

1 **Biodegradation of animal fats in a co-composting process with wastewater sludge**

2
3
4 Luz Ruggieri, Adriana Artola, Teresa Gea*, Antoni Sánchez

5
6
7
8 Composting Research Group
9 Department of Chemical Engineering
10 Escola Tècnica Superior d'Enginyeria
11 Universitat Autònoma de Barcelona
12 Bellaterra (Cerdanyola, 08193-Barcelona, Spain)

13
14
15 * Corresponding author:
16 Dr. Teresa Gea
17 Phone: 34-935811879
18 Fax: 34-935812013
19 Email: teresa.gea@uab.cat

20
21
22
23
24
25 Scientific relevance:

26
27 This paper includes a study about the biodegradation of fats in high proportion (40-50%) by
28 means of a composting process where wastewater sludge is used as co-substrate. Several
29 aspects of the composting process and their relation with biodegradation of fats are analysed
30 in detail, including composting routine parameters (temperature, moisture content and
31 porosity) and biochemical parameters related to fats biodegradation, such as respiration
32 activity and lipolytic activity exhibited by microbial populations. The effects of turning the
33 material are also reported. The results obtained can be used for the fats recycling to obtain a
34 stabilised compost with agronomical value.

35
36
37 Pre-print of Ruggiere, Luz et al. «Biodegradation of animal fats in a co-composting
38 process with wastewater sludge» in International Biodeterioration & Biodegradation
39 (Elsevier), Vol. 62, Issue 3, (October 2008), p. 297-303. The final version is available at
40 DOI [10.1016/j.ibiod.2008.02.004](https://doi.org/10.1016/j.ibiod.2008.02.004)

1 **Abstract**

2
3 A composting process was proposed as an effective technology for the biodegradation
4 of fats in a proportion of 40-50%. Anaerobically digested sludge was used as co-substrate for
5 animal fats to balance the C/N ratio of the composting mixture and to provide additional
6 biodegradable organic matter and active biomass. Two different strategies were studied: static
7 pile and dynamic turned pile. Air-filled porosity was initially adjusted to 40% for both
8 experiments. It was observed that non-turned strategy increases the formation of material
9 agglomerates which derived in a non-homogeneous fat distribution, the development of
10 filamentous fungi, and a considerable increase in the amount of leachate generated. Turning
11 the composting material resulted in the best results for composting fat enriched wastes,
12 preventing the formation of agglomerates. An effective biodegradation up to 92% of the fats
13 was observed under these conditions. Besides, the addition of fats increased significantly the
14 duration of the thermophilic period of the composting process.

15
16
17 **Key words:** Animal fats; Composting; Porosity; Turning; Wastewater sludge.

1 **1. Introduction**

2
3 Fats and oils are among the main components of organic matter in wastewater (Saatci
4 et al., 2001) and solid wastes, especially those produced by the food industry (Galli et al.,
5 1997; Mari et al., 2003). Fats and oils are essentially triglycerides consisting of linear fatty
6 acids attached, as esters, to glycerol. The biodegradation of fats begins with the enzymatic
7 hydrolysis of the ester bond carried out by lipases, followed by the consumption of glycerol
8 and beta-oxidation of fatty acids (Lalman and Bagley, 2000). However, fat biodegradation
9 can be limited by its insolubility in water (Lefebvre et al., 1998).

10 Among the available technologies to recycle organic solid wastes, composting is often
11 presented as a low-technology and low-investment process to convert organic solid wastes to
12 a soil amendment known as compost. Composting is a biotechnological process by which
13 different microbial communities initially biodegrade organic matter into simpler nutrients
14 and, in a second stage, form complex organic macromolecules such as humic acids (Hsu and
15 Lo, 1999). The composting process is characterized by the rapid development of thermophilic
16 temperatures due to the heat generation in the first degradation stage in which a significant
17 pathogen content reduction is also achieved. Composting is an aerobic microbial process that
18 requires optimal moisture and porosity (Haug, 1993). Temperature, oxygen and moisture
19 content are often selected as control variables of the composting process along with other
20 chemical, biochemical or microbiological properties such as enzymatic activity or respiration
21 indices (Barrena et al., 2005; Saviozzi et al., 2004; Tiquia, 2005).

22 References can be found in literature regarding the composting of fat-enriched wastes.
23 Special attention has been given to wastes derived from the olive oil industry (Albuquerque et
24 al., 2006a; Cegarra et al., 2006). In general, routine composting parameters show that
25 composting fat-enriched wastes is possible with low percentages of fats (5-15%) in the initial
26 mixture (Filippi et al., 2002; García-Gómez et al., 2003; Lemus and Lau, 2002), which causes
27 a longer thermophilic phase attributed to the high chemical energy content of fats (Nakano
28 and Matsumura, 2001). On the other hand, reported values of lipid degradation are usually
29 high, within 80-90% (Baeta-Hall et al., 2005; García-Gómez et al., 2003; Lemus and Lau,
30 2002) and up to 97% when long composting periods are considered (Baddi et al., 2004).
31 However, references to the degradation of fats of animal origin or organic matrices with high
32 fat content (over 15%) are scarce.

