



Genome-wide association study of nevirapine hypersensitivity in a sub-Saharan African HIV-infected population.

Carr, DF; Bourgeois, S; Chaponda, M; Takeshita, LY; Morris, AP; Castro, EMC; Alfirevic, A; Jones, AR; Rigden, DJ; Haldenby, S; Khoo, S; Lalloo, DG; Heyderman, RS; Dandara, C; Kampira, E; van Oosterhout, JJ; Ssali, F; Munderi, P; Novelli, G; Borgiani, P; Nelson, MR; Holden, A; Deloukas, P; Pirmohamed, M

(c) The Author 2017.

CC BY

For additional information about this publication click this link.

<http://qmro.qmul.ac.uk/xmlui/handle/123456789/19421>

Information about this research object was correct at the time of download; we occasionally make corrections to records, please therefore check the published record when citing. For more information contact scholarlycommunications@qmul.ac.uk

Genome-wide association study of nevirapine hypersensitivity in a sub-Saharan African HIV-infected population

Daniel F. Carr^{1*†}, Stephane Bourgeois^{2†}, Mas Chaponda^{1,3}, Louise Y. Takeshita⁴, Andrew P. Morris^{1,5}, Elena M. Cornejo Castro¹, Ana Alfirevic¹, Andrew R. Jones⁴, Daniel J. Rigden⁴, Sam Haldenby⁶, Saye Khoo¹, David G. Lalloo⁷, Robert S. Heyderman^{3,8}, Collet Dandara⁹, Elizabeth Kampira⁹, Joep J. van Oosterhout^{3,10}, Francis Ssali¹¹, Paula Munderi¹², Giuseppe Novelli¹³, Paola Borgiani¹³, Matthew R. Nelson¹⁴, Arthur Holden¹⁵, Panos Deloukas^{2,16,17} and Munir Pirmohamed¹

¹Department of Molecular and Clinical Pharmacology, University of Liverpool, Liverpool, UK; ²William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, UK; ³Malawi-Liverpool-Wellcome Trust Clinical Research Programme, College of Medicine, University of Malawi, Malawi; ⁴Institute of Integrative Biology, University of Liverpool, Liverpool, UK; ⁵Department of Biostatistics, University of Liverpool, Liverpool, UK; ⁶Centre for Genomic Research, University of Liverpool, Liverpool, UK; ⁷Liverpool School of Tropical Medicine, Liverpool, UK; ⁸Division of Infection and Immunity, University College London, London, UK; ⁹Division of Human Genetics, University of Cape Town, Cape Town, South Africa; ¹⁰Dignitas International, Zomba, Malawi; ¹¹Joint Clinical Research Centre, Headquarters, Kampala, Uganda; ¹²UVRI/MRC Uganda Research Unit on AIDS, Entebbe, Uganda; ¹³Department of Biomedicine and Prevention, University of Rome 'Tor Vergata', Rome, Italy; ¹⁴GlaxoSmithKline, Research Triangle Park, NC, USA; ¹⁵SAEC Consortium, Ltd, Chicago, IL, USA; ¹⁶Wellcome Trust Sanger Institute, Hinxton, Cambridge, UK; ¹⁷Princess Al-Jawhara

Al-Brahim Centre of Excellence in Research of Hereditary Disorders (PACER-HD), King Abdulaziz University, Jeddah, 21589, Saudi Arabia

*Corresponding author. Tel: +44-151-795-5392; E-mail: d.carr@liv.ac.uk

†Authors contributed equally.

Received 15 August 2016; returned 10 October 2016; revised 13 October 2016; accepted 20 November 2016

Background: The antiretroviral nevirapine is associated with hypersensitivity reactions in 6%–10% of patients, including hepatotoxicity, maculopapular exanthema, Stevens–Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN).

Objectives: To undertake a genome-wide association study (GWAS) to identify genetic predisposing factors for the different clinical phenotypes associated with nevirapine hypersensitivity.

Methods: A GWAS was undertaken in a discovery cohort of 151 nevirapine-hypersensitive and 182 tolerant, HIV-infected Malawian adults. Replication of signals was determined in a cohort of 116 cases and 68 controls obtained from Malawi, Uganda and Mozambique. Interaction with *ERAP* genes was determined in patients positive for *HLA-C*04:01*. *In silico* docking studies were also performed for *HLA-C*04:01*.

Results: Fifteen SNPs demonstrated nominal significance ($P < 1 \times 10^{-5}$) with one or more of the hypersensitivity phenotypes. The most promising signal was seen in SJS/TEN, where rs5010528 (*HLA-C* locus) approached genome-wide significance ($P < 8.5 \times 10^{-8}$) and was below *HLA*-wide significance ($P < 2.5 \times 10^{-4}$) in the meta-analysis of discovery and replication cohorts [OR 4.84 (95% CI 2.71–8.61)]. rs5010528 is a strong proxy for *HLA-C*04:01* carriage: *in silico* docking showed that two residues (33 and 123) in the B pocket were the most likely nevirapine interactors. There was no interaction between *HLA-C*04:01* and *ERAP1*, but there is a potential protective effect with *ERAP2* [$P = 0.019$, OR 0.43 (95% CI 0.21–0.87)].

Conclusions: *HLA-C*04:01* predisposes to nevirapine-induced SJS/TEN in sub-Saharan Africans, but not to other hypersensitivity phenotypes. This is likely to be mediated via binding to the B pocket of the *HLA-C* peptide. Whether this risk is modulated by *ERAP2* variants requires further study.

Introduction

Nevirapine, an NNRTI used for HIV¹ infection is effective² as part of combination antiretroviral therapy, but causes hypersensitivity in 6%–10% of patients.^{3,4} This can manifest in various ways, ranging from nevirapine-induced rash (NIR) (i.e. a maculopapular exanthema without any systemic manifestations), hypersensitivity syndrome (HSS) to severe blistering skin reactions such as Stevens–Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN)⁵ (1–2 per 1000 exposed individuals⁶). Extra-cutaneous involvement typically manifests as hepatotoxicity.⁷

Identification of the genetic risk factors for nevirapine hypersensitivity has focused on candidate gene approaches. Nevirapine is primarily metabolized by the hepatic cytochrome P450s 2B6 (CYP2B6) and 3A4 (CYP3A4).⁸ The exon 4 variant in *CYP2B6* (c.516G>T), which encodes a non-synonymous amino acid substitution (Gln172His) (rs3745274), leads to loss of function,^{9,10} with the variant T allele resulting in higher nevirapine plasma concentrations in both Caucasian¹¹ and sub-Saharan¹² adult patients. The associations with *CYP2B6* polymorphisms are rather confusing with the *CYP2B6* c.516G>T SNP associated with nevirapine-induced cutaneous adverse events in black and white populations¹³ but not with nevirapine-induced hepatotoxicity.¹⁴ The association with HLA alleles is even more complex, with *HLA-DRB1*01:01* (Caucasian^{13,15,16}), *HLA-C*04* (Thai,¹⁷ Chinese¹⁸ and Black¹³), *HLA-C*08* (Japanese¹⁹) and *HLA-B*35:05* (Thai^{13,20}) acting as predisposing alleles for nevirapine hypersensitivity. Our own previous study within a subset of patients from the Malawian HIV population described in this paper identified an association between *HLA-C*04:01* and nevirapine-induced SJS.²¹

In this study, in order to overcome some of the issues associated with candidate gene analysis, we have undertaken a genome-wide association study (GWAS) in a Malawian HIV cohort of nevirapine-exposed patients in order to identify genetic biomarkers of nevirapine hypersensitivity in an unbiased manner. We have also investigated whether there is any interaction between *HLA-C*04:01* in SJS/TEN patients and the endoplasmic reticulum aminopeptidase genes (*ERAP1* and *ERAP2*), which have been shown to modulate the risk of various immune diseases, in particular ankylosing spondylitis.²²

Methods

Patients

Discovery cohort

Antiretroviral-naïve patients ($n = 1117$) were prospectively recruited as previously described²¹ (Figure 1) from the Queen Elizabeth Central Hospital (QECH), Blantyre, Malawi, between March 2007 and December 2008. All were self-reported black African, over the age of 16, and had no baseline jaundice. CD4+ counts and liver function tests were monitored at 0, 2, 6, 10, 14, 18 and 22 weeks. Fifty-seven patients from this prospective cohort had nevirapine-induced hypersensitivity fulfilling the criteria of one or more of the following phenotypes:

- NIR: widespread maculopapular exanthema with no systemic manifestations but which worsened on treatment continuation.
- HSS: widespread rash with systemic manifestations (i.e. fever, cough or abnormal liver function tests). This is also known as DRESS (drug reaction with eosinophilia and systemic symptoms).

