1	The effect of fungal pathogens on the water and carbon economy of trees:
2	implications for drought-induced mortality
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10	Number of words: 3317 words
11	2 colour figures
12	Drought-induced forest mortality is emerging as a widespread phenomenon with potentially large
13	implications for forest function and dynamics (Allen et al., 2010; Anderegg et al., 2012; Martínez-
14	Vilalta et al., 2012). Although the physiological mechanisms underlying tree mortality are still not
15	completely understood, there is agreement that they involve the storage and transport systems of water
16	and carbohydrates (McDowell et al., 2008; Sala et al., 2010; McDowell, 2011). The xylem of plants is
17	susceptible to drought-induced embolism and severe water deficits may result in the complete loss of
18	xylem hydraulic conductivity and cause tree mortality (hydraulic failure; cf. Tyree & Sperry, 1988;
19	McDowell et al., 2008; Choat et al., 2012). Drought also has detrimental effects on the carbon
20	economy of plants, and it has been hypothesized that reduced assimilation due to stomatal closure
21	may lead to a depletion of stored carbon reserves and, eventually, to tree death due to carbon
22	starvation (Waring, 1987; Martínez-Vilalta et al., 2002; Bréda et al., 2006; McDowell et al., 2008).
23	However, only in recent studies has a direct link between reduced carbon reserves and tree mortality
24	been established (Adams et al., 2009; Galiano et al., 2011; Adams et al., 2013; Hartmann et al., 2013;
25	Mitchell et al., 2013; Quirk et al., 2013; Sevanto et al., 2014). Finally, phloem transport could also
26	become impaired due to the inability of plants to maintain phloem turgor under extremely low xylem
27	water potentials, limiting the local availability of carbohydrates for metabolic functions (Sala et al.,
28	2010; Sevanto <i>et al.</i> , 2014).
29	We postulate that tree mortality research has suffered from a false dichotomy of drought versus biotic
30	attack (McDowell et al., 2013). Pests and pathogens cause tree mortality and it is well known that
31	drought may predispose forests to attacks by insects (Mattson & Haack, 1987; Gaylord et al., 2013)
32	and fungal pathogens (Desprez-Loustau et al., 2006; La Porta et al., 2008). The interaction between
33	drought stress and the damage caused by forest pests and pathogens has been addressed in a recent

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- 34 meta-analysis (Jactel et al., 2012), and the connection between the physiological status of the tree and
- disease development has motivated a number of reviews in the past (Schoeneweiss, 1975; Boyer,
- 36 1995). Biotic agents have also been included in theoretical models for drought induced mortality
- 37 (Martínez-Vilalta et al., 2002; McDowell et al., 2008; McDowell et al., 2011). However, previous
- 38 reports have not fully acknowledged the diversity of trophic interactions that microorganisms
- 39 establish with the host trees and how this diversity has direct consequences in terms of the
- 40 physiological mechanisms leading to mortality. Tree mortality can result directly from a toxic effect
- 41 from metabolites produced by pathogens, but pathogens can also disrupt the xylem and phloem of the
- 42 infected hosts and affect their carbon economy through the consumption of carbon reserves and the
- 43 induction of carbon-expensive defences. Here, we develop a new framework that brings together the
- 44 effects of pathogens and drought on the water and carbon economy of trees, and explore the
- 45 implications for the process of drought-induced mortality.

46 A new framework based on trophic interactions

- 47 We argue that predictions of drought-induced mortality under pathogen attack can be improved by
- 48 taking into account the type of trophic interaction that the pathogen establishes with the host. Three
- 49 main types of trophic interactions can be distinguished amongst tree pathogens: biotrophs,
- 50 necrotrophs and vascular wilts (Deacon, 1997) (Fig. 1). In general terms, biotrophs drain carbon and
- 51 nutrients from living cells the host response is based on recognition followed by programmed cell
- 52 death (Glazebrook, 2005). Necrotrophs instead interact with the host through the defence response
- 53 and get carbon and nutrients from dead cells the host response is based on carbon-based constitutive
- 54 and induced responses from living cells surrounding the infection (Glazebrook, 2005). A third
- 55 category includes vascular fungi that colonize the vascular system systemically, often aided by toxins
- 56 (Yadeta & Thomma, 2013) the host responses are based on blocking vertical and lateral spread in
- 57 the xylem.
- 58 In this letter, we describe how each of these pathogen types interact with the water and carbon
- 59 transport systems of trees, and by which mechanisms they may contribute to drought-induced
- 60 mortality (Fig. 2). As a basis for our rationale, we use the mechanistic model of McDowell *et al.*
- 61 (2011) to represent the mortality process of trees subjected to drought stress. We show that during
- 62 drought pathogens may disrupt the carbon balance of trees through three non-exclusive processes: (i)
- 63 by directly depleting non-structural carbohydrate (NSC) reserves, (ii) by forcing consumption of NSC
- reserves by the host or (iii) by increasing repair costs (Fig. 2a). Our model makes explicit predictions
- on the changes in photosynthesis, growth and respiration; as well as on the impacts on the NSC
- budget, phloem, and xylem transport during a drought episode leading to tree death. The amount of
- 67 carbon allocated to defence and the impact on a biotic agent's biomass are also included. Tree death is

68 represented as the point in which no carbon for sustaining the basic metabolism is available (i.e., zero 69 available carbon point), regardless of the process leading to this point. Death occurs when carbon 70 available falls below (intersects) the amount of carbon needed for osmotic adjustment and 71 maintenance of phloem and xylem transport. By considering different trophic interactions, two novel 72 perspectives for current mortality models are put forward: (i) we show the fundamental differences 73 among the *mechanisms* leading to tree mortality between biotrophs, necrotrophs and vascular wilt 74 pathogens, and (ii) we predict how different type of pathogens affect the timing of the zero available 75 *carbon* point, and consequently whether they contribute or not to drought-induced mortality. We have 76 considered the timing of the interaction between drought and pathogens in two ways. Either the 77 pathogen acts simultaneously with drought, as an opportunistic agent taking advantage of the effects 78 of reduced water availability on the host (inciting or contributing factor following Manion's (1981) 79 theory of decline); or else acts prior to the drought episode, causing a long-term effect weakening the 80 tree (predisposing factor). Our framework focuses on drought as the stress condition of the host, and 81 we do not discuss drought as the weather phenomenon that could facilitate/impair the pathogen spore 82 dispersal or germination and competition with other microorganisms.

