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## Measurement of hydrogen peroxide influx into cells: Preparation for measurement using on chip microelectrode array

Hannah R. Kriscovich<sup>a</sup>, Sarah M. Libring<sup>b</sup>, Siddarth V. Sridharan<sup>c</sup>, James K. Nolan<sup>d</sup>, Jose F. Rivera<sup>c</sup>, Jenna L. Rickus<sup>d, e</sup>, David B. Janes<sup>c</sup>

<sup>a</sup> Wallace H. Coulter Department of Biomedical Engineering, Georgia Institute of Technology

<sup>b</sup> Department of Biomedical Engineering, Rutgers University

<sup>c</sup> School of Electrical and Computer Engineering, Purdue University

<sup>d</sup> Agricultural & Biological Engineering, Purdue University

<sup>e</sup> Weldon School of Biomedical Engineering, Purdue University

### ABSTRACT

Hydrogen peroxide ( $H_2O_2$ ) is commonly known as a toxic reactive oxidative species (ROS) for cells. Recent studies have found evidence that  $H_2O_2$  is also an important cellular signalling molecule. Quantifying cellular influx of  $H_2O_2$  will contribute to researchers' understanding of the role  $H_2O_2$  plays in healthy cells and cells involved in the progression of cancers and degenerative diseases. This work utilizes an assay kit and fluorescence techniques to evaluate cell lines and conditions to create a model biological system for measuring cellular  $H_2O_2$  consumption. Pancreatic beta cells (MIN6), astrocytes, and glioblastoma cells (GBM43 and GBAM1) were placed in 10  $\mu M$  and 20  $\mu M$   $H_2O_2$  solutions for up to 5 hours. The consumption of  $H_2O_2$  was measured using an Amplex Red Hydrogen Peroxide/Peroxidase Assay Kit (Molecular Probes/Invitrogen). GBAM1 cells exposed to 20  $\mu M$   $H_2O_2$  displayed the fastest rate of  $H_2O_2$  consumption ( $4.8 \pm 1.2$  nmol  $H_2O_2$ /min/ $10^6$  cells), followed by GBM43 cells ( $1.5 \pm 0.46$ ), astrocytes ( $1.1 \pm 0.24$ ), and MIN6 cells ( $0.29 \pm 0.075$ ). Additionally, the rate of consumption increased with increases in  $H_2O_2$  concentration. In the future, an on-chip micro-electrode array (MEA) will be used for real-time electrochemical experiments to measure influx of  $H_2O_2$  by astrocytes and GBAM1 cells with spatio-temporal resolution that the current techniques lack. The results from the electrochemical experiments will be compared to results from the assay kit to determine the ability of the MEA to accurately measure  $H_2O_2$  concentration and flux. The MEA can be extended to a wide variety of cellular environments for analysis of additional real-time biological events.

### KEYWORDS

Hydrogen peroxide, Biosensors, Microelectrode arrays (MEA), Real-time flux, Astrocytes