- SJS: blistering eruption affecting <10% of body surface area with two or more mucous membranes involved.
- TEN: blistering rash affecting >30% of body surface area and two or more mucous membranes. Patients with overlap syndrome had 10%–30% of their body surface area affected.
- Drug-induced liver injury (DILI): jaundice and abnormal ALT.

In addition, a total of 149 cases of nevirapine-induced hypersensitivity were recruited prospectively from QECH separately from the study described above, and a further 28 were identified retrospectively from patient records at the same centre. Out of a total of 234 hypersensitive cases, 159 where sufficient genomic DNA was available were included, along with 193/1060 of the nevirapine-treated age- and gender-matched controls (352 in total), in the discovery GWAS. Numbers of tolerant controls included were constrained by DNA quality and quantity.

Replication cohort

We recruited a number of patients with nevirapine hypersensitivity, with different phenotypes, from a number of centres (Table 1) to replicate our findings:

- Thirty nevirapine-hypersensitive patients and matched (age and gender) HIV-positive nevirapine-treated controls from Malawi. All controls and eight of the cases were from the original study but not included in the initial GWAS due to DNA quantity restraints. The other 22 cases presenting with the hypersensitivity phenotype according to the above criteria were identified from the QECH after the conclusion of the initial recruitment phase (December 2008).
- Thirty-two nevirapine-hypersensitive cases and age- and gender-matched controls identified in Uganda from the DART study cohort.²³ Cases were defined according to available patient records and subsequently categorized into the sub-phenotypes described above.
- Twenty-seven pregnant female patients with nevirapine-induced hepatotoxicity and 10 nevirapine-tolerant pregnant controls from Mozambique. Cases were defined as previously stated,²⁴ and included patients who discontinued nevirapine due to increased liver enzymes (grade 3/4). Controls were excluded if ALT/AST levels exceeded the median value observed in the case cohort.
- Twenty-seven female patients with nevirapine-induced SJS/TEN from Mozambique.²⁵ In this instance SJS/TEN was defined as development of exanthema and blistering starting mainly on the trunk, involving $\geq 10\%$ of the body surface with mucosal involvement.

Ethics

Full ethics approval for the study was received from the Liverpool School of Tropical Medicine Research Ethics Committee (Liverpool, UK), the College of Medicine Research and Ethics Committee, University of Malawi (Blantyre, Malawi) and the Uganda National Council for Science and Technology. All patients gave their written informed consent and those who met the criteria for a case had nevirapine withdrawn in accordance with Malawian National Treatment Guidelines. Local ethics approval was obtained for the DART study as previously described²³ with subsequent ethics approval for a pharmacogenetic sub-study also obtained.²⁶

DNA extraction

Genomic DNA was extracted from whole blood for the discovery cohort²¹ and replication cohorts as previously described.^{24,25}

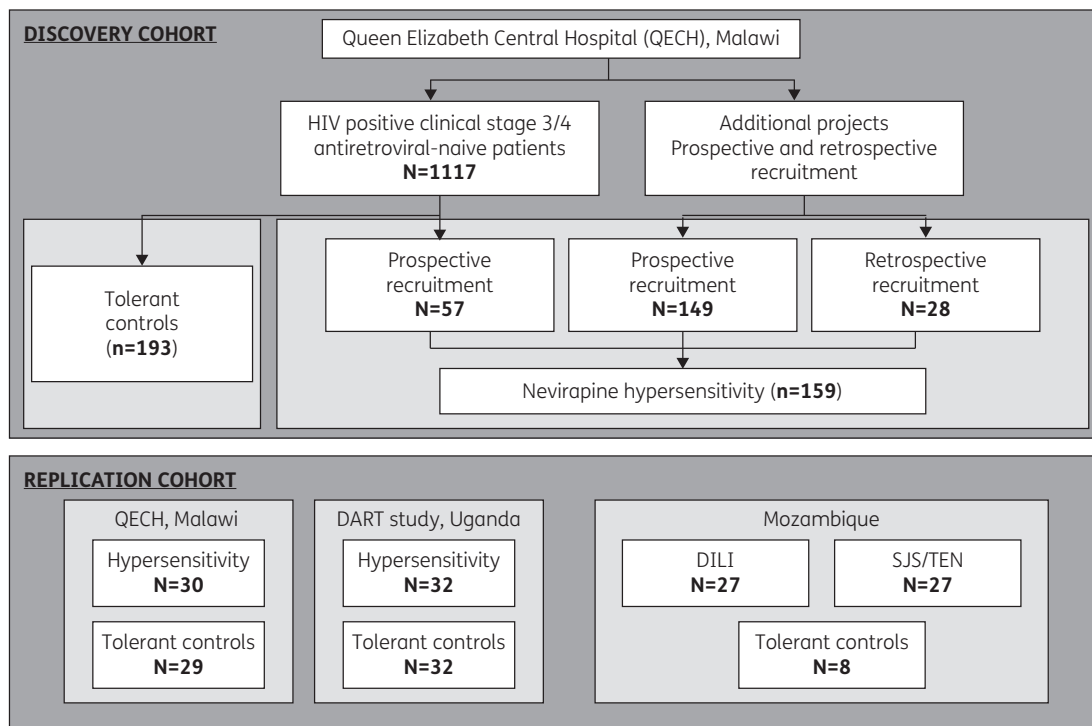


Figure 1. Schematic representation of the source of both nevirapine-hypersensitive and -tolerant patients for the GWAS discovery and replication cohorts.

Discovery cohort genotyping and sample quality control (QC)

A total of 352 samples were genotyped for 1 048 713 variants using the HumanOmni1-Quad_v1 chip (Illumina). Variants were excluded from analysis if their minor allele frequency (MAF) was $<1\%$, the call rate was $<99\%$ for an MAF between 1% and 3%, the call rate was $<98\%$ for an MAF of $>3\%$, or if Hardy–Weinberg expectations were not satisfied ($P < 10^{-4}$).

Individuals were excluded if the sample call rate was $<95\%$, the assigned gender contradicted genetic information from the X chromosome heterozygosity, or if they appeared to be duplicates, or related to other individuals in the study (as measured by identity by state using PLINK²⁷). Multidimensional scaling analysis of genotype data was undertaken by merging the data with HapMap3 cohort data and using the mds function in PLINK²⁷ in order to determine population stratification (Figure S1, available as Supplementary data at JAC Online).

Discovery cohort imputation

Imputation of genotypes, after phasing of each chromosome using ShapeIT²⁸, was carried out using IMPUTE V2.3.1,²⁹ 1000G phase 1 integrated v3 macGT1 reference panel haplotypes (March 2012).³⁰ After imputation, SNPs with an information measure (info score) <0.8 were discarded, and a threshold of 0.5 was applied on genotype uncertainty. Imputed variants with an MAF $<1\%$ were then excluded.

Discovery cohort association analysis

Univariate logistic regression analysis of non-genetic covariates (age, gender, BMI, CD4+ cell count) was undertaken for each hypersensitivity phenotype. Statistically significant variables ($P < 0.05$) were included in the subsequent logistic regressions to test for the association of each hypersensitivity phenotype with each SNP passing QC. All statistical analyses were

undertaken using PLINK²⁷ and R.³¹ Given prior associations between nevirapine hypersensitivity and HLA allele associations, it was felt reasonable to specify a Bonferroni-corrected HLA-wide significance threshold of $P < 2.5 \times 10^{-4}$, based on the presumption that there are usually <200 effective HLA allele tests.

Replication cohort genotyping, QC and association analysis

SNPs determined to have a nominally significant association with a nevirapine hypersensitivity phenotype ($P < 1 \times 10^{-5}$) in the discovery cohort were subsequently typed in the replication cohort using either the Sequenom MassArray iPLEX platform (Sequenom Inc., San Diego, CA, USA) or custom TaqMan real-time PCR SNP genotyping assays (Life Technologies, Paisley, UK) according to the manufacturer's protocols. SNPs were excluded if they failed to meet the genotype QC thresholds as outlined for the discovery cohort or if assay design software parameters prohibited their inclusion.

Logistic regression analysis of the replication cohort, including and excluding CD4+ count as a covariate, where appropriate (as determined in the discovery cohort), was carried out. Meta-analysis of combined discovery and replication cohorts was undertaken using a fixed-effects model with inverse-variance effect size weighting in GWAMA.³²

Imputation of HLA allelotype and MHC locus

Imputation of HLA-C allelotype from the discovery cohort SNP array data was undertaken using HLA*IMP:02³³ (see Supplementary data).

HLA-C and ERAP gene–gene interactions

In cases and tolerant controls positive for carriage of the rs5010528 G allele, which was used as a proxy for HLA-C*04:01, we investigated both ERAP1 (rs10050860 and rs30187) and ERAP2 (rs2248374, rs2549782) SNPs

Table 1. Non-genetic data for nevirapine-tolerant and -hypersensitive patient cohorts included in both the main and replication analyses after sample exclusion based on genotyping QC criteria

	Cases [median (range)]						Controls [median (range)]						Phenotype (n)							
	n	age	female	BMI	CD4+	n	age	female	BMI	CD4+	n	age	female	BMI	CD4+	NIR	HSS	SJS/TEN	DILI	
Discovery cohort ^a	151	35 (17–69)	63%	20.5 (14.6–41.4)	235 (3–906)	182	35 (17–63)	60%	20.2 (13.7–36.4)	170 (2–677)	56	23	51	21						
Replication cohort																				
Malawi ^b	30	36 (24–58)	48%	19.8 (13.1–26.8)	207 (15–560)	29	36 (24–56)	47%	19.9 (14.9–30.3)	145 (25–314)	13	3	9 ^c	6 ^c						
Uganda	32	37 (24–62)	73%	21.1 (8.9–39.6)	73 (17–201)	31	37 (25–53)	71%	21.3 (11.5–35.6)	66 (11–196)	14	8	2	10						
Mozambique (SJS/TEN) ^d	27	31 (21–52)	100%	22.8 (12.4–35.0)	467 (107–906)	8	31 (23–41)	100%	27.5 (26.1–35.0)	556 (244–682)	–	–	–	–	–	–	–	–	–	
Mozambique (DILI)	27	32 (23–43)	100%	23.7 (13.7–34.0)	324 (23–2305)	–	–	–	–	–	–	–	–	–	–	–	–	–	–	
combined	116	34 (21–62)	78%	21.7 (8.9–39.6)	198 (15–2305)	68	36 (23–56)	64%	20.9 (11.5–35.6)	102 (11–682)	27	10	38	42						

^aCD4+ data missing for six cases.^bCD4+ data missing for four hypersensitive patients and BMI data missing for one hypersensitive patient.^cOne patient presented with SJS/TEN and DILI phenotype.^dCD4+ data missing for three hypersensitive and four tolerant patients, and BMI data missing for six hypersensitive and four tolerant patients.

(using data from the Illumina array), which have previously been shown to interact with HLA-mediated immune diseases. Association of *ERAP1* and *ERAP2* SNPs with SJS/TEN risk was determined in the *HLA-C*04:01*-positive cohort (cases and controls) by logistic regression with CD4+ cell count as a covariate using PLINK.²⁷ A Bonferroni adjustment for multiple testing was applied with a significance threshold of $P = 0.125$.

Targeted sequencing of MHC region

Sixteen genomic DNA samples from nevirapine-induced SJS/TEN and 16 age- and gender-matched tolerant controls were carried forward for MHC-targeted sequencing. The methodology is detailed in the Supplementary data.

Allelotyping

HLA allelotyping was performed from raw FASTQ data files using Omixon Target v1.81 HLA Typing software and utilized the HLA database version 3.15.0. (Omixon Ltd, Budapest, Hungary).

In silico docking

In order to predict possible modes of interaction between nevirapine and *HLA-C*04:01*, *in silico* docking was undertaken. The methodology is detailed in the Supplementary data.

Results

Discovery cohort

A total of 333 samples (151 cases and 182 controls) out of 352 passed QC. Of the 19 excluded samples, 9 failed heterozygosity checks (outliers by >3 SD), 8 failed identity checks and 2 failed the call rate threshold. Multi-dimensional analysis for population stratification (Figure S1) demonstrated no population outliers. In total, 817 728 SNPs passed QC and were carried over for imputation with the 1000 genomes panel. Imputation produced a dataset of 1 421 851 11 variants. Cohort characteristics are shown in Table 1. We considered five nevirapine-induced hypersensitivity phenotypes for analysis—NIR, HSS, SJS/TEN, DILI (Table 1)—and also combined these different phenotypes into an overall hypersensitivity group.

Univariate logistic regression analysis showed CD4+ cell count to be a statistically significant variable for NIR ($P = 0.016$), SJS ($P = 0.003$) and all hypersensitivity cases ($P = 2.5 \times 10^{-6}$). Therefore, we included CD4+ cell count as a covariate in the SNP logistic regression model for these three phenotypes. Multidimensional scaling (MDS) variables were not included as covariates in the logistic regression since the population stratification analysis suggested that the cohort was homogeneous and genomic control was unnecessary (Figure S1). From the genome-wide logistic regression analyses, we identified 15 SNPs with $P < 1 \times 10^{-5}$, with at least one of the five different phenotypes analysed (Figure 2; summarized in Table 2). No variant reached genome-wide significance.

Replication cohort

Of the 15 SNPs considered for replication, one (rs150223496) could not be typed due to proximal sequence constraints of the Sequenom assay design process, and QC failure for TaqMan genotyping Hardy-Weinberg equilibrium (HWE)

