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Delineating the factors and cellular mechanisms involved in the survival of
Cerebellar Granule neurons

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CONFLICT OF INTEREST STATEMENT.

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2 The authors declare that they have no competing interests.
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ABSTRACT

Cerebellar granule neurons (CGNs) constitute the most abundant neuronal population in the mammalian brain. Their postnatal generation and the feasibility to induce their apoptotic death *in vitro* make them an excellent model to study the effect of several neurotransmitters and neurotrophins. Here, we first review which factors are involved in the generation and proliferation of CGNs in the external granule layer (EGL) and in the regulation of their differentiation and migration to internal granule layer (IGL). Special attention was given to the role of several neurotrophins and the NMDA subtype of glutamate receptor. Then, using the paradigm of potassium deprivation in cultured CGNs, we address several extracellular factors that promote the survival of CGNs, with particular emphasis on the cellular mechanisms. The role of specific protein kinases leading to the regulation of transcription factors and recent data involving the small G protein family is also discussed. Finally, the participation of some members of Bcl-2 family and the inhibition of mitochondria-related apoptotic pathway is also considered. Altogether, these studies evidence that CGNs are a key model to understand the development and the survival of neuronal populations.

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Keywords:

Cerebellar granule neurons, development, differentiation, neuroprotection, survival pathways

Introduction

1 During the last decades the cerebellum has been studied in great detail, not only to understand its important
2 role in the control of motor coordination, but also because its laminated structure and limited number of cell
3 types have offered a model system to study neuronal development, differentiation and survival.
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6 Cerebellar granule neurons (CGNs) are the most abundant class of central nervous system neurons. Their
7 progenitors arise prenatally from the rhombic lip, in the boundary of the mesencephalon and metencephalon
8 (mes/met region), to form the external granule layer (EGL). In rodents, the neuroblasts of the EGL
9 proliferate during the first postnatal week. During the second postnatal week, neurons differentiate during
10 their migration through the molecular and Purkinje layers to reach their mature state in the internal granule
11 layer (IGL) by the end of the third postnatal week.¹ During their migration from the EGL towards the IGL,
12 CGNs neurons that fail to receive excitatory inputs from mossy fibres will die by apoptosis.^{2,3}
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15 16 **Extracellular factors involved in the proliferation, differentiation and survival of CGNs**

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18 The study of the mechanisms and factors involved in CGNs proliferation, differentiation and survival has
19 been facilitated by the possibility to maintain these neurons in culture in the presence of a depolarizing
20 medium. The combined use of CGNs primary cultures together with *in vivo* models has provided a deep
21 knowledge of the main factors sequentially involved in CGNs proliferation in germinal layers and their
22 differentiation and survival during migration to the IGL.
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26 Bone morphogenetic proteins (BMPs) seem to be the main factors for the generation of CGNs progenitors in
27 the rhombic lip. Work by Hatten and co-workers⁴ has shown that BMP6, BMP7 and Growth Differentiation
28 Factor 7 induce the expression of *Math1*, a transcription factor required for the specification of CGNs
29 progenitors in the prenatal cerebellum. Moreover, they demonstrated that BMP-treated explants from the
30 mes/met region were able to generate CGNs when transplanted to postnatal cerebellum. CGNs progenitors
31 proliferate postnatally in the EGL. Several factors have been implicated in their proliferation and in the exit
32 from the proliferative state to start differentiation. Numerous reports have shown that Sonic Hedgehog (Shh),
33 secreted by Purkinje cells, is the main factor mediating the proliferation of CGNs progenitors⁵, probably by
34 *Nmyc1* activation.⁶ However, other factors, such as insulin-like growth factor (IGF) 1 and 2, have also been
35 implicated in the maintenance of the proliferative state of CGNs progenitors. Activation of IGF1 receptor by
36 IGF1 and IGF2 enhances Shh-mediated proliferation of purified CGNs progenitors.⁷ The synergy between
37 Shh and IGFs could be related to the fact that *Igf2* expression is modulated by Shh.⁸ However, IGF1 and
38 IGF2 are still able to stimulate the proliferation of CGNs progenitors (although to a lesser extent) when Shh
39 signaling is impaired⁷ indicating that proliferation of CGNs progenitors in the EGL does not only depend on
40 the presence of pro-mitogenic factors such as Shh, and that other mechanisms are involved. In fact it is
41 known that transition from proliferation to differentiation in the EGL starts even in the continuous presence
42 of Shh. Thus, it should be the presence of other factors (anti-mitogens) that drives CGNs progenitors towards
43 differentiation. For example, IGF binding protein 5 is expressed when progenitors start to proliferate (second
44 postnatal week) and it blocks IGF and Shh-mediated proliferation of purified CGNs progenitors.⁷ Another
45 factor that has been reported to induce cell cycle exit in CGNs progenitors is BMP4. The expression levels of
46 BMP4 and Smad1 (an intracellular protein needed for BMP signaling) increase in the EGL during the second
47 postnatal week and BMP4 promotes neuronal cell differentiation in CGNs cultures.⁹ Other factors are also
48 known to promote CGNs differentiation such as pituitary adenylate cyclase-activating polypeptide (PACAP),
49 vitronectin, Wnt3 or activation of GPR3 receptors, which shows the complexity of the mechanisms involved
50 in this process at the end of the first postnatal week in the EGL.¹⁰⁻¹³
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1 Differentiation and maturation of CGNs start during the second postnatal week and occur in parallel to their
2 migration from the EGL towards the IGL.¹ During their migration, stimulation of glutamatergic synapses
3 between mossy fibres and CGNs are responsible for their survival.^{2,3} It has been reported *in vivo* that
4 activation of NMDA receptors (NMDAR) is necessary for CGNs survival since pharmacological blockade of
5 these receptors produce an increase in the apoptotic rate.¹⁴ Similar requirement has been observed *in vitro*.
6 Cultured CGNs die from apoptosis when potassium chloride (KCl) concentration changes from 25 mM
7 (K25) to 5 mM (K5). The addition of NMDA rescues CGNs from K5-mediated apoptosis.^{15,16} Moreover, it
8 has been reported that a 24 hr-exposure to NMDA in immature CGNs cultures (2 DIV) is enough to promote
9 a long-lasting neuroprotective effect since cells survive for up to 8 DIV in K5.¹⁷ Although different signal
10 transduction pathways have been related to the protective effect of NMDA on differentiating CGNs (see
11 below), several studies have shown that NMDA-dependent release of the neurotrophin BDNF is a major
12 factor in CGNs survival. Blocking TrkB receptor activation, by either antagonists or BDNF antibodies,
13 produces an important reduction in NMDA-mediated neuroprotection.¹⁷⁻¹⁹
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16 CGNs primary cultures have also been used to search other pro-survival factors. Interestingly, some of the
17 factors that are involved in the proliferation of progenitor cells were also related to survival of CGNs during
18 their migration towards the IGL. For example, IGF1 is involved in the proliferation of CGNs progenitors
19 (see above) but it also promotes the survival of CGNs, both *in vitro* and *in vivo*, by down-regulating pro-
20 apoptotic factors such as Bax, Bim and Bad, and up-regulating Bcl-x(L) and Bcl-2.²⁰⁻²² Also PACAP, which
21 inhibits the proliferation of progenitors in the EGL¹¹, acts as a pro-survival factor for CGNs.^{23,24} Altogether,
22 it seems that an orchestra of several extracellular factors plays an important, and complementary, role to
23 glutamatergic inputs from mossy fibres in the control of CGNs survival during their migration towards the
24 IGL. Then, we examine by which molecular mechanisms NMDA and the pro-survival factors induce the
25 survival of CGNs in the paradigm of KCl deprivation in cultured CGNs.
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31 **Molecular pathways related to the neuroprotection of CGNs**