83 Biotrophs and their direct dependence from the carbon in the infected tissue

84 Biotrophic pathogens have evolved mechanisms to derive carbon directly from living cells with

- 85 specialised structures named haustoria, which tap into host cells and create a local carbon sink (Fig.
- 1). Some well-studied biotrophic pathogens are *Erysiphe alphitoides*, *Phaeocryptopus gaeumannii*,
- 87 and rust pathogens like for instance *Peridermium pini* or *Melampsora* spp. Trees have evolved

88 defence mechanisms that shut the flow of carbon towards the pathogen. The defence is based on a fast

89 recognition of the threatening agent that triggers a programmed cell death (PCD) that kills the infected

- cells and withdraws the carbon and nutrients before they are assimilated by the pathogen (Fig. 1)
- 91 (Glazebrook, 2005). Together with PCD, trees also trigger salicylic-acid mediated defence responses
- 92 (Fig. 2d). With effector molecules, biotrophs manipulate the defence machinery of the host in order to
- 93 delay defence responses in order to gain enough time to multiply and spread into neighbouring cells
- 94 (Fig. 2e). Biotrophs mainly affect the carbon cycle by reducing assimilation and, compared with
- 95 hemi-biotrophs and necrotrophs, they produce little disruption of the water and carbon transport
- 96 systems of the host (Fig. 2c) (Bassanezi et al., 2002). Known mechanisms of reduction of
- 97 photosynthetic capacity involve the reduction of stomatal conductance by physically occluding of
- 98 stomata with mycelia or fruiting bodies, as well as other not-yet-understood mechanisms of fungal
- 99 interference with RuBisCO activity (Manter et al., 2000; Hajji et al., 2009). During fungal
- 100 establishment and especially when fruiting bodies are produced, carbon is drained from the leaves,
- 101 which become carbon sinks (Hewitt & Ayres, 1976), hence early leaf-shedding is a common tree
- 102 reaction to reduce carbon losses (Manter et al., 2003). In those cases in which stomatal functions are

103 heavily impaired (Manter *et al.*, 2000), damages can be very severe, leading to significant growth

- 104 reductions of infected trees (Kimberley et al., 2010). Damages can also accumulate over several years
- by, for instance, eliciting recurrent early leaf-shedding processes, reducing NSC reserves and
- 106 increasing the chances of death in the long run (Marcais & Bréda, 2006).

107 During acute drought carbon assimilation decreases and leaf cells may decrease their non-structural 108 carbon reserves (e.g., Adams et al. 2013). Increasing demands of sucrose by leaves decrease the influx 109 of carbon into the biotrophs, which cannot compete for sugars with living cells under drought 110 conditions (Wyness & Ayres, 1987). Low carbon accessibility during drought slows down fungal 111 multiplication, and deters further damages (Fig. 2e). Sporulation and mycelial growth, for example, 112 has been shown to be negatively affected by previous water stress (Ayres, 1977; Woolacott & Ayres, 113 1984), and the link between low disease levels of biotrophs and low NSC carbon availability has been 114 established in model plant systems (Engelsdorf et al., 2013). The strong connection between the 115 nutritional status of the host and the pathogen makes us hypothesize that drought will negatively 116 affect biotrophs during pathogen attack and therefore no worsening effects on tree death are 117 anticipated. As shown in our framework, no significant changes on the timing of the zero carbon available point are predicted (Fig. 2b). Our hypothesis is supported by the fact that biotrophs tend to 118 119 be more prevalent in well watered and fertilized sites (Toome et al., 2010), and are expected to 120 decrease in current climate scenarios including increased drought conditions (Desprez-Loustau et al., 2007; La Porta et al., 2008; Sturrock et al., 2011; Marçais & Desprez-Loustau, 2012). Still, we predict 121 worsening effects of drought on host survival in those cases in which biotrophs attain significant 122 123 population levels on the tree prior to the drought onset (Fig. 2b). Depleted carbohydrate reserves may 124 impair the subsequent capacity of trees to cope with water stress. Furthermore, if early leaf-shedding

125 has followed the biotroph attack, a carbon-expensive crown restoration may also accelerate tree death.

126 Necrotrophic pathogens and the importance of carbon for defense and pathogenicity

127 Necrotrophic pathogens obtain nutrients from dead cells and from structural carbon sources such as

128 cellulose and hemicellulose. Necrotrophs can attack leaves, twigs, branches, the stem or the root

129 system where they can destroy cambium and the vascular tissue and hence affect both carbon and

- 130 water transport systems. Tree defence is activated upon pathogen contact with living cells and is
- 131 mainly directed at compartmentalizing the pathogen within carbon-expensive barriers (Fig. 1).
- 132 Compartmentalization also implies the sacrificial conversion of vascular tissues in the sapwood (Oliva
- 133 *et al.*, 2012), and, in the case of pathogens causing cankers, in the cambial zone and the phloem.
- 134 Necrotrophs neutralize tree defences and kill living cells by secreting enzymes and toxins (Fig. 1).
- 135 Some well-known necrotrophic pathogens include many root rots such as *Heterobasidion annosum* or
- 136 Armillaria sp. and canker pathogens such as Cryphonectria parasitica or Cytospora chrysosperma.