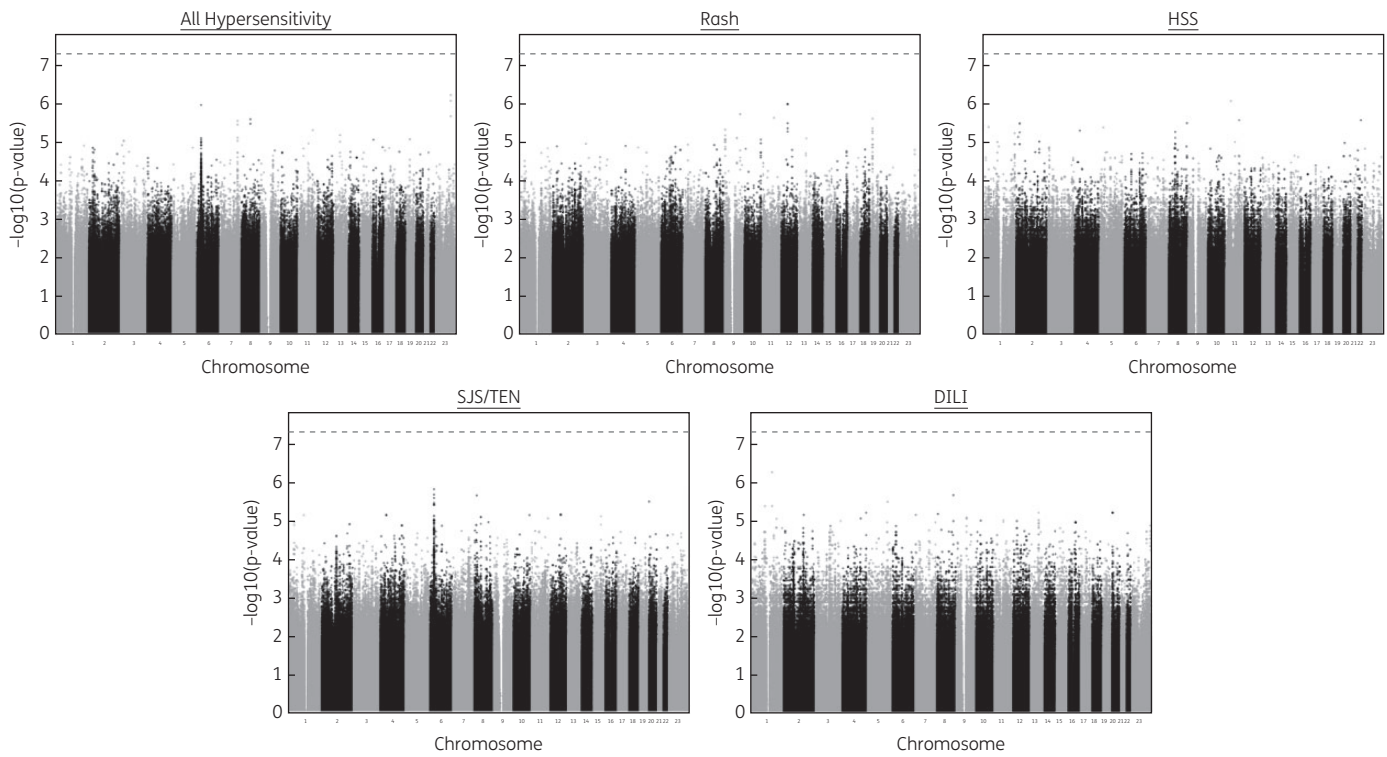


Figure 2. Manhattan plots of association for logistic regression SNP analysis for the five defined nevirapine hypersensitivity phenotypes. The phenotypes, all hypersensitivity, NIR and SJS/TEN show P values from logistic regression incorporating CD4+ count as a covariate. For HSS and DILI no covariates were incorporated. The broken line indicates genome-wide significance ($P = 5 \times 10^{-8}$).

Table 2. Top SNPs identified in association with a nevirapine hypersensitivity phenotype within the main cohort analysis

Phenotype	SNP	Chr	Position (GRCh37 .p13)	Reference allele	Associated allele	Gene	Typed/ imputed	Logistic regression	
								P value	OR (95% CI)
All hypersensitivity (n = 151)	rs34213790	6	31318579	G	A	3' of <i>HLA-B</i>	imputed	1.15×10^{-6}	2.34 (1.66–3.30)
	rs11764223	7	137742056	T	G	<i>AKR1D1</i>	imputed	2.99×10^{-6}	0.45 (0.33–0.63)
	rs11988543	8	68596362	T	A	<i>CPA6</i>	imputed	2.69×10^{-6}	3.37 (2.03–5.60)
	rs9527426	13	56208216	T	C	<i>MIR5007</i>	typed	9.86×10^{-6}	2.25 (1.58–3.21)
	rs35990155	23	11827017	G	T	<i>KIAA1210</i>	imputed	6.33×10^{-7}	5.65 (2.86–11.17)
Rash (n = 56)	rs10815440	9	661057	G	A	<i>KANK1</i>	typed	8.86×10^{-6}	5.80 (2.68–12.67)
	rs115848367	10	128045499	A	C	<i>ADAM12</i>	imputed	8.83×10^{-7}	4.70 (2.38–9.30)
	rs74373347	12	48919428	A	G	<i>OR8S1</i>	imputed	1.08×10^{-6}	11.05 (4.12–29.04)
	rs6511720	19	11202306	G	T	<i>LDLR</i>	typed	5.49×10^{-6}	4.66 (2.40–9.05)
SJS/TEN (n = 51)	rs5010528	6	31241032	A	G	<i>HLA-C</i>	typed	4.13×10^{-6}	4.75 (2.45–9.22)
	rs150223496	15	76421885	G	A	<i>C15orf27</i>	imputed	8.20×10^{-6}	17.34 (4.95–60.74)
DILI (n = 21)	rs147773805	1	107741393	A	T	<i>NTNG1</i>	imputed	4.47×10^{-6}	11.05 (3.96–30.83)
	rs114693001	9	76074363	A	T	intergenic	imputed	8.82×10^{-6}	9.35 (3.49–25.06)
	rs142213069	13	77962182	G	C	5' of <i>MYCBP2</i>	imputed	6.64×10^{-6}	14.42 (4.51–46.04)
	rs6139258	20	3958616	T	C	<i>RNF24</i>	typed	6.64×10^{-6}	14.42 (4.52–46.04)

P values and ORs were determined by logistic regression with CD4+ cell count as a covariate (except for HSS and DILI).

Table 3. Logistic regression analysis of candidate SNPs in the nevirapine hypersensitivity replication cohort

Phenotype	SNP	Chr	Position (GRCh37 .p13)	Genotyping platform	Reference allele	Associated allele	Gene	Logistic regression	
								P value	OR (95% CI)
All hypersensitivity (n = 62)	rs34213790	6	31318579	iPLEX	G	A	3' of HLA-B	0.10	1.56 (0.94–2.60)
	^a rs12112517	7	137743167	iPLEX	A	C	AKR1D1	0.21	1.42 (0.82–2.46)
	rs11988543	8	68596362	TaqMan	T	A	CPA6	0.77	1.13 (0.54–2.28)
	rs9527426	13	56208216	iPLEX	T	C	MIR5007	0.83	1.06 (0.62–1.80)
	rs35990155	23	11827017	iPLEX	G	T	KIAA1210	0.86	0.94 (0.48–1.86)
Rash (n = 27)	rs10815440	9	661057	iPLEX	G	A	KANK1	0.99	3.3 × 10 ⁻¹⁰ (0–∞)
	rs115848367	10	128045499	iPLEX	A	C	ADAM12	0.70	1.18 (0.51–2.70)
	rs74373347	12	48919428	iPLEX	A	G	OR8S1	0.76	0.81 (0.21–3.11)
	rs6511720	19	11202306	iPLEX	G	T	LDLR	0.86	1.07 (0.51–2.24)
SJS/TEN (n = 38)	rs5010528	6	31241032	TaqMan	A	G	HLA-C	0.02	5.33 (1.37–20.80)
	^b rs150223496	15	76421885	NA	G	A	C15orf27	NA	NA
DILI (n = 42)	rs1730858	1	107619244	iPLEX	G	A	NTNG1	0.61	1.57 (0.27–8.97)
	^c rs114693001	9	76074363	TaqMan	A	T	intergenic	0.99	2.6 × 10 ⁻⁹ (0–∞)
	^c rs142213069	13	77962182	iPLEX	G	C	5' of MYCBP2	0.99	2.4 × 10 ⁻⁹ (0–∞)
	rs6139258	20	3958616	iPLEX	T	C	RNF24	0.03	11.17 (1.29–96.38)

SNP associations below arbitrary statistical significance ($P < 0.1$) are highlighted in bold. NA denotes statistical analysis not applicable.

^aSNPs within the association signal that are substituted from the discovery cohort SNP (high LD) due to genotyping assay design constraints.

^bSNP signal where replication in the replication cohort was not possible.

^cSNPs that were not be typed in the Mozambique DILI cohort (n = 15 cases). Mozambique individuals were also omitted from the 'all hypersensitivity' analysis.