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33 It has been described that specific protein kinase cascades involving phosphoinositide 3-kinase (PI3K)/Akt,
34 extracellular-signal regulated kinase (ERK), protein kinase A (PKA) or calcium/calmodulin kinase IV
35 (CaMKIV) promote the neuroprotective activity of neurotransmitters and pro-survival factors by the increase
36 of transcriptional activity. IGF-1 exerts its neuroprotective effect through the activity of PI3K²⁵ and the
37 phosphorylation of Akt.²⁶ The activation of PI3K/Akt decreases the activity of Forkhead transcription
38 factors, favoring the survival role of IGF-1.²² Other transcription factors, such as the family of Myocyte
39 enhanced factor-2, also participate in IGF-1-mediated survival effect.²⁷
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43 The requirement of PI3K/Akt pathway was also observed in the neuroprotective effect of NMDA in
44 immature¹⁷ and mature^{28,29} CGNs. But this effect is not restricted to the activation of PI3K/Akt pathway as
45 other protein kinases have also been involved. In mature CGNs, it has been showed that neuroprotection by
46 NMDA is mediated by ERK activation.²⁹ However, other studies reported that inhibition of ERK pathway
47 does not prevent the neuroprotective activity of NMDA.^{18,28} We recently reported that the survival role of
48 PI3K/Akt and ERK pathways in CGNs depends on the small G proteins acting upstream.³⁰ We demonstrated
49 that both pathways are involved in NMDA-mediated neuroprotection when they are activated by Ras. In
50 contrast, the stimulation of the ERK pathway by other members of the small G proteins family, such as
51 Rap1, is not sufficient to promote protection. Our results showed that the biological significance of NMDA-
52 mediated activity of PI3K/Akt and ERK pathway depends on which monomeric-G protein is acting
53 upstream. Furthermore, the requirement of Ras to activate these pathways also occurs for other protective
54 factors.³¹ Thus, these results support a central role for Ras in the survival of CGNs. NMDA-mediated
55 activation of PI3K/Akt and ERK pathways stimulates the activation of CREB, an important transcription
56 factor for the pro-survival effect of NMDA.³² As indicated above, the neuroprotective effect of NMDA is
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1 mediated by the release of BDNF.³³ We recently described that the interaction of CREB with the *Bdnf*
2 promoter is enhanced by potassium depolarization, whereas it is not improved by NMDA treatment.¹⁹ We
3 demonstrated that the stimulation of NMDAR triggers the CREB-dependent *Nurr1* activation, which results
4 in BDNF up-regulation. Moreover, we have characterized *Nurr1* as a key factor in NMDA-dependent
5 survival of CGNs. Other factors mediating the survival of CGNs are related to the activation of ERK-CREB
6 pathway. Interestingly, BMP-6 has been described to promote CGNs survival in culture through the
7 stimulation of MEK-ERK-CREB pathway.³⁴ Thus, differential pathway activation by BMP-6 could be
8 related to its function promoting the formation of CGNs progenitors (Smad-dependent) or the survival of
9 CGNs (Smad-independent) during cerebellum development.

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11 PACAP exerts a potent neuroprotective effect on cultured CGNs.^{35, 37} More recently, it has shown in mice
12 deficient for PAC1, the high-affinity PACAP receptor, that endogenous PACAP is crucial for the survival of
13 CGNs.³⁸ The neuroprotective effect of PACAP is mediated through the activation of ERK²⁴ and the cAMP-
14 dependent PKA^{39,40}. The increased activity of these pathways leads to the stimulation of prosurvival gene
15 expression, such as *c-fos* or *Bcl-2*.^{41,42} In addition, it has been described that PACAP mediates its
16 neuroprotective effect, in part, by inhibition of delayed rectifier K(+) current (I(K)) via cAMP/PKA
17 transduction pathway.^{43,44} Moreover, the survival effect of PACAP could be mediated by others pathways. It
18 has been demonstrated that PACAP promotes the increase of intracellular calcium from intracellular stores
19 and calcium influx through calcium channels.^{45,46} The mobilization of calcium allows the activation of
20 CaMKIV, and then the increase of CREB activity.^{47,48} More recently, it has been shown that PACAP-
21 mediated protective effect is also through the release of tissue plasminogen activator.⁴⁹

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23 The pivotal role of the Bcl-2 family proteins and the mitochondrial pathway in the apoptosis of CGNs has
24 been extensively documented. Bax and other members of the Bcl-2 family are sufficient to promote
25 apoptosis of CGNs.^{50,51} The reduction of these proteins is an important step in the neuroprotection of CGNs.
26 It has been shown that neuroprotection mediated by trophic factors is associated with the reduction of Bax,
27 Bad and Bim levels.^{21,22} On the other hand, the increased expression of anti-apoptotic members of Bcl-2
28 family has been related to the neuroprotective effect of IGF-1, PACAP, NMDA and others.^{21,42,52} The
29 regulation of the Bcl-2 members allows a reduction of the apoptotic mitochondrial activity, reducing
30 cytochrome c release^{52,53} and the activation of caspases.^{17,21,54,55} The inhibition of other apoptotic pathways is
31 also an important step in the survival of CGNs. For instance, the inhibition of JNK prevents the apoptotic
32 death of CGNs¹⁷, and the long-lasting neuroprotective effect of NMDA was also related to the inhibition of
33 JNK and the phosphorylation of c-jun.⁵² Taken together, the survival of CGNs mediated by NMDAR and
34 neurotrophin receptors results in the stimulation of common survival mechanisms and suppression of
35 apoptotic pathways.

36 37 38 39 40 41 42 43 44 45 **Conclusion**

46
47 The data presented here support that CGNs are a key model to decipher which factors and cellular
48 mechanisms underlie neuronal development and survival. Recent publications unraveled the action of
49 numerous neurotrophic factors and stimulation of NMDAR in promoting the activity of CGNs at postnatally
50 stages. Moreover, significant works revealed the similarity between these two, not only in the promotion of
51 the survival, but also in terms of the signaling pathways that they activate in CGNs.

52 53 54 55 **Acknowledgments**

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References

1. Altman J. Morphological development of the rat cerebellum and some of its mechanisms. *Exp Brain Res* 1982; Suppl.6:8-49.
2. Burgoyne RD., Cambray-Deakin MA. The cellular neurobiology of neuronal development: the cerebellar granule cell. *Brain Res* 1988;472:77-101.
3. Wood KA., Dipasquale B., Youle RJ. In situ labeling of granule cells for apoptosis-associated DNA fragmentation reveals different mechanisms of cell loss in developing cerebellum. *Neuron* 1993;11:621-32.
4. Alder J., Lee KJ., Jessell TM., Hatten ME. Generation of cerebellar granule neurons in vivo by transplantation of BMP-treated neural progenitor cells. *Nat Neurosci* 1999;2:535-40.
5. Dahmane N., Ruiz i Altaba A. Sonic hedgehog regulates the growth and patterning of the cerebellum. *Development* 1999;126:3089-100.
6. Kenney AM., Widlund HR., Rowitch DH. Hedgehog and PI-3 kinase signaling converge on Nmyc1 to promote cell cycle progression in cerebellar neuronal precursors. *Development* 2004;131:217-28.
7. Fernández C., Tatard VM., Bertrand N., Dahmane N. Differential modulation of Sonic-hedgehog-induced cerebellar granule cell precursor proliferation by the IGF signaling network. *Dev Neurosci* 2010;32:59-70.
8. Hahn H., Wojnowski L., Specht K., Kappler R., Calzada-Wack J., Potter D., et al. Patched target *Igf2* is indispensable for the formation of medulloblastoma and rhabdomyosarcoma. *J Biol Chem* 2000;275:28341-4.
9. Angley C., Kumar M., Dinsio KJ., Hall AK., Siegel RE. Signaling by bone morphogenetic proteins and *Smad1* modulates the postnatal differentiation of cerebellar cells. *J Neurosci* 2003;23:260-8.
10. Pons S., Trejo JL., Martínez-Morales JR., Martí E. Vitronectin regulates Sonic hedgehog activity during cerebellum development through CREB phosphorylation. *Development* 2001;128:1481-92.
11. Allais A., Burel D., Isaac ER., Gray SL., Basille M., Ravni A., et al Altered cerebellar development in mice lacking pituitary adenylate cyclase-activating polypeptide. *Eur J Neurosci* 2007;25:2604-18.
12. Tanaka S., Shaikh IM., Chiocca EA., Saeki Y. The Gs-linked receptor GPR3 inhibits the proliferation of cerebellar granule cells during postnatal development. *PLoS One* 2009;4:e5922.
13. Anne SL., Govak EE., Ayrault O., Kim JH., Zhu X., Murphy DA., et al. WNT3 inhibits cerebellar granule neuron progenitor proliferation and medulloblastoma formation via MAPK activation. *PLoS One* 2013;8:e81769
14. Monti B., Contestabile A. Blockade of the NMDA receptor increases developmental apoptotic elimination of granule neurons and activates caspases in the rat cerebellum. *Eur J Neurosci* 2000;12:3117-23.
15. Balázs R., Jorgensen OS., Hack N. N-methyl-D-aspartate promotes the survival of cerebellar granule cells in culture. *Neuroscience* 1988;27:437-51.
16. Moran J., Patel AJ. Stimulation of the N-methyl-D-aspartate receptor promotes the biochemical differentiation of cerebellar granule neurons and not astrocytes. *Brain Res* 1989 ;486 :15-25.
17. Xifro X., Malagelada C., Miñano A., Rodríguez-Alvarez J. Brief exposure to NMDA produces long-term protection of cerebellar granule cells from apoptosis. *Eur J Neurosci* 2005;21:827-40.
18. Bhave SV., Ghoda L., Hoffman PL. Brain-derived neurotrophic factor mediates the anti-apoptotic effect of NMDA in cerebellar granule neurons: signal transduction cascades and site of ethanol action. *J Neurosci* 1999;19:3277-86.
19. Barneda-Zahonero B., Servitja JM., Badiola N., Miñano-Molina AJ., Fadó R., Saura CA., et al. *Nurr1* protein is required for N-methyl-D-aspartic acid (NMDA) receptor-mediated neuronal survival. *J Biol Chem* 2012;287:11351-62.

20. D’Mello SR., Galli C., Ciotti T., Calissano P. Induction of apoptosis in cerebellar granule neurons by low potassium: inhibition of death by insulin-like growth factor I and cAMP. *Proc Natl Acad Sci USA* 1993;90:10989-93.
21. Chrysis D., Calikoglu AS., Ye P., D’Ercole AJ. Insulin-like growth factor-I overexpression attenuates cerebellar apoptosis by altering the expression of Bcl family proteins in a developmentally specific manner. *J Neurosci* 2001;21:1481-9.
22. Linseman DA., Phelps RA., Bouchard RJ., Le SS., Laessig TA., McClure ML., et al. Insulin-like growth factor-I blocks Bcl-2 interacting mediator of cell death (Bim) induction and intrinsic death signaling in cerebellar granule neurons. *J Neurosci* 2002;22:9287-97.
23. Gonzalez BJ., Basille M., Vaudry D., Fournier A., Vaudry H. Pituitary adenylate cyclase-activating polypeptide promotes cell survival and neurite outgrowth in rat cerebellar neuroblasts. *Neuroscience* 1997;78:419-30.
24. Villalba M., Bockaert J., Journot L. Pituitary adenylate cyclase-activating polypeptide (PACAP-38) protects cerebellar granule neurons from apoptosis by activating the mitogen-activated protein kinase (MAP kinase) pathway. *J Neurosci* 1997;17:83-90.
25. D’Mello SR., Borodezt K., Soltoff SP. Insulin-like growth factor and potassium depolarization maintain neuronal survival by distinct pathways: possible involvement of PI 3-kinase in IGF-I signaling. *J Neurosci* 1997;17:1548-60.
26. Dudek H., Datta SR., Franke TF., Birnbaum MJ., Yao R., Cooper GM., et al. Regulation of neuronal survival by the serine-threonine protein kinase Akt. *Science* 1997;275:661-5.
27. Wiedmann M., Wang X., Tang X., Han M., Li M., Mao Z. PI3K/Akt-dependent regulation of the transcription factor myocyte enhancer factor-2 in insulin-like growth factor-1- and membrane depolarization-mediated survival of cerebellar granule neurons. *J Neurosci Res* 2005;81:226-34.
28. Zhang FX., Rubin R., Rooney TA. N-Methyl-D-aspartate inhibits apoptosis through activation of phosphatidylinositol 3-kinase in cerebellar granule neurons. A role for insulin receptor substrate-1 in the neurotrophic action of n-methyl-D-aspartate and its inhibition by ethanol. *J Biol Chem* 1998;273:26596-602.
29. Lafon-Cazal M., Perez V., Bockaert J., Marin P. Akt mediates the anti-apoptotic effect of NMDA but not that induced by potassium depolarization in cultured cerebellar granule cells. *Eur J Neurosci* 2002;16:575-83.
30. Xifró X., Miñano-Molina AJ., Saura CA., Rodríguez-Álvarez J. Ras protein activation is a key event in activity-dependent survival of cerebellar granule neurons. *J Biol Chem* 2014;289:8462-72.
31. Obara Y., Horgan AM., Stork PJ. The requirement of Ras and Rap1 for the activation of ERKs by cAMP, PACAP and KCl in cerebellar granule cells. *J Neurochem* 2007;101:470-82.
32. Monti B., Marri L., Contestabile A. NMDA receptor-dependent CREB activation in survival of cerebellar granule cells during in vivo and in vitro development. *Eur J Neurosci* 2002;16:1490-8.
33. Marini AM., Rabin SJ., Lipsky RH., Mocchetti I. Activity-dependent release of brain-derived neurotrophic factor underlies the neuroprotective effect of N-methyl-D-aspartate. *J Biol Chem* 1998;273:29394-9.
34. Barneda-Zahonero B., Miñano-Molina AJ., Badiola N., Fadó R., Xifró X., Saura CA., et al. Bone morphogenetic protein-6 promotes cerebellar granule neurons survival by activation of the MEK/ERK/CREB pathway. *Mol Biol Cell* 2009;20:5051-63.
35. Journot L., Villalba M., Bockaert J. PACAP-38 protects cerebellar granule cells from apoptosis. *Ann N Y Acad Sci* 1998;865:100-10.
36. Botia B., Basille M., Allais A., Raoult E., Falluel-Morel A., Galas L., et al. Neurotrophic effects of PACAP in the cerebellar cortex. *Peptides* 2007;28:1746-52.
37. Falluel-Morel A., Aubert N., Vaudry D., Desfeux A., Allais A., Burel D., et al. Interactions of PACAP and ceramides in the control of granule cell apoptosis during cerebellar development. *J Mol Neurosci* 2008;36:8-15.

