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Phytoplankton structure, biochemical, stoichiometry and elemental composition in Lake Nasser, Egypt.

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Abstract

The relationship between environmental variables and elemental, biochemical composition of phytoplankton species in Lake Nasser were studied. In the lake 130 phytoplankton species were recorded, the lake is dominated mainly by *Cyclotella glomerata* (Bacillariophyceae), *Planktolyngbya limnetica*, *Eucapsis minuta* (Cyanophyceae). Phytoplankton organic carbon content at the lake was 32%. The elevation of NO₃ (365 µg/l) and CO₃ (21 mg/l) at the east of the lake was concurrently with the highest proportion of inorganic C, N and algal protein content. C/N ratio decreased thus Redfield ratio indicating a high growth rate of phytoplankton with increasing protein content. C/H and O/C ratio for natural phytoplankton samples were less than one, which means unsaturated aliphatic compounds at metabolites categories of phytoplankton. RDA analysis cleared that, *Cyclotella glomerata* (dominant at the north area) was sensitive to flushing, tolerant to nutrient deficiency and had higher protein content. *Planktolyngbya limnetica* (dominant at the middle) was more sensitive to pH, phosphorus is non-limiting factor and characterized by elevation in lipid content. *Eucapsis minuta* (dominant at the south) was tolerant to mild light deficiency and contains a maximum value of carbohydrates and Chlorophyll *a*.

Keywords: Algae, FGs codons, Stoichiometry,

Introduction

Lake Nasser represents the main national freshwater reservoir of Egypt and the main strategic importance for sustaining Egypt's water policy [1]. Phytoplankton has been used as indicators of environmental [67] and climatic changes [44]. It constitutes the basis of food chain in aquatic environment [49]. Phytoplankton growth is regulated by essential resources such as light and nutrients [23]. Phytoplankton communities in Lake Nasser are rich with density and biomass [41] and increases to southwards [10]. The taxonomic group of bacillariophyceae was the most important group and key players in the global carbon cycle in Lake

Nasser [70], followed by cyanophyceae [29]. Phytoplankton classes varied significantly in their content of protein, carbohydrate and lipid. Chlorophyceae and Prasinophyceae were normally richer in carbohydrate than species from other classes [13]. Carbon is the principal structural component of phytoplankton ($24\pm 80\%$ of dry weight) and used as standard element in quantifying plant biomass production and bioenergetics [6]. The other important macroelements in algae are nitrogen and phosphorous, which are the main components for the growth rate process [66]. There are few studies concerning the qualitative and quantitative information of the elements in algae and what their role in the different species [32]. The elemental composition of microalgae is a fusion of the evolutionary history of plastid, resulting in modifications in genetic constraints and selection pressures associated with environmental variables [51]. Ecological stoichiometry shows that not only the carbon content, but also the carbon-to-nutrient ratio is important in producing high plant biomass [61]. Algal food quality and its biochemical content may also vary with algal stoichiometry [24].

The objective of this study was to determine how the elemental composition of phytoplankton and deviations from the Redfield ratio at Lake Nasser changed due to phytoplankton structure. Investigate the effect of physico-chemical variables on dominant phytoplankton species and its biochemical content in the littoral area of the Lake.

Material and Methods

Lake Nasser is a vast reservoir in southern Egypt and one of the largest man-made lakes in the world [1]. The Lake extends about 350 km are within the Egyptian territory between $22^{\circ} 00'$ to $23^{\circ} 58'$ N and $31^{\circ} 19'$ to $33^{\circ} 15'$ E. The total surface area of the Lake is 5237 km^2 at 180 m over mean Sea level [4] and [35]. The length of eastern shoreline is almost double that of the western shoreline [40] and [42] (Fig.1.) Seasonally, water samples were collected using 1.5 L Ruttner sampler from 9 sites at both east (E) and west (W) area of Lake Nasser during 2013 (Table 1, Fig.1). Chemical analysis and phytoplankton differentiation were determined at each sample subsurface (S) and near bottom (B). Biochemical contents of phytoplankton and stoichiometry were subsurface.



Figure 1: The location of the selected littoral sites in Lake Nasser at the north (N), middle (M) and south (S) part of the lake.

Table 1: The sampling locations in Lake Nasser.

Code	Sites	location	Latitude	Longitude	Distance from High Dam (Km)
1	High Dam		23° 56.24'	32° 51.89'	2.9
2	Dahmit	North	23° 51.1'	32° 53.78'	21
3	Kalabsha		23° 33.41'	32° 52.04'	47.7
4	Wadi-Abyad		23° 20.08'	32° 56.12'	74.3
5	El-Madiq	Middle	22° 56.47'	32° 37.68'	130.1
6	Korosko		22° 37.65'	32° 24.14'	167.2
7	Amada		22° 43.97'	32° 5.45'	199
8	Tushka	South	22° 36.32'	31° 55.21'	240
9	Abu-Simble		22° 19.71'	31° 37.14'	268.8

Electrical conductivity (mS cm^{-1}) was measured by using a conductivity meter (S.C.T.33 YSI), Transparency (cm) by Secchi disc and water temperature by an ordinary thermometer. pH by using Orion Research Ion Analyzer 399A pH meter.

CO_3^{2-} and HCO_3^- were measured titrimetrically against standard H_2SO_4 (0.02 N) using phenolphthalein and methyl orange indicators. Dissolved oxygen (DO) was determined by azide modification method. Concentrations of nitrite (NO_2), nitrate (NO_3), ammonium (NH_4) and orthophosphate (PO_4) in water samples were determined using the colorimetric techniques [8].

For phytoplankton analysis water samples collected from the Ruttner sampler (500 mL) was preserved in 4% neutral formalin [43]. Uttermohl method was applied for counting (until 400 counted individual) and identification of phytoplankton species [9] using inverted microscope (ZEISS 1M4738). The main references used in identification were: [33, 22, 12, 60, 50, 62 and 19].