137 The accessibility to carbon by both the tree and the pathogen determines the outcome of the 138 interaction by simultaneously affecting the capacity of the pathogen to build up further inoculum and 139 counteract tree defences, and the capacity of the tree to build up a sufficiently strong response (Fig. 140 2h). Some necrotrophic root pathogens gain access by themselves to the carbon sources within the 141 host by degrading constitutive and induced defence barriers, like bark or lignin. In these cases 142 pathogens use carbon from external sources like neighbouring infected or dead trees (Stenlid, 1987; 143 Cleary et al., 2012). In the case of necrotrophs affecting branches or the main stem, the pathogen must 144 gain access to carbon rich tissues of the phloem passively, either via airborne infection of wounded 145 tissues or by entering the tree as endophytes (Manion & French, 1967). In any case, the outcome of the interaction depends on the host's carbon availability in order to react fast and compartmentalize 146 147 the pathogen (Guyon et al., 1996). The magnitude of carbon needed for defence is large and it has 148 been shown to have a negative impact on tree radial growth (Bendz-Hellgren & Stenlid, 1995; 149 Krokene et al., 2008; Cruickshank et al., 2011; Oliva et al., 2012). By forcing the tree to invest carbon 150 in defence, necrotrophs affect water transport and storage indirectly by inducing low growth, which 151 results in lowering the overall conductivity of diseased tissues (Joseph et al., 1998) and reducing sapwood storage (Oliva et al., 2012). Necrotrophs can also destroy functional tissues in leaves, stem 152 and roots, which may require repair, and thus they can increase further the carbon needs from the 153 host. Under favourable conditions for the host, necrotrophic interactions may persist for decades until 154 155 trees ultimately die (Cherubini et al., 2002). Indeed, large cankers are often seen in trees and are the 156 result of many years of seasonal variations in the capacity of the tree to prevent the pathogen advance

- 157 (Manion, 1981; Solla et al., 2006) (Fig. 2i).
- 158 The outcome of necrotrophic interactions is influenced by external stress factors such as drought 159 affecting carbon availability in the host. Severe and prolonged drought periods usually reduce carbon
- reserves (Galiano *et al.*, 2011; McDowell, 2011; Galiano *et al.*, 2012), limiting the availability of
- 161 carbon to support defences and preventing the establishment or the expansion of previously
- 162 established necrotrophs [e.g., Kane and Kolb (2010), Gaylord et al. (2013), Anderegg and Anderegg
- 163 (2013)]. Decreased tree defences facilitate the access of necrotrophic pathogens to carbon sources,
- 164 from which they build up further inoculum and produce further damages (Fig. 2) (Manion & French,
- 165 1967; Lygis *et al.*, 2005; Marcais & Bréda, 2006). Defoliation frequently occurs during drought
- 166 periods, and degradation of starch into readily usable/transportable sugar compounds to restore the
- 167 crown can also facilitate carbon access to necrotrophic root pathogens (Wargo, 1972). As lesions
- 168 enlarge, the size of the front, where host and pathogen interact, increases and with it the carbon costs
- 169 to contain the pathogen's progression (Fig. 2h). As with biotrophs, carbon used prior to the drought
- 170 for repairing infected tissues or for building up defences can also contribute to accelerating tree
- 171 mortality (Fig. 2f). Overall, necrotrophs accelerate drought-induced mortality (Fig. 2f) either by

- depleting resources and creating repair needs in advance or by making trees run out of carbon at a
- 173 faster rate. Consistent with our framework, increased damages have often been observed/expected
- under drought conditions by necrotrophic canker (Luque *et al.*, 2000; Desprez-Loustau *et al.*, 2006;
- 175 Waldboth & Oberhuber, 2009) and root rot pathogens (La Porta *et al.*, 2008; Sturrock *et al.*, 2011).

176 Vascular wilts and the destruction of the water transport system

- 177 Vascular wilt pathogens thrive inside xylem conduits, releasing toxic compounds and disturbing water 178 transport (Fig. 1). Some examples of vascular wilt pathogens include some Ophiostoma species, 179 remarkably O. novo-ulmi and also several Ceratocystis and Leptographium species. These type of 180 pathogens feed on xylem sap sugars, carbon leakages, defence compounds and sugars from cell-wall 181 degradation processes (Hammerbacher et al., 2013; Yadeta & Thomma, 2013). Trees block vertical 182 spread by clogging the conduits with tyloses, while lateral spread is prevented by *in situ* synthesis of 183 carbon compounds and barrier structures to compartmentalize the infection (Shigo & Tippett, 1981; 184 Bonsen et al., 1985; Yadeta & Thomma, 2013). Defence can be carbon expensive (Guérard et al., 185 2007) and result in a reduction of sugars in the vicinity of the lesion (Viiri et al., 2001). Investment in defence can be at the expense of radial growth (Krokene et al., 2008) and also imply a sacrificial loss 186 187 of conductive tissue (Joseph et al., 1998). In contrast to necrotrophs, vascular wilt pathogens have 188 significant direct effects on water transport and storage in trees (Fig. 2). Xylem disruption has 189 immediate effects and may cause sudden mortality on adult trees (Tyree & Zimmermann, 2002). 190 Conduit clogging results in foliage wilting that impacts current and future carbon reserves by cutting 191 downstream carbon supply and by reducing autumn re-assimilation of nutrients from leaves. Under 192 these conditions, xylem, phloem and foliage damage become very costly to repair (Fig. 2j). Wilt 193 diseases are often associated with bark beetles that feed on the phloem, increasing even further the 194 costs of repair and reducing the capacity to allocate carbon to the crown and restore foliage. 195 Nevertheless, insect phloem damage has been shown to be of lesser importance compared with xylem dysfunction induced by insect-vectored wilt pathogens (Hubbard et al., 2013), although in some cases 196 197 disruption of the water balance of the tree is not a pre-requisite for the success of the bark beetles 198 (Wullschleger et al., 2004).
- In contrast to carbon starvation-driven mortality in the case of necrotrophs, mortality in trees infected by vascular wilt pathogens seems to be triggered by hydraulic failure (Fig. 2k). Disruption of the vascular system is fast and permanent, hence rapid mortality of the corresponding areas of the crown or the whole tree can be observed. Increased damages by insect bark beetles and their associated vascular wilt pathogens are associated with dryer climatic conditions (Williams *et al.*, 2010), but, contrarily to necrotrophs, drought during the infection/attack may be more important than previous drought events (Croisé *et al.*, 2001). The availability of carbon for defence at the moment of attack is