Table 4. Meta-analysis of significantly associated SNPs in the discovery and replication cohorts

SNP	Gene	Phenotype	Cohort	Case/ control	n	MAF	Logistic regression		Meta-analysis	
							P	OR (95% CI)	P	OR (95% CI)
rs34213790	3' of HLA-B	all hypersensitivity	discovery	case	151	0.45	1.15 × 10 ⁻⁶	2.34 (1.66–3.30)	–	–
			replication	control	182	0.37				
rs5010528	HLA-C	SJS/TEN	discovery	case	27	0.50	0.10	1.56 (0.94–2.60)	6.69 × 10 ⁻⁷	2.06 (1.55–2.74)
				control	60	0.38				
			replication	case	51	0.36	4.13 × 10 ⁻⁶	4.75 (2.45–9.22)	–	–
				control	182	0.14				
rs6139258	RNF24	DILI	discovery	case	38	0.38	6.03 × 10 ⁻³	5.12 (1.60–16.41)	8.47 × 10 ⁻⁸	4.84 (2.71–8.61)
				control	68	0.12				
			replication	case	21	0.19	6.64 × 10 ⁻⁶	14.42 (4.52–46.04)	–	–
				control	182	0.02				
			replication	case	42	0.07	0.03	11.17 (1.29–96.38)	5.66 × 10 ⁻⁷	13.62 (4.90–37.84)
				control	68	0.01				

Data shown are for analysis undertaken with covariates (CD4+ cell count) included in the regression model (except DILI) for both the discovery and replication cohorts (± supplementary cohort for SJS/TEN and DILI).

P value >0.0001, call rate >90%. Thus, 14 SNPs were carried forward for analysis. A total of 59 Malawian samples (30 cases and 29 controls) and 63 Ugandan samples (32 cases and 31 controls) passed QC (call rate >90%) as described in Table 1. Due to sample constraints, SNP signals identified in the discovery cohort for the 'all hypersensitivity' phenotype were not typed in the samples from Mozambique in the replication cohort.

For nevirapine-induced DILI (Table 3), combining the discovery and replication cohorts for the SNP rs6139258 in the RNF24 locus strengthened the association [$P = 5.7 \times 10^{-7}$, OR 13.62 (95% CI 4.90–37.84)] (Table 4). A single SNP (rs5010528 in the HLA-C locus) showed an association with SJS/TEN in the replication cohort (38 cases, 59 controls) [$P = 0.006$; OR 5.12 (95% CI 1.60–16.42)]. Combining the discovery and replication cohorts strengthened the

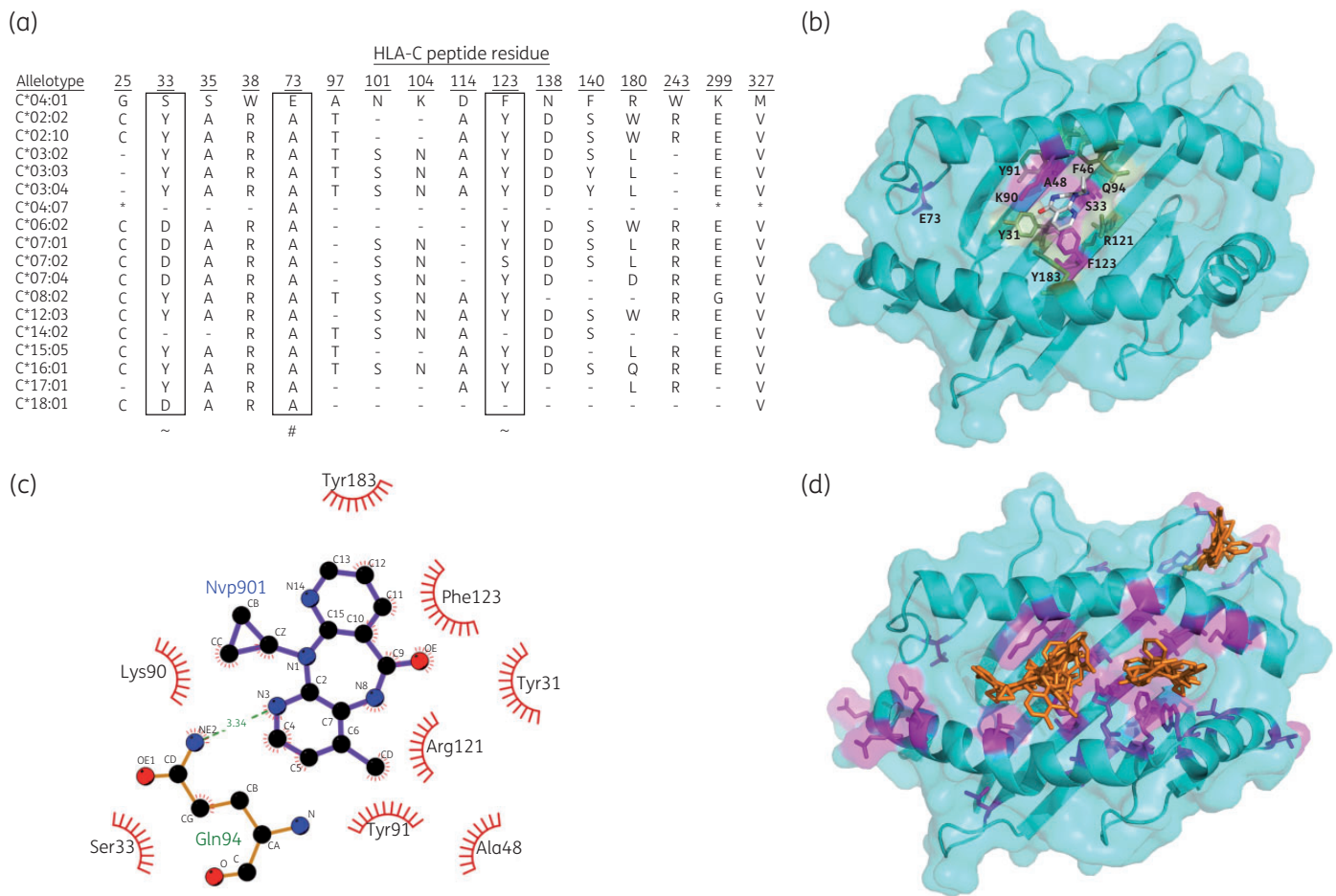


Figure 3. *In silico* docking of HLA-C*04:01 and nevirapine. (a) Specific peptide residues present in the different HLA-C allelotypes previously identified in the Malawian cohort ($n = 116$). - shows continuity with the reference peptide (C*04:01) and * identifies peptides not sequenced in an allelotypes. ~ denotes putative nevirapine interaction and # denotes residue substituted by SNP (rs1050409) identified in GWAS. Residue reference numbers are as defined by the *in silico* model. (b) HLA-C*04:01/nevirapine docking mode of conformation with the highest predicted affinity (lowest score) as produced using the PyMOL software. Key interacting residues are highlighted. (c) A LIGPLOT+⁴⁰ 2D schematic representation of the interaction of nevirapine with HLA-C*04:01 PBD residues in the highest affinity predicted docking conformation. The broken line indicates a hydrogen bond and radiating lines indicate hydrophobic interactions. (d) HLA-C*04:01/nevirapine (orange) docking mode of conformation for the 20 highest predicted affinity (lowest scores) as produced using the PyMOL software.

overall association, which approached genome-wide significance [$P = 8.5 \times 10^{-8}$, OR 4.84 (95% CI 2.71–8.61)]. No positive signals were identified for the ‘all hypersensitivity’ phenotype (62 cases, 59 controls).

HLA-C allelotype imputation

Overall allelotype imputation from the SNP array data using HLA*IMP demonstrated 71.5% concordance with the HLA typing obtained for 116 of our patients using the sequence-based methodology. However the ability of the imputation to correctly call HLA-C*04:01 alleles was 90%.

Within the 116 patients for which HLA allelotyping and SNP array genotype data were available, HLA-C-04:01 allele carriage co-occurred with the rs5010528 G allele in 112/116 cases (96.5%). For the imputed HLA allelotype data, C*04:01 co-occurred with rs5010528 G in 303/333 cases (91%).

The initial discovery logistic regression analysis demonstrated two non-synonymous SNPs in the HLA-C locus associated with SJS/TEN that were in absolute linkage disequilibrium (LD) with rs5010528 (Table 5). The first SNP (rs146911342) encodes a valine-to-methionine amino acid substitution at residue 327 and the second (rs1050409) encodes an alanine to glutamic acid at residue 73 (close to the peptide binding domain), which was also associated with SJS/TEN [$P = 4.1 \times 10^{-6}$, OR 4.75 (95% CI 2.45–9.23)]. Both are key defining residues of the HLA-C*04 allelotypes as defined in the HLA-IMGT database³⁴ and within the Malawian cohort (Figure 3). Verification of either SNP in our discovery or replication cohorts via other genotyping methodologies was not possible due to sequence constraints in assay design (the proximal nucleotide sequence for primer design was not sufficiently unique or contained a restrictive number of other genetic variants).

