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Ghler, A; Trung, TT; Hopf, V; Kohler, C; Hartleib, J; Wuthiekannun, V; Peacock, SJ; Limmathurotsakul, D; Tuanyok, A; Steinmetz, I (2017) Multitarget Quantitative PCR Improves Detection and Predicts Cultivability of the Pathogen *Burkholderia pseudomallei*. *Applied and environmental microbiology*, 83 (8). ISSN 0099-2240 DOI: <https://doi.org/10.1128/AEM.03212-16>

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Multitarget Quantitative PCR Improves Detection and Predicts Cultivability of the Pathogen *Burkholderia pseudomallei*

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ABSTRACT *Burkholderia pseudomallei* is present in the environment in many parts of the world and causes the often-fatal disease melioidosis. The sensitive detection and quantification of *B. pseudomallei* in the environment are a prerequisite for assessing the risk of infection. We recently reported the direct detection of *B. pseudomallei* in soil samples using a quantitative PCR (qPCR) targeting a single type three secretion system 1 (TTSS1) gene. Here, we extend the qPCR-based analysis of *B. pseudomallei* in soil by validating novel qPCR gene targets selected from a comparative genomic analysis. Two hundred soil samples from two rice paddies in northeast Thailand were evaluated, of which 47% (94/200) were *B. pseudomallei* culture positive. The TTSS1 qPCR and two novel qPCR assays that targeted open reading frames (ORFs) BPSS0087 and BPSS0745 exhibited detection rates of 76.5% (153/200), 34.5% (69/200), and 74.5% (150/200), respectively. The combination of TTSS1 and BPSS0745 qPCR increased the detection rate to 90% (180/200). Combining the results of the three qPCR assays and the BPSS1187 nested PCR previously published, all 200 samples were positive by at least one PCR assay. Samples positive by either TTSS1 ($n = 153$) or BPSS0745 ($n = 150$) qPCR were more likely to be direct-culture positive, with odds ratios of 4.0 (95% confidence interval [CI], 1.7 to 9.5; $P < 0.001$) and 9.0 (95% CI, 3.1 to 26.4; $P < 0.001$), respectively. High *B. pseudomallei* genome equivalents correlated with high CFU counts by culture. In conclusion, multitarget qPCR improved the *B. pseudomallei* detection rate in soil samples and predicted culture positivity. This approach has the potential for use as a sensitive environmental screening method for *B. pseudomallei*.

IMPORTANCE The worldwide environmental distribution of the soil bacterium *Burkholderia pseudomallei* remains to be determined. So far, most environmental studies have relied on culture-based approaches to detect this pathogen. Since current culture methods are laborious, are time consuming, and have limited sensitivity, culture-independent and more sensitive methods are needed. In this study, we show that a *B. pseudomallei*-specific qPCR approach can detect significantly higher numbers of *B. pseudomallei*-positive soil samples from areas where it is endemic compared with that from culture. The use of multiple independent *B. pseudomallei*-specific qPCR targets further increased the detection rate of *B. pseudomallei* compared with that from single targets. Samples with a high molecular *B. pseudomallei* load were more likely to be culture positive. We conclude that our quantitative multitarget approach

Received 12 December 2016 **Accepted** 1 February 2017

Accepted manuscript posted online 10 February 2017

Citation Göhler A, Trung TT, Hopf V, Kohler C, Hartleib J, Wuthiekanun V, Peacock SJ, Limmathurotsakul D, Tuanyok A, Steinmetz I. 2017. Multitarget quantitative PCR improves detection and predicts cultivability of the pathogen *Burkholderia pseudomallei*. *Appl Environ Microbiol* 83:e03212-16. <https://doi.org/10.1128/AEM.03212-16>.

Editor Christopher A. Elkins, FDA Center for Food Safety and Applied Nutrition

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might be useful in defining areas where there is a risk of *B. pseudomallei* infections in different parts of the world.

KEYWORDS *Burkholderia pseudomallei*, melioidosis, Thailand, qPCR, rice field, soil

B*urkholderia pseudomallei* causes melioidosis, an infectious disease with high case fatality rates. This bacterium is a tier 1 select agent and a natural inhabitant of soil and surface waters in tropical and subtropical regions where it is endemic (1–5). Melioidosis is acquired through inoculation, aerosols, or ingestion (6, 7). Most clinical cases of melioidosis are reported from northeast (NE) Thailand and northern Australia (8). In NE Thailand, melioidosis is the third most frequent cause of death due to infectious diseases (8). In northern Australia, it is the most common cause of fatal community-acquired pneumonia (9–11). Melioidosis is not restricted to these regions and is known to occur in other parts of Asia, America, and Africa (3, 4, 12, 13), where the epidemiological situation is less well defined. Moreover, a recently published model predicted an alarming number of 165,000 cases of human melioidosis per year worldwide, from which 89,000 people are predicted to die (14). This study illustrates the fact that major regions of the world have never been examined for the occurrence of *B. pseudomallei* in the environment. Areas where it is highly endemic and where diagnostic microbiology capabilities are not available may go unrecognized, and environmental surveillance for *B. pseudomallei* can provide an early warning to clinicians and local authorities. Moreover, precise quantitative environmental detection is also fundamental for shedding light on the ecology of this pathogen.

The majority of previous studies have used culture-based approaches to detect and quantify *B. pseudomallei* from the environment (4, 5, 12, 13, 15, 16). Culture from soil is influenced by numerous factors, including soil sampling depth (17), detachment of bacteria from the soil matrix (18), soil volume, incubation temperature, and the culture media used (12). Guidelines have been published that standardize environmental sampling of *B. pseudomallei* for culture techniques (15), although there is still a need to define the optima for a number of culture parameters, such as medium composition and incubation temperature, through relevant studies. The current methodology also lacks selectivity and cannot reliably prevent overgrowth of *B. pseudomallei* by other bacteria, which can lead to false-negative results. Given that culture methods are labor intensive, are time consuming, and have limited sensitivity, a more sensitive molecular screening of environmental samples is needed to complement culture approaches.

In a previous study, we established a protocol for detecting and quantifying *B. pseudomallei* DNA directly in soil samples using a single-copy fragment of the type three secretion system 1 (TTSS1) as a quantitative PCR (qPCR) target. Using this, we detected significantly higher numbers of *B. pseudomallei* genome equivalents (GE) than CFU using culture in a number of samples (19). The question remains as to whether qPCR-based detection of *B. pseudomallei* has the potential to significantly increase the detection rate in culture-negative samples. Here, we describe the development of a molecular assay using novel *B. pseudomallei*-specific gene targets, in which we compared single- and multitarget-based qPCR approaches for directly detecting and quantifying *B. pseudomallei* in 200 soil samples from rice paddies in northeast Thailand with previously published cultural data (12). Furthermore, we applied our molecular approach to soil samples from southern Vietnam where *B. pseudomallei* was isolated from soil more than 16 years ago (20). The combination of multiple qPCR targets increased the detection rate compared with those from single targets in samples with low *B. pseudomallei* counts. Our results demonstrate that multitarget qPCR screening can be used to predict growth of *B. pseudomallei* from soil samples.

RESULTS

***In silico* identification of novel *B. pseudomallei* qPCR target regions.** To date, qPCR-based environmental detection of *B. pseudomallei* has primarily relied on the TTSS1 target. We investigated the value of additional molecular targets for detecting *B.*

TABLE 1 Oligonucleotides used in this study

Purpose	Primer or probe	Oligonucleotide sequence ^a (5' to 3')	Concn (nM)	Amplified DNA fragment and length	Reference or source
qPCR target <i>Bacteria</i>	16S forward	TGGAGCATGTGGTTAATTCGA	250	16S rRNA, 159 bp	41
	16S reverse	TGCGGGACTTAACCCAACA	100		
	16S probe	FAM-CACGAGCTGACGACARCCATGCA-BHQ1	100		
Detection of PCR inhibitors	IAC2 forward	GAATTCGCCCTTATTAGCCGAC	400	Insertion sequence of pCR2.1-IAC, 166 bp	19
	IAC2 reverse	GGAATTCGCCCTTAATGCGC	400		
	CMV3 probe	HEX-TGATCGGGCTTATCGGTTCTTGATC-BHQ2	260		
qPCR targets for <i>B. pseudomallei</i>	BpTT4176 forward	CGTCTCTATACTGTCGAGCAATCG	400	TTSS 1 gene, 5' region (−82 to +34) of BPSS1407/ <i>sctD</i> , 115 bp	42
	BpTT4290 reverse	CGTGACACCGGTCAGTATC	400		
	BpTT4208 probe	FAM-CCGGAATCTGGATCACCACCACCTTCC-BHQ1	260		
	BPSS0087-Lfw	GAATGCGTGC GCGAGCA	500	BPSS0087 coding region, ^b 88 bp	This study
	BPSS0087-Brev	CTCGGCCGGTCCGGAAT	400		
	BPSS0087-P2	HEX-AGTCGTACGCAGCGCGCG—BHQ2	200	BPSS0745 coding region, ^b 132 bp	This study
	BPSS0745-Afw	GCGAGCAAATCCGTCTCTCA	500		
	BPSS0745-Fsrev	ATGCCAGGGCACATGGCTA	400		
	BPSS0745-P2	FAM-ATCATTAGGCGGGTGCCTG-BHQ1	200		
BPSS1187 nested PCR for <i>B. pseudomallei</i>	174 outer forward	ACCTTCGTCGGCATGGTAG	400	BPSS 1187 coding region, ^b 552 bp	19
	725 outer reverse	GCCCCGTTCTGGTCTTTATTC	400		
	8653 inner forward	ATCGAATCAGGGCGTTCAAG	400	BPSS 1187 coding region, ^b 81 bp	43
	8653 inner reverse	CATTCGGTGACGACACGACC	400		
	8653 inner probe	FAM-CGCCGCAAGACGCCATCGTTCAT-BHQ1	260		

^aFAM, 6-carboxyfluorescein; HEX, hexachloro-6-carboxyfluorescein; Cy5, indodicarbocyanine; BHQ1, black hole quencher 1; BHQ2, black hole quencher 2.

^bCoding sequence denoted for the complete genome sequence of *B. pseudomallei* K96243 that encodes a hypothetical protein.

pseudomallei in soil. Using whole-genome sequence data, we identified 11 coding sequences (CDS) that were unique to *B. pseudomallei*. These CDS had no BLAST hits or hits with an E value less than $1e-10$ in other tested *Burkholderia* genomes (data not shown). Initial SYBR green real-time PCR assays were developed based on these gene sequences, and were then tested with a small panel (Biowatch panel) of DNA samples ($n = 60$) from *B. pseudomallei* and its near-neighbor species. Four conserved regions, BPSL0092, BPSS0087, BPSS0135, and BPSS0745, were proven to be specific to *B. pseudomallei*. Two of the four gene targets, namely, BPSS0087 and BPSS0745, were selected for qPCR assay development because initial experiments revealed a lower limit of detection (LOD) than BPSL0092 and BPSS0135 (data not shown). Both open reading frames (ORFs) code for hypothetical proteins.

Development of highly specific duplex qPCR assay using BPSS0087 and BPSS0745. We established qPCR probe assays targeting the two single-copy genes BPSS0087 and BPSS0745. Primer and probe sets were created and optimized using genomic DNA of *B. pseudomallei* K96243 (<https://www.ncbi.nlm.nih.gov/genome/?term=K96243>) (Table 1). Each single assay and the BPSS0087/BPSS0745 duplex assay showed linearity over several orders of magnitude (see Fig. S1 in the supplemental material). The LODs for BPSS0087 and BPSS0745 targets in the duplex assay were ~ 7.0 GE and ~ 2.5 GE, respectively, which are in a range similar to the LOD of 3 GE of the previously described TTSS1 qPCR. To verify the specificity of the two gene targets, we tested the genomic DNA of 85 *B. pseudomallei* and 42 non-*B. pseudomallei* isolates from different origins (Table 2; see also Table S2) using the BPSS0087/BPSS0745 duplex qPCR and compared the results to those from the TTSS1 assay. All *B. pseudomallei* were identified and no false-positive amplification products were detected in the non-*B. pseudomallei* strains using each of the three qPCR assays. Moreover, spiking experiments revealed that neither humic acids nor artificially added soil DNA had an inhibitory effect on assay performance (see Fig. S4 and S5).

Combination of multiple qPCR targets increases the *B. pseudomallei* detection rates in soil samples from northeast Thailand. We then analyzed 200 soil samples from rice paddies from NE Thailand with the two novel qPCRs and compared the results with those from the established TTSS1 qPCR assay. The same samples were used before

TABLE 2 Specificities of the qPCR assays^a

Species	No. of samples	No. with positive reaction (%)		
		BPSS0087 qPCR	BPSS0745 qPCR	TTSS1 qPCR
<i>B. pseudomallei</i>	85	85 (100)	85 (100)	85 (100)
<i>B. thailandensis</i>	6	0 (0)	0 (0)	0 (0)
<i>Burkholderia cepacia</i> complex species	30	0 (0)	0 (0)	0 (0)
Non- <i>Burkholderia</i> species ^b	6	0 (0)	0 (0)	0 (0)

^a5 ng of each genomic DNA was used.

^bIncluding two *Achromobacter* species, one *Escherichia coli*, and three *Ralstonia pickettii* isolates (see Table S2 in the supplemental material for a complete list).

to validate different *B. pseudomallei* cultivation methods as described previously (12). That study by Limmathurotsakul and coworkers used a panel of different culture techniques, including enrichment cultures, and reported 47% ($n = 94$) culture-positive samples, of which 35% ($n = 70$) were positive on direct culture (12). Higher detection rates were observed for the TTSS1 qPCR with 76.5% ($n = 153$) and for the BPSS0745 qPCR with 75% ($n = 150$). Although the analytical sensitivities of the three qPCR assays were comparable (see above and Fig. S1), only 34.5% ($n = 69$) of samples were detected by the BPSS0087 qPCR (Table 3). We found that 90% ($n = 180$) of samples contained BPSS0745 and/or TTSS1 as *B. pseudomallei*-specific sequences (Fig. 1). When the results from all three qPCR assays were combined, there was only a marginal increase to 91% ($n = 182$) in the overall detection rate, indicating that 97% (67 from 69) of samples that were BPSS0087 positive were also detected by either the BPSS0745 or TTSS1 qPCR (Fig. 1). If the remaining 18 qPCR-negative samples were subjected to the previously described BPSS1187 nested PCR, the positivity rate reached 100% (Table 1). As pictured in Fig. 1, 61 (30.5%) samples were detected by each of the three qPCR targets, and 130 (65%) samples were detected by at least two qPCR targets.

To verify positive qPCR results from samples with results that were discrepant between different qPCRs (see Fig. S2) and to exclude false-positive detection in single-target positive samples, we cloned and sequenced amplicons from selected samples (see Text S1). We found that sequenced amplicons of either BPSS0087, BPSS0745, or TTSS1 from qPCR-positive samples clearly indicate *B. pseudomallei* sequences (Text S1). However, we also found evidence that some samples contained yet unknown sequences where primers but not the probes for TTSS1 and BPSS0745 bound (or formed dimers), and thereby either might lower the sensitivity for the respective

TABLE 3 Detection of *B. pseudomallei* in 200 soil samples from two Thai rice fields

Result	No. of samples			Sensitivity (% [95% CI])
	Overall	Field 1	Field 2	
Negative	0	0	0	
Positive by culture or at least one PCR assay	200	100	100	100
Culture positive ^a	94	47	47	47 (40.2–53.9)
Direct-culture positive ^b	70	40	30	35.0 (28.7–41.8)
PCR positive ^c	200	100	100	100
qPCR positive ^d	182	94	88	91 (86.2–94.3)
TTSS1 positive	153	80	73	76.5 (70.1–81.9)
BPSS0087 positive	69	32	37	34.5 (28.2–41.3)
BPSS0745 positive	150	79	70	75 (68.6–80.5)

^aSamples in which *B. pseudomallei* growth was detected by any culture method used by Limmathurotsakul and colleagues (12).

^bSamples in which *B. pseudomallei* was detected without enrichment according to Limmathurotsakul and colleagues (12).

^cIncludes all qPCR assays and BPSS1187 nested PCR.

^d*B. pseudomallei*-positive samples by any qPCR assay (BPSS0087, BPSS0745, and TTSS1 qPCR).

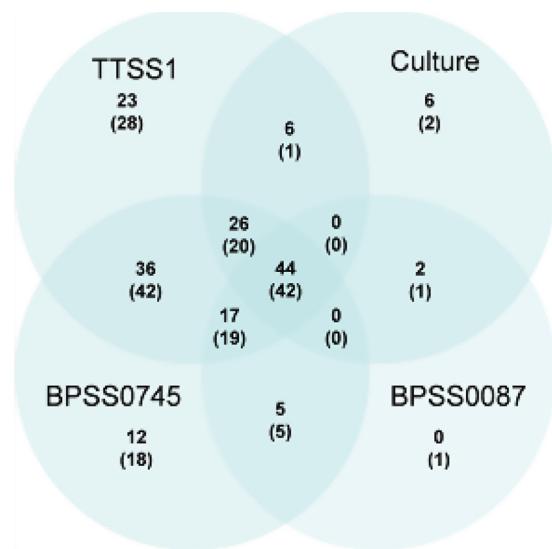


FIG 1 Venn diagram of *B. pseudomallei* detection in soil samples through a combination of different molecular (qPCR) assays and culture. Ten samples that were only positive by BPSS0745 qPCR and direct culture and one sample that was only positive by BPSS0087 and TTSS1 are not displayed. All six culture-positive and qPCR-negative samples were positive by BPSS1187 nested PCR (19).

target or might even lead to false-negative results. These findings are likely to explain the different GE numbers determined by the three qPCR targets in some samples (Fig. S2) and the increased detection rate observed using multiple qPCR targets (Fig. 1).

The odds of BPSS0087, BPSS0745, and TTSS1 qPCR-positive samples also being direct-culture positive were higher than qPCR-negative samples based on odds ratios (OR) of 6.4 (95% confidence interval [CI], 3.3 to 12.1; $P < 0.001$), 9 (95% CI, 3.1 to 26.4; $P < 0.001$), and 4 (95% CI, 1.7 to 9.5; $P < 0.001$), respectively. In other words, a sample being positive by qPCR was more likely to be direct-culture positive. Samples being positive by at least two qPCR assays ($n = 130$) were 7.1 times more likely to be direct-culture positive (95% CI, 3.1 to 15.9; $P < 0.001$) and 2.2 times more likely to show *B. pseudomallei* growth, including enrichment (95% CI, 1.2 to 4.1; $P = 0.009$) than samples with one positive or all negative qPCR-assay(s) (see Table S5). This indicates that single-target-positive samples have lower *B. pseudomallei* GE counts compared with those of multiple-target-positive samples (Fig. 2), which are more likely to be culture positive. Conversely, direct-culture-negative samples exhibited significantly lower *B. pseudomallei* GE than direct-culture-positive ones (Fig. 3). This effect was significant (Mann-Whitney U test, $P < 0.001$) for each of the qPCR assays. The GE values obtained with the BPSS0087 qPCR exhibited a gap with very few positive samples between 0 and $\sim 10^4$ GE per g soil (Fig. 3), pointing to a higher LOD in nonartificial DNA mixtures than in pure genomic *B. pseudomallei* DNA. The *B. pseudomallei* counts obtained with BPSS0745 or TTSS1 qPCR did not differ significantly (Wilcoxon signed-rank test, $P = 0.32$). These two qPCR assays exhibited significantly higher (Wilcoxon signed-rank test, $P < 0.001$) *B. pseudomallei* GE counts compared with those from BPSS0087 qPCR and direct culture (*B. pseudomallei* CFU).

Each of the qPCR assays showed a significant correlation with CFU values as given by Spearman's rho (ρ_{sp}) correlation coefficients above 0.46 (see Table S3). Furthermore, the qPCR assays correlated with each other (Fig. S2). As already indicated from Fig. 2 and 3, the BPSS0087 qPCR had decreased sensitivities in samples with a low *B. pseudomallei* GE compared with those from the other two assays (Fig. S2).

High molecular *B. pseudomallei* GE and abundance correlate with cultivability.

To examine any potential association between the total bacterial load and the presence of *B. pseudomallei* in the same sample, we determined the *B. pseudomallei* proportion (abundance). The median of all samples was 9.7×10^7 (range, 1.3×10^7 to 1.9×10^{10})

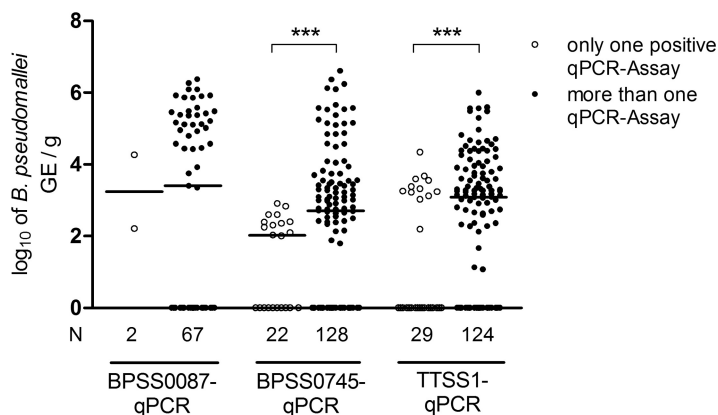


FIG 2 Comparison of *B. pseudomallei* GE in soil samples with only one and more than one positive qPCR assay(s). Scatter plot (with medians) comparison of \log_{10} -transformed GE as *B. pseudomallei* count per gram soil. *B. pseudomallei* GE of qPCR-positive samples were grouped (for each assay) in samples with one or more positive qPCR assays. Samples with a value of zero are the result of positive and negative subsamples in single qPCRs (see Materials and Methods), leading to a median of zero. N denotes the number of samples per group. ***, $P < 0.001$ as determined by a Mann-Whitney U test.

bacterial 16S rRNA genes per gram soil. The abundance in one gram of soil, calculated from data of the two sensitive qPCR assays (BPSS00745 and TTSS1), ranged from 0 to 2.7×10^{-2} for BPSS0745 qPCR and 0 to 8.86×10^{-3} for TTSS1 qPCR *B. pseudomallei* GE per bacterial 16S rRNA gene (Fig. 4). The median abundance, calculated from these two assays, was 3.15×10^{-7} (range, 0 to 7.36×10^{-3}) *B. pseudomallei* GE per bacterial 16S rRNA gene.

The variations in *B. pseudomallei* abundance were mainly related to alterations in the *B. pseudomallei* count rather than changes of total bacterial load. Therefore, we correlated the *B. pseudomallei* GE to the number of culture-positive samples (Fig. 5A). For instance, only 31% of samples (15 of 48) were culture positive in the first and lowest GE count quartile, whereas 78% of culture-positive samples (36 of 46) were found in samples with the highest GE count (4th quartile). This trend was significant for both culture (P for trend < 0.01) and direct culture (P for trend < 0.001). Moreover, there was a correlation between the GE, and also for abundance, and the number of *B. pseudomallei* colonies on agar after direct culture (Fig. 5B; see also Table S4 and Fig. S3). However, a multivariable model is needed to evaluate whether abundance is indepen-

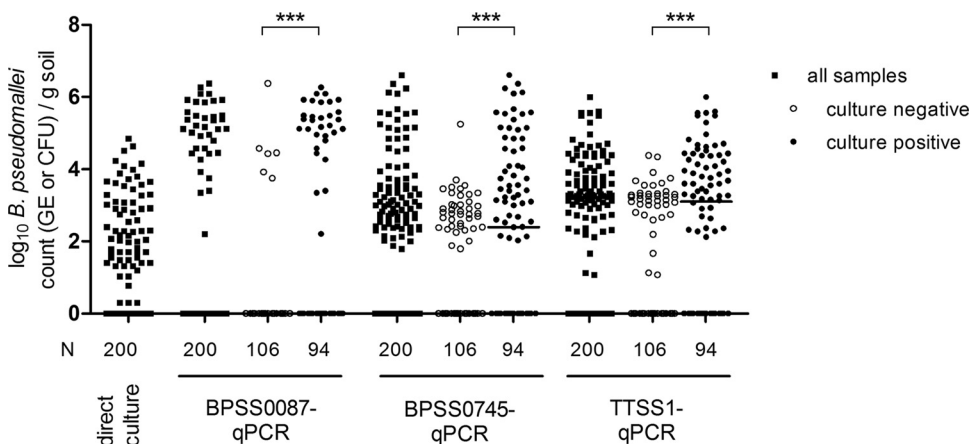


FIG 3 Comparison of *B. pseudomallei* GE detected by the various assays of culture-positive and -negative samples. Scatter plot (with medians) comparison shows \log_{10} -transformed *B. pseudomallei* CFU per gram soil as determined by direct culture and \log_{10} -transformed *B. pseudomallei* GE per gram soil as determined by qPCR assays. N denotes number of samples per group. ***, $P < 0.001$ by a Mann-Whitney U test.

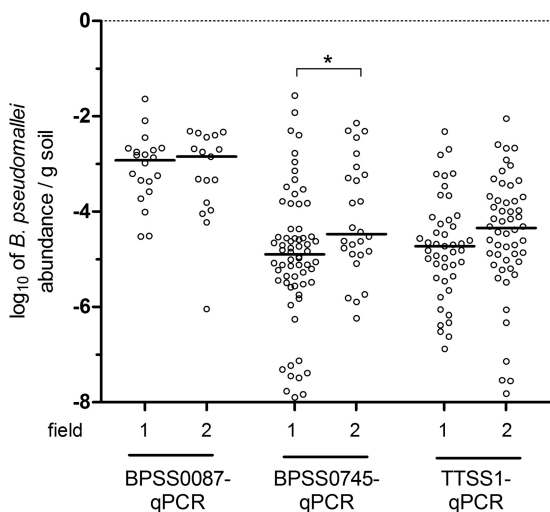


FIG 4 Abundance of *B. pseudomallei* in single soil samples on fields 1 and 2. The scatter plot (with medians) of *B. pseudomallei* abundance is given as log₁₀ of GE (TTSS1, BPSS0087, and BPSS0745 qPCR) per 16S rRNA gene. *, *P* < 0.05 by a Mann-Whitney U test.

dently associated with cultivability. In conclusion, the means of the GE calculated from the BPSS0745 and TTSS1 assays serve as predictors for (direct) cultivability in this study.

***B. pseudomallei* spatial distribution within rice fields.** The spatial distribution of *B. pseudomallei* in the two rice fields was visualized for the data obtained from the combination of the two qPCR methods as well as for the culture results (Fig. 6). The molecular and cultural approaches demonstrated similar distribution patterns of *B. pseudomallei*. As shown in Fig. 3, most samples exhibited significantly higher GE values compared with CFU values. However, 10% of all samples showed CFU values that were >50-fold higher than GE values, as pictured in Fig. 6E and F. It seems likely that a heterogeneity in the microdistribution of *B. pseudomallei* within single samples (19) is responsible for this phenomenon.

Application of the multiple qPCR target approach in soil samples from southern Vietnam. To further test the general applicability of our three-*B. pseudomallei*-target approach in soil samples from other areas where it is endemic, we analyzed 42

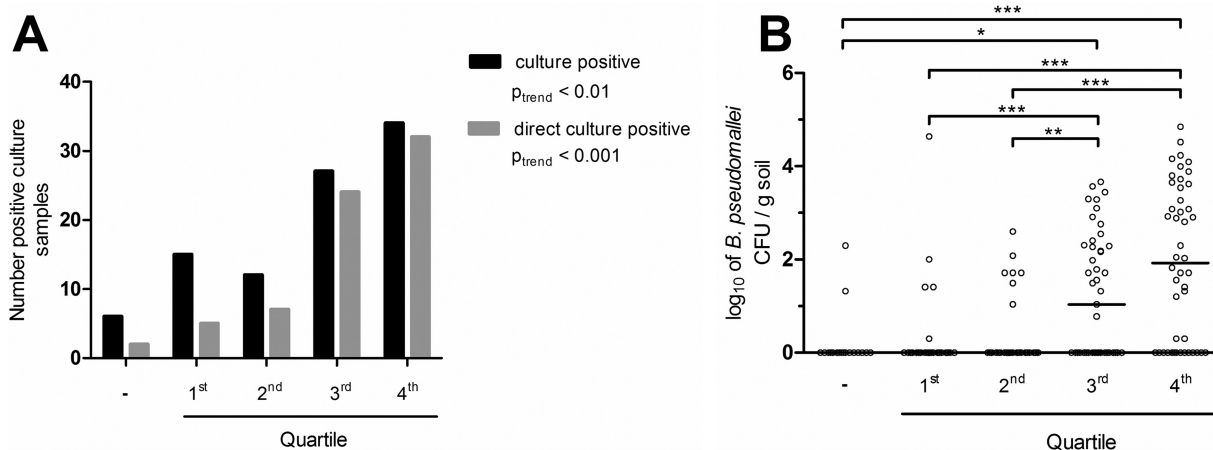


FIG 5 Correlations between (A) culture and direct culture positivity rates and (B) numbers of *B. pseudomallei* CFU with increasing genome equivalents (GE) of *B. pseudomallei*. Results were grouped according to calculated means from GE data of BPSS0745 and TTSS1. (B) Data are presented as a scatter plot (with medians); samples were divided between qPCR negative (“-”; *n* = 18) and samples exhibiting positive qPCR assays (*n* = 182), which were grouped in quartiles: 1st, 0 GE/g soil (*n* = 48); 2nd, 0 to $\leq 4.21 \times 10^1$ GE/g soil (*n* = 43); 3rd, 4.21×10^1 to $\leq 8.37 \times 10^2$ GE/g soil (*n* = 45); 4th, $> 8.37 \times 10^2$ GE/g soil (*n* = 46). Trend analysis in panel A was conducted with a chi-square test. *, **, and *** represent *P* values of <0.05, <0.01, and <0.001, respectively, by a Kruskal-Wallis test.

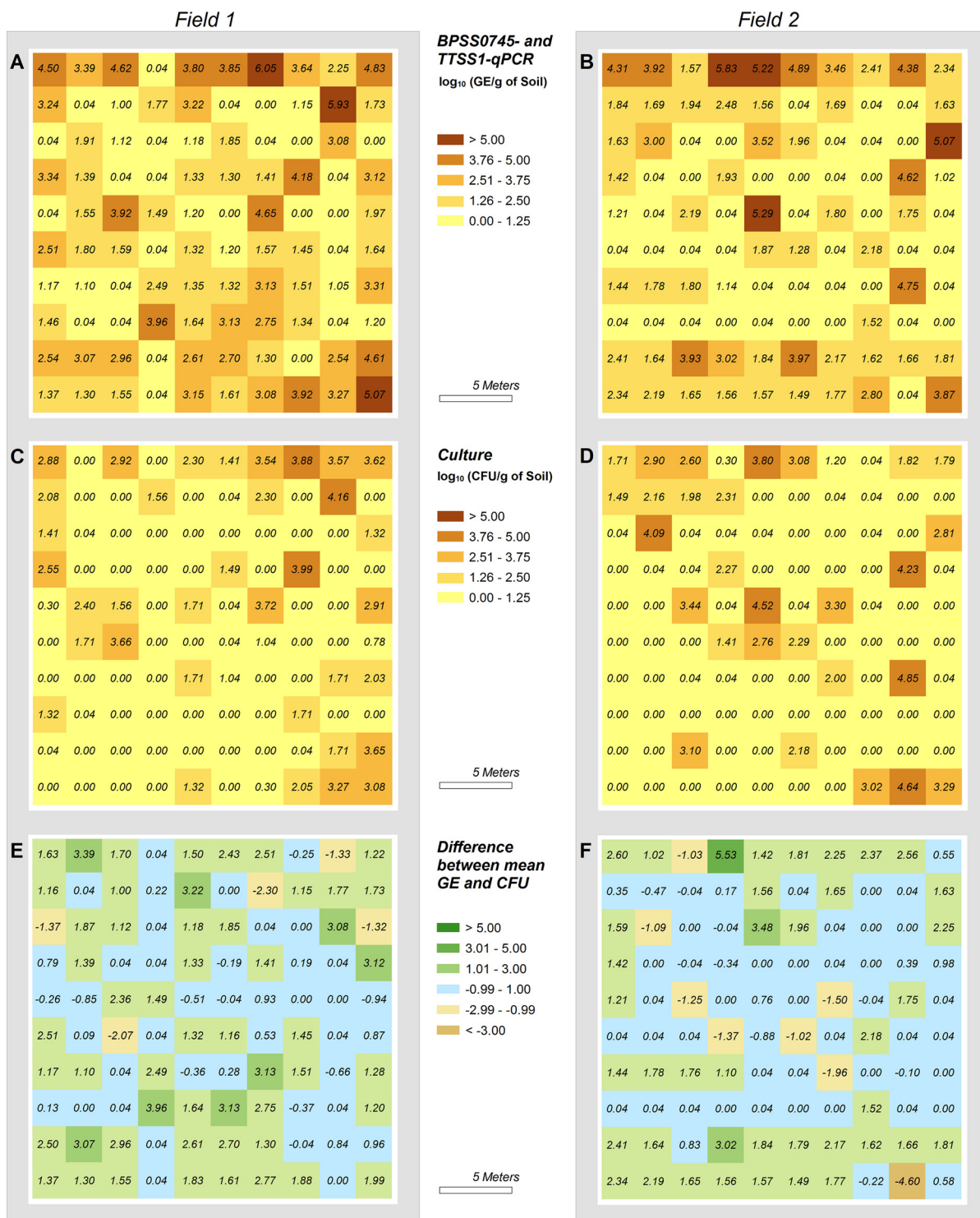


FIG 6 Spatial distributions of *B. pseudomallei* on both rice fields tested. Field 1 is displayed on panels A, C, and E. Field 2 is displayed on panels B, D, and F. *B. pseudomallei* counts are given as log₁₀ of mean GEs for qPCR (calculated from TTSS1 and BPSS0745) (A, B) or as CFU (for direct-culture method) per gram soil (C, D). Artificial values (0.1 GE or CFU/g soil) were assigned to samples only positive in enrichment culture and to qPCR-positive samples with a calculated median of 0 for differentiation to qPCR- or culture-negative samples. (E, F) Differences between culture and molecular methods are visualized as differences between the log₁₀ of the mean GE and CFU values.

soil samples from 28 sampling sites taken in southern Vietnam. Among those samples, 6 (14.3%) were culture positive and 35 (83.3%) were positive by at least one of the qPCR targets (see Fig. S6), confirming the higher detection rate as seen with the soil samples from Thailand. Five of the six culture-positive samples were positive for all three of the

qPCR targets, whereas one sample was negative by TTSS1 but positive by the other two targets (Table S6 and Fig. S6). These results confirm our previous observation that a single-qPCR-target approach might lead to false-negative results. The *B. pseudomallei* load varied between 0 and 4.71×10^4 GE/g soil (Table S6). Among the qPCR-positive samples, the median load was approximately 10 times higher in culture-positive samples than in culture-negative ones (Mann-Whitney U test, $P < 0.001$). This observation confirms our results obtained with the Thai soil samples (see above), where a higher molecular *B. pseudomallei* load was also linked to culture positivity.

DISCUSSION

Most studies on the environmental prevalence of *B. pseudomallei* in different regions of the world have relied on culture-based methods (4, 12, 13, 15, 16). However, such methods are laborious and time consuming, especially for large-scale screening. This acts as a barrier to mapping the worldwide distribution of *B. pseudomallei* (3). Even in single rice fields that are highly culture positive across numerous sampling points, significant numbers of adjacent culture-negative samples can be found (21, 22). Although this might reflect spatial heterogeneity, it may also indicate that current protocols do not reliably detect cultivable *B. pseudomallei* and/or miss viable but noncultivable states of this pathogen. This suggestion is supported by a recent study from Laos, showing that the molecular detection of *B. pseudomallei* in enrichment cultures of soil samples was higher than the detection of *B. pseudomallei* growth in the respective subcultures (23). For large-scale environmental screening, a completely culture-independent and sensitive method is desirable. Such a method might be useful for identifying samples for subsequent targeted attempts to culture *B. pseudomallei* strains.

In a previous study, we showed that direct TTSS1 target-based qPCR from soil samples detected significantly higher numbers of *B. pseudomallei* genome equivalents than culture-based CFU (19). This study focused on developing direct molecular *B. pseudomallei* quantification from soil samples by using a limited number of samples, which were mainly highly culture positive. It remains to be shown that direct qPCR-based detection also leads to increased qualitative sensitivity, namely, an increased detection rate of *B. pseudomallei* in otherwise culture-negative samples.

The results of our present study clearly show that TTSS1-based qPCR detected significantly higher numbers of *B. pseudomallei*-positive Thai soil samples (76.5%) than did culture (positivity rate, 47%). Interestingly, the detection rate was further increased to 90% by a combination of the TTSS1 qPCR with the BPSS0745-based qPCR. Our data suggest that this increase is due to the additional detection of samples with low GE counts, which are not reliably detected by a single qPCR assay, whereby samples that were false negative by a single qPCR target became positive using another qPCR target. The variable performance of the single qPCRs and the much higher sensitivity of the multitarget approach compared with that of culture were also confirmed in the 42 soil samples tested from southern Vietnam (Fig. S6 in the supplemental material).

As shown by Price et al. with genomic DNA from pure *B. pseudomallei* cultures, the TTSS1 qPCR can lead to false-negative results (24). The fact that we found culture-positive TTSS1 qPCR-negative samples from Thailand and Vietnam that were positive by BPSS0745 qPCR or BPSS0087 qPCR clearly shows the usefulness of alternative qPCR targets. Our multiple-target approach is further supported by our findings that individual primers bound to yet unknown sequences of non-*B. pseudomallei* soil microbes, and thereby most likely lowered the sensitivity of single qPCRs (see Text S1), which might even lead to false-negative results. We observed this potential bias with the recently published *B. pseudomallei*-specific 122018 assay (24). This assay discriminated between *B. pseudomallei* and *B. thailandensis*, but exhibited a significant decrease in sensitivity in the presence of large amounts of *B. thailandensis* DNA (see Fig. S4). Since *B. thailandensis* is likely to be encountered in environmental samples of *B. pseudomallei* from regions where it is endemic, this assay would probably lead to false-negative results. A similar effect with *B. thailandensis* was not found for the BPSS0087 or the

BPSS0745 qPCR assay (see Fig. S5). Another recently published *B. pseudomallei*-specific assay (25) may also not be useful for environmental studies, since the target gene *wcbG* was shown to be present in the genomic DNA of *B. thailandensis* E555.

Despite these differences, the overall results of the BPSS0087, BPSS0745, and TTSS1 qPCR were correlated. Moreover, results from each of the qPCR assays correlated with the number of *B. pseudomallei* detected by direct culture in Thai soil samples. BPSS0745 and TTSS1 PCR *B. pseudomallei* counts were, on average, 5.6 to 7.2 times higher than CFU values. This value was slightly lower than that from our previous study using the TTSS1 qPCR in which we found 10.6-fold higher GE values compared with CFU values in a limited number of mostly highly culture-positive soil samples (19).

In contrast to the relatively constant total bacterial load observed, the *B. pseudomallei* counts exhibited a strong variation leading to fluctuating abundances ranging from zero to 0.74% of *B. pseudomallei* in the two tested rice fields. This uneven spread might reflect a microdistribution due to variable biotic and abiotic conditions, which are known to influence the soil microbiome in rice paddies (26–33). By using molecular and cultural methods, we found that in the two rice fields tested, this observed uneven distribution seems to reflect *B. pseudomallei* presence rather than cultivability.

Another important outcome of our study is that our molecular assays predicted cultivability of *B. pseudomallei* in Thai and Vietnamese soils. This was shown for qualitative and quantitative results from Thai soil. Samples that are positive for single qPCR or for a combination are more likely to be culture positive. The higher the molecular abundance of *B. pseudomallei* that is detected, the more likely it is to isolate *B. pseudomallei* strains. In conclusion, our sensitive multitarget screening and direct quantification might guide future studies on the ecology of *B. pseudomallei* in different parts of the world.

MATERIALS AND METHODS

Bacterial strains and growth conditions. All bacterial isolates used in this study are listed in Table S2 in the supplemental material. *B. pseudomallei* K96243 was used as the reference strain. Isolates were stored at -80°C with 20% (vol/vol) glycerol in tryptic soy broth (TSB), were streaked onto Columbia agar plates supplemented with 5% sheep blood (Becton Dickinson, Heidelberg, Germany), and were incubated overnight under aerobic conditions at 37°C . Fresh cultures grown overnight in Lennox broth or tryptic soy broth (TSB) were harvested for bacterial DNA extraction using a QIAamp DNA purification kit (Qiagen, Hildesheim, Germany).

In silico identification of new *B. pseudomallei*-specific molecular targets. To identify new specific gene targets of *B. pseudomallei*, we generated a list of *B. pseudomallei* conserved genes using comparative genomic analysis. Thirty available *B. pseudomallei* genomes were used in this study (see Table S1). All genomes were annotated using myRAST (34), and homologs were grouped on the basis of their amino acid sequence similarity using orthoMCL (35, 36). For each gene, the homolog from K96243 was chosen to represent the homology group. These gene sequences were compared against the genome sequences of other *Burkholderia* species, such as *B. thailandensis* ($n = 17$), *B. mallei* ($n = 17$), *B. multivorans* ($n = 4$), *B. ubonensis* ($n = 4$), *B. oklahomensis* ($n = 2$), and *B. thailandensis*-like sp. ($n = 3$) (37), to identify *B. pseudomallei*-specific genes using BLAST.

Soil samples. Soil samples from two rice fields were taken in Ubon Ratchathani province, NE Thailand, during March (pre-rainy season) 2011 and consisted of sandy loam (12). Each field was divided into a grid system of 10 by 10 (in total 100) sampling points 2.5 m apart, as described before (12). Aliquots of the same samples were used previously to evaluate a simplified culture method for the isolation of *B. pseudomallei* (12). Here, we refer to CFU data obtained by direct subculture of a soil suspension as a part of the standard method without enrichment (termed “direct culture”) (12). We use the term “culture positive” for any sample that previously showed *B. pseudomallei* colonies (12) after either direct culture or any enrichment culture in TBSS-C50 (threonine-basal salt solution plus colistin at 50 mg/liter) broth (38), according to published guidelines (12, 39). A 50-g aliquot of each original 200-g sample was shipped at ambient temperature to the laboratory in Germany for molecular analysis. Ten soil samples were taken around Greifswald, Germany, as negative controls from an area where the bacterium is not endemic to confirm the specificity of the PCR methods and to test for cross-contamination. Moreover, 42 soil samples collected at 28 sampling sites in 13 out of 19 provinces in southern Vietnam were tested. The sampling was part of a nationwide *B. pseudomallei* environmental surveillance carried out in September 2015 (to be published elsewhere). The distance between two consecutive sites ranged from 6 to 153 km (mean ~ 44 km). All samples were collected in rice fields at a 30-cm soil depth. At each sampling site, the distance between two consecutive samples ranged from 20 to 30 m. One to three samples from each site were randomly chosen for testing. Cultures were performed using 10 g of soil in 20 ml of Galimand's broth (40). After 2 days of standing incubation at 40°C , 10-fold serial dilutions of the supernatants were spread on Ashdown agar and incubated at 40°C for 5 days. *B. pseudomallei* was identified on the basis of its characteristic colony morphology and *recA* sequence analysis.

Extraction of nucleic acids from soil samples and removal of PCR inhibitors. Upon arrival in Greifswald, Thai soil samples and German negative samples were divided and processed. DNA was extracted at least three independent times per sample using 1-g subsamples. The detailed extraction procedure based on a modified protocol using cetyltrimethylammonium bromide (CTAB) was published before (19). The soil samples from Vietnam (0.5 g) were extracted using the InnuSPEED soil DNA kit (Analytik Jena, Germany). Reextractions of DNA were performed from soil samples that had been maintained at -80°C .

Standards for qPCR. As the standard for analysis of the total bacterial load, a 159-bp fragment of the bacterial 16S rRNA gene was cloned into the pSC-B-amp/kan vector (named pSC-16SB), as described elsewhere (41). The inhibition control plasmid pCR2.1-IAC was described before (19) and consisted of an artificial synthesized DNA fragment of 135 bp within a pCR2.1 plasmid (MWG-Biotech, Ebersberg, Germany), exhibiting the same thermal conditions as the TTSS1 target amplification. Each qPCR assay (BPSS0087, BPSS0745, and TTSS1 qPCR) included a standard curve made by serial dilutions of *B. pseudomallei* K96243 gDNA over five orders of magnitude to ensure and control optimal qPCR efficiencies and to calculate the copy number of the respective target. Since these assays target a single-copy region, the number of amplicon copies can directly be converted into genome equivalents (GE) (19). Bacterial loads of the samples were calculated by using the number of 16S rRNA genes as described previously (41).

qPCR for determination of inhibition, bacterial load, and *B. pseudomallei*. Regions coding for BPSS0087 and BPSS0745 were selected as qPCR targets (see Results). To create a qPCR duplex assay for BPSS0087 and BPSS0745 with optimal conditions, different sets of primers were designed. Final concentrations of the primer/probe combinations are provided in Table 1. Oligonucleotides were purchased from TIB Molbiol (Berlin, Germany), MWG-Eurofins (Ebersberg, Germany), or Microsynth (Balgach, Switzerland). The qPCRs for TTSS1, bacterial load (16S rRNA genes), and inhibition (IAC) were performed as described elsewhere (19, 41). The BPSS0087/BPSS0745 qPCR was performed in parallel as a duplex assay with the following temperature protocol: 50°C for 2 min, 95°C for 10 min, 50 cycles of 95°C for 15 s, 57°C for 30 s, and 72°C for 30 s. The BPSS1187 nested PCR for detecting *B. pseudomallei* was performed as described for TTSS1-negative samples (19). All qPCRs from duplicates were analyzed using the Light Cycler 480 instrument II (Roche, Mannheim, Germany). For quantifications, the second derivative maximum algorithm was applied (Light Cycler 480 software release 1.5.0). At least six individual standard curves were used per qPCR assay to determine the limit of detection (LOD) according to the work of Price and colleagues (24).

qPCR quality characteristics and PCR-positive samples. To ensure sufficient quality of the subsamples, several criteria had to be fulfilled. First, no inhibition of the qPCR was detectable; the difference between the quantification cycle (C_q) value of the sample + 100 copies pCR2.1-IAC and pure pCR2.1-IAC was below a value of 2. Second, more than 10^6 16S rRNA genes per PCR were detectable. If one of the three 1-g subsamples per sample did not fulfil these criteria, another 1-g subsample was extracted. A sample was defined as PCR positive if at least one replicate of the three subsamples revealed a C_q value above the limit of detection. Furthermore, all 10 soil sample from a non-endemic region, namely Greifswald, Germany, proved to be negative by *B. pseudomallei*-specific qPCRs. PCR amplicons from selected samples and qPCR targets were cloned using the Zero blunt cloning kit (Thermo Fisher Scientific, Braunschweig, Germany), were introduced into *E. coli* DH5a according to the manufacturer's instructions, and were subsequently sequenced.

Statistical analysis. The total bacterial loads were used to calculate the percentage of *B. pseudomallei* within the bacterial community (abundance) after fulfilling the quality criteria (see above). For each single qPCR assay and 1-g subsample, *B. pseudomallei* GE and the number of rRNA genes per g were calculated as means of the technical duplicates. The medians calculated from the three independent subsamples represented the *B. pseudomallei* and total bacterial loads per sample and qPCR target, respectively. The abundance of each sample was calculated as the mean GE count of the BPSS0745 and TTSS1 qPCR divided by the bacterial load. GraphPad Prism software version 4.0 for Windows (GraphPad Software, San Diego, CA, USA) was used for calculations and statistical analyses (Wilcoxon signed-rank test, Mann-Whitney U test, chi-square-trend analyses, Kruskal-Wallis test, and correlations by Spearman's rho analyses). For visualizing the distribution of *B. pseudomallei*, ArcGIS for desktop (version 10.3.1.4959; Environmental Systems Research Institute, Inc.) was used. To differentiate true negative samples from positive samples with a calculated median of 0 GE or CFU in graphical representations, an artificial value of 0.1 CFU or GE per g soil was used.

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <https://doi.org/10.1128/AEM.03212-16>.

SUPPLEMENTAL FILE 1, PDF file, 0.4 MB.

ACKNOWLEDGMENTS

This work was coordinately supported by the Vietnamese Ministry of Science and Technology and the German Federal Ministry of Education and Research through the project RENOMAB (Research Network on Melioidosis and *Burkholderia pseudomallei*; reference number 01DP13007).

REFERENCES

- White NJ. 2003. Melioidosis. *Lancet* 361:1715–1722.
- Cheng AC, Currie BJ. 2005. Melioidosis: epidemiology, pathophysiology, and management. *Clin Microbiol Rev* 18:383–416. <https://doi.org/10.1128/CMR.18.2.383-416.2005>.
- Currie BJ, Dance DA, Cheng AC. 2008. The global distribution of *Burkholderia pseudomallei* and melioidosis: an update. *Trans R Soc Trop Med Hyg* 102(Suppl 1):S1–S4. [https://doi.org/10.1016/S0035-9203\(08\)70002-6](https://doi.org/10.1016/S0035-9203(08)70002-6).
- Wiersinga WJ, Birnie E, Weehuizen TA, Alabi AS, Huson MA, Huis in 't Veld RA, Mabala HK, Adzoda GK, Raczynski-Henk Y, Esen M, Lell B, Kreamsner PG, Visser CE, Wuthiekanun V, Peacock SJ, van der Ende A, Limmathurotsakul D, Grobusch MP. 2015. Clinical, environmental, and serologic surveillance studies of melioidosis in Gabon, 2012–2013. *Emerg Infect Dis* 21:40–47. <https://doi.org/10.3201/eid2101.140762>.
- Chen YL, Yen YC, Yang CY, Lee MS, Ho CK, Mena KD, Wang PY, Chen PS. 2014. The concentrations of ambient *Burkholderia pseudomallei* during typhoon season in endemic area of melioidosis in Taiwan. *PLoS Negl Trop Dis* 8:e2877. <https://doi.org/10.1371/journal.pntd.0002877>.
- Currie BJ, Fisher DA, Howard DM, Burrow JN, Selvanayagam S, Snelling PL, Anstey NM, Mayo MJ. 2000. The epidemiology of melioidosis in Australia and Papua New Guinea. *Acta Trop* 74:121–127. [https://doi.org/10.1016/S0001-706X\(99\)00060-1](https://doi.org/10.1016/S0001-706X(99)00060-1).
- Gibney KB, Cheng AC, Currie BJ. 2008. Cutaneous melioidosis in the tropical top end of Australia: a prospective study and review of the literature. *Clin Infect Dis* 47:603–609. <https://doi.org/10.1086/590931>.
- Limmathurotsakul D, Wongratanacheewin S, Teerawattanasook N, Wongsuvan G, Chaisuksant S, Chetchotisakd P, Chaowagul W, Day NP, Peacock SJ. 2010. Increasing incidence of human melioidosis in northeast Thailand. *Am J Trop Med Hyg* 82:1113–1117. <https://doi.org/10.4269/ajtmh.2010.10-0038>.
- Currie BJ. 2015. Melioidosis: evolving concepts in epidemiology, pathogenesis, and treatment. *Semin Respir Crit Care Med* 36:111–125. <https://doi.org/10.1055/s-0034-1398389>.
- Douglas MW, Lum G, Roy J, Fisher DA, Anstey NM, Currie BJ. 2004. Epidemiology of community-acquired and nosocomial bloodstream infections in tropical Australia: a 12-month prospective study. *Trop Med Int Health* 9:795–804. <https://doi.org/10.1111/j.1365-3156.2004.01269.x>.
- Meumann EM, Cheng AC, Ward L, Currie BJ. 2012. Clinical features and epidemiology of melioidosis pneumonia: results from a 21-year study and review of the literature. *Clin Infect Dis* 54:362–369. <https://doi.org/10.1093/cid/cir808>.
- Limmathurotsakul D, Wuthiekanun V, Amornchai P, Wongsuvan G, Day NP, Peacock SJ. 2012. Effectiveness of a simplified method for isolation of *Burkholderia pseudomallei* from soil. *Appl Environ Microbiol* 78:876–877. <https://doi.org/10.1128/AEM.07039-11>.
- Rolim DB, Rocha MF, Brilhante RS, Cordeiro RA, Leitao NP, Jr, Inglis TJ, Sidrim JJ. 2009. Environmental isolates of *Burkholderia pseudomallei* in Ceara State, northeastern Brazil. *Appl Environ Microbiol* 75:1215–1218. <https://doi.org/10.1128/AEM.01953-08>.
- Limmathurotsakul D, Golding N, Dance DA, Messina JP, Pigott DM, Moyes CL, Rolim DB, Bertherat E, Day NP, Peacock SJ, Hay SI. 2016. Predicted global distribution of *Burkholderia pseudomallei* and burden of melioidosis. *Nat Microbiol* 1:15008. <https://doi.org/10.1038/nmicrobiol.2015.8>.
- Limmathurotsakul D, Dance DA, Wuthiekanun V, Kaestli M, Mayo M, Warner J, Wagner DM, Tuanyok A, Wertheim H, Yoke Cheng T, Mukhopadhyay C, Puthucherry S, Day NP, Steinmetz I, Currie BJ, Peacock SJ. 2013. Systematic review and consensus guidelines for environmental sampling of *Burkholderia pseudomallei*. *PLoS Negl Trop Dis* 7:e2105. <https://doi.org/10.1371/journal.pntd.0002105>.
- Rattanavong S, Wuthiekanun V, Langla S, Amornchai P, Sirisouk J, Phetsouvanh R, Moore CE, Peacock SJ, Buisson Y, Newton PN. 2011. Randomized soil survey of the distribution of *Burkholderia pseudomallei* in rice fields in Laos. *Appl Environ Microbiol* 77:532–536. <https://doi.org/10.1128/AEM.01822-10>.
- Thomas AD, Forbes-Faulkner J, Parker M. 1979. Isolation of *Pseudomonas pseudomallei* from clay layers at defined depths. *Am J Epidemiol* 110:515–521.
- Trung TT, Hetzer A, Topfstedt E, Gohler A, Limmathurotsakul D, Wuthiekanun V, Peacock SJ, Steinmetz I. 2011. Improved culture-based detection and quantification of *Burkholderia pseudomallei* from soil. *Trans R Soc Trop Med Hyg* 105:346–351. <https://doi.org/10.1016/j.trstmh.2011.03.004>.
- Trung TT, Hetzer A, Gