplocheilus latipes* Temmick and Schlegel, with special reference to the sex-linked inheritance. *Genetics* 6: 554–573.
35. Sarre SD, Georges A, Quinn A (2004) The ends of a continuum: genetic and temperature-dependent sex determination in reptiles. *Bioessays* 26: 639–645.
36. West SA, Griffin AS, Gardner A (2007) Social semantics: altruism, cooperation, mutualism, strong reciprocity and group selection. *Journal of Evolutionary Biology* 20: 415–432.