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OroSTIFF: Face-referenced measurement of perioral stiffness in

health and disease

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Abstract

A new device and automated measurement technology known as OroSTIFF is described to characterize non-participatory perioral stiffness in healthy adults for eventual application to patients with orofacial movement disorders associated with neuromotor disease, traumatic injury, or congenital clefts of the upper lip. Previous studies of perioral biomechanics required head stabilization for extended periods of time during measurement which precluded sampling patients with involuntary body/head movements (dyskinesias), or pediatric subjects. The OroSTIFF device is face-referenced and avoids the complications associated with head-restraint. Supporting data of non-participatory perioral tissue stiffness using OroSTIFF are included from 10 male and 10 female healthy subjects. The OroSTIFF device incorporates a pneumatic glass air cylinder actuator instrumented for pressure, and an integrated subminiature displacement sensor to encode lip aperture. Perioral electromyograms were simultaneously sampled to confirm passive muscle state for the superior and inferior divisions of the orbicularis oris muscles. Perioral stiffness, derived as a quotient from resultant force (ΔF) and interangle span (ΔX), was modeled with multilevel regression techniques. Real-time calculation of the perioral stiffness function demonstrated a significant quadratic relation between imposed interangle stretch and resultant force. This stiffness growth function also differed significantly between males and females. This study demonstrates the

Conflict of interest

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OroSTIFF '*proof-of-concept*' for cost-effective non-invasive stimulus generation and derivation of perioral stiffness in a group of healthy unrestrained adults, and a case study to illustrate the dose-dependent effects of Levodopa on perioral stiffness in an individual with advanced Parkinson's disease who exhibited marked dyskinesia and rigidity.

Keywords

upper lip; lower lip; biomechanics; force; displacement; elastic recoil

1. Introduction

Limb and orofacial stiffness are modulated by central descending inputs to lower motor neurons, reflex gain, muscle and connective tissue properties, postural orientation to gravitational loads, and geometry of muscle attachments (Barlow & Müller, 1991; Oluwatosin & Oluwatosin, 1998; Shiller et al., 2002). Quantifying stiffness typically involves imposing a specific displacement (ΔX) on a muscle-tissue system (e.g., whole body, limb, jaw) and measuring the resultant force (ΔF). The ratio of the resultant force to displacement yields a stiffness quotient ($\Delta F/\Delta X$). Muscle rigidity, regarded a clinical correlate of stiffness, is used to evaluate neurologic status, pharmacological efficacy (Caligiuri & Galasko, 1992), and neurosurgical intervention (Prochazka et al., 1997; Barlow et al., 1998; Sepehri et al., 2007).

Although biomechanical studies of limb rigidity have provided valuable insight into the neural regulation of limb movement disorders (Webster, 1964; Rushworth, 1964; Houk, 1979), similar application to the perioral system has been tenuous due to inadequate methods of transduction. Perioral anatomy is complex with its constituent muscles arranged in four layers according to their origins (Frellinger et al., 1987). The upper lip is associated with at least six muscles, including the orbicularis oris superior, depressor septi, levator labii superioris, levator labii superioris alaeque nasi, zygomaticus major, and levator anguli oris muscles. The lower lip is associated with four, including the orbicularis oris inferior, depressor labii inferioris, depressor anguli oris, and mentalis muscles (Zemlin, 1998). Several muscles converge just lateral to the buccal angle and interlace to form a dense, three-dimensionally mobile, fibromuscular mass known as the modiolus (Al-Hoqail & Meguid, 2009). Nearly all muscles converging toward the modiolus have dermal terminations. The modiolus supports decussation between fibers of the orbicularis oris and labial tractors terminating in the modiolus, and is involved with all forces acting on the oral angle (Pellisier et al., 2000; Zufferey, 2002). Several types of deep and superficial fascia also converge on the modiolus (Mitz & Peyronie, 1976) and function to organize and stabilize muscles, nerves, and vessels (Dzubow, 1986).

Eric Müller (Müller et al., 1985), a physicist and pioneer for speech biomechanics, developed the first mass-spring based three-dimensional space-frame model of the perioral system to simulate and test hypotheses concerning articulatory performance dynamics. He suggested that assessment of perioral stiffness would provide a potentially useful set of biomarkers to clarify the effects of progressive neuromotor disease, traumatic brain injury, pharmacologic treatment, neurosurgical, or maxillofacial surgical intervention on facial animation and speech motor control.