- also of a lesser importance in comparison with necrotrophs (Christiansen & Ericsson, 1986). We thus
- 207 postulate that vascular wilt pathogens accelerate drought-induced mortality under drought mostly by
- 208 damaging the xylem vascular system and subsequently causing phloem impairment and foliage
- 209 wilting. Of special importance is the rapid escalation of repair costs as the attack builds up (Fig. 2j).
- 210 While carbon reserves can be reasonably high at the onset of a drought event, they may still not be
- 211 enough for rebuilding a sufficient amount of foliage, phloem and xylem for tree survival. By
- 212 increasing repair costs, wilt pathogens can also accelerate drought induced mortality processes (Fig.
- 213 2j).

214 Concluding remarks

215 The presented framework sets the ground for predicting the role of pathogens on tree mortality under 216 drought based on the type of trophic interaction established with the host. Although most pathogens 217 fall within the three categories described in the previous sections, some might establish more than one 218 type of trophic interaction. This is the case of the so-called hemibiotrophs, a category that includes 219 many *Phytophthora* species that share characteristics with both biotrophs and necrotrophs. In these 220 cases, we suggest that the type of trophic interaction that contributes more to the pathogen's inoculum 221 build-up should be considered. Would pre-inoculation water stress (Marçais et al., 1993) or carbon 222 starvation (Engelsdorf et al., 2013) favour disease development, these pathogens should be considered 223 for their necrotrophic phase and thus be expected to accelerate drought-induced-mortality. Other 224 pathogens can display a behaviour in between a wilt pathogen and a necrotroph. These pathogens are 225 typically secondary pathogens affecting woody tissues, like shoots and twigs (Jactel et al., 2012), and 226 while they can cause disease under negative water potentials, tree resistance is typically restored when 227 water stress is remediated (Crist & Schoeneweiss, 1975; Schoeneweiss, 1975; Johnson et al., 1997). 228 The fact that the pathogenicity of these fungi is strongly dependent on xylem colonisation (Luchi et 229 al., 2005), and that the necrotrophic phase precedes the wilting of the infected tissue, makes them 230 similar to the "vascular wilt pathogens" in our framework. The same reasoning can be applied to 231 similar pathogens for which pre-inoculation water stress and carbon limitation would contribute little 232 to host susceptibility (Madar et al., 1989).

Future climate scenarios predict an impact on water and carbon balance of trees (Wang *et al.*, 2012).

At the same time, forest pathogens are pervasive in forest ecosystems all over the globe and are

known to cause tree mortality and have a major role in forest dynamics (Worrall et al., 2005). Carbon

- and water systems are inevitably connected and both are affected by drought and by pathogens.
- 237 Pathogens can accelerate drought-induced mortality by directly depleting NSC, accelerating NSC
- consumption by the host or by increasing repairing costs (Fig. 2a). These three processes are tightly
- connected with the type of trophic interactions established between the host and the pathogen. We

- 240 describe how these types of pathogens would interact with the host, and by which mechanisms would
- 241 cause the death of the tree. This theoretical framework allows us to predict that some pathogens such
- 242 as necrotrophs or vascular wilts can benefit from drought events, and thus contribute to drought
- 243 induced mortality; and that some, like biotrophs are very unlikely to cause significant damages under
- 244 drought. Considering their different effects on the host and the contrasted interaction with drought,
- 245 determining under what environmental conditions the previous trophic interactions will be favoured
- 246 (or disfavoured) is pivotal to predictions of how forests will respond to warmer and drier conditions in
- 247 the future. Future research needs to quantify the contribution of pathogens to direct drought effects in
- the context of drought-induced tree mortality. Manipulative experiments controlling both drought and 248
- 249 pathogen inoculum can be used to assess the extent to which pathogens accelerate mortality by
- 250 comparing the time needed to kill trees under drought with and without specific pathogens (Fig. 2a).