For biochemical analysis water sample was sieved and filtered through zooplankton net (100 μm mesh size) to separate macrozooplankton. Defined volume of water sample was refiltered on glass microfiber filter GF/F. Biuret method [20] was used for protein analysis which involves the use of a Biuret reagent, subsequent absorbance measurements at 550 nm and comparison to a bovine standard absorbance at the same wavelength. Total carbohydrate content was determined by the sulphuric acid hydrolysis method, glucose as a standard [25]. Lipid extraction was performed using Sulphophosphovanillin procedure [16]. Chlorophyll *a* (Chl. *a*) was extracted in acetone (90%) overnight, prepared and measured spectrophotometry and calculated applying trichromatic equation [8]. Trophic state index was calculated according to [14]. The elemental composition was based on the calculations of [28].

Total caloric content was calculated on the proximate biochemical composition according to [68], by the relative caloric coefficients (5.56 for proteins, 4.1 for carbohydrates and 9.45 for lipids).

Statistical analysis: Redundancy analysis (RDA) was used as method to extract and summaries the variation in between biotic factors as dominant species, biochemical content and elemental composition of phytoplankton and abiotic physico-chemical parameters in the different sites. RDA is a direct gradient analysis technique which summarises linear relationships between components of response variables. Data were analyzed using CANOCO 5.0 [64] and Statistica 8.0.

Results and Discussion

The aquatic environment as physical and chemical characteristics of Lake Nasser has been depending on flood events. During the last few decades, there has been much interest in the routes influencing the development of phytoplankton populations, primarily in relation to physico-chemical factors [45]. The chemistry of water describes the general hydrobiological relationship [59]. Water in the east side of the Lake was more transparent than the west one and transparency was gradually decreased southward in both sides (Table 2). The southern sites were generally affected by the turbidity or abundant suspended organic materials than the northern sites. Water temperature fluctuated between 20.5 °C to 25.5 °C at east side of the lake

and from 18.7 °C to 26.3 °C at the west side (Table 2). Temperature has a significant positive correlation with Chl. *a* (R= 0.42). Light transparency and temperature are the most important physical variables controlling on the aquatic environment [3]. pH is normally alkaline with a slight local variations at the lake and it increased in the bottom layers than the surface. pH was correlated with Chl. *a* and Cyanophyceae abundance (r =0.23 and 0.56, P < 0.001). Gharib and [29] also showed that there was a close linear correlation between pH, DO and Chl. *a*. The highest conductivity value of 226 µS/cm was recorded in the north at the west area. This result coincided with [65]. DO fluctuate from 4.4 to 6.8 mg l⁻¹ at both sides during the study period. Alkalinity of the lake was within the allowable limits of Bureau of Indian Standards BIS (200 mg/lit) [48]. N:P ratios give valuable information on nutrient limitation. N: P < 7 was most frequently noted in the Lake water, indicating N is the limiting phytoplankton growth [69].

Table 2: Annual average of physico-chemical variables studied in the littoral area of the Lake.

Parameters	Littoral area	Sites									
		North			Middle			South			
		St_1	St_2	St_3	St_4	St_5	St_6	St_7	St_8	St_9	
Physical parameters	Temp.	East	23.6	22.5	24.3	24.7	20.5	23.9	25.5	25.4	25.0
		West	23.4	23.5	23.4	24.5	21.9	25.6	26.3	24.7	18.7
	pH	East	7.9	8.0	8.2	8.3	8.3	8.4	8.6	8.4	8.3
		West	8.1	8.1	8.3	8.1	8.4	8.5	8.5	8.3	8.5
	EC	East	218	215	210	205	206	205	212	210	200
		West	226	208	208	205	201	210	212	209	
Chemical parameters	Trans. Cm	East	313	385	345	348	295	215	208	250	180
		West	363	357	353	320	245	223	167	230	260
	PO ₄ (µg L ⁻¹)	East	78	84	79	71	23	24	39	39	115
		West	74	73	67	71	20	34	35	95	35
	NO ₂ (µg L ⁻¹)	East	14	38	9	5	7	9	16	16	31
		West	12	10	12	4	5	9	21	26	60
	NH ₄ (µg L ⁻¹)	East	73	105	65	48	67	77	58	73	42
		West	151	94	162	38	83	162	47	93	41
	NO ₃ (µg L ⁻¹)	East	213	151	211	365	323	238	295	286	323
		West	358	198	155	198	334	402	453	350	518
	CO ₂ (mg L ⁻¹)	East	21	12	19	11	19	20	16	14	14
		West	11	17	16	12	10	14	9	10	13
	HCO ₃ (mg L ⁻¹)	East	120	115	110	113	99	95	111	110	107
		West	132	115	127	109	102	112	112	110	108
	DO (mg L ⁻¹)	East	5.0	4.4	4.7	5.3	5.3	5.0	5.5	5.9	6.1
		West	5.3	4.9	4.7	4.6	5.2	5.1	5.8	5.5	6.8

Phytoplankton was chosen as the most sensitive indicator of changing nutrient conditions because of the potential rapidity of population responses [15] and plays a significant role in maintaining the biological balance in the aquatic environment [63]. Microscopic examination of phytoplankton during study period revealed that a total of 130 species were obtained in the littoral area of the Lake and belonging to 7 classes. Phytoplankton community structure is mainly composed of Bacillariophyceae (42 sp.) representing about 51.7% and Cyanophyceae (37 sp.) constituting 31.7% then Chlorophyceae (36 sp.) forming 13.4%, this result coincided with [2]. Cyanophyceae was observed during the study as first predominant class in summer [30]. While, Dinophyceae (6 sp.), Chrysophyceae (5 sp.), Creptophyceae (3 sp.) and Euglenophyceae (1 sp.) persisted as frequent or rare forms. Bacillariophyceae density was higher in the east than the west side, in reversely with Cyanophyceae and Chlorophyceae. Bacillariophyceae was positively associated with NO_3 ($R=0.17$). [5] stated that Bacillariophyceae are favoured in nutrients rich environment mainly nitrates. Growing of diatoms are declining with water temperature and PO_4 where $R=-0.43$ and -0.24 , respectively [55]. Phytoplankton assemblage structure are considered as important indicators and widely used for ecological status assessment [38]. The highest phytoplankton densities at the east area was 1800×10^4 cell/L in winter and reduced to 35×10^4 cell/L in spring, while the maximum in the west was 1475×10^4 cell/L in winter and minimized to 25×10^4 cell/L in spring. Seasonal phytoplankton crop reached its maxima (751×10^4 cell/L) in the west area during winter. The highest annual value of 530×10^4 cell/L was noted at middle area of the east. Annual trend line of phytoplankton crop increase southward in the surface water layer, but it reverse in the bottom layer as shown in Fig (2). Regional variation of phytoplankton reflects its response to changing environmental conditions and grazing [17]. The highest species diversity (87 spp) was recorded during spring while the lowest (45 spp) was obtained during autumn. Phytoplankton species number decreased to half and responds rapidly to changes in the aquatic environment [34]. Phytoplankton was more abundant in the littoral area than in the middle of the open area [37] in press.