Previous studies have primarily focused on active rigidity during voluntary movement (Caligiuri & Galasko, 1992; Webster, 1966). However, some neurological disorders manifest elevated (e.g., Parkinson's disease - PD) or decreased muscle activity (e.g., cerebellar disease) at rest. This nonparticipatory 'resting' state is known as passive rigidity. In patients with idiopathic PD, labial stiffness was positively correlated with perioral muscle activity and inversely related to the magnitude of lip movements during speech (Hunker et al., 1982).

Another report challenged this finding and noted no clear statistical relation between lip stiffness and lip kinematics but observed that PD speakers became more hypokinetic as a function of speech rate (Caliguiri, 1989). The precise effects of rigidity or increased stiffness on lip displacement amplitude, velocity, and speaking rate remain poorly understood in this neurologic disorder.

Active and passive force were measured in the perioral system of healthy young adults (Barlow & Müller, 1991) in order to determine the relation between muscle length (interangle span) and *in vivo* resultant force in the perioral region. Active force during maximum voluntary contraction (MVC) of the orbicularis oris muscles increased quadratically with increases in the span (aperture) between the corners of the mouth, and were significantly greater for male than female subjects, whereas passive tissue forces were nearly equal as a function of span. In healthy adults, the lower lip is also stiffer than its upper lip counterpart, and may be due in part to its antigravity role in postural control.

Stiffness as a regulated variable has been hypothesized to play a significant role in movement, associated with equilibrium position and end-point accuracy (Shadmehr, 1993; Feldman & Levin, 2009), force recruitment, and velocity scaling among articulators during speech production (Gracco, 1994; Löfqvist & Gracco, 1997; Shaiman & Gracco, 2002). Separate cortical control systems have been hypothesized for movement and stiffness regulation related to reciprocal activation and coactivation of antagonist muscle groups (Humphrey & Reed, 1983). Precise regulation of lip stiffness is essential for accurate production of fricative sounds such as [f] and [v] (Ito et al., 2004) and the duration of lip closure during consonant production (Löfqvist, 2005). Altered stiffness regulation *between* articulators is another feature of orofacial motor control which could impact facial movements. For example, jaw perturbation experiments revealed that stiffness is up-regulated between the upper lip and jaw in order to maintain the constriction area between lips during the production of a fricative consonant (Gomi et al., 2002). Increased jaw stiffness is associated with a decrease in the variability of speech kinematics (Shiller et al., 2002). Stiffness regulation appears to play a central role for speech motor learning and adaptation (Nasir & Ostry, 2006; Tremblay et al., 2008).

Recently, perioral tissue stiffness has been measured using a wall-mounted linear actuator operating under position feedback to sequentially impose step increases in interangle lip span in healthy female (Seibel & Barlow, 2007) and male adults (Chu et al., 2009) sampled under a passive (no-contraction) condition. The methods used however required head stabilization in a cephalostat for several minutes during the presentation of lip displacements. This approach precludes study in clinical populations, including pediatric patients (i.e., orofacial cleft), or in cases where dyskinesia is present (e.g., progressive neuromotor disease, traumatic brain injury).

To solve this limitation, a new face-referenced interangle device known as OroSTIFF was developed in our bioengineering laboratory to examine the feasibility of real-time perioral stiffness measurements in unrestrained participants in health and disease. It was hypothesized that passive 'non-participatory' perioral stiffness would vary quadratically as a function of interangle span displacement. Sex was also regarded as a potentially significant independent variable because of known differences in lip dynamics (Barlow & Rath, 1985; Barlow & Müller, 1991). The OroSTIFF device application and related test results are expected to provide clinicians a new method for evaluating the efficacy of pharmacological and surgical intervention in the perioral system of patients with neuromotor disease (e.g., Parkinson's disease), craniofacial anomalies (e.g., cleft lip), lip/face carcinoma, or traumatic injury to the face and/or brain centers involved in facial animation (e.g., bomb blast, missile wounds, vehicular).

2. Materials and methods

2.1 Subjects

Twenty healthy adults (10 males, 10 females), 19 to 31 years of age with no prior history of neurological and craniofacial disorder, and/or speech impairment participated in this study (Table 1). Informed consent, approved by the University of Kansas Human Subjects Internal Review Board, was obtained from all subjects.