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255 References

- Adams HD, Germino MJ, Breshears DD, Barron-Gafford GA, Guardiola-Claramonte M, Zou CB,
 Huxman TE. 2013. Nonstructural leaf carbohydrate dynamics of *Pinus edulis* during drought induced tree mortality reveal role for carbon metabolism in mortality mechanism. *New Phytologist* 197(4): 1142-1151.
- Adams HD, Guardiola-Claramonte M, Barron-Gafford GA, Villegas JC, Breshears DD, Zou CB, Troch
 PA, Huxman TE. 2009. Temperature sensitivity of drought-induced tree mortality portends
 increased regional die-off under global-change-type drought. *Proceedings of the National* Academy of Sciences 106(17): 7063-7066.
- Allen CD, Macalady AK, Chenchouni H, Bachelet D, McDowell N, Vennetier M, Kitzberger T, Rigling
 A, Breshears DD, Hogg EH, et al. 2010. A global overview of drought and heat-induced tree
 mortality reveals emerging climate change risks for forests. *Forest Ecology and Management* 267 259(4): 660-684.
- Anderegg WRL, Anderegg LDL. 2013. Hydraulic and carbohydrate changes in experimental drought induced mortality of saplings in two conifer species. *Tree Physiology* 33(3): 252-260.
- Anderegg WRL, Anderegg LDL, Sherman C, Karp DS. 2012. Effects of widespread drought-induced
 aspen mortality on understory plants. *Conservation Biology* 26(6): 1082-1090.
- Ayres PG. 1977. Effect of water potential of pea leaves on spore production by *Erysiphe pisi* (powdery mildew). *Transactions of the British Mycological Society* 68(1): 97-100.
- Bassanezi RB, Amorim L, Filho AB, Berger RD. 2002. Gas exchange and emission of chlorophyll
 fluorescence during the monocycle of rust, angular leaf spot and anthracnose on bean
 leaves as a function of their trophic characteristics. *Journal of Phytopathology* 150(1): 37-47.
- Bendz-Hellgren M, Stenlid J. 1995. Long-term reduction in the diameter growth of butt rot affected
 Norway spruce, *Picea abies. Forest Ecology and Management* 74(1-3): 239-243.
- Bonsen KJM, Scheffer RJ, Elgersma DM. 1985. Barrier zone formation as a resistance mechanism of
 elms to dutch elm disease. *IAWA Bulletin* 6: 71-76.
- Boyer JS. 1995. Biochemical and biophysical aspects of water deficits and the predisposition to
 disease. Annual Review of Phytopathology 33(1): 251-274.
- Bréda N, Huc R, Granier A, Dreyer E. 2006. Temperate forest trees and stands under severe drought:
 a review of ecophysiological responses, adaptation processes and long-term consequences.
 Ann. For. Sci. 63(6): 625-644.
- Cherubini P, Fontana G, Rigling D, Dobbertin M, Brang P, Innes JL. 2002. Tree-life history prior to
 death: two fungal root pathogens affect tree-ring growth differently. *Journal of Ecology* 90(5): 839-850.
- Choat B, Jansen S, Brodribb TJ, Cochard H, Delzon S, Bhaskar R, Bucci SJ, Feild TS, Gleason SM,
 Hacke UG, et al. 2012. Global convergence in the vulnerability of forests to drought. *Nature* 491(7426): 752-755.
- 292 Christiansen E, Ericsson A. 1986. Starch reserves in *Picea abies* in relation to defence reaction
 293 against a bark beetle transmitted blue-stain fungus, *Ceratocystis polonica*. *Canadian Journal* 294 of Forest Research 16(1): 78-83.
- Cleary MR, van der Kamp BJ, Morrison DJ. 2012. Pathogenicity and virulence of Armillaria sinapina
 and host response to infection in Douglas-fir, western hemlock and western redcedar in the
 southern Interior of British Columbia. *Forest Pathology* 42(6): 481-491.
- 298 Crist CR, Schoeneweiss DF. 1975. The influence of controlled stresses on susceptibility of European
 299 white birch stems to attack by *Botryosphaeria dothidea*. *Phytopathology* 65: 369-373
- 300 Croisé L, Lieutier F, Cochard H, Dreyer E. 2001. Effects of drought stress and high density stem
 301 inoculations with *Leptographium wingfieldii* on hydraulic properties of young Scots pine
 302 trees. *Tree Physiology* 21(7): 427-436.

- 303 Cruickshank MG, Morrison DJ, Lalumière A. 2011. Site, plot, and individual tree yield reduction of
 304 interior Douglas-fir associated with non-lethal infection by Armillaria root disease in
 305 southern British Columbia. *Forest Ecology and Management* 261(2): 297-307.
- 306 Deacon J. 1997. Modern mycology: Blackwell Science Ltd.

307 **Desprez-Loustau M-L, Marçais B, Nageleisen L-M, Piou D, Vannini A. 2006.** Interactive effects of 308 drought and pathogens in forest trees. *Annals of Forest Science* **63**(6): 597-612.