Most dominant phytoplankton species were identified in to functional groups (FGs), named alphanumerical codes, according to their sensitivities and tolerances as described by [4, 54, 56 and 46], with its percentage abundance in the two littoral area of the Lake (Table 3).

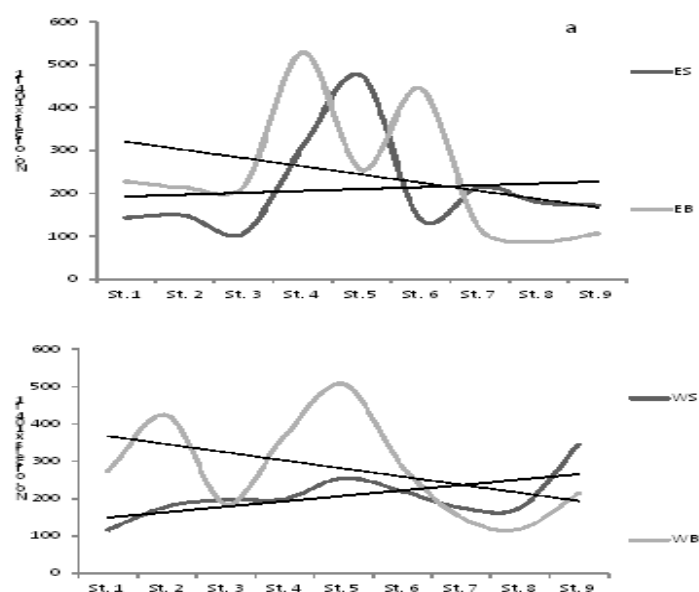


Figure 2ab: Temporal, spatial distributions of phytoplankton crop at the littoral area of Lake Nasser. ES: East surface, EB: east bottom WS: west surface, WB: west bottom.

Table 3: Trait of the dominant phytoplankton functional groups according to their sensitivities and tolerances (Reynolds et al., 2002; Reynolds 2006 and Padisák et al. 2009) and their percentage in the different zones of the Lake

Code	Dominant phytoplankton species in Lake Nasser	Annual percentage in the two littoral area		Tolerances	Sensitivities
		Eastern Lake side	Western Lake side		
A	<i>Cyclotella glomerata</i>	25.6%	25.2%	Nutrient deficiency	pH rise
D	<i>Cyclotella ocellata</i>	6.7%	6.9%	Flushing	Nutrient depletion
	<i>Aulacoseira granulata</i>	13.9%	10.6%	Mild light and C deficiency	Stratification Si depletion
P	<i>Stauroneis parallelum</i>	7.8%	15.1%	Mild light and C deficiency	Stratification Si depletion
XI	<i>Actina rodanensis fusiformis</i>	16.4%	17%	Stratification	Nutrient depletion
S1	<i>Planktolyngbya limnetica</i>	58.3%	39.4%	Highly light deficient	Flushing
L0	<i>Merismopedia punctata</i>	-	12.5%	Segregated nutrients	Prolonged or deep mixing
N	<i>Cosmarium</i> sp.	5%	-	Nutrient deficiency	Stratification, pH rise
	<i>Coclastrum microborum</i>	7%	-	-	Settling into low light
	<i>Scenedesmus quadricauda</i>	-	12.7%	-	Settling into low light
	<i>Pediastrum simplex</i>	3.3%	-	-	Settling into low light
J	<i>Cocystis zoea toris</i>	7.2%	-	Low nutrients High turbidity	CO ₂ deficiency
	<i>Dityosphaerium pulchellum</i>	4.1%	6.7%	Low nutrients High turbidity	CO ₂ deficiency

Cellular composition is used as an important biomarker tool in clarifying the taxonomy of algae [36]. Protein peak was found at the north of the two littoral areas where *Cyclotella glomerata* (Bacillariophyceae) represent the dominant species 48.6 %. Annual trend line of protein refers to its contents were decreased southward in the studied region (Fig. 3). *Cyclotella cryptica* contain 260 mg/g protein used as an alternative feed for aquaculture [47].

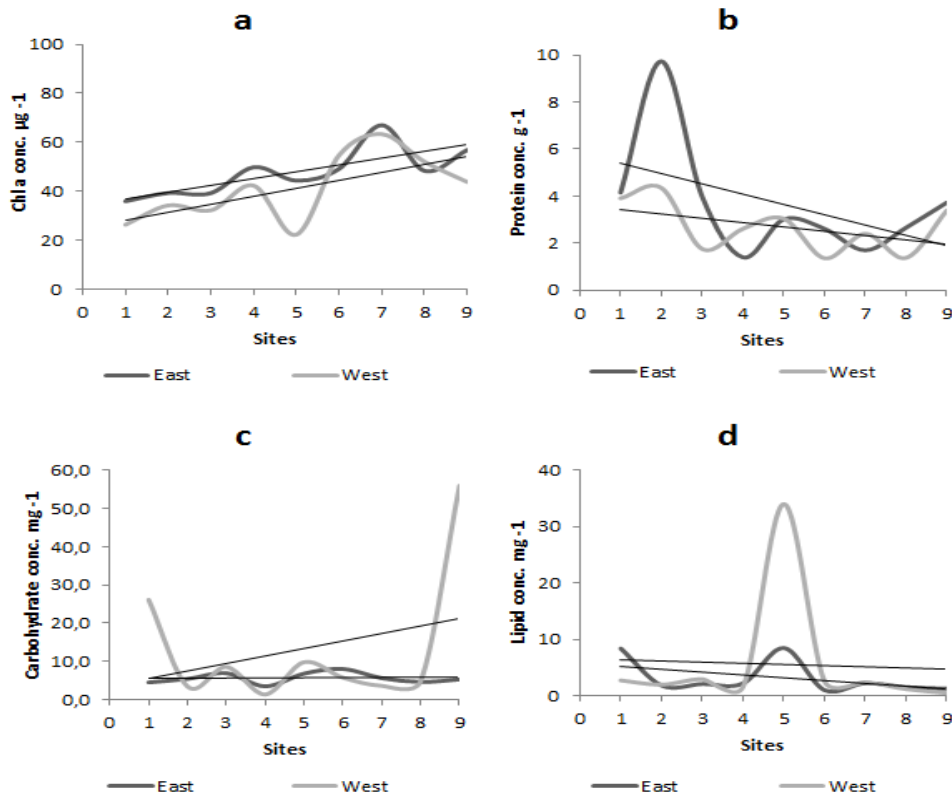


Figure 3a, 3b, 3c,3d : Fluctuations in biochemical contents and chlorophyll *a* of phytoplankton at the two littoral area of Lake Nasser

Phytoplankton at the north of the lake attained the highest calorific value, especially at the east side 34 Kcal/l. currently with the elevation of the mean value of protein contents (4.6 g/l) and Chl. *a* (51 $\mu\text{g}/\text{l}$) at the same sector. Seasonal variations in calorific value of the studied phytoplankton indicated that winter as well as spring had the highest energy value of 21 Kcal/l in the algal biochemical structure which attributed to the maximum protein compositions during both seasons (4 g/L) and increasing in the carbohydrates (17.25 mg/L), lipids (7 mg/L) and phytoplankton standing crop of 393×10^4 cell/L during winter. The environmental variables interacted with the observed variations in energy content of the phytoplankton in fresh water lake, the quality of phytoplankton varied with energy content in the engulfing phytoplankton [18]. Microalgae varied significantly in their content of protein,

carbohydrate and lipid which are important indicators for ecological status assessment [38]. In our study, Chl. *a* concentrations was located in the range of 5.0-140 µg/l for both littoral sides. The Lake can be described as eutrophic according to TSI where TSI was fluctuated between 59.1 (W) and 60.4 (E) in the lake [14] and [26]. Protein is one of the most abundant macromolecules in the studied algal community (3.7 g/L at E and 2.7g/L at W) as shown in Table 4. Protein constituting a major part of the cell mass under nutrient enrichment conditions [71]. The elevation of the mean value CO₃ (16 mg/L) at the east of the lake, indicated that the largest proportion of inorganic C in water body were incorporated into algal protein. Carbon fixed in photosynthesis incorporated rapidly into amino acids [27]. In contrast, the proportion of storage compounds as carbohydrates (5.6 and 13.3 mg/L) and lipids (3.2 and 5.6 mg/L) for east and west area decreases in determined algal community. Protein ratio decreases when the phytoplankton community is in stationary phase or cells become destroyed [57].

Table 4: Mean elemental formulae for the primary metabolites and stoichiometric ratio of phytoplankton community at the littoral area of Lake Nasser.

Formula		East	West
Chlorophyll <i>a</i> (µg/l)	[C ₄₆ H ₅₂ O ₅ N ₄ Mg]	51.03	41.36
Proteins (g/l)	[C ₁₃₉ H ₂₁₇ O ₄₅ N ₃₉ S]	3.7	2.7
Carbohydrates (mg/l)	[C ₁₇ H ₂₈ O ₁₄]	5.6	13.3
Lipids (mg/l)	[C ₅₃ H ₃₉ O ₆]	3.2	5.6
C/N		3.6	3.8
C/H		0.6	0.33
O/C		0.33	0.31

The elemental phytoplankton compositions C, N, H and O were higher at the east side of the lake than the west one, this may related to phytoplankton community was dominated by Bacillariophyceae which represent 53.1% (E) and thus would be expected to have a relatively large amount of allocation to elemental compositions than the other classes. Organic carbon content of phytoplankton in investigated area was 32% of the elemental compositions. Phytoplankton contributes most of the organic carbon available to pelagic food webs [54]. The primary importance of H and O content are estimate their relationship with the amount of oxygen consumed during formation of phytoplankton organic material [7]. The usual average C/N ratio according to Redfield et al. (1963) is 6.6. However, microalgae at the study area had a high progress rate where the C/N ratio decreased to 3.6 (E) and 3.8 (W) at the lake

(Table 4), as a result of increasing the proportion of proteins at the same area thus support the idea that these algae were not nitrogen-limited.

Table 5: RDA Analysis 'Var-part-3groups-Simple-effects-tested', results. Total variation is 1558.143, explanatory variables account for 71.0% (adjusted explained variation is 51.7%)

Summary Table:				
Statistic	Axis 1	Axis 2	Axis 3	Axis 4
Eigenvalues	0.4798	0.1417	0.0478	0.0289
Explained variation (cumulative)	47.98	62.15	66.93	69.82
Pseudo-canonical correlation	0.9432	0.8194	0.6108	0.6265
Explained fitted variation (cumulative)	67.55	87.50	94.24	98.31
Permutation Test Results:				
On All Axes	pseudo-F=3.7, P=0.002			

The result of the RDA clearly show the seasonal differences, in winter the plankton assemblages clearly separate from other season and plankton is mainly dominated by *Cyloellata* species, while in summer *Planktolyngbya limnetica* and *Aulacoseira* became dominated.(Fig 4) When variance partitioning was also included in the analysis the temporal, environmental and biochemical parameters allow us the separate analysis of these factors. The 6 environmental variables and 5 dominant phytoplankton species at the different selected sites, as in Figure 5. Analysis indicated that, the most important factors affecting on phytoplankton distribution are NO₃, NO₂, PO₄, pH, Transparency and NH₄. RDA cleared that, *Planktolyngbya limnetica* was dominated at the middle area, more related to pH while phosphorus is non-limiting factor for its growth. Both *Aulacoseira granulate* and *Eucapsis minuta* are tolerant to mild light deficiency and more dominated at the south area. *Cyclotella glomerata* and *C. operculata* are dominated at the north area, sensitive to flushing and tolerant to nutrient deficiency especially (NO₂, NO₃ and PO₄), as shown in table 3. The species compositions are influenced by a change of environmental variables especially nutrient, transparency and temperature [21]. RDA (Fig, 5) showed an elevation in lipid at the middle of the littoral area of the lake which probably due to increasing of annual phytoplankton crop (508 x 10⁴ cell/L) where the most leading species was *Planktolyngbya limnetica* Lemmer contains a high level of saturated fatty acids (39.7%) [52]. While the maximum value of carbohydrate and Chl. *a* were obtained at the southern area of the lake (Fig. 5), which accompanied with the maximum ratio of 34.8 and 13.4% for both *Eucapsis minuta* and *Aulacoseira granulata*.