2.2 Protocol

Subjects were seated in a comfortable chair and instructed to remain speechless and motionless, and relax facial muscles during the 2 minute sampling (Figure 1). To ensure subjects' nonparticipation during the imposed perioral stretch, Ag/AgCl 4-mm diameter bipolar electrodes (2-cm interelectrode distance) were placed over the orbicularis oris superior (OOS*m*) and orbicularis oris inferior (OOI*m*) muscles. Biopotentials were conditioned with Grass P511 bioamplifiers (30Hz-1kHz bandpass, Gain=20K). A 1-cm interincisal bite block was molded (KERR Xtrude-XPTM) for each subject to stabilize the mandible during stiffness sampling.

The thin wall tubular stainless steel face-referenced OroSTIFF device (mass = 40.7gm) was coupled bilaterally to the oral angles via lip saddles and supported on the mental symphysis with a double-adhesive tape collar for vertical stabilization (see *chin anchor* on Figure 1). The device incorporates an Airpel[®] custom microminiature pneumatic glass-cylinder actuator instrumented for pressure (Honeywell #26PCCFAG, +/- 15 psi) and an integrated custom subminiature displacement sensor (differential variable reluctance transformer [DVRT], MicroStrain[®], Inc) to encode lip aperture. The pneumatic actuator was manually pressurized with a 10-cc syringe which in turn imposed an interangle stretch of approximately 20 mm. A 30-gauge blunt tip cannula, vented to atmosphere, was coupled in parallel with the OroSTIFF pneumatic system. This cannula provided a fixed resistive load, essentially a controlled air leak, upon which the perioral recoil force would act to allow the equal-arm scissor cantilevers to return to their initial lip aperture resting position (L₀ + 15 mm). A block diagram of the instrumentation is shown in Figure 2.

Interangle oral aperture at rest provides an estimate of resting muscle length (L_0) and was measured with a digital caliper for each subject. The OroSTIFF interangle span was initialized to [$L_0 + 15$ mm] for all subjects. A series of 5 interangle stretch trials were completed while sampling force, displacement, and electromyograms (EMG) from bipolar electrodes placed on the upper lip (OOS*m*) and lower lip (OOI*m*) in real-time with custom software (OroSTIFF v. 3.0.5, LabVIEWTM 8.0). Individual interangle stretch trials were completed within 10 seconds, and the perioral stiffness protocol was completed within 2 minutes for each participant.

2.3 Data acquisition and analysis

2.3.1 Identifying nonlinear segment of force-displacement curve—Air pressure within the microminiature pneumatic glass-cylinder actuator and the displacement signal from the DVRT were digitized at 2 ksamples/sec at 16-bits resolution. These waveforms were down-sampled to yield 100 pressure and position samples which were digitally low-pass filtered ($f_{lp} = 30$ Hz, 2-pole Butterworth), and subsequently averaged in 10 bins of 10, yielding an effective sample rate of 200 Hz for real-time calculation and display of force, displacement, and derived stiffness.

Stiffness coefficients (N/mm) were automatically calculated in real time during the phase of elastic recoil for each of 5 trials as the low-mass interangle yokes of the OroSTIFF device returned to the participant's interangle rest position. The stiffness coefficient was calculated

as the change in force over a 1 mm change in interangle span and sequentially evaluated at 1 mm intervals. Real-time display of stiffness coefficient versus span began when 3 conditions were met simultaneously: span > 0.5 mm, force decreasing, and a positive slope for a 10-point linear fit of force versus span. Graphic display continues until span < 0.5 mm (see Figure 3, points D to E). The absolute number of stiffness points along the recoil trajectory depends on the maximum interangle span achieved. To determine stiffness for a specific span a 100-point running cubic spline was evaluated at 0.5 mm above and below the desired span (i.e., force was evaluated at 19.5 and 18.5 mm to calculate stiffness for a nominal span of 19 mm). The cubic spline allows force to be determined at regular displacement intervals.

2.3.2 Determine muscle activity pattern during non-passive stretch—The root-mean-square (RMS) of the upper lip (OOS*m*) and lower lip (OOI*m*) EMG signals were computed during the phase of elastic recoil for each of 5 trials with an increment of 10 samples at 2000Hz (averaging time of 5 ms) to quantitatively verify non-participation of perioral muscles.

