

PATHOGENICITY OF A BOVINE VIRAL DIARRHOEA VIRUS STRAIN IN PREGNANT SOWS: SHORT COMMUNICATION

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The biological properties of bovine viral diarrhoea virus (BVDV) strain Oregon C24V were studied after intranasal and subcutaneous infection of pregnant sows. This virus strain is widely used in Hungary for immunising cattle against bovine viral diarrhoea (BVD). Based upon the results of the clinical, gross pathological, histopathological and virological examinations it can be established that the given strain caused asymptomatic infection and serological conversion in sows that were in the second third of gestation. The virus caused clinically apparent disease in some of the piglets born at term, which indicates that it had crossed the placenta. More than half (57%) of the live-born piglets died within 60 days of birth. The sows and their progeny did not shed the virus. BVDV infection has great differential diagnostic importance in pigs, as classical swine fever (CSF) virus strains of reduced virulence cause similar clinical symptoms and gross and histopathological changes.

Key words: Pestivirus, classical swine fever virus, bovine viral diarrhoea virus, pregnant sow, fetopathic effect

Bovine viral diarrhoea virus (BVDV) belongs to the *Pestivirus* genus of the *Flaviviridae* family, together with classical swine fever virus (CSFV) and border disease virus (BDV) of sheep.

The infection of pigs with BVDV under natural conditions was first described in Australia (Flynn and Jones, 1964), then it was reported from the Netherlands, Germany, France, Great Britain, Denmark and the United States. Serological surveys indicate that the prevalence of infection in pig herds varies between 3 and 40%, depending on the age of the animals and the degree of their contact with cattle (Stewart et al., 1971; Carbrey et al., 1976; Terpstra and Wensvoort, 1988; Vannier and Leforban, 1992). Cattle shedding the virus serve as the main source of infection, but different live virus vaccines contaminated with BVDV may also transmit the virus. In 1973, the virus was successfully is o-

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lated from swine (Fernelius et al., 1973). Clinical symptoms usually do not develop after the infection, but in pregnant sows occasionally a fetopathic effect may occur. The newborn piglets are weak, retarded in growth and show diarrhoea and respiratory symptoms. Affected animals usually die within a few weeks (Carbrey et al., 1976; Terpstra and Wensvoort, 1991; Liess and Moennig, 1990; Terpstra and Wensvoort, 1991; Paton and Done, 1994). The condition could not be consistently reproduced by the experimental infection of pregnant sows (Fernelius et al., 1973; Mengeling, 1988; Leforban et al., 1990). The consequences of infection depend on the infective virus strain and the stage of gestation of the sow (Vannier and Leforban, 1992). The NADL strain did not cause disease in piglets, while the Singer strain adapted to cell lines of porcine origin were found to be severely fetopathic (Mengeling, 1988; Leforban et al., 1990). The Oregon C24V strain induced an antibody response in piglets, which protected them against CSFV infection (Snowdon and French, 1968). Other authors (Simonyi and Bíró, 1967) found that the antibodies elicited by the Oregon C24V strain did not protect the animals against CSFV infection. The most severe reproductive symptoms develop if the sows are infected between day 25 and 41 of gestation (Mengeling, 1988; Leforban et al., 1990).

Due to the close antigenic relatedness of pestiviruses, BVDV gives cross-reaction with CSFV in conventional serological tests. A further diagnostic difficulty results from the fact that by clinicopathological examinations the disease produced by CSFV strains of reduced virulence cannot be distinguished from the condition caused by BVDV (Loken, 1995).

In this study, the clinical signs, gross and histopathological changes and the kinetics of virus shedding and the immune response were studied in pregnant sows experimentally infected with BVDV by the intranasal and subcutaneous route.

Materials and methods

Four Hungarian Large White sows being in the second third of gestation (days 54–73) and free from virus-neutralising antibodies to CSFV and BVDV were used in the experiment. Two sows were inoculated with BVDV strain Oregon C24V: one of them received 2000 TCID₅₀ virus intranasally while the other one subcutaneously. Two sows were kept isolated as uninfected negative controls.

The animals were observed daily for clinical signs and their rectal temperature was recorded. After infection, nasal and rectal swabs as well as urine and blood samples were taken from the sows and the piglets weekly over a period of 16 weeks.

The animals that died during the experiment and those killed in week 16 after experimental infection were examined for gross pathological changes, and organ samples (spleen, lungs, kidney, brain, tonsil, mesenteric and submandibular

lar lymph node) were collected from them for histopathological examination. The samples were fixed in 10% neutral buffered formalin, embedded in paraffin, and the sections were stained with haematoxylin and eosin.

Detection of antibodies to BVDV virus in the serum was performed by the virus neutralisation test, in secondary calf testicle cell cultures, using the Oregon C24V strain.

Virus isolation was done in secondary calf testicle cell cultures.

Results and discussion

Both infected sows remained clinically healthy throughout the period of observation, and did not shed the virus. From postinfection week 4 up to the end of the experiment, virus-neutralising antibodies to BVDV could be detected in a titre of 1:80–1:240 in both animals. Gross pathological changes were not found in the sows killed at the end of the experiment. The histopathological findings included focal interstitial nephritis and follicular hyperplasia in the lymphoid organs.

The intranasally infected sow delivered 13 live piglets on day 115 of gestation. In that litter, eight piglets were weak and had ruffled hair coat and showed splayleg, trembling, myoclonus, diarrhoea, and fever (40.2–40.9 °C). The affected piglets died within 40 days of birth. The subcutaneously infected sow delivered eight live-born and one stillborn fetuses on day 113 of gestation. Four out of the live-born fetuses fell ill, showing clinical signs similar to those described above, and died within 60 days of birth. Attempts to isolate virus from the piglets failed. The serum of all piglets contained antibodies to BVDV in titres gradually decreasing over time. In the 8th week of life, the piglets of the intranasally infected sow no longer had serum antibodies in titres detectable by the VN test. In the piglets of the subcutaneously infected sow the antibodies had a higher baseline titre (1:240) and persisted longer: they were still detectable in week 9 after birth.

In the live-born piglets that subsequently died there were petechial haemorrhages at the predilection sites and lymphoid hyperplasia in the spleen and lymph nodes. Three dead piglets had interstitial pneumonia and acute gastroenteritis. The clinically healthy piglets euthanised at the end of the experiments were free from gross and histopathological changes.

The negative control sows and their offspring remained healthy throughout the experiment.

In this study, sows being in the second third of gestation were successfully infected by the subcutaneous and intranasal route with a pestivirus of ruminant origin, strain Oregon C24V used also for vaccine production. Successful infection was confirmed serologically. The virus crossed the placenta and caused clinically apparent disease in the progeny. Perinatal mortality was substantial: 57% of the live-born piglets died. The sows and their piglets did not shed the virus, which ob-

ervation is consistent with the findings of Leforban et al. (1992). The decreasing antibody titre suggests that the piglets did not develop active immunity; rather, antibodies of maternal origin were detected in their serum.

The clinical symptoms and gross pathological changes found in the piglets are consistent with the symptoms and lesions produced by CSFV strains of reduced virulence. This fact underscores the differential diagnostic importance of BVDV infection.

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