However, targeted sequencing of the HLA locus in 16 SJS/TEN and 16 tolerant controls (Table 5) suggested that both the

Table 5. Association of nevirapine-induced SJS/TEN and imputed SNPs of the HLA-C locus from the main cohort and the targeted sequencing cohort

SNP	GWAS (discovery cohort)										Targeted sequencing cohort				LD with rs5010528 (<i>D'</i>)
	bp (GRCh37.p13)	A ₁ /A ₂	Amino acid substitution	typed/imputed	SJS/TEN MAF (n=51)	control MAF (n=182)	<i>P</i>	OR (95% CI)	SJS/TEN MAF (n=16)	control MAF (n=16)	<i>P</i>	OR (95% CI)			
rs146911342	31 237 779	C/T	p.V327M	imputed	0.36	0.15	4.13×10^{-6}	4.75 (2.45-9.23)	0.34	0.13	0.013	34.14 (2.07-561.6)	1		
rs41562714	31 238 538	G/C		imputed	0.36	0.15	4.13×10^{-6}	4.75 (2.45-9.23)	0.34	0.13	0.013	34.14 (2.07-561.6)	1		
rs1050409	31 239 501	G/T	p.A73E	imputed	0.36	0.15	4.13×10^{-6}	4.75 (2.45-9.23)	0.34	0.13	0.013	34.14 (2.07-561.6)	1		
rs41553018	31 239 742	G/C		imputed	0.36	0.15	3.71×10^{-6}	4.75 (2.45-9.23)	0.34	0.13	0.013	34.14 (2.07-561.6)	1		
rs4361609	31 240 635	G/C		imputed	0.36	0.15	4.13×10^{-6}	4.75 (2.45-9.23)	0.34	0.13	0.013	34.14 (2.07-561.6)	1		
rs5010528	31 241 032	A/G		typed	0.36	0.15	4.13×10^{-6}	4.75 (2.45-9.23)	0.34	0.13	0.013	34.14 (2.07-561.6)	-		
rs58019823	31 241 215	A/G		imputed	0.36	0.15	4.13×10^{-6}	4.75 (2.45-9.23)	-	-	-	-	NA		
^a rs2524087	31241 294	C/G		imputed	0.37	0.15	3.71×10^{-6}	4.73 (2.45-9.14)	-	-	-	-	NA		
^a rs59103503	31 287 944	G/T		imputed	0.37	0.15	2.75×10^{-6}	5.00 (2.55-9.80)	-	-	-	-	NA		
^a rs77641320	31 298 229	C/G		imputed	0.35	0.14	1.64×10^{-6}	5.53 (2.75-11.12)	-	-	-	-	NA		
^a rs9468965	31 300 247	T/A		imputed	0.44	0.19	2.24×10^{-6}	4.61 (2.24-8.69)	-	-	-	-	NA		
^a rs33564987	31 309 423	T/C		imputed	0.49	0.27	8.01×10^{-6}	3.76 (2.10-6.74)	-	-	-	-	NA		
^a rs35435945	31 311 374	T/G		imputed	0.49	0.27	9.83×10^{-6}	3.72 (2.08-6.67)	-	-	-	-	NA		
^a rs35278939	31 319 780	G/A		imputed	0.47	0.23	8.01×10^{-6}	4.04 (2.19-7.47)	-	-	-	-	NA		

The list comprises all SNPs with a *P* value of $<1 \times 10^{-5}$ in the main cohort analysis. Statistical significance and OR (95% CI) for the targeted sequencing cohort is determined by logistic regression with CD4+ cell count as covariate. NA denotes LD indeterminate in targeted sequencing cohort as SNP typing not available.

^aSNP was imputed in the discovery cohort analysis but not detected and/or called in the sequencing data.

imputed non-synonymous SNPs may be in absolute LD with the original discovery cohort signal SNP (rs5010528), again demonstrating a significant association with nevirapine-induced SJS/TEN. Allelotype inference from the targeted sequencing SNP data also confirmed that both non-synonymous SNPs were in 100% co-occurrence with *HLA-C*04:01* (data not shown). Our data do not show an association between any of the other *HLA* gene loci and other nevirapine-induced hypersensitivity phenotypes.

HLA-C*04:01 and ERAP1 and ERAP2 SNP interactions

Given the previously reported interactions between ERAP genes and HLA class I-mediated diseases, in particular ankylosing spondylitis,³⁵ we determined whether there was an interaction with the carriage of *HLA-C*04:01* using the rs5010528 G allele as a proxy SNP (Table S1). There was no significant association ($P > 0.05$) between the *ERAP1* variants and SJS/TEN risk in carriers of *HLA-C*04:01*. However, both *ERAP2* variants showed a nominal association with SJS/TEN risk [$P = 0.019$, OR 0.43 (95% CI 0.21–0.87)], though this did not pass the Bonferroni threshold for multiple testing ($P = 0.0125$).

In silico docking

In light of the association observed between nevirapine-induced SJS/TEN and an SNP (rs1050409) encoding an amino acid substitution at residue 73 of HLA-C (p.A73E), *in silico* docking was undertaken to determine the possible effect of the residue substitution on nevirapine binding. The data suggest that none of the predicted modes of nevirapine docking conformation interact with residue 73, which appears to be on the periphery of the peptide-binding domain (Figure 3). The lowest scoring (predicted highest affinity) mode highlights an interaction between nevirapine and residues 33 and 123 in the B pocket (Figure 3). In *HLA-C*04:01*, residues 33 and 123 are serine and phenylalanine respectively (Figure 3). The majority of other allelotypes do not possess these particular residues (with the exception of *C*04:07* and *C*14:02*). Docking of the metabolite 12-hydroxy-nevirapine was also undertaken, since it has also been suggested as potentially responsible for nevirapine-induced adverse drug reactions,³⁶ these were in general agreement with those for nevirapine, in that docking seems to take place around the B pocket (e.g. near residues 33 and 123), but with more variability in the different modes predicted than for nevirapine. None of the predicted modes interacted with residue 73 (data not shown). Taken together, the docking results suggest that binding of either nevirapine or 12-hydroxy-nevirapine around the centre of the peptide-binding regions is likely to be important in the mechanism of the immune-mediated reaction.

Discussion

The investigation of genetic factors predisposing to serious adverse drug reactions is challenging because of their rarity. Despite this, we have assembled one of the largest cohorts of patients with clinically well-characterized nevirapine hypersensitivity, including SJS/TEN. GWAS analysis of our Malawian discovery cohort ($n = 333$) identified 15 polymorphisms having a suggestive association with nevirapine hypersensitivity (Table 2). Subsequent analysis of these variants in our replication cohort suggested that

three of the SNPs may be potential risk factors (Table 3): rs34213790 3' of the *HLA-B* gene locus with all hypersensitivity phenotypes; rs5010528 in the *HLA-C* gene locus with SJS/TEN, and rs6139258 in the *RNF24* gene locus with DILI. The weakest of the above three association signals, SNP rs34213790, is unlikely to be an independent marker of nevirapine hypersensitivity in general, and its association may be due to a haplotype effect between *HLA-C*04:01* (rs5010528; see below) and B allelotypes.

SNP rs6139258 in the *RNF24* gene locus only marginally failed to pass the Bonferroni threshold of significance in the replication cohort. Very little is known regarding the function of *RNF24*. However, it is known that it is a protein that interacts with transient receptor potential cation channel 6 (TRPC6),³⁷ a receptor-activated channel, expressed in liver cells,³⁸ which plays a role in cellular calcium homeostasis. *TRPC6* has been suggested to play a role in hepatoma cell-line proliferation, possibly via a cyclin D-modulated mechanism.³⁸ Thus *RNF24* may have some biological plausibility in the pathogenesis of nevirapine-induced liver injury, and merits further investigation in additional patients with nevirapine-induced DILI, and functional work to uncover the possible mechanisms (if any) of the association.