2.3.3 Calibration of the OroSTIFF device—The DVRT factory calibration data as well as digital caliper measurements were used to determine the ratio of DVRT position-to-interangle span. Force was calibrated with a load cell placed between the stainless steel interangle lip saddles of the OroSTIFF device. Device stiffness was determined by clamping the stainless steel interangle lip saddles and measuring position and force while modulating pressure with the 10-cc Becton syringe. The device functionality was verified using a precision linear spring.

After OroSTIFF program initialization, voltage offsets were determined with pressure vented to atmosphere, position set to zero, and EMG disconnected. All four signals were scaled linearly using these offsets and previously determined calibration slopes to yield force (N), displacement (mm), and EMG (uV). The DVRT position signal was converted to interangle span by multiplying by a constant to account for differences in the scissor-equal arm cantilever lengths on opposite sides of the central pivot needle bearing, and by correcting for device stiffness. Measured force is divided by this effective device stiffness and subtracted from position to yield interangle span. The negative slope seen in figure 3 between points A and B, and C and D represents the effective device stiffness.

2.3.4 Data analysis of force vs. displacement (\Delta F/\Delta X)—Given the hierarchically nested design of the data, in which the perioral stiffness was measured through a series of 5 interangle stretch trials (level-1) for each participant (level-2), multilevel regression analysis was conducted using SAS Version 9.1 (SAS Institute, 2004). First, an unconditional means model (i.e., null model) was fit in order to determine the random variance components. Then, level-1 and level-2 predictors and cross-level interaction terms were introduced into the null model with their significant random effects. The level-1 predictors represented the linear and quadratic regression slopes of the interangle span on the perioral stiffness. The cross-level interaction terms represented the sex differences in the linear and quadratic regression slopes.

3. Results

3.1 The regression result of force vs. displacement ($\Delta F/\Delta X$)

The fitted null model is given by the expression $Y_{ij} = \gamma_{00} + u_{oj} + r_{ij}$, where $u_{oj} \sim N(0, \tau_{00})$ and $r_{ij} \sim N(0, \sigma^2)$ for trial *i* and participant *j*. This model expresses the stiffness scores as the sum of an overall mean (γ_{00}), a series of random deviations from that mean (u_{oj}), and a random error (r_{ij}) associated with the *i*th trial in the *j*th participant. The null model also showed that the estimated mean stiffness score across all trials and all participants was .073 N/mm (*SE* = .002,

t [19] = 40.90, p<.01). The estimated variances of level-1 and level-2 residual errors were . 00180 (SE = .00006, z = 30.28.002, p<.01) and .00005 (SE = .00002, z = 2.17, p<.05), respectively. The intra-class correlation (ICC) indicated that most of the variability (97.6%) in the stiffness scores occurred within participants. These estimates suggested that the stiffness scores do differ within participants and there is less but significant variation between participants.

The final model included two level-1 predictors that represent the linear and quadratic regression slopes of the interangle span on the perioral stiffness. Two cross-level interaction terms were also included to test whether the regression slopes differ between males and females. This model can be written by

 $Stiffness_{ij} = \gamma_{00} + \gamma_{01}Sex_j + \gamma_{10}Span_{ij} + \gamma_{20}Span_{ij}^2 + \gamma_{11}(Sex_j \times Span_{ij}) + \gamma_{21}(Sex_j \times Span_{ij}^2) + u_{1j}Span_{ii} + u_{0j} + r_{ij},$

where $\begin{pmatrix} u_{oj} \\ u_{1j} \end{pmatrix} \sim N \begin{bmatrix} 0 \\ 0 \end{bmatrix}, \begin{pmatrix} \tau_{00} & \tau_{01} \\ \tau_{10} & \tau_{11} \end{bmatrix}$ and $r_{ij} \sim N(0, \sigma^2)$ for trial *i* and participant *j*. The random

effect (u_{2j}) for the $Span_{ij}^2$ was not included because its estimation made Newton-Raphson algorithm for optimizing residual likelihood function (REML) to yield a non-positive definite G matrix. The parameter estimates from the fitted final model are shown in Table 2. It was shown that the perioral stiffness increased as a quadratic function of the interangle span, $\hat{\gamma}_{10}$ = .00054, t(1811) = 37.28, p < .01. More importantly, this quadratic slope significantly differed between males and females, $\hat{\gamma}_{21} = .00005$, t(1811) = 2.31, p < .05 (see Figure 4). Although the linear function of the interangle span was also significant, there was no sex difference in this linear slope.