- 309 Desprez-Loustau M-L, Robin C, Reynaud G, Déqué M, Badeau V, Piou D, Husson C, Marçais B. 2007.
 310 Simulating the effects of a climate-change scenario on the geographical range and activity of 311 forest-pathogenic fungi. *Canadian Journal of Plant Pathology* 29(2): 101-120.
- Engelsdorf T, Horst RJ, Pröls R, Pröschel M, Dietz F, Hückelhoven R, Voll LM. 2013. Reduced
 carbohydrate availability enhances the susceptibility of *Arabidopsis* toward *Colletotrichum higginsianum. Plant Physiology* 162(1): 225-238.
- Galiano L, Martínez-Vilalta J, Lloret F. 2011. Carbon reserves and canopy defoliation determine the
 recovery of Scots pine 4 yr after a drought episode. *New Phytologist* 190(3): 750-759.
- Galiano L, Martínez-Vilalta J, Sabaté S, Lloret F. 2012. Determinants of drought effects on crown
 condition and their relationship with depletion of carbon reserves in a Mediterranean holm
 oak forest. *Tree Physiology* 32(4): 478-489.
- Gaylord ML, Kolb TE, Pockman WT, Plaut JA, Yepez EA, Macalady AK, Pangle RE, McDowell NG.
 2013. Drought predisposes piñon-juniper woodlands to insect attacks and mortality. *New* Phytologist 198(2): 567-578.
- Glazebrook J. 2005. Contrasting mechanisms of defense against biotrophic and necrotrophic
 pathogens. Annual Review of Phytopathology 43(1): 205-227.
- Guérard N, Maillard P, Bréchet C, Lieutier F, Dreyer E. 2007. Do trees use reserve or newly
 assimilated carbon for their defense reactions? A 13C labeling approach with young Scots
 pines inoculated with a bark-beetle-associated fungus (*Ophiostoma brunneo ciliatum*).
 Annals of Forest Science 64(6): 601-608.
- Guyon J, Jacobi W, McIntyre G. 1996. Effects of environmental stress on the development of
 Cytospora canker of aspen. *Plant Disease* 80: 1320-1326.
- Hajji M, Dreyer E, Marçais B. 2009. Impact of *Erysiphe alphitoides* on transpiration and
 photosynthesis in *Quercus robur* leaves. *European Journal of Plant Pathology* 125(1): 63-72.
- Hammerbacher A, Schmidt A, Wadke N, Wright LP, Schneider B, Bohlmann J, Brand WA, Fenning
 TM, Gershenzon J, Paetz C. 2013. A Common Fungal Associate of the Spruce Bark Beetle
 Metabolizes the Stilbene Defenses of Norway Spruce. *Plant Physiology* 162(3): 1324-1336.
- Hartmann H, Ziegler W, Trumbore S. 2013. Lethal drought leads to reduction in nonstructural
 carbohydrates in Norway spruce tree roots but not in the canopy. *Functional Ecology* 27(2):
 413-427.
- Hewitt HG, Ayres PG. 1976. Effect of infection by *Microsphaera alphitoides* (powdery mildew) on
 carbohybrate levels and translocation in seedlings of *Quercus robur*. *New Phytologist* 77(2):
 379-390.
- Hubbard RM, Rhoades CC, Elder K, Negron J. 2013. Changes in transpiration and foliage growth in
 lodgepole pine trees following mountain pine beetle attack and mechanical girdling. *Forest Ecology and Management* 289(0): 312-317.
- Jactel H, Petit J, Desprez-Loustau M-L, Delzon S, Piou D, Battisti A, Koricheva J. 2012. Drought
 effects on damage by forest insects and pathogens: a meta-analysis. *Global Change Biology* 18(1): 267-276.
- Johnson JW, Gleason ML, Parker SK, Provin EB, Iles JK. 1997. Duration of water stress affects
 development of *Sphaeropsis* canker on Scots pine. *Journal of Arboriculture* 23: 73-76.
- Joseph G, Kelsey RG, Thies WG. 1998. Hydraulic conductivity in roots of ponderosa pine infected
 with black-stain (*Leptographium wageneri*) or annosus (*Heterobasidion annosum*) root
 disease. *Tree Physiology* 18(5): 333-339.

- Kane JM, Kolb TE. 2010. Importance of resin ducts in reducing ponderosa pine mortality from bark
 beetle attack. *Oecologia* 164(3): 601-609.
- 355 Kimberley MO, Hood IA, Knowles RL. 2010. Impact of Swiss needle-cast on growth of Douglas-fir.
 356 Phytopathology 101(5): 583-593.
- Krokene P, Nagy NE, Solheim H. 2008. Methyl jasmonate and oxalic acid treatment of Norway
 spruce: anatomically based defense responses and increased resistance against fungal
 infection. *Tree Physiology* 28(1): 29-35.
- La Porta N, Capretti P, Thomsen IM, Kasanen R, Hietala AM, Von Weissenberg K. 2008. Forest
 pathogens with higher damage potential due to climate change in Europe. *Canadian Journal* of Plant Pathology 30(2): 177-195.
- Luchi N, Ma R, Capretti P, Bonello P. 2005. Systemic induction of traumatic resin ducts and resin
 flow in Austrian pine by wounding and inoculation with *Sphaeropsis sapinea* and *Diplodia scrobiculata*. *Planta* 221(1): 75-84.
- Luque J, Parlade J, Pera J. 2000. Pathogenicity of fungi isolated from *Quercus suber* in Catalonia (NE
 Spain). Forest Pathology 30(5): 247-263.
- Lygis V, Vasiliauskas R, Larsson K-H, Stenlid J. 2005. Wood-inhabiting fungi in stems of *Fraxinus excelsior* in declining ash stands of northern Lithuania, with particular reference to *Armillaria cepistipes*. *Scandinavian Journal of Forest Research* 20(4): 337-346.
- 371Madar ZZ, Solel Z, Kimchi M. 1989. Effect of water stress in cypress on the development of cankers372caused by Diplodia pinea f. sp. cupressi and Seiridium cardinale. Plant Disease 73(484-486).
- 373 **Manion PD. 1981.** *Tree disease concepts*. Englewood Cliffs, NJ.: Prentice Hall.
- Manion PD, French DW. 1967. Nectria galligena and Ceratocystis fimbriata cankers of aspen in
 Minnesota. Forest Science 13(1): 23-28.
- Manter DK, Bond BJ, Kavanagh KL, Rosso PH, Filip GM. 2000. Pseudothecia of Swiss needle cast
 fungus, *Phaeocryptopus gaeumannii*, physically block stomata of Douglas fir, reducing CO2
 assimilation. *New Phytologist* 148(3): 481-491.
- Manter DK, Bond BJ, Kavanagh KL, Stone JK, Filip GM. 2003. Modelling the impacts of the foliar
 pathogen, *Phaeocryptopus gaeumannii*, on Douglas-fir physiology: net canopy carbon
 assimilation, needle abscission and growth. *Ecological Modelling* 164(2–3): 211-226.
- 382 Marcais B, Bréda N. 2006. Role of an opportunistic pathogen in the decline of stressed oak trees.
 383 Journal of Ecology 94(6): 1214-1223.
- Marçais B, Desprez-Loustau M-L. 2012. European oak powdery mildew: impact on trees, effects of
 environmental factors, and potential effects of climate change. *Annals of Forest Science*: 1 10.
- Marçais B, Dupuis F, Desprez-Loustau ML. 1993. Influence of water stress on susceptibility of red
 oak (*Quercus rubra*) to *Phytophthora cinnamomi*. *European Journal of Forest Pathology* 23(5): 295-305.
- Martínez-Vilalta J, Lloret F, Breshears DD. 2012. Drought-induced forest decline: causes, scope and
 implications. *Biology Letters* 8(5): 689-691.
- Martínez-Vilalta J, Prat E, Oliveras I, Piñol J. 2002. Xylem hydraulic properties of roots and stems of
 nine Mediterranean woody species. *Oecologia* 133(1): 19-29.
- 394 Mattson WJ, Haack RA. 1987. The role of drought in outbreaks of plant-eating insects. *BioScience* 395 37(2): 110-118.
- McDowell N, Pockman WT, Allen CD, Breshears DD, Cobb N, Kolb T, Plaut J, Sperry J, West A,
 Williams DG, et al. 2008. Mechanisms of plant survival and mortality during drought: why do
 some plants survive while others succumb to drought? *New Phytologist* 178(4): 719-739.
- 399 McDowell NG. 2011. Mechanisms linking drought, hydraulics, carbon metabolism, and vegetation
 400 mortality. *Plant Physiology* 155(3): 1051-1059.