33 Composting of fats is inherently difficult due to their nutrients deficiency, with
34 especially low nitrogen and phosphorous content relative to high carbon content (Sasaki et al.,
35 2003). This fact usually implies the utilization of a co-substrate to compensate the C/N ratio
36 of the initial mixture and to act as inoculum. Different types of sludge, due to their typical low
37 C/N ratio, are considered suitable for being composted with fats or fat-enriched wastes
38 (Wakelin and Forster, 1997).

39 Water and oxygen are necessary for the biological activity of microorganisms and
40 their availability is directly related to the total porosity and the air-filled porosity (AFP) of the
41 material. These physical properties considerably affect the biological activity of a composting
42 matrix (Malinska and Richard 2006). Annan and White (1998) suggested an AFP range
43 between 30-60% as optimal, depending on the nature of the material to be composted. The
44 use of a bulking agent is required in sludge and fat composting due to the low AFP of both
45 materials. A long list of waste materials have been proposed as bulking agents in several
46 studies. The most widely used materials are wood chips and sawdust (Larsen and McCartney,
47 2000; Wong and Fang, 2000). Also the type and proportion of bulking agent has been
48 highlighted as being responsible for the thermophilic phase duration and rate of oil
49 biodegradation (Manios et al., 2006).

1 Our previous work (Gea et al., 2007a) showed that animal fat could be successfully
2 composted with sludge at high ratios to obtain a stable and sanitized product. Composting of
3 mixtures with a fat content of up to 50% was possible although a maximum content of 30%
4 was recommended to obtain high fat degradation (85%) and to avoid excessively long
5 composting periods. In addition, this work displayed the importance of bulking agent
6 selection.

7 The main objective of the present work is to study the biodegradation of animal fats
8 present in a high proportion in a composting process at laboratory scale with digested sewage
9 sludge used as co-substrate and to determine the influence of the strategy used for composting
10 (static or dynamic). The effects of initial adjustment of AFP are also studied. Chemical and
11 physical properties of the materials were monitored during the process, also the lipolytic
12 activity and the respiration index were used to measure the biological activity.

15 **2. Materials and Methods**

17 *2.1. Composting materials*

18
19 Animal fat (Trg Debo Fancy, KAO Corporation S.A., Spain) collected from a cow
20 slaughterhouse was mixed with anaerobically digested sewage sludge from the wastewater
21 treatment plant of Granollers (Barcelona, Spain). Long chain fatty acids (LCFA) profile of
22 animal fat used was: 3% myristic acid (C14:0), 30% palmitic acid (C16:0), 17% stearic acid
23 (C18:0), 38% oleic acid (C18:1), 6% linoleic acid (C18:2), 6% other LCFA. Wood chips from
24 a local carpentry were used as bulking agent. Initial characteristics of composted materials are
25 summarized in Table 1.

27 *2.2 Composting mixtures*

28
29 Two composting strategies were tested: static system and dynamic turned system. In
30 both experiments, a high animal fat content (40-50%) was used. The initial characteristics of
31 the composted mixtures are summarized in Table 2.

32 Bulking agent particle size was in the range of 0-10 mm. A mixing ratio of bulking
33 agent to sludge/fat mixture was calculated to adjust initial the AFP to 40% as porosity
34 requirement (Eftoda and McCartney, 2004). The approximate volumetric ratio to achieve the
35 porosity requirements was in the range of 2:1-1:1 bulking agent:sludge.

37 *2.3 Composting experiments*

38
39 The experiments were carried out using a 30 l insulated vessel conditioned for
40 composting with an O₂ control system. The online parameters recorded were temperature and
41 O₂ content. Gas content was measured with an O₂ sensor (Sensox 6C, Sensotran, Spain). One
42 Pt-100 sensor (SR-NOH, Desin, Spain) inserted at the centre of the reactor was used for
43 temperature monitoring. All sensors were connected to a self-made data acquisition system
44 implemented in a personal computer. Oxygen control was performed by a feedback oxygen
45 control that automatically supplied fresh air (room temperature) to the reactor by means of a
46 flow meter (Sensotran mod. MR3A18SVVT) when oxygen concentration was under 10%.
47 Water was manually added to the composting mixture when necessary to maintain moisture
48 content in the optimal range for composting (40-60%) (Haug, 1993).

49 Two strategies were used in the composting experiments. Static composting was
50 carried out without homogenization of the composting mass, whereas in dynamic composting

1 a complete manual mixing and homogenization of the material was carried out at the moment
2 of sampling. This procedure was used to simulate static (non-turned) and dynamic (turned)
3 composting conditions.

4 5 *2.4. Air-filled porosity*

6
7 Air-filled porosity is expressed as the ratio of gas-filled pore volume of the sample to
8 total sample volume. AFP was measured using a self made constant volume air pycnometer
9 according to the description of Annan and White (1998) and Oppenheimer et al., (1997) with
10 an effective sample chamber volume of 1.65 l and using an initial pressure of 6 bar. AFP
11 determinations were carried out in triplicate. The average value of AFP was calculated.

12 *2.5. Sampling procedure*

13
14 For each analyzed sample, a representative sample (2 l) was carefully taken out from
15 the core of the reactor to minimize the possible alterations of the composting mass for AFP
16 determination. After analyzing AFP a mixed sub-sample of approximately 700 ml was used to
17 determine the rest of the parameters studied during the experiments, whereas the remaining
18 sample volume was returned to the reactor. It was assumed that these high sample volumes
19 were representative of the whole composting mass in the reactor.