38. Falluel-Morel A., Tascau LI., Sokolowski K., Brabet P., DiCicco-Bloom E. Granule cell survival is deficient in PAC1^{-/-} mutant cerebellum. *J Mol Neurosci* 2008;36:38-44.
39. Chang JY., Korolev VV., Wang JZ. Cyclic AMP and pituitary adenylate cyclase-activating polypeptide (PACAP) prevent programmed cell death of cultured rat cerebellar granule cells. *Neurosci Lett* 1996 ;206 :181-4.
40. Campard PK., Crochemore C., Rene F., Monnier D., Koch B., Loeffler JP. PACAP type I receptor activation promotes cerebellar neuron survival through the cAMP/PKA signaling pathway. *DNA Cell Biol* 1997;16:323-33.
41. Vaudry D., Gonzalez BJ., Basille M., Anouar Y., Fournier A., Vaudry H. Pituitary adenylate cyclase-activating polypeptide stimulates both c-fos gene expression and cell survival in rat cerebellar granule neurons through activation of the protein kinase A pathway. *Neuroscience* 1998;84:801-12.
42. Aubert N., Falluel-Morel A., Vaudry D., Xifró X., Rodríguez-Álvarez J., Fisch C., et al. PACAP and C2-ceramide generate different AP-1 complexes through a MAP-kinase-dependent pathway: involvement of c-Fos in PACAP-induced Bcl-2 expression. *J Neurochem* 2006;99:1237-50.
43. Mei Y.A., Vaudry D., Basille M., Castel H., Fournier A., Vaudry H., et al. PACAP inhibits delayed rectifier potassium current via a cAMP/PKA transduction pathway: evidence for the involvement of I_k in the anti-apoptotic action of PACAP. *Eur J Neurosci* 2004;19:1446-58.
44. Castel H., Vaudry D., Mei Y.A., Lefebvre T., Basille M., Desrues L., et al. The delayed rectifier channel current I_k plays a key role in the control of programmed cell death by PACAP and ethanol in cerebellar granule neurons. *Ann N Y Acad Sci* 2006;1070:173-9.
45. Tabuchi A., Koizumi M., Nakatsubo J., Yaguchi T., Tsuda M. Involvement of endogenous PACAP expression in the activity-dependent survival of mouse cerebellar granule cells. *Neurosci Res* 2001;39:85-93.
46. Basille-Dugay M., Vaudry H., Fournier A., Gonzalez B., Vaudry D. Activation of PAC1 Receptors in Rat Cerebellar Granule Cells Stimulates Both Calcium Mobilization from Intracellular Stores and Calcium Influx through N-Type Calcium Channels. *Front Endocrinol* 2013;4:56.
47. Sée V., Boutillier AL., Bito H., Loeffler JP. Calcium/calmodulin-dependent protein kinase type IV (CaMKIV) inhibits apoptosis induced by potassium deprivation in cerebellar granule neurons. *FASEB J* 2001;15:134-44.
48. Kokubo M., Nishio M., Ribar TJ., Anderson KA., West AE., Means AR. BDNF-mediated cerebellar granule cell development is impaired in mice null for CaMKK2 or CaMKIV. *J Neurosci* 2009;29:8901-13.
49. Raoult E., Roussel BD., Bénard M., Lefebvre T., Ravní A., Ali C., et al. Pituitary adenylate cyclase-activating polypeptide (PACAP) stimulates the expression and the release of tissue plasminogen activator (tPA) in neuronal cells: involvement of tPA in the neuroprotective effect of PACAP. *J Neurochem* 2011;119:920-31.
50. Miller TM., Moulder KL., Knudson CM., Creedon DJ., Deshmukh M., Korsmeyer SJ., et al. Bax deletion further orders the cell death pathway in cerebellar granule cells and suggests a caspase-independent pathway to cell death. *J Cell Biol* 1997;139:205-17.
51. Zhokhov SS., Desfeux A., Aubert N., Falluel-Morel A., Fournier A., Laundenbach V., et al. Bax siRNA promotes survival of cultured and allografted granule cell precursors through blockade of caspase-3 cleavage. *Cell Death Differ* 2008;15:1042-53.
52. Xifró X., Falluel-Morel A., Miñano A., Aubert N., Fadó R., Malagelada C., et al. N-methyl-D-aspartate blocks activation of JNK and mitochondrial apoptotic pathway induced by potassium deprivation in cerebellar granule cells. *J Biol Chem* 2006;281:6801-12.
53. Falluel-Morel A., Aubert N., Vaudry D., Basille M., Fontaine M., Fournier A., et al. Opposite regulation of the mitochondrial apoptotic pathway by C2-ceramide and PACAP through a MAP-kinase-dependent mechanism in cerebellar granule cells. *J Neurochem* 2004;91:1231-43.