- 401 McDowell NG, Beerling DJ, Breshears DD, Fisher RA, Raffa KF, Stitt M. 2011. The interdependence
 402 of mechanisms underlying climate-driven vegetation mortality. *Trends in Ecology and* 403 *Evolution* 26(10): 523-532.
- 404 McDowell NG, Ryan MG, Zeppel MJB, Tissue DT. 2013. Feature: Improving our knowledge of
 405 drought-induced forest mortality through experiments, observations, and modeling. New
 406 Phytologist 200(2): 289-293.
- 407 Mitchell PJ, O'Grady AP, Tissue DT, White DA, Ottenschlaeger ML, Pinkard EA. 2013. Drought
 408 response strategies define the relative contributions of hydraulic dysfunction and
 409 carbohydrate depletion during tree mortality. *New Phytologist* 197(3): 862-872.
- 410 Oliva J, Camarero JJ, Stenlid J. 2012. Understanding the role of sapwood loss and reaction zone
 411 formation on radial growth of Norway spruce (*Picea abies*) trees decayed by *Heterobasidion* 412 *annosum s.l. Forest Ecology and Management* 274(0): 201-209.
- 413 Quirk J, McDowell NG, Leake JR, Hudson PJ, Beerling DJ. 2013. Increased susceptibility to drought 414 induced mortality in Sequoia sempervirens (Cupressaceae) trees under Cenozoic
 415 atmospheric carbon dioxide starvation. American Journal of Botany 100(3): 582-591.
- 416 Sala A, Piper F, Hoch G. 2010. Physiological mechanisms of drought-induced tree mortality are far
 417 from being resolved. *New Phytologist* 186(2): 274-281.
- 418 Schoeneweiss DF. 1975. Predisposition, stress, and plant disease. Annual Review of Phytopathology
 419 13(1): 193-211.
- Sevanto S, McDowell NG, Dickman LT, Pangle R, Pockman WT. 2014. How do trees die? A test of
 the hydraulic failure and carbon starvation hypotheses. *Plant, Cell & Environment* 37(1): 153 161.
- 423 Shigo A, Tippett JT. 1981. Compartimentalization of American elm tissues infected by *Ceratocystis* 424 *ulmi. Plant Disease* 65: 715-718.
- Solla A, Sánchez-Miranda Á, Camarero JJ. 2006. Radial-growth and wood anatomical changes in
 Abies alba infected by Melampsorella caryophyllacearum: a dendroecological assessment of
 fungal damage. Annals of Forest Science 63(3): 293- 300.
- 428 Stenlid J. 1987. Controlling and predicting the spread of *Heterobasidion annosum* from infected
 429 stumps and trees of *Picea abies. Scandinavian Journal of Forest Research* 2(1): 187 198.
- 430 Sturrock RN, Frankel SJ, Brown AV, Hennon PE, Kliejunas JT, Lewis KJ, Worrall JJ, Woods AJ. 2011.
 431 Climate change and forest diseases. *Plant Pathology* 60(1): 133-149.
- 432 Toome M, Heinsoo K, Luik A. 2010. Relation between leaf rust (*Melampsora epitea*) severity and the
 433 specific leaf area in short rotation coppice willows. *European Journal of Plant Pathology* 434 126(4): 583-588.
- 435 **Tyree M, Zimmermann M. 2002.** *Xylem structure and the ascent of sap.* Berlin: Springer.
- 436 Tyree MT, Sperry JS. 1988. Do woody plants operate near the point of catastrophic xylem
 437 dysfunction caused by dynamic water stress? *Plant Physiology* 88(3): 574–580.
- Viiri H, Niemelä P, Kitunen V, Annila E. 2001. Soluble carbohydrates, radial growth and vigour of
 fertilized Norway spruce after inoculation with blue-stain fungus, *Ceratocystis polonica*.
 Trees Structure and Function 15(6): 327-334.
- Waldboth M, Oberhuber W. 2009. Synergistic effect of drought and chestnut blight (*Cryphonectria parasitica*) on growth decline of European chestnut (*Castanea sativa*). Forest Pathology
 39(1): 43-55.
- Wang W, Peng C, Kneeshaw DD, Larocque GR, Luo Z. 2012. Drought-induced tree mortality:
 ecological consequences, causes, and modeling. *Environmental Reviews* 20(2): 109-121.
- Wargo PM. 1972. Defoliation-induced chemical changes in sugar maple roots stimulate growth of
 Armillaria mellea. Phytopathology 62: 1278-1283.
- 448 Waring RH. 1987. Characteristics of trees predisposed to die. *Bioscience* 37: 569-577.