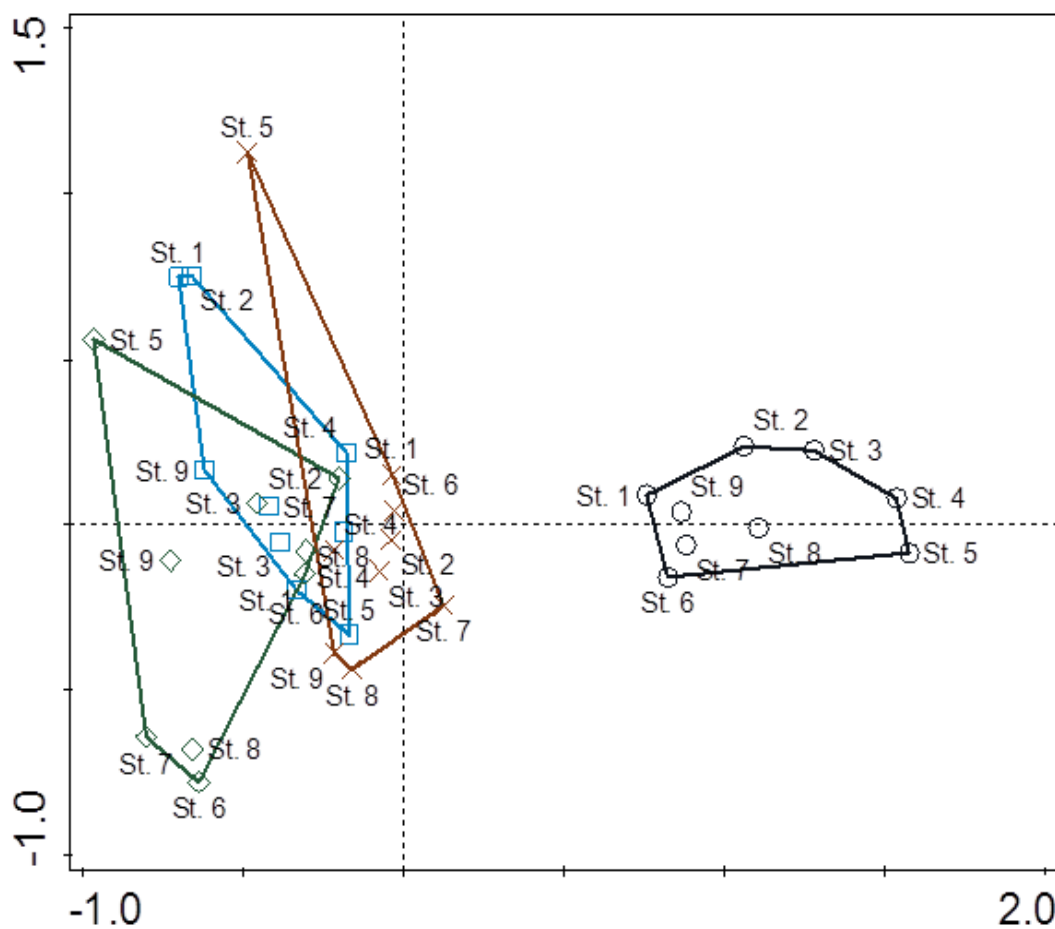


Figure 4: RDA unconstrained ordination of the dominant phytoplankton species (Objects) and selected environmental (Variables) in the different sites of the Lake.

RDA (Fig. 5) cleared that protein is highly coordinate with the elemental compositions of studied phytoplankton. The stoichiometry and diversity of phytoplankton frequently returns the supply of nutrients and light [31]. The stoichiometry of phytoplankton depends on the physiological state of the cell [11]. C/H and O/C ratio for natural phytoplankton samples were less than 1. Which pointed to the metabolites categories of studied phytoplankton contain unsaturated aliphatic compounds such as alkenes and aliphatic tail of fatty acids [58].