The residual ICC showed that 43.1% of total residual variance occurred between participants. The squared multiple correlation indicated that approximately 89.3% of level-1 residual variance ($\hat{\sigma}^2$) was accounted for by this model. Since the estimated level-2 residual variance ($\hat{\tau}_{00}$) increased with the random slope, the squared multiple correlation for this level could not be obtained. The likelihood-ratio (LR) test suggested that both the random intercept (LR $\chi^2 = 232.10, p < .01$) and the random linear slope (LR $\chi^2 = 1439.00, p < .01$) were tenable.

3.2 The muscle activity pattern during non-passive stretch

Perioral muscle activity remained remarkably constant as interangle span was increased confirming the non-participatory nature of the experimental task. The distribution of EMG RMS values for the OOS and OOI muscle recording sites pooled among subjects is shown in Figure 5a and 5b. A one-way ANOVA for EMG RMS versus interangle span indicated non-significance for both the upper lip (F=0.80, p=.737, R-sq (adj) =.00%), and lower lip (F=0.54, p=.967, R-sq (adj) =.00%) and confirmed the non-participatory nature of the stiffness sampling protocol.

4. Discussion

This present study demonstrated a non-invasive and rapid (< 2 mins) method for real-time data acquisition and analysis of perioral tissue stiffness without head restraint. The derived regression function for non-participatory perioral stiffness using OroSTIFF is consistent with previous reports in healthy participants using costly servo-controlled linear actuators that required head restraint (Seibel & Barlow, 2007). The results indicated that intersubject variability in stiffness growth functions increased as the interangle span increased (Figure 4). This is likely due to individual differences in perioral anatomy. Male subjects yielded

(1)

significantly higher stiffness coefficients than female subjects. A similar trend was noted in stiffness measures sampled at lip midline (Ho et al., 1982). This may be due in part to undocumented differences in perioral anatomy, including the size and orientation of muscles, connective tissue, and vessels.

Sample Clinical Application

The clinical utility of the OroSTIFF application was put to the test in a participant with a significant movement disorder, a 68-year old male with a 12-year history of idiopathic Parkinson's disease (PD). He exhibited moderate-severe dyskinesia of head, trunk, and extremities under the prescribed dosage of anti-PD medications and this is designated as the 'ON' state. He exhibited reduced dyskinesia, but increased dystonia and rigidity in the 'OFF' condition (unmedicated condition). This individual consented to refrain from taking his anti-PD medications from the previous evening for a period of 12-hours, and subsequently arrived at the laboratory at 9AM in the 'OFF' condition. An incisal bite block was molded, and the OroSTIFF protocol was completed in less than 2 minutes. The participant was given his prescribed dose of anti-PD medications with a glass of water. After 50 minutes, the participant was in the 'ON' state, and the OroSTIFF protocol was repeated once again to assess the effects of L-Dopa on perioral stiffness functions.

Several striking features are apparent from the PD stiffness plots given in Figure 6. First, in the 'OFF' state, perioral stiffness is approximately 7 times greater than normal, and because of this high level of tonic muscle activity, the imposed displacements generated by the OroSTIFF device were limited to approximately 10 mm beyond resting span (L_0+15 mm). Administration of the anti-PD medications had a significant effect on reducing the slope and offset of the quadratic stiffness function, however perioral stiffness remained significantly elevated over normal levels. A video clip is included with this research report to demonstrate the OroSTIFF sampling protocol for this PD participant. Future studies will focus on the relation between perioral stiffness and orofacial hypokinesia during speech.

The current instantiation of the OroSTIFF device is limited to sampling relatively low levels of interangle lip force associated with non-participatory (passive) conditions in health and disease. High stiffness, short-throw titanium cantilevers are under development to reduce the size and mass of the OroSTIFF device, and permit measurement of active force production up to 25 N among pediatric and adult populations. The current instrument design also precludes unilateral assessment of stiffness in the perioral region. This feature would be useful for patients who have sustained unilateral injury to the lower face, undergoing lip revision surgery due to uni- or bilateral congenital cleft of the upper lip, manifest laterality differences in perioral stiffness due to a central brain lesion or progressive neuromotor disease (e.g., Parkinson's disease, cerebrovascular stroke), or peripheral cranial mononeuropathy (Bell's palsy). One possible solution for unilateral measurement is to mechanically reference lip stiffness measures sampled from either the right or left oral angle to a midline transducer voke positioned in a custom dental impression mold within the occlusal plane. Such a configuration has been used successfully to sample maximum voluntary contraction (tetany) and controlled force dynamics in a midline vertical lip compression task in young adults with moderate-to-severe congenital forms of upper motor neuron syndrome (Barlow & Abbs, 1986).