- Williams AP, Allen CD, Millar Cl, Swetnam TW, Michaelsen J, Still CJ, Leavitt SW. 2010. Forest
 responses to increasing aridity and warmth in the southwestern United States. *Proceedings* of the National Academy of Sciences.
- 452 Woolacott B, Ayres PG. 1984. Effects of plant age and water stress on production of conidia by
 453 *Erysiphe graminis* f.sp. *hordei* examined by non-destructive sampling. *Transactions of the* 454 *British Mycological Society* 82(3): 449-454.
- Worrall JJ, Lee TD, Harrington TC. 2005. Forest dynamics and agents that initiate and expand canopy
 gaps in *Picea-Abies* forests of Crawford Notch, New Hampshire, USA. *J Ecology* 93(1): 178 190.
- Wullschleger SD, McLaughlin SB, Ayres MP. 2004. High-resolution analysis of stem increment and
 sap flow for loblolly pine trees attacked by southern pine beetle. *Canadian Journal Forest Research* 34: 2387–2393.
- Wyness LE, Ayres PG. 1987. Plant-fungus water relations affect carbohydrate transport from pea
 leaf to powdery mildew (*Erysiphe pisi*) mycelium. *Transactions of the British Mycological* Society 88(1): 97-104.
- 464 Yadeta K, Thomma B. 2013. The xylem as battleground for plant hosts and vascular wilt pathogens.
 465 Frontiers in Plant Science 4.

Accepted version

467 Figure captions

468 Figure 1. Carbon fluxes between host and pathogen depending on the type of trophic interaction occurring in xylem and phloem. Three types of pathogens are represented: biotrophs, necrotrophs 469 470 and vascular wilts. Biotrophic pathogens derive carbon directly from living cells. Although chemical 471 tree responses are triggered (not represented), the defence system of the tree is based on a 472 programmed cell death that removes the carbon from the infected cells and stops the flow of carbon to the pathogen. Vascular wilt pathogens thrive inside xylem conduits where they feed on carbon leaking 473 474 from cells killed by means of toxins/enzymes and on carbon from xylem sap. They are also able to 475 metabolize defence compounds and can obtain sugars by degrading the xylem cell wall. Damages in 476 the xylem elicit carbon based defence responses. Necrotrophic pathogens kill living cells by secreting 477 enzymes and toxins and obtain nutrients from dead cells and from structural carbon sources such as 478 cellulose. Like vascular wilts, necrotrophs may also feed on tree defence compounds (not 479 represented). Trees defend themselves by compartmentalizing the pathogen within carbon-expensive 480 barriers.

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Figure 2. Hypothetical mechanisms of mortality for biotrophic, necrotrophic and vascular wilt 482 pathogens under drought conditions. Drought induced mortality is based on the model by 483 McDowell et al. (2011) and it is assumed to happen when the amount of NSC is smaller than the 484 amount of carbon needed to maintain osmotic regulation and xylem and phloem transport (marked as 485 *, in panels "a", "b", "f" and "j"). Pathogens affect NSC reserves differently depending on the type of 486 trophic interaction (Fig. 1), by directly depleting NSC reserves, accelerating NSC consumption and 487 increasing repair costs (a). The mechanism of mortality is shown for each type of pathogen and 488 489 separately depending on whether the attack occurs during drought (solid red line) or before the 490 drought period (dashed red line). For the sake of comparison, the process of mortality under drought 491 conditions without biotic agents is also shown (thick black line). In general, low water availability 492 causes stomatal closure and lower carbon assimilation. Under prolonged drought conditions the tree 493 may require using NSC reserves to maintain tissue growth and respiration. NSC may also be used to 494 restore the hydraulic system of the tree by refilling embolized conduits. If under pathogen attack, and 495 depending on the type of trophic interaction, trees elicit different responses impacting the acquisition, 496 storage and transport of water and carbon in the tree. The main effects of the drought pathogen 497 interaction on NSC, xylem and phloem transport and induced defence are represented. Biotrophs feed 498 directly on NSC reserves (b) without affecting phloem and xylem (c). The direct use of NSC by the 499 pathogen translates directly into pathogen population growth, which accelerates NSC depletion in a 500 positive feedback (e). In the case of drought, biotrophs cannot compete with plant tissues for carbon; 501 hence pathogen biomass decreases (e). Previous depletion of NSC by the biotroph can accelerate

502	mortality due to lower reserves at the onset of the drought (b). A minimal biotroph establishment is
503	predicted during the drought owing the declining availability of carbon as water deficit intensifies (e),
504	and no interactive effects with drought are predicted for infections establishing during drought (b).
505	Necrotrophs find their main carbon source on phloem elements in leaves, stem, and roots (Fig. 1).
506	Under normal conditions they establish a long-term interaction requiring carbon investment from the
507	host into defence (\mathbf{f}). Following phloem and xylem destruction, the host compartmentalizes the
508	pathogen further sacrificing more phloem and xylem (g). Eventual defence failures allow the
509	pathogen access to living cells and structural carbon sources (Fig. 1), from which it builds up further
510	biomass (i). Under drought, carbon for defence becomes scarce (\mathbf{f}) allowing previous infections to
511	expand (i) causing further damage to phloem and xylem (g), increasing further the costs of
512	compartmentalization (h). As a result, we predict that necrotrophs exacerbate drought effects,
513	accelerating tree mortality (\mathbf{f}). A similar outcome is expected for necrotroph infections occurring
514	during drought (f). Pathogen biomass remains after tree death (h) due to the capacity of this type of
515	pathogens to survive on dead tissues. Vascular wilt pathogens thrive in the xylem. Trees block
516	vertical pathogen spread by clogging the conduits (Fig. 1), with consequences for both xylem
517	transport (\mathbf{k}) and NSC (\mathbf{j}) . Negative water potentials allow them to colonise large parts of the xylem
518	(\mathbf{m}, \mathbf{j}) , causing foliage wilting and mortality of phloem tissues (k).
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