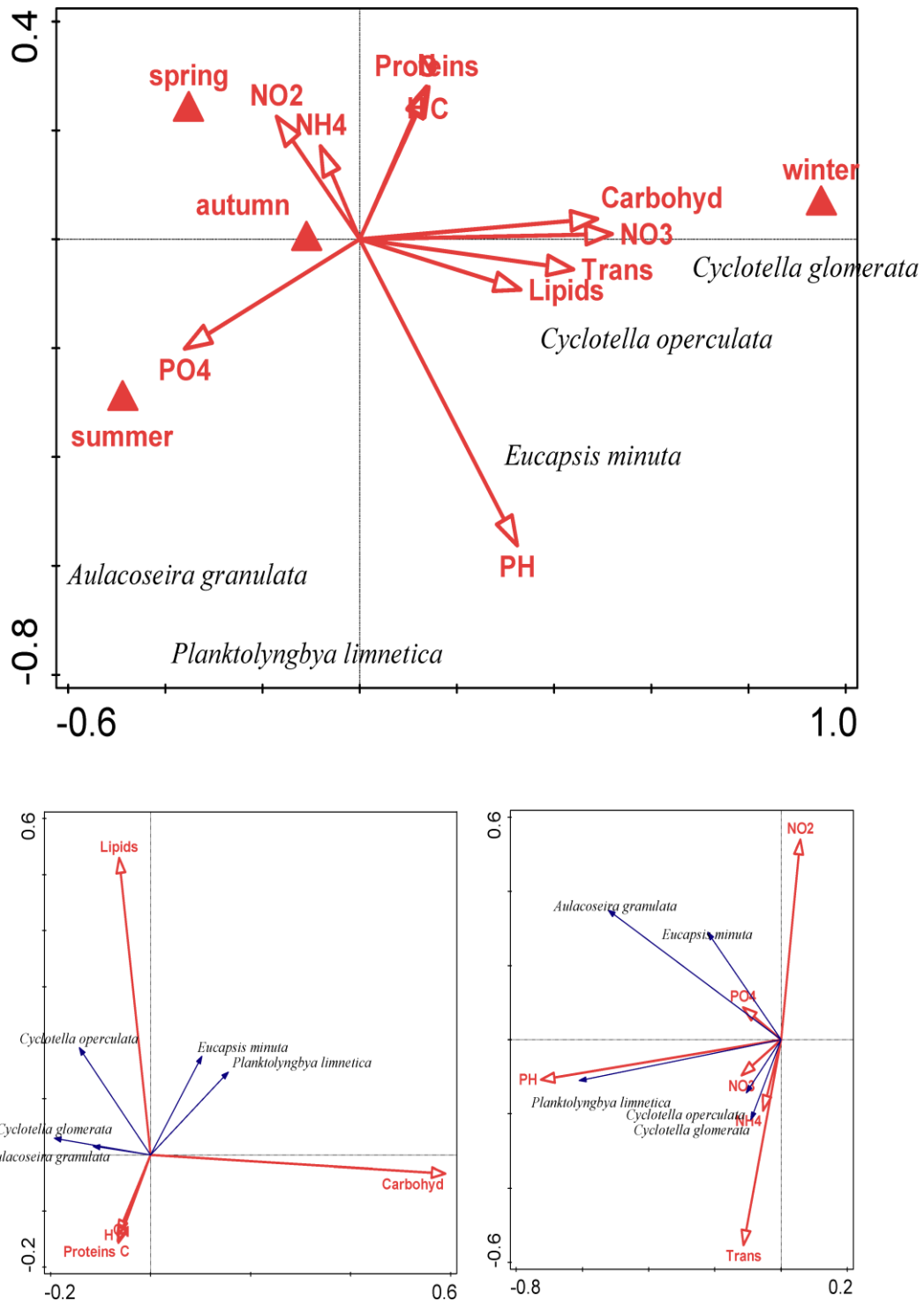


Figure 5: RDA unconstrained ordination of the dominant phytoplankton species (Objects) and selected environmental (Variables) in the different sites of the Lake, for the temporal, environmental and biochemical parameters

Conclusion

Examination of elemental contents and biochemical parameters in natural algal communities can yield important insights into the effects of physico-chemical data and nutrient composition of water body on the energetic value of algae. Bacillariophyceae can be classified as high elemental and biochemical composition group at the lateral area of Lake Nasser. A continuous monitoring should be organized to analyze the elemental composition of species from diverse taxonomic groups under a variety of environmental conditions. Furthermore, results of the study may be an alarm for all stakeholders, who are concerned about the lake changes as a result of the construction of the Renaissance Dam in Ethiopia.

References

- [1] Abd Ellah, R. G., "Thermal Stratification in Lake Nasser, Egypt Using Field Measurements" *World Applied Sciences Journal* 6 (4): 546-549, 2009.
- [2] Abdel-Monem, A. M., "Spatial distribution of phytoplankton and primary productivity in Lake Nasser" Ph.D. Thesis, University College for Girls, Ain Sham University. 161 pp, 1995.
- [3] Abd El-Monem, A.M., "Radio photosynthesis and Assimilation Rate of ¹⁴C Uptake By Phytoplankton In Closed, Turbid, Saline Lake, Lake Qarun-Egypt" *Arab J. Nucl. Sci. Appl.* 34(2): 243-250, 2001.
- [4] Abou El Ella, S.M., El Samman, T.A., "Ecosystem status of the north part of Lake Nubia" *African J. Biol. Sci.*, 6 (2), pp. 7–21, 2010.
- [5] Adesalu, T. A., Bagbe, M., Keyede, D., "Hydrochemistry and phytoplankton composition of two tidal creeks in south-western Nigeria" *Intern. J. of Trop. Biol.* 58(3): 827-840, 2010.
- [6] Andersen, T., Elser, J.J. and Hessen, D.O., "Stoichiometry and population dynamics" *Ecology Letters* 7:884-900, 2004.
- [7] Anderson, L.A., "n the hydrogen and oxygen content of marine phytoplankton" *Deep-Sea Research* 1, 42 (9).1675-1680, 1995.
- [8] APHA (American Public Health Association), "Standard methods for the examination of wastewater" 18th edition. 1015 Fifteen street. NW. Washington, DC 20005, 1996.
- [9] APHA (American Public Health Association), "Standard Methods for the Examination of Water and Wastewater" 20th ed., Inc. New York, 1998.
- [10] Bishai, H.M., Abdel Malek, S.A., Khalil, M.T., "Lake Nasser" *Publication of National Biodiversity Unit* No.11: 500-577pp, 2000.
- [11] Bonachela, J.A., Allison, S.D., Martiny, A.C., Levin, S.A., "A model for variable phytoplankton stoichiometry based on cell protein regulation" *Biogeosciences*, 10, 4341–4356, 2013.
- [12] Burrelly, P., "Les algues deau douce 11 les Algues jaunes et bruns N" *Boubee and cia, Paris*, pp. 438, 1968.