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54. Vaudry D., Cottet-Rousselle C., Basille M., Falluel-Morel A., Fournier A., Vaudry H., et al. Pituitary adenylate cyclase-activating polypeptide inhibits caspase-3 activity but does not protect cerebellar granule neurons against beta-amyloid (25-35)-induced apoptosis. *Regul Pept* 2004;123:43-9.
55. Subramaniam S., Shahani N., Strelau J., Laliberté C., Brandt R., Kaplan D., et al. Insulin-like growth factor 1 inhibits extracellular signal-regulated kinase to promote neuronal survival via the phosphatidylinositol 3-kinase/protein kinase A/c-Raf pathway. *J Neurosci* 2005;25:2838-52.

10 **Figure legends**

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12 **Figure 1: Extracellular factors involved in the proliferation, differentiation and protection of CGNs at postnatal stages.** Main factors introduced in the text involved in the proliferation (prolifer.), differentiation (differ.) and protection (antiapop.) of CGNs at different postnatal stages, from the external granule layer (EGL) to the internal granule layer (IGL). Dashed arrow indicates the migration of CGNs from EGL to IGL. In brackets, the action of glutamate is through the stimulation of NMDA receptor (NMDAR). ML indicates molecular layer, PL, Purkinje layer and WM, white matter.

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21 **Figure 2: Cellular mechanisms involved in the neuroprotection of CGNs.** We introduced the signaling pathways involved in the survival of CGNs promoted by some neurotrophins, neuropeptides and the NMDAR stimulation in the paradigm of potassium deprivation. Bold arrows indicate activation and dashed arrows denote inhibition.

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FIGURE 1

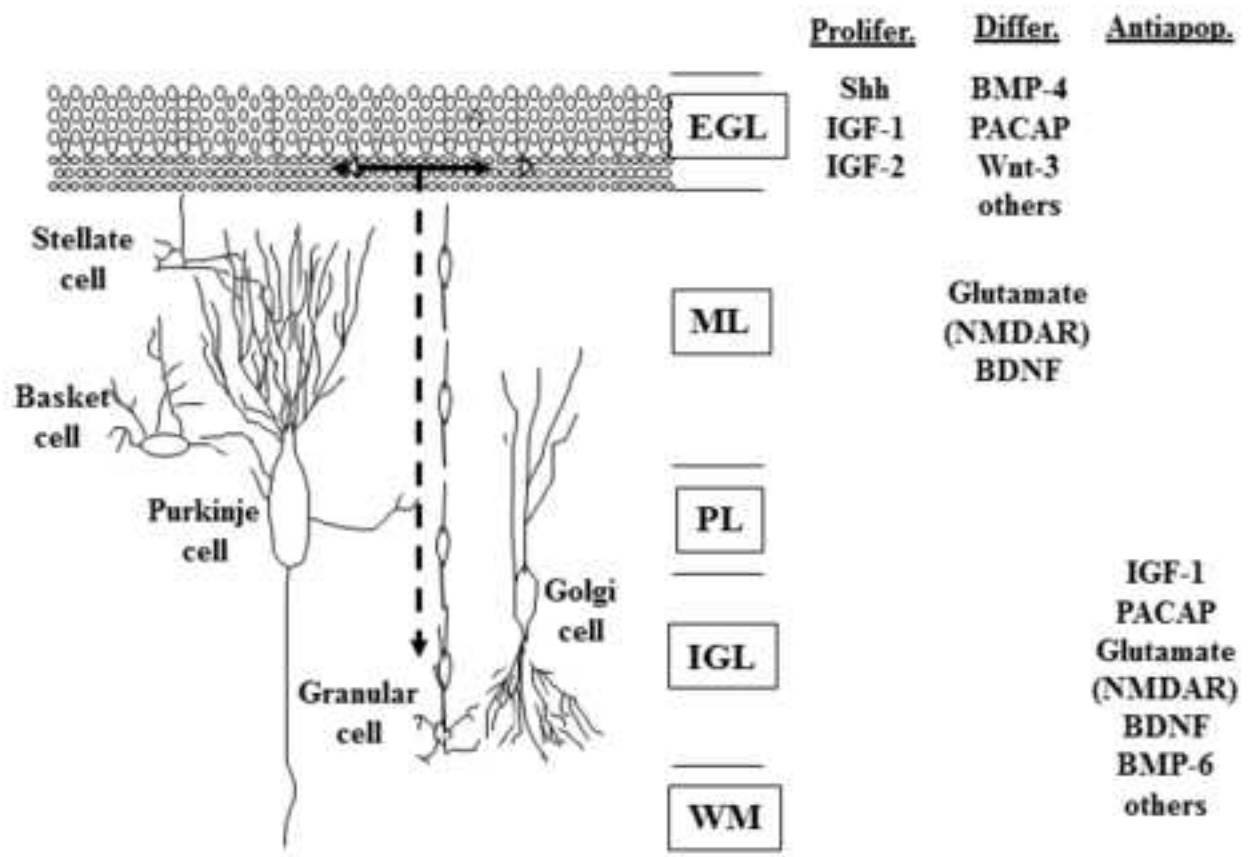


FIGURE 2