Supplementary Material

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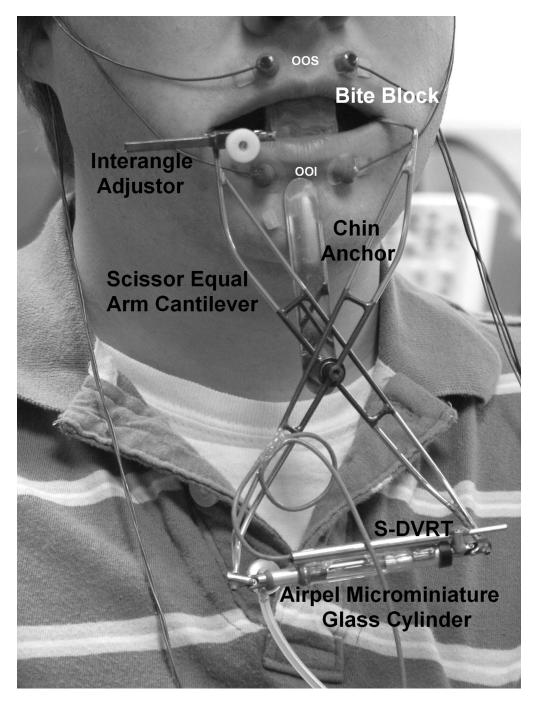


Figure 1.

Instrumentation and configuration to assess the force-displacement relation (stiffness) for the perioral tissues. The equal-arm scissor cantilever and stainless steel lip saddles (hooks) positioned for imposing increases in interangle span and automated stiffness sampling. A lip span adjustor accommodates to individual differences in lip aperture and used to set initial position ($L_0 + 15$ mm). The stainless steel 0.020" shimstock chin anchor articulated on a needle bearing and was positioned over the subject's mental symphysis for vertical stabilization. InVivo Metrics 4 mm Ag/AgCl surface electrodes placed over orbicularis oris superior (OOS) and orbicularis oris inferior (OOI) muscle recording sites.

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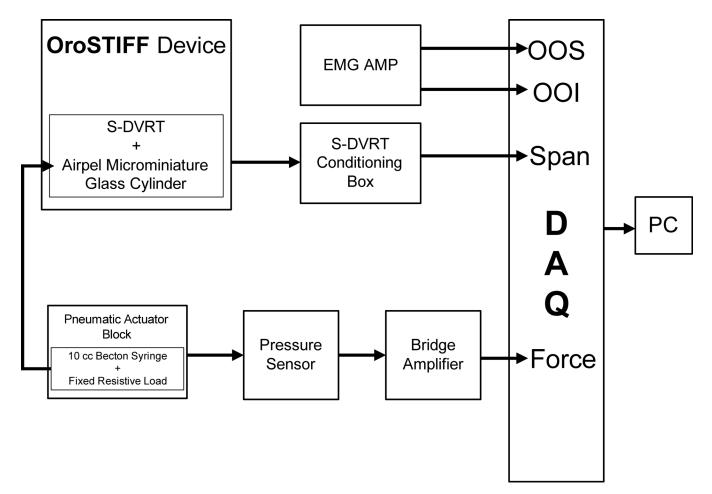


Figure 2.

A schematic block diagram of the OroSTIFF system. The device is equipped with a subminiature differential variable reluctance transducer to measure displacement of the lips, and a pressure transducer to derive the reactive force associated with perioral tissue recoil. A fixed pneumatic resistive load, in the form of 30 gauge blunt tip cannula, permits the moveable cantilevers of the OroSTIFF device to return to its initial position $[L_0 + 15 \text{ mm}]$ following each imposed stretch trial.