- [13] Brown, M.R., Jeffrey, S.W., Volkman, J.K., Dunstan, G.A., "Nutritional properties of microalgae for mariculture" *Aquaculture* 151: 315-331, 1997.
- [14] Carlson, R.E., "A trophic state index for lakes" *Limnology and Oceanography* v.22, n.2, pp.361-369, 1977.
- [15] Carvalho, L., Bennion, H., Dawson, H., Furse, M., Gunn, I., Hughes, R., Johnston, A., Luckes, S., Maitland, P., May, L., Monteith, D., Taylor, R., Trimmer, M., Winder, J., "Nutrient conditions for different levels of ecological status and biological quality in surface waters (Phase 1)" Environment Agency R&D Technical Report P2-260/4/TR, Bristol, 2002.
- [16] Chabrol, E., Castellano, A., "SPV method for estimation of total serum lipid" *J. Lab. Clin. Med.* 57, 300, 1961.
- [17] Chang, F. H., Bradford, J. M., "Standing stocks and productivity of phytoplankton off Wasteland, New Zealand" *Jour. Marine and freshwater research* 19: 193-211, 1985.
- [18] Chattopadhyay, C., "Polyphenolics and Energy Content in Phytoplankton" Evidence from a Freshwater Lake Proceedings of the Zoological Society. 67: pp 18-27, 2014.
- [19] Compère, P., "Contribution á L'étude des algues du sènègal (1)" *Algues de Lac et du Bas-Sènègal. Bull. Jard. Bct. Nat. Bclg.*, 61: 171-267, 1991.
- [20] David, J.H., Hazel, P., "Analytical biochemistry" *Hand Book*, 18 ed. 497p, 1993.
- [21] Dayala, V.T., Salas, P.M., Sujatha, C. H., "Spatial and seasonal variations of phytoplankton species and their relationship to physicochemical variables in the Cochin estuarine waters, Southwest Coast of India" *Indian Journal of Geo-Marine Sciences*, Vol. 43 (6), pp. 937-947, 2014.
- [22] Deskachary, T.V., "Cyanophyta" First ed. Indian Council of Agricultural Research. New Delhi. 686 pp., 1959.
- [23] Diehl, S., Erger, B., Wohrl, R., "Flexible nutrient stoichiometry mediates environmental influences, on phytoplankton and its resources" *Ecology*, 86:2931–2945, 2005.
- [24] Diekmann, A.B.S., Peck, M.A., Holste, L., St John, M.A., Campbell, R.W., "Variation in diatom biochemical composition during a simulated bloom and its effect on copepod production" *Journal of Plankton Research*. 31(11). 1391–1405, 2009.
- [25] Dubois, M., Gilles, K.A., Hamilton, J. K., Rebers, P.A., Smith, F., "Colorimetric method for the determination of sugars and related substances" *Anal. Chem.* 28, 350-356, 1956.
- [26] El Shahat, M.M., "Lake Nasser overview. In: J.F. Craig (Editor), Sustainable fish production in Lake Nasser: Ecological basis and management policy" International Center for Living Aquatic Resources Managemen (ICLARM), Penang, Malaysia, 2000.
- [27] Fogg, E.G., Thake, B., "Algal cultures and phytoplankton ecology" 3rd edition. Wisconsin, 273pp., 1987.

- [28] Fraga, F., Pérez, F.F., "Transformaciones entre composición química del fitoplancton, composición elemental y relación de Redfield" *Sci. Mar.*, 54: 69-76, 1990.
- [29] Gharib, S., Abd El-Halim, A., "Spatial variation of phytoplankton and some physic-chemical variables during the highest flood season in Lake Nasser (Egypt)" *Egyptian Journal of Aquatic Research*, VOL. 32 NO. 1, 246-263, 2006.
- [30] Habib, O.A., "Primary production of the phytoplankton" J.F. Carig (ed.) *Sustainable fish production in Lake Nasser: Ecological basis and Management policy. ICLARM Conf. Proc.* 61, 184p., 2000.
- [31] Hall S.R., Leibold, M.A., Lytle, D.A., Smith, V.H., "Grazers, producer stoichiometry, and the light/nutrient hypothesis revisited" *Ecology*, 88:1142–1152, 2007.
- [32] Hamdy, A., "Biosorption of heavy metals by marine algae" *Curr Microbiol.* 41:232-238, 2000.
- [33] Hannford, L.T., Britton, M.E., "The algae of Illinois. The Univ. of Chicago Press, Chicago, Illinois, U.S.A. 407 pp., 1952.
- [34] Harper, D.M., "The effects of artificial enrichment up on the planktonic and benthic communities in a mesotrophic to hypertrophic loch series in Scotland" *Hydrobiologia* 137:9-19, 1986.
- [35] Heikal, M.T., "Impact of water level fluctuation on water quality and trophic state of Lake Nasser and its Khors" *Egypt, Egypt. J. Aquat. Biol. Fish.*, 14 (1), pp. 75–86, 2010.
- [36] Huss, V.A.R., Frank, C., Hartmann, E.C., Hirmer, M., Kloboucek, A., Seidel, B.M., Wenzeler, P., Kessler, E., "Biochemical taxonomy and molecular phylogeny of the genus *Chlorella* Sensu Lato (Chlorophyta)" *J. Phycol.* 35: 587–598, 1999.
- [37] Hussian, A. M., Napiórkowska-Krzebietke, A., Mohamed, A.F., Toufeek, M. A., Abd El-Monem, A. M., Morsi, H. H., " Phytoplankton response to changes of physicochemical variables in Lake Nasser, Egypt" *Elementology Journal*, in press, 2015.
- [38] Hutorowicz A., Napiórkowska-Krzebietke, A., Pasztaleniec, A., Hutorowicz, J., LycheSolheim, A., Skjelbred, B., "Phytoplankton – In: Ecological status assessment of the waters in the Wel River catchment" *Guidelines for integrated assessment of ecological status of rivers and lakes to support river basin management plans* (Ed.) H. Soszka, Wyd. IRS, Olsztyn: 143-168, 2011.
- [39] Ishiwata, Y., Ohi, N., Obata, M., Taguchi, S., "Carbon to volume relationship of *Isochrysis galbana* (Prymnesiophyceae) during cell divisions" *Plankton Benthos Res* 8(4): 178–185, 2013.
- [40] Latif, A. F. A., "Fisheries of Lake Nasser and Lake Nubia. In: "Report on trip to Lake Nasser and Lake Nubia" Entz, B. A. & Latif, A.F.A. (edits.), Aswan Reg. Plan., Lake Nasser Develop. Cent, A.R.E. pp. 46-137, 1974.