20 21 *2.6. Analytical Methods*

22
23 Fat content was determined by a standard Soxhlet method (U.S. Environmental
24 Protection Agency, 1998) using n-heptane as organic solvent. Additional analytical methods
25 to determine moisture content, volatile organic matter (OM), nitrogen Kjeldhal and pH were
26 carried out according to the standard procedures (U.S. Department of Agriculture and U.S.
27 Composting Council, 2001).

28 29 *2.7. Static Respiration Index*

30
31 Static Respiration Index (SRI) was determined in a static respirometer according to the
32 original model described by Ianotti et al. (1993) and following the modifications and
33 recommendations given by the U.S. Department of Agriculture and U.S. Composting Council
34 (2001). Assays were run at process temperature at the moment of sampling. A complete
35 description of the equipment and procedure can be found in Barrena et al. (2005). Three
36 replicates were used in each case and the average value was recorded. SRI is expressed as mg
37 O₂ g OM⁻¹ h⁻¹.

38 39 *2.8. Lipolytic Activity*

40
41 Lipolytic activity was determined using a commercial kit (Roche/Hitachi Lip num
42 1821792) as described by Lopez et al. (2002). Lipases were extracted from 5 g of a
43 representative sample of the composting material using 50 ml of 400 mM tris-HCl buffer with
44 10 mM CaCl₂ (pH 8). Triton X-100 (Panreac, Barcelona, Spain) at 5% (w/w) was added in
45 order to assure a quantitative extraction of lipases. It was assumed that lipolytic activity is
46 mainly non-water extractable, for that reason its presence in leachate was considered
47 negligible (Gessesse et al., 2003). After 30 minutes of extraction using a magnetic stirrer,
48 supernatant was centrifuged (25 min, 7000g) and filtered (0.45 μm) to remove biomass and
49 suspended solids. This sample was used for lipolytic activity determination. Standard lipolytic

1 activity assays were run at 30°C. Lipolytic activity was expressed as activity units per gram of
2 dry matter (AU g⁻¹). One activity unit was defined as the quantity of enzyme necessary to
3 release 1 µmol of fatty acid per minute under the specified conditions.

4 5 *2.9. Long chain fatty acids (LCFA)*

6
7 50 ml of heptane (99% purity) were added to 5 g of sample and mixed in a magnetic
8 stirrer for 30 minutes to extract LCFA. Afterwards, the suspension was centrifuged (30 min,
9 7000g) and the resulting supernatant filtered through a Millipore Millex FGS filter (0.2 µm).
10 This extract was used for free LCFA determination by gas chromatography using a Perkin-
11 Elmer AutoSystem XL Gas Chromatograph with a flame ionization detector (FID) and a HP
12 Innowax 30 m x 0.25 x 0.25 µm column. The carrier gas was Helium and a split ratio of 13
13 was used. An initial temperature of 120°C was kept for 1 min; then, it was increased up to
14 250°C at 8°C min⁻¹, and maintained at this temperature for 7 min. The system was calibrated
15 with different LCFA standards (including lauric, myristic, palmitic, stearic, oleic and linoleic
16 acid from Sigma, Spain) of concentrations in the range of 0-100 mg l⁻¹.

17 18 19 **3. Results**

20 21 *3.1. Initial porosity adjustment*

22
23 For the reported composting experiments, air-filled porosity of initial samples was
24 experimentally measured by air pycnometry and adjusted to 40% by adding bulking agent.
25 This is a new procedure to adjust porosity in composting mixtures, since the typical recipes
26 for composting are only based on volumetric ratios of bulking agent:substrate (Haug, 1993),
27 which do not directly consider the available porosity. In low-porosity materials, such as
28 wastewater sludge and fats, an experimental measure of porosity should be recommended for
29 an enhanced biodegradation. However, despite the significance of AFP in the composting
30 process, only few publications reflect its measurement and evolution along the composting
31 process (Eftoda and McCartney, 2004; Su et al., 2006).

32 In relation to the optimal values of porosity, previous experiments (data not shown)
33 with an initial AFP value of 30% did not follow the expected evolution and thermophilic
34 range of temperatures was not reached, being the biodegradation of fats negligible. Thus, an
35 initial AFP of 40% was selected in further experiments and could be considered as a
36 minimum porosity requirement in the composting of mixtures of low-porosity materials.

37 38 *3.2. Static composting*

39
40 Figures 1a, 1b and 1c show the results of different physical, chemical and biological
41 parameters monitored during the static composting experiment (without turning). The fat
42 content in the initial organic matrix was 43.7% (Table 2).

43 Figure 1a shows the temperature and SRI profiles obtained. Thermophilic
44 temperatures were reached at the second day of composting and maintained during 25 days.
45 Afterwards, a cooling phase and a mesophilic maturation stage were observed. SRI registered
46 the highest value on day 11 (6.36 mg O₂ g OM⁻¹ h⁻¹), which indicates a high metabolic
47 activity at that moment, coinciding with temperatures above 60°C. On day 30 the temperature
48 and the SRI decreased considerably, which indicates a reduction of the biological activity.
49 SRI followed the same pattern as temperature and has proved to be a reliable measure of
50 biological activity, biodegradability and stability of the material (Barrena et al., 2005). Similar

1 maximum values of SRI have been obtained at the initial thermophilic stage in composting
2 experiments with different types of wastes such as paper sludge, hair wastes and anaerobically
3 digested sludge. Higher values ($12 \text{ mg O}_2 \text{ g OM}^{-1} \text{ h}^{-1}$) have been observed for more readily
4 biodegradable materials as the organic fraction of municipal solid waste and raw sludge from
5 municipal wastewater treatment plant (Barrena et al., 2007; Gea et al., 2004).