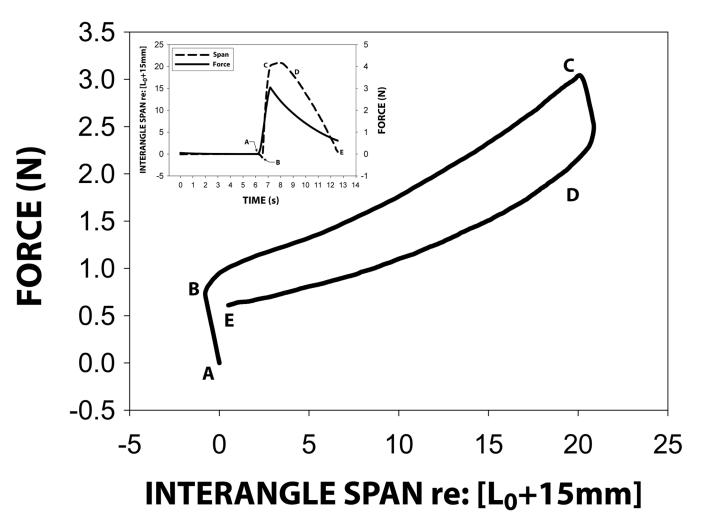


Figure 3.

Typical force-span hysteresis curve sampled from a normal adult subject with graphic insert showing span-time (black line) and force-time (dashed line) plots. Point A: preload condition of the OroSTIFF device on a subject's face; B: onset of interangle stretch phase ; C: peak interangle stretch; the recoil phase (D–E) during which stiffness is calculated (Δ Force/ Δ Span). Regions from A–E and just after C reflect bending (elasticity) of the OroSTIFF device without change in S-DVRT position output.

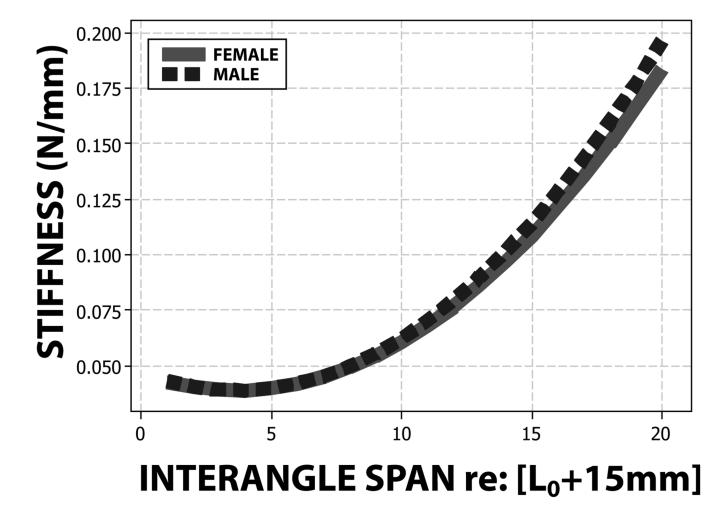


Figure 4.

Regression functions for male (dotted black line) and female subjects (solid gray line).

Male regression equation: $\widehat{Y}_{ij} = .04682 - .00442^* Span_{ii} + .00059 Span_{ii}^2$

Female regression equation: $\widehat{Y}_{ij} = .04583 - .00396^* Span_{ii} + .00054 Span_{ii}^2$

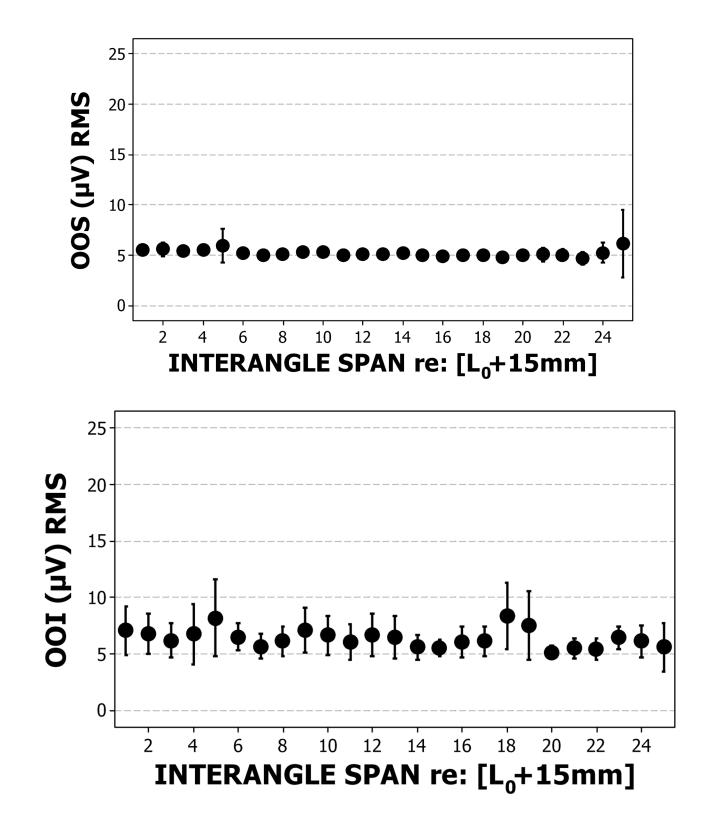


Figure 5a & 5b.

The distribution of mean and standard error of EMG RMS values (μ V) for upper lip (5a) and lower lip (5b) recording sites for all participants during the 'face-relaxed' non-participatory conditions. Eighty-five observations at each interangle span except at 25 mm, where there were only 3 observations.

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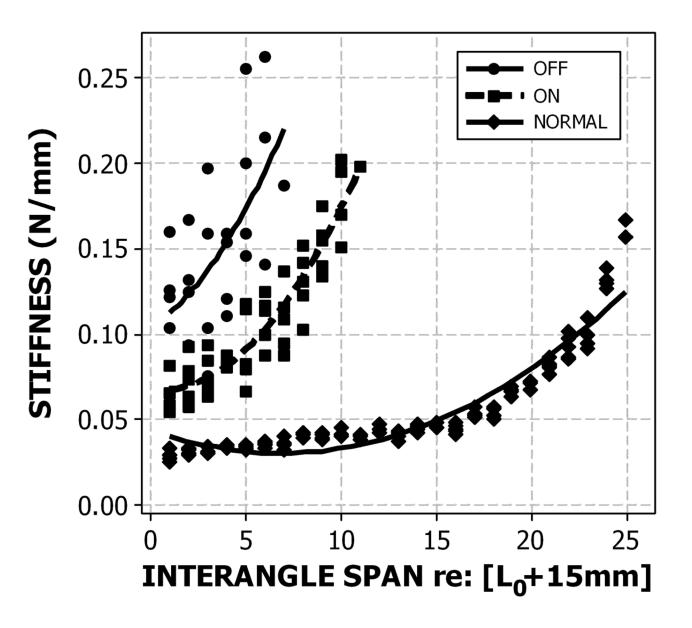


Figure 6.

Perioral stiffness data and quadratic functions for a 68-year old male with Parkinson's disease in the ON (with anti-PD meds) and OFF (without meds) conditions plotted relative to an ageand sex-matched healthy control.

Table 1

Physical characteristics of participants.

	Male	Female	
Number of Subjects	10	10	
Age (yrs)	24.59 (SD=3.07)	23.18 (SD=2.76)	
Weight (kg)*	67.78 (SD=7.35)	57.52 (SD=8.44)	
Height (cm)*	174.22 (SD=11.12)	164.74 (SD=4.22)	
Head circumference (cm)	57.05 (SD=1.66)	55.69 (SD=2.07)	
Nasion-to-inion (cm)	39.22 (SD=2.23)	37.54 (SD=1.75)	
Lip Resting Span (mm)	47.32 (SD=2.82)	46.46 (SD=2.16)	

*Significant difference at p<.05

Table 2

Multilevel regression parameter estimates of between- and within- level components.

Fixed Effect	Estimate	SE	t	р
Intercept	.04583	.00408	11.23	<.01
Sex	.00099	.00579	.17	.86
Span	00396	.00107	-3.70	< .01
Span ²	.00054	.00002	37.28	< .01
$\mathbf{Sex}\times\mathbf{Span}$	00046	.00152	30	.76
$\text{Sex}\times\text{Span}^2$.00005	.00002	2.31	< .05
Random Effect	Estimate	SE	z	р
σ^2	.00019	.00000	30.09	< .01
τ_{00}	.00015	.00005	2.82	< .01
τ_{11}	.00001	.00000	2.97	< .01
τ_{01}	00004	.00001	-2.81	< .01
ICC	.431			
R_{trial}^2	.893			
R ² participant	-			
-2lnL	-10360.1			
AIC	-10352.1			