The most compelling of the three signals, rs5010528, gave an OR of 4.84 for nevirapine-induced SJS/TEN, and was replicated in patients from three countries (Malawi, Uganda and Mozambique) at the Bonferroni threshold ($P < 0.05$), approaching genome-wide significance in the combined analysis ($P = 8.5 \times 10^{-8}$) (Table 4). SNP rs5010528 is located within the *HLA-C* gene locus. High co-occurrence of rs5010528 with *HLA-C*04:01* was observed (96.5%) in 116 patients within this study who had previously been HLA typed by sequence-based methods. The association between nevirapine and rs5010528 (as a proxy for *C*04:01*) can be considered statistically significant when applying an HLA-wide significance threshold of $P < 2.5 \times 10^{-4}$. Additionally, *HLA-C* allelotypes imputed from the SNP array data also showed a high co-occurrence (91%) in the main study cohort, suggesting rs5010528 may be a good proxy for *HLA-C*04:01*. Thus, the GWAS data appear to confirm our previous finding²¹ associating *HLA-C*04:01* with nevirapine-induced SJS/TEN. Of note, the association of rs5010528 with other hypersensitivity phenotypes was not as strong, suggesting that the risk conferred by rs5010528 and thus *HLA-C*04:01* is specific for nevirapine-induced SJS/TEN. The reason for this is unclear, and will require further investigation.

In terms of clinical utility, rs5010528 appears to have little potential as a pre-emptive genetic test. Indeed, based on a prevalence of SJS/TEN in our prospective cohort of 1.07% and assuming a dominant mode of inheritance, the positive (PPV) and negative (NPV) predictive values were 2.8% and 42.4% respectively. For the *RNF24* variant (rs6139258) the PPV, based on a prevalence of DILI of 0.63%, is also very low (0.2%).

Only one previous GWAS investigating nevirapine hypersensitivity has been reported, but in a smaller Thai population (72 cases, 77 controls).³⁹ Patients had a wide variety of rashes, with only 11 grade 4 cases (6.9%), which would be equivalent to our cases with SJS/TEN. The SNP rs9461684 in the *HLA-C* locus was significantly associated with nevirapine rash, but no *HLA* allelotype imputation or *HLA* sequencing was carried out. In our data, rs9461684 is in high LD with our top SNP, rs5010528 ($D' = 1.0$, $r^2 = 0.972$). The discrepancy may be a result of the different LD patterns in the

different ethnic groups studied, as well as the much lower numbers of patients with serious skin reactions in the Thai study.

From the imputed SNP data of the discovery cohort and targeted resequencing data (Table 5), it is clear that rs5010528 is in LD with a functional non-synonymous SNP (rs1050409) that leads to an alanine-to-glutamic acid substitution at residue 73, which lies near to the peptide-binding domain of the HLA-C protein. However, *in silico* modelling suggested that this residue does not interact with nevirapine in any docking conformation. Two other residues of *HLA-C*04:01* (33 and 123 in the model) appear to be the key interactors in the majority of the predicted modes of nevirapine docking (Figure 3), including the most favoured. However, it should be noted that this is a predictive model and further analysis of the HLA-C/nevirapine complex is needed to further elucidate the potential for docking. The association signal at residue 73 (rs1050409) is likely to be a proxy for the 33 and 123 residues also present in *HLA-C*04:01*. However, this work has provided the first evidence that nevirapine binds to the B pocket of *HLA-C*04:01*.

ERAP gene variants interact in a protective manner in HLA-mediated diseases such as ankylosing spondylitis in individuals who carry the risk *HLA* alleles.³⁵ *ERAP1* and *ERAP2* are enzymes involved in antigenic peptide precursor trimming prior to loading into HLA class I molecules (and may thus alter the peptidome) and may potentially also alter the expression of the risk HLA class I allele. To our knowledge, this is one of the first examinations of whether there is interaction between drug-induced HLA disease and the *ERAP* genes. We were, however, unable to detect an interaction between *ERAP1* variants and *HLA-C*04:01* in African patients with SJS/TEN. However, a nominal association ($P=0.019$) was observed for both *ERAP2* SNPs (Table S1). A limitation of our analysis is the small sample size, particularly given the much larger numbers that have been studied in ankylosing spondylitis. Nevertheless, the possibility of an association with *ERAP2* is intriguing, and needs further investigation not only with nevirapine-induced hypersensitivity, but also with other HLA-related adverse drug reactions.

In identifying an SNP in the *HLA-C* locus that appears to be a proxy for the *HLA-C*04:01* allele, as a risk factor for nevirapine-induced SJS/TEN, this study has added further weight to existing evidence. The data generated also suggest that, in sub-Saharan African HIV patients, no other strong, significant genetic risk factors for nevirapine hypersensitivity exist that could be utilized as clinical predictive markers. However, the data are valuable in terms of the mechanistic insights they provide. Additionally, *in silico* analysis has identified two putative HLA-C peptide residues that are predicted to be key for the binding of nevirapine, which warrant further investigation as to their role in the pathogenesis of SJS/TEN. Further work is also needed to determine the reasons for organ-specific toxicities in different patients.

Acknowledgements

We thank the patients and staff of the ART clinic of Queen Elizabeth Central Hospital, Blantyre, in particular Mr S. Kaunda, Clinical Officer. We also thank Dr Christiane Hertz-Fowler, Dr Margaret Hughes, Dr Lisa Olohan and Dr Anita Lucaci from the Centre for Genomic Research for undertaking the library preparation and sequencing of the MHC region in the cohort of 32 DNA samples.

Funding

The initial GWAS was funded by the International Serious Adverse Events Consortium (iSAEC). The iSAEC is a non-profit organization dedicated to identifying and validating DNA variants useful in predicting the risk of drug-related serious adverse events. The Consortium brings together the pharmaceutical industry, regulatory authorities and academic centres to address clinical and scientific issues associated with the genetics of drug-related serious adverse events. The iSAEC's current funding members include: Abbott, Amgen, AstraZeneca, Daiichi Sankyo, GlaxoSmithKline, Merck, Novartis, Pfizer, Takeda and the Wellcome Trust. Mas Chavonda was funded by a 3 year Wellcome Trust training fellowship WT078857MA administered through the University of Liverpool. Malawi-Liverpool-Wellcome Trust Clinical Research Programme is funded through a Core Programme Grant award from the Wellcome Trust. Munir Pirmohamed is a National Institute for Health Research Senior Investigator, and also wishes to thank the MRC Centre for Drug Safety Science for support.

The DART study was supported by the UK Medical Research Council (grant number G0600344), the UK Department for International Development and the Rockefeller Foundation.

Andrew P. Morris is a Wellcome Trust Senior Research Fellow in Basic Biomedical Science (grant number WT098017).

Louise Y. Takeshita is funded by a PhD fellowship from CNPq (National Council for Scientific and Technological Development, Brazil).

Panos Deloukas' work forms part of the research themes contributing to the translational research portfolio of Barts Cardiovascular Biomedical Research Unit which is supported and funded by the National Institute for Health Research.

Transparency declarations

All authors: no conflicts of interest to declare.

GlaxoSmithKline, Gilead Sciences, Boehringer-Ingelheim and AbbVie donated drugs for the DART study.

Supplementary data

Figure S1 and Table S1 are available as Supplementary data at JAC Online.

References

- van Leth F, Phanuphak P, Ruxrungtham K *et al.* Comparison of first-line antiretroviral therapy with regimens including nevirapine, efavirenz, or both drugs, plus stavudine and lamivudine: a randomised open-label trial, the 2NN Study. *Lancet* 2004; **363**: 1253–63.
- Siegfried NL, Van Deventer PJ, Mahomed FA *et al.* Stavudine, lamivudine and nevirapine combination therapy for treatment of HIV infection and AIDS in adults. *Cochrane Database Syst Rev* 2006; **19**: issue CD004535.
- Phillips E, Gutierrez S, Jahnke N *et al.* Determinants of nevirapine hypersensitivity and its effect on the association between hepatitis C status and mortality in antiretroviral drug-naïve HIV-positive patients. *AIDS* 2007; **21**: 1561–8.
- Wit FW, Kesselring AM, Gras L *et al.* Discontinuation of nevirapine because of hypersensitivity reactions in patients with prior treatment experience, compared with treatment-naïve patients: the ATHENA cohort study. *Clin Infect Dis* 2008; **46**: 933–40.
- Fagot JP, Mockenhaupt M, Bouwes-Bavinck JN *et al.* Nevirapine and the risk of Stevens-Johnson syndrome or toxic epidermal necrolysis. *AIDS* 2001; **15**: 1843–8.