6 Figure 1b shows moisture content and AFP profiles. Additional watering of the matrix
7 was required in several occasions during the process to maintain moisture in the
8 recommended range for composting (40-60%, Haug, 1993). When the reactor was opened for
9 sampling, water was homogeneously added on the surface of the composting mass without
10 additional mixing. However, due to the hydrophobic nature of the initial organic matrix, one
11 hour after watering approximately the 50% of the added water was released as leachate. This
12 fact highlights the difficulty of water content control in hydrophobic matrices under static
13 conditions.

14 Different stages can be observed in AFP and moisture content evolution during the
15 process (Fig 1b). Firstly, AFP increased as moisture decreased due to aeration and high
16 temperatures. Secondly, after watering (day 4), AFP decreased, as the added water filled the
17 free air pores. This inverse relationship was expected as total porosity (pores occupied by
18 water plus AFP) remains constant assuming that no physical or structural changes occur. In
19 the second stage (days 7-15), AFP decreased as moisture decreased, reflecting an overall
20 decrease of total porosity. This could be the result of fat melting due to the high temperatures
21 of the composting mass. During sampling, it was observed that fat, initially homogeneously
22 integrated with sludge, melted and flowed from it, filling the macro pores, resulting in the
23 compaction of the organic matrix and a lower value of AFP (Figure 2a). From day 15 to 40,
24 AFP and moisture content increased expectedly upon the biodegradation of organic matter,
25 which was confirmed by the high values of final AFP and moisture content.

26 Agglomeration was observed when temperature decreased in the cooling phase prior
27 to the maturation stage of composting (day 25). Figure 2b shows the agglomeration of organic
28 material due to filamentous fungi. Rojas-Avelizapa et al. (2007) reported an increase in fungal
29 biomass over bacterial biomass in the final stage of aerobic treatment of fat-enriched wastes.
30 The formation of agglomerates caused larger intra particular pores (Ahn et al., 2007) that
31 contributed to the increase in observed AFP values. Consequently, preferential paths for water
32 circulation appeared at the inner section of the mass and an increase in leachate generation
33 was observed due to the hydrophobic characteristics of the substrate. The lack of mixing
34 favoured agglomeration. The need for turning fat-enriched matrices in a composting process
35 has been highlighted as a measure to reduce compaction and to provide homogenization and
36 re-inoculation (Albuquerque et al., 2006a, 2006b).

37 Figure 1c shows fat content, organic matter and lipolytic activity profiles. The most
38 significant reduction was observed in the first days of process (31% fat content at day 11) and
39 from that moment until the end of the process only a slight reduction occurred.

40 On the other hand, high temperatures and high SRI values (Figure 1a) were registered
41 until day 25. This demonstrates that biological activity was not inhibited. Lipolytic activity
42 (Figure 1c) was detected during the second week of process and increased progressively
43 during the thermophilic phase to reach and maintain maximum levels in the mesophilic
44 cooling and maturation stages. This profile is opposite to temperature and SRI profiles, which
45 was probably due to an accumulation of lipolytic activity.

46 Table 3 shows the concentration of free LCFA in three different moments of the
47 process. It can be observed how LCFA concentration increased considerably from day 14 to
48 day 25. From this data, it can be deduced that in the initial stage of the composting process,
49 fat hydrolysis to LCFA was not limited, and the LCFA consumption was the rate limiting step
50 in biological activity, which was confirmed by the presence of lipolytic activity (Figure 1c).

1 Different mechanisms for LCFA and lipid hydrolysis inhibition have been reported in aerobic
2 and anaerobic biodegradation processes (Alves et al., 2001; Angelidaki et al., 1999; Becker et
3 al., 1999; Fernández et al., 2005; Hanaki et al., 1981; Lalman and Bagley 2001; Loperena et
4 al., 2006). LCFA concentrations found in this experiment are much higher than those found as
5 inhibitory in other references. It is thus possible that an inhibition of the degradation of
6 triglycerides, due to LCFA accumulation, provoked a diminution of fat degradation.

7 8 3.3. *Dynamic (turned) composting* 9

10 Figures 3a, 3b and 3c show the results obtained for composting under dynamic
11 (turned) conditions. Initial organic matrix used in this experiment presented a fat content of
12 47.5%. This value is slightly higher than that of static composting experiment. However, it
13 should be noted that it is very difficult to adjust the fat content of the mixture. In any cases,
14 both experiments (static and dynamic), fat content can be considered as very high since
15 reported values are in the range of 5-15% (Filippi et al., 2002; García-Gómez et al., 2003;
16 Lemus and Lau, 2002).

17 In the dynamic experiment, the composting mass was mixed at the moment of
18 sampling to emulate the homogenization provided by a dynamic composting system.
19 Agglomerates did also appear but were in fewer amount and smaller in size than those formed
20 using a static strategy. Consequently, the development of fungal communities and
21 agglomeration was limited as it was previously observed (Albuquerque et al., 2006b; Cegarra
22 et al., 2006).

23 Figure 3a shows the temperature and SRI profiles obtained in the composting process,
24 where thermophilic temperatures were reached on day 3 and maintained during 21 days. SRI
25 followed the same profile than temperature and showed an activity decrease on day 21. In
26 general SRI values in thermophilic phase were higher than those of static material indicating a
27 slightly more active process (maximum value $6.95 \text{ mg O}_2 \text{ g OM}^{-1} \text{ h}^{-1}$ at day 19).

28 Figure 3b shows moisture content and AFP profiles. Only two moisture adjustments
29 were necessary during the process to maintain moisture content within the range
30 recommended for the composting process (Haug, 1993). This important difference from static
31 experiment was attributed to the fact that water was fully integrated in the mixture. As a
32 result, the leachate volume generated was inferior to the volume generated in static conditions
33 in a 38%. AFP increased during the process as organic matter biodegraded. On day 19, a large
34 increase in AFP occurred, along with the end of the thermophilic phase. The high values of
35 AFP (around 70%) registered until day 33 coincided with the highest rate of fat
36 biodegradation during the process, from 31% to 7% (figure 3c). During the last stage of the
37 process (days 33 to 50) AFP decreased to 55%. This can be attributed to the compaction
38 phenomena observed in highly degraded organic wastes (Mohee and Mudho, 2005).

39 Figure 3c shows fat content, organic matter and lipolytic activity. Both fat content and
40 organic matter decreased considerably during all the process. No LCFA accumulation was
41 observed during the composting process, which confirms the hypothesis that periodical
42 homogenization of the composting mass and the agglomerates destruction favoured the
43 biological activity. Moreover, fat content decreased progressively during the composting
44 process reaching a final value of 5.4%, which implies a global fat biodegradation of 92%.

45 Although an important fat content reduction was observed, no lipolytic activity was
46 detected until the second week of process. As observed under static conditions, lipolytic
47 activity presented an increasing trend through the process. However, under dynamic
48 conditions the values of lipolytic activity were more erratic, which suggested a possible
49 analytical problem, for instance, in the lipase extraction method used for solid samples.
50 Further research is required in order to establish an effective method for enzyme extraction

1 and enzymatic activity determination in composting environments, especially when
2 quantitative determinations are necessary. A possible alternative for lipolytic activity analysis
3 could be the determination on a solid sample finely dispersed without solid-liquid extraction.
4 Although other enzyme activities such as dehydrogenase have been measured using this
5 approach (Barrena et al., 2008), no results for lipase activity have been reported, which is
6 probably due to the inherent interface catalytic mechanism of lipases involving mass transfer
7 limitations (Verger, 1997). Anyway, a procedure of lipase extraction for composting samples
8 could be the aim of future works.

10 **4. Discussion**

12 Table 4 shows the reduction of weight, dry mass, organic matter and fat content
13 obtained for the experiments carried out. Composting under dynamic conditions showed the
14 highest reduction of weight, dry mass, organic matter and fat content. Final fat content was
15 5% under these conditions. This indicates that there is a residual fraction of fats present in the
16 anaerobically digested sludge, which is not biodegradable under composting process
17 conditions. This fact has been reported previously (Réveillé *et al.*, 2003). However, it can be
18 concluded that sewage sludge acts as an adequate cosubstrate in fats composting in terms of
19 nitrogen source and providing additional biodegradable organic matter and active biomass.

20 From Figures 1a and 3a it can be deduced that a high fat content in the composting
21 mass leads to a longer thermophilic stage compared to those typically reported for laboratory
22 scale sewage sludge composting experiments (Gea et al, 2004), which confirms previous
23 observations (Gea et al., 2007a). This fact could be an inconvenient for the management of
24 composting plants when dealing with fat-enriched wastes (Manios et al., 2006). However,
25 fats can be useful as co-substrate in the case of composting low energy content wastes to fulfil
26 the international requirements on compost sanitation (temperature above 55°C for a total
27 period of 2 weeks, U.S. Environmental Protection Agency, 1995). Lipids contain twice the
28 energy of other organic materials like sugars and starch (Fernandes et al., 1988; Viel et al.,
29 1987). This high-energy content represents a clear advantage for processes where
30 thermophilic temperatures are desirable.

31 On the other hand, it has been demonstrated that an initial AFP adjustment of 40%
32 permits to reach the thermophilic range of temperatures. However, only considering the initial
33 AFP value does not guarantee the desired development of the biodegradation process.
34 Monitoring AFP along the composting time can ensure that this parameter does not fall under
35 the recommended values for aerobic microbial activity, but AFP does not give information
36 about porosity distribution. A visual observation of the composting matrix is additionally
37 necessary when composting particular types of wastes to detect structural changes of the
38 material as the agglomerates formation. Material turning should complement adequate AFP
39 values in those cases to achieve a correct porosity distribution.

40 Finally, it should be highlighted that moisture requirements and leachate generation
41 were lower under dynamic conditions because the formation of agglomerates was avoided.
42 These are important facts in considering the cost and the environmental impacts of the waste
43 management used for fats biodegradation (Wei et al., 2001). A mechanical turning system
44 without forced ventilation has been suggested as optimal for fat-enriched wastes from olive
45 oil mills (Cayueta et al., 2006, Cegarra et al., 2006).

48 **Acknowledgements**

1 The authors wish to thank KAO Corporation for the fat supply. Financial support was
2 provided by the Spanish Ministerio de Educación y Ciencia (Project CTM2006-00315).
3

1 References

- 2
3
4 Ahn, H.K., Richard, T.L., Glanville, T.D., 2007. Laboratory determination of compost
5 physical parameters for modeling of airflow characteristics. *Waste Management*. In
6 press.
- 7 Alburquerque, J.A., González, J., García, D., Cegarra, J., 2006a. Measuring detoxification
8 and maturity in compost made from “alperujo”, the solid by-product of extracting
9 olive oil by the two-phase centrifugation system. *Chemosphere* 64, 470–477.
- 10 Alburquerque, J.A., González, J., García, D., Cegarra, J., 2006b. Effects of bulking agent on
11 the composting of “alperujo”, the solid by-product of the two-phase centrifugation
12 method for olive oil extraction. *Process Biochemistry* 41, 127–132.
- 13 Alves, M., Mota, J.A., Álvares, M.A., Pereira, M., Mota, M., 2001. Effects of lipids and oleic
14 acid on biomass development in anaerobic fixed-bed reactors. Part II: Oleic acid
15 toxicity and biodegradability. *Water Research* 35, 264-270.
- 16 Angelidaki, I., Ellegaard, L., Ahring, B.K., 1999. A comprehensive model of anaerobic
17 bioconversion of complex substrates to biogas. *Biotechnology and Bioengineering* 63,
18 363-372.
- 19 Annan, J., White, R., 1998. Evaluation of techniques for measuring air-filled porosity in
20 compost of municipal biosolid and wood chips. In: Das, K.C., Graves, E.F., (Eds.),
21 *Composting in the Southeast – Proceedings of the 1998 conference*, Athens, Georgia,
22 pp. 88-96.
- 23 Baddi, G.A., Alburquerque, J. A., González, J., Cegarra, J., Hafidi, M., 2004. Chemical and
24 spectroscopic analyses of organic matter transformations during composting of olive
25 mill wastes. *International Biodeterioration and Biodegradation* 54, 39-44.
- 26 Baeta-Hall, L., Saagua, M.C., Bartolomeu, M.L., Anselmo, A.M., Rosa, M.F., 2005.
27 Biodegradation of olive oil husks in composting aerated piles. *Bioresource*
28 *Technology* 96, 69-78.
- 29 Barrena, R., Vázquez, F., Gordillo, M.A., Gea, T., Sánchez, A., 2005. Respirometric assays at
30 fixed and process temperatures to monitor composting process. *Bioresource*
31 *Technology* 96, 1153-1159.
- 32 Barrena, R., Pagans, E., Artola, A., Vázquez, F., Sánchez, A., 2007. Co-composting of hair
33 waste from the tanning industry with de-inking and municipal wastewater sludges.
34 *Biodegradation* 18, 257-268.
- 35 Barrena, R., Vázquez, F., Sánchez, A., 2008. Dehydrogenase activity as a method for
36 monitoring the composting process. *Bioresource Technology* 99, 905–908.
- 37 Becker, P., Koster, D., Popov, M.N., Markossian, S., Antranikian, G., Markl, H., 1999. The
38 biodegradation of olive oil and the treatment of lipid-rich wool scouring wastewater
39 under aerobic thermophilic conditions. *Water Research* 33, 653-660.
- 40 Cayuela, M. L., Sanchez-Monedero, M. A., Roig, A., 2006. Evaluation of two different
41 aeration systems for composting two-phase olive mill wastes. *Process Biochemistry*,
42 41, 616-623.
- 43 Cegarra, J., Alburquerque, J.A., González, J., Tortosa, G., Chaw, D., 2006. Effects of the
44 forced ventilation on composting of a solid olive-mill by-product (“alperujo”)
45 managed by mechanical turning. *Waste Management* 26, 1377-1383.
- 46 Eftoda, G., McCartney, D., 2004. Determining the critical bulking agent requirement for
47 municipal biosolid composting. *Compost Science and Utilization* 12, 208-218.
- 48 Fernandes, F., Viel, M., Sayag, D., André, L., 1988. Microbial breakdown of fats through in-
49 vessel co-composting of agricultural and urban wastes. *Biological Wastes* 26, 33-48.

- 1 Fernández, A., Sánchez, A., Font, X., 2005. Anaerobic co-digestion of a simulated organic
2 fraction of municipal solid wastes and fats of animal and vegetable origin.
3 Biochemical Engineering Journal 26, 22-28.
- 4 Filippi, C., Benidi, S., Levi-Minzi, R., Cardelli, R., Saviozzi, A., 2002. Co-composting of
5 olive oil mill by-products: chemical and microbiological evaluations. Compost
6 Science and Utilization 10, 63-71.
- 7 Galli, E., Pasetti, L., Fiorelli, F., Tomati, U., 1997. Olive-mill wastewater composting:
8 microbiological aspects. Waste Management and Research 15, 323-330.
- 9 García-Gómez, A., Roig, A., Bernal, M.P., 2003. Composting of the solid fraction of olive
10 mill wastewater with olive leaves: organic matter degradation and biological activity.
11 Bioresource Technology 86, 59-64.
- 12 Gea, T., Barrena, R., Artola, A., Sánchez, A., 2004. Monitoring the biological activity of the
13 composting process: Oxygen Uptake Rate (OUR), Respirometric Index (RI), and
14 Respiratory Quotient. Biotechnology and Bioengineering 88, 520-527.
- 15 Gea, T., Ferrer, P., Alvaro, G., Valero, F., Artola, A., Sánchez, A. 2007a. Co-composting of
16 sewage sludge:fats mixtures and characteristics of the lipases involved. Biochemical
17 Engineering Journal 33, 275-283.
- 18 Gea, T., Barrena, R., Artola, A., Sánchez, A., 2007b. Optimal bulking agent particle size and
19 usage for heat retention and disinfection in domestic wastewater sludge composting.
20 Waste Management 27, 1108-1116.
- 21 Gessesse, A., Dueholm, T., Petersen, S.B., Nielsen, P.H., 2003. Lipase and protease
22 extraction from activated sludge. Water Research 37, 3652-3657.
- 23 Hanaki, K., Matsuo, T., Nagase, M., 1981. Mechanism of inhibition by Long-Chain Fatty
24 Acids in Anaerobic Digestion Process. Biotechnology and Bioengineering 23, 1591-
25 1610.
- 26 Haug, R.T., 1993. The practical Handbook of Compost Engineering. Lewis Publishers, Boca
27 Raton.
- 28 Hsu, J., Lo, S., 1999. Chemical and spectroscopic analysis of organic matter transformations
29 during composting of pig manure. Environmental Pollution 104, 189-196.
- 30 Iannotti, D., Pang, B., Toth, D., Elwell, H., Keener, M., Hoitink, H., 1993. A Quantitative
31 Respirometric Method for Monitoring Compost Stability. Compost Science and
32 Utilization 1, 181-190.
- 33 Lalman, J.A., Bagley, D.M., 2000. Anaerobic degradation and inhibitory effects of linoleic
34 acid. Water Research 34, 4220-4228.
- 35 Lalman, J.A., Bagley, D.M., 2001. Anaerobic degradation and methanogenic inhibitory
36 effects of oleic and stearic acids by a mixed culture. Water Research 35, 2975-2983.
- 37 Larsen, K.L., McCartney, D.M., 2000. Effect of C:N ratio on microbial activity and N
38 retention: bench-scale study using pulp and paper biosolids. Compost Science and
39 Utilization 8, 147-159.
- 40 Lefebvre, X., Paul, E., Mauret, M., Baptiste, P., Capdeville, B., 1998. Kinetic characterization
41 of saponified domestic lipid residues aerobic biodegradation. Water Research 32,
42 3031-3038.
- 43 Lemus, G.R., Lau, A.K., 2002. Biodegradation of lipidic compounds in synthetic food wastes
44 during composting. Canadian Biosystems Engineering 44, 33-39.
- 45 Loperena, L., Saravia, V., Murro, D., Ferrari, M.D., Lareo, C., 2006. Kinetic properties of a
46 commercial and a native inoculum for aerobic milk fat degradation. Bioresource
47 Technology 97, 2160-2165.
- 48 López, N., Pérez, R., Vázquez, F., Valero, F., Sánchez, A., 2002. Immobilization of different
49 *Candida rugosa* lipases by adsorption onto polypropylene powder. Application to

- 1 chiral synthesis of ibuprofen and trans-2-phenyl-1-cyclohexanol esters. *Journal of*
2 *Chemical Technology and Biotechnology* 77, 175-182.
- 3 Malinska, K.A., Richard, T.L., 2006. The impact of physical properties and compaction on
4 biodegradation kinetics during composting. In: Kraft, E., Bidlingmaier, W., Bertoldi,
5 M., Diaz, L.F., Barth, J., (Eds.), *Proceedings of 5th International Conference ORBIT*
6 *2006, Weimar*, pp. 125-132.
- 7 Manios, T., Maniadakis, K., Kalogeraki, M., Mari, E., Stratakis, E., Terzakis, S., Boytzakis,
8 P., Naziridis, Y., Zampetakis, L., 2006. Efforts to explain and control the prolonged
9 thermophilic period in two-phase olive oil mill sludge composting. *Biodegradation* 17,
10 285-292.
- 11 Mari, I., Ehaliotis, C., Kotsou, M., Balis, C., Georgakakis, D., 2003. Respiration profiles in
12 monitoring the composting of by-products from the olive oil agro-industry.
13 *Bioresource Technology* 87, 331-336.
- 14 Mohee, R., Mudhoo, A., 2005. Analysis of the physical properties of an in-vessel composting
15 matrix. *Powder Technology* 155, 92-99.
- 16 Nakano, K., Matsumura, M., 2001. Improvement of treatment efficiency of thermophilic oxic
17 process for highly concentrated lipid wastes by nutrient supplementation. *Journal of*
18 *Bioscience and Bioengineering* 92, 532-538.
- 19 Oppenheimer, J., Martin, J., Walker, L., 1997. Measurements of air-filled porosity in
20 unsaturated organic matrices using a pycnometer. *Bioresource Technology* 59, 241-
21 247.
- 22 Réveillé, V., Mansuy, L., Jardé, E., Garnier-Sillam, E., 2003. Characterisation of sewage
23 sludge-derived organic matter: lipids and humic acids. *Organic Geochemistry* 34, 615-
24 627.
- 25 Rojas-Avelizapa, N.G., Roldán-Carrillo, T., Zegarra-Martínez, H., Muñoz-Colunga, A.M.,
26 Fernández-Linares, L.C., 2007. A field trial for an ex-situ bioremediation of a drilling
27 mud-polluted site. *Chemosphere* 66, 1595-1600.
- 28 Saatci, Y., Arslan, E.I., Konar, V., 2001. Removal of total lipids and fatty acids from
29 sunflower oil factory effluent by UASB reactor. *Bioresource Technology* 87, 269-272.
- 30 Sasaki, N., Suehara, K.I., Kohda, J., Nakano, Y., Yang, T., 2003. Effects of C/N ratio and pH
31 of raw materials on oil degradation efficiency in a compost fermentation process
32 *Journal of Bioscience and Bioengineering* 96, 47-52.
- 33 Saviozzi, A., Cardelli, R., Levi-Minzi, R., Riffaldi, R., 2004. Evolution of biochemical
34 parameters during composting of urban wastes. *Compost Science and Utilization* 12,
35 153-160.
- 36 Su, D., McCartney, D., Wang, Q., 2006. Comparison of Free Air Space Test Methods.
37 *Compost Science and Utilization* 14, 103-113.
- 38 Tiquia, S.M., 2005. Microbiological parameters as indicators of compost maturity. *Journal of*
39 *Applied Microbiology* 99, 816-828.
- 40 US Department of Agriculture and US Composting Council., 2001. Test methods for the
41 examination of composting and compost. Edaphos International, Houston.
- 42 US Environmental Protection Agency, 1995. A Guide to the Biosolids Risk Assessments for
43 the EPA Part 503 Rule. On line at: <http://www.epa.gov/owm/mtb/biosolids/503rule/>
- 44 US Environmental Protection Agency., 1998. Method 9071B. On line at:
45 <http://www.epa.gov/SW-846/pdfs/9071b.pdf>
- 46 Verger, R., 1997. 'Interfacial activation' of lipases: Facts and artifacts. *Trends in*
47 *Biotechnology* 15, 32-38.
- 48 Viel, M., Sayag, D., Peyre, A., André, L., 1987. Optimization of In-Vessel Co-Composting
49 Through Heat Recovery. *Biological Wastes* 20, 167-185.

1 Wakelin, N.G., Forster, C.F., 1997. An investigation into microbial removal of fats, oils and
2 greases. *Bioresource Technology* 59, 37-43.
3 Wei, Y.S., Fan, Y.B., Wang, M.J., 2001. A cost analysis of sewage sludge composting for
4 small and mid-scale municipal wastewater treatment plants. *Resources Conservation
5 and Recycling* 33, 203-216.
6 Wong, J.W.C., Fang, M., 2000. Effects of lime addition on sewage sludge composting
7 process. *Water Research* 34, 3691-3698.
8
9
10

1 **Figure Legends**

2

3

4

5 **Fig. 1.** Composting results for static experiment: a) Temperature and Static Respiration Index;
6 b) Moisture Content and Air filled Porosity (sampling and watering points are also indicated);
7 c) Organic Matter, Fat Content and Lipolytic Activity.

8

9 **Fig. 2.** Composting material in static experiment.

10

11 **Fig. 3.** Composting results for turned experiment. a) Temperature and Static Respiration
12 Index; b) Moisture Content and Air filled Porosity (sampling and watering points are also
13 indicated); c) Organic Matter, Fat Content and Lipolytic Activity.

14

15

16

17

18

19

20

21

1 **Table 1:** Main initial characteristics of composted materials.
2
3
4
5
6
7

Parameter	Wastewater sludge	Wood Chips	Animal Fat
Moisture content (%)	72.7	11.8	<2
Organic matter (% dry basis)	61.5	95.3	> 99
Fat content (% dry basis)	8.0	2.9	> 99
N-Kjeldhal (% dry basis)	2.6	0.1	<0.02
C/N ratio	8	500	>4000
pH	7.6	-	-
Air-filled porosity (%)	30.0	87.6	3.0

8

9

10

1 **Table 2:** Initial characteristics of the mixtures in static and turned strategies.
2
3
4
5
6
7

Parameter	Static	Turned
Total mass (kg)	15.3	12.8
Moisture content (% wet basis)	52.0	44.3
Organic matter (% dry basis)	75.2	72.6
Fat content (% dry basis)	43.7	47.5
Air-filled porosity (%)	40	40
pH	7.2	7.0

8
9
10
11

1 **Table 3:** LCFA concentration (mg g^{-1} , dry basis) in different days of the composting process
2 for static material.

3
4
5
6
7

Day of process	Palmitic (C16:0)	Stearic (C18:0)	Oleic (C18:1)	Total LCFA
14 (thermophilic)	7.2	2.4	9.7	19.3
25 (cooling phase)	25.5	13.4	72.1	111
39 (maturation)	12.7	5.0	40.3	58

8
9
10

1 **Table 4:** Total mass, dry mass, organic matter and fat content reductions (in percentage)
2 obtained in the composting processes.

3
4
5
6
7

Parameter	Static	Turned
Total mass	16.8	24.5
Dry mass	30.1 ± 9.9	34.9 ± 10.0
Organic matter	36.8 ± 10.4	43.5 ± 11.1
Fat content	56.5 ± 18.1	92.6 ± 19.2

8
9
10