- [41] Latif, A. F. A., "Lake Nasser" In: "Status of African Reservoirs Fisheries", Kapetsky, J. M. & Petr, T. (edits.) pp 193-246. CIFA Technical Paper -No. 10, 1984.
- [42] Mageed, A.A.A., Heikal, M.T., "Factors affecting seasonal patterns in epilimnion zooplankton community in one of the largest man-made lakes in Africa (Lake Nasser, Egypt)" *Limnol. - Ecol. Manag. Inl. Waters* 36, 91–97, 2006.
- [43] Margalef, R., "A Manual on Method for Measuring Primary Production in Aquatic Environments (Ed. R. A. Vollenweider)" *IBP Handbood.* 12: 7-14, 1974.
- [44] Marinov, I., Doney, S. C., Lima, I.D., "Response of ocean phytoplankton community structure to climate change over the 21st century: partitioning the effects of nutrients, temperature and light" *Biogeosciences*, 7, 3941–3959, 2010.
- [45] Nassar, M.Z., Mohamed, H.R., Khiray, H.M., Rashedy, S.H., "Seasonal fluctuations of phytoplankton community and physico-chemical parameters of the north western part of the Red Sea, Egypt" *Egyptian Journal of Aquatic Research.* 40, 395–403, 2014.
- [46] Padisak, J., Crossetti, L. O., Naselli-Flores, L., "Use and misuse in the application of the phytoplankton functional classification: a critical review with updates" *Hydrobiologia*, 621: 1-19, 2009.
- [47] Pahl, S.L., Lewis, D., Chen, F., King, K., "Growth dynamics and the proximate biochemical composition and fatty acid profile of the heterotrophically grown diatom *Cyclotella cryptica*" *J. Appl. Phycol.* ; 22(2):165-171, 2010.
- [48] PatilShilpa, G., ChondeSonal, G., JadhavAasawari, S., RautPrakash, D., "Impact of Physico-Chemical Characteristics of Shivaji University lakes on Phytoplankton Communities, Kolhapur, India" *Research Journal of Recent Sciences*, Vol. 1(2), 56-60, 2012.
- [49] Perumal, N.V., Rajkumar, M., Perumal, P., Rajasekar, K.T., "Seasonal variations of plankton diversity in the Kaduviyar estuary, Nagapattinam, India" *J Environ Biol* 30:1035-1046, 2009.
- [50] Prescott, A.G.W., "How to know the fresh water algae" (third edition) 293pp., 1978.
- [51] Quigg, A., Irwin, A.J., Finke, Z.V., "Evolutionary inheritance of elemental stoichiometry in phytoplankton" *Proc. R. Soc. B.* 1-9, 2010.
- [52] Rajeshwari, K. R., Rajashekhar, M., "Biochemical Composition of Seven Species of Cyanobacteria Isolated from Different Aquatic Habitats of Western Ghats, Southern India" *An International Journal, Brazilian Archives of Biology and Technology*, Vol.54, n. 5: pp. 849-857, 2011.
- [53] Redfield, A.C., Ketchum, B.H., Richard, F.A., "The influence of organisms on the composition of sea-water" in *The Sea*, Vol. 2, M.N. Hill, John Wiley and Sons, Inc. Ed. New York, 26-77 pp., 1963.

- [54] Reynolds, C.S., "The ecology of phytoplankton" Cambridge University Press, 535 pp., 2006.
- [55] Reynolds, C.S., "The ecology of freshwater phytoplankton" Cambridge Univ. Press. 384 pp., 1986.
- [56] Reynolds, C.S., Huszar, V.L., Naselli-Flores, L. and Melo, S., "Towards a functional classification of the freshwater phytoplankton", *J. Phytopl. Res.*, 24: 417-428, 2002.
- [57] Ríos, A.F., Fraga, F., Pérez, F.F., Figueiras, F.G., "Chemical composition of phytoplankton and Particulate Organic Matter in the Ría de Vigo (NW Spain)" *Sci. Mar.*, 62 (3): 257-271, 1998.
- [58] Romano, S. T., Dittmar, V., Bondarev, R. J. M., Weber, M. R., Viant, H. N., Schulz-Vogt, "Exo-Metabolome of *Pseudovibrio* sp. FO-BEG1 Analyzed by Ultra-High Resolution Mass Spectrometry and the Effect of Phosphate Limitation" *Plos One*. 9: 1-11, 2014.
- [59] Sithik, A. M., Thirumaran, G., Arumugam R., Kannan, R. R., Anantharaman, P., "Physico-Chemical Parameters of Holy Places Agnitheertham and Kothandaramar Temple; Southeast Coast of India" *Am.-Eurasian J. Sci. Res.*, 4 (2): 108-116, 2009.
- [60] Starmach, K., "Flora Slodkowodna Polski. Tom 4. Creptophyceae Dinophyceae Raphidophyceae" Krakow. 519 pp., 1974.
- [61] Sudhakar, K., Premalatha, M., "Theoretical assessment of algal biomass potential for carbon mitigation and biofuel production" *Iranica J. Energy & Environ.*, 3 (3): 232-240, 2012.
- [62] Tikkanen, T., "Kasviplanktonopas" Helsinki. 278 pp., 1986.
- [63] Tiwari, A.1., Chauhan, S.V., "Seasonal phytoplanktonic diversity of Kitham Lake, Agra" *J Environ Biol* 27: 35-38, 2006.
- [64] Ter Braak C.J.F. & Šmilauer P., "Canoco reference manual and user's guide: software for ordination, version 5.0" Microcomputer Power, Ithaca, USA, 496 pp., 2012.
- [65] Touliabah, H.E., Abdel-Monem, A.M., Gaballah, M.M., "Statistical analysis of phytoplankton structure and environmental variables in recent, subtropical, man-made lake (Lake Nasser – Egypt) 1- Along the main channel" *Az. J. Microbiol.* (49): 189-202, 2000.
- [66] Van de Waal, D.B., Verschoor, A.M., Verspagen, J.M., Donk D.V., Huisman J., "Climate-driven changes in the ecological stoichiometry of aquatic ecosystems" *Front. Ecol. Environ.* 8, 145–152, 2010.
- [67] Varadharajan D., Soundarapandian P., "Effect of Physico-Chemical Parameters on Species Biodiversity with Special Reference to the Phytoplankton from Muthupettai, South East Coast of India" *J Earth Sci Clim Change* 5:5, 2014.
- [68] Vollenweider, R.A., "Concerning calculation methods and limitations of proxy-estimates of proteins, carbohydrates and lipids in crustacean

- zooplankton from CHN analyses: some comments" *J. Limnol.*, 59(2): 170-178, 2000.
- [69] Vrede, T., Tranvik, L.J., "Iron constraints on planktonic primary production in oligotrophic lakes" *Ecosystems* 9: 1094–1105, 2006.
- [70] Litchman E., Klausmeier C. and Yoshiyama K., "Contrasting size evolution in marine and freshwater diatoms", *Proc Natl Acad Sci U S A*, 24; 106(8): 2665–2670, 2009.
- [71] Geider, R. J. and LA Roche J., "Redfield revisited: Variability of C: N: P in marine microalgae and its biochemical basis", *Eur. J. Phycol.* 37: 1–17, 2002.