- 6** Mittmann N, Knowles SR, Koo M *et al.* Incidence of toxic epidermal necrolysis and Stevens-Johnson syndrome in an HIV cohort: an observational, retrospective case series study. *Am J Clin Dermatol* 2012; **13**: 49–54.
- 7** De Maat MM, Mathot RA, Veldkamp AI *et al.* Hepatotoxicity following nevirapine-containing regimens in HIV-1-infected individuals. *Pharmacol Res* 2002; **46**: 295–300.
- 8** Riska P, Lamson M, MacGregor T *et al.* Disposition and biotransformation of the antiretroviral drug nevirapine in humans. *Drug Metab Dispos* 1999; **27**: 895–901.
- 9** Jinno H, Tanaka-Kagawa T, Ohno A *et al.* Functional characterization of cytochrome P450 2B6 allelic variants. *Drug Metab Dispos* 2003; **31**: 398–403.
- 10** Lang T, Klein K, Fischer J *et al.* Extensive genetic polymorphism in the human CYP2B6 gene with impact on expression and function in human liver. *Pharmacogenetics* 2001; **11**: 399–415.
- 11** Rotger M, Colombo S, Furrer H *et al.* Influence of CYP2B6 polymorphism on plasma and intracellular concentrations and toxicity of efavirenz and nevirapine in HIV-infected patients. *Pharmacogenet Genomics* 2005; **15**: 1–5.
- 12** Penzak SR, Kabuye G, Mugenyi P *et al.* Cytochrome P450 2B6 (CYP2B6) G516T influences nevirapine plasma concentrations in HIV-infected patients in Uganda. *HIV Med* 2007; **8**: 86–91.
- 13** Yuan J, Guo S, Hall D *et al.* Toxicogenomics of nevirapine-associated cutaneous and hepatic adverse events among populations of African, Asian, and European descent. *AIDS* 2011; **25**: 1271–80.
- 14** Haas DW, Bartlett JA, Andersen JW *et al.* Pharmacogenetics of nevirapine-associated hepatotoxicity: an Adult AIDS Clinical Trials Group collaboration. *Clin Infect Dis* 2006; **43**: 783.
- 15** Martin AM, Nolan D, James I *et al.* Predisposition to nevirapine hypersensitivity associated with HLA-DRB1*0101 and abrogated by low CD4 T-cell counts. *AIDS* 2005; **19**: 97–9.
- 16** Vitezica ZG, Milpied B, Lonjou C *et al.* HLA-DRB1*01 associated with cutaneous hypersensitivity induced by nevirapine and efavirenz. *AIDS* 2008; **22**: 540–1.
- 17** Likanonsakul S, Rattanatham T, Feangvad S *et al.* HLA-Cw*04 allele associated with nevirapine-induced rash in HIV-infected Thai patients. *AIDS Res Ther* 2009; **6**: 22.
- 18** Gao S, Gui XE, Liang K *et al.* HLA-dependent hypersensitivity reaction to nevirapine in Chinese Han HIV-infected patients. *AIDS Res Hum Retroviruses* 2011; **28**: 540–3.
- 19** Gatanaga H, Yazaki H, Tanuma J *et al.* HLA-Cw8 primarily associated with hypersensitivity to nevirapine. *AIDS* 2007; **21**: 264–5.
- 20** Chantarangsu S, Mushiroda T, Mahasirimongkol S *et al.* HLA-B*3505 allele is a strong predictor for nevirapine-induced skin adverse drug reactions in HIV-infected Thai patients. *Pharmacogenet Genomics* 2009; **19**: 139–46.
- 21** Carr DF, Chaponda M, Jorgensen AL *et al.* Association of human leukocyte antigen alleles and nevirapine hypersensitivity in a Malawian HIV-infected population. *Clin Infect Dis* 2013; **56**: 1330.
- 22** Kenna TJ, Robinson PC, Haroon N. Endoplasmic reticulum aminopeptidases in the pathogenesis of ankylosing spondylitis. *Rheumatology (Oxford)* 2015; **54**: 1549–56.
- 23** Mugenyi P, Walker AS, Hakim J *et al.* Routine versus clinically driven laboratory monitoring of HIV antiretroviral therapy in Africa (DART): a randomised non-inferiority trial. *Lancet* 2010; **375**: 123–31.
- 24** Ciccacci C, Borgiani P, Ceffa S *et al.* Nevirapine-induced hepatotoxicity and pharmacogenetics: a retrospective study in a population from Mozambique. *Pharmacogenomics* 2010; **11**: 23–31.
- 25** Ciccacci C, Di Fusco D, Marazzi MC *et al.* Association between CYP2B6 polymorphisms and nevirapine-induced SJS/TEN: a pharmacogenetics study. *Eur J Clin Pharmacol* 2013; **69**: 1909–16.
- 26** Munderi P, Snowden WB, Walker AS *et al.* Distribution of HLA-B alleles in a Ugandan HIV-infected adult population: NORA pharmacogenetic substudy of DART. *Trop Med Int Health* 2011; **16**: 200–4.
- 27** Purcell S, Neale B, Todd-Brown K *et al.* PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007; **81**: 559–75.
- 28** Delaneau O, Zagury JF, Marchini J. Improved whole-chromosome phasing for disease and population genetic studies. *Nat Methods* 2013; **10**: 5–6.
- 29** Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet* 2009; **5**: e1000529.
- 30** Marchini J, Howie B. Genotype imputation for genome-wide association studies. *Nat Rev Genet* 2010; **11**: 499–511.
- 31** R Core Team (2013). *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing. ISBN 3-900051-07-0. <http://www.R-project.org>.
- 32** Magi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. *BMC Bioinformatics* 2010; **11**: 288.
- 33** Dilthey A, Leslie S, Moutsianas L *et al.* Multi-population classical HLA type imputation. *PLoS Comput Biol* 2013; **9**: e1002877.
- 34** Robinson J, Halliwell JA, Hayhurst JD *et al.* The IPD and IMGT/HLA database: allele variant databases. *Nucleic Acids Res* 2015; **43**: D423–31.
- 35** Evans DM, Spencer CC, Poinon JJ *et al.* Interaction between ERAP1 and HLA-B27 in ankylosing spondylitis implicates peptide handling in the mechanism for HLA-B27 in disease susceptibility. *Nat Genet* 2011; **43**: 761–7.
- 36** Sharma AM, Novalen M, Tanino T *et al.* 12-OH-nevirapine sulfate, formed in the skin, is responsible for nevirapine-induced skin rash. *Chem Res Toxicol* 2013; **26**: 817–27.
- 37** Lussier MP, Lepage PK, Bousquet SM *et al.* RNF24, a new TRPC interacting protein, causes the intracellular retention of TRPC. *Cell Calcium* 2008; **43**: 432–43.
- 38** El Boustany C, Bidaux G, Enfissi A *et al.* Capacitative calcium entry and transient receptor potential canonical 6 expression control human hepatoma cell proliferation. *Hepatology* 2008; **47**: 2068–77.
- 39** Chantarangsu S, Mushiroda T, Mahasirimongkol S *et al.* Genome-wide association study identifies variations in 6p21.3 associated with nevirapine-induced rash. *Clin Infect Dis* 2011; **53**: 341–8.
- 40** Laskowski RA, Swindells MB. LigPlot+: multiple ligand-protein interaction diagrams for drug discovery. *J Chem Inf Model* 2011; **51**: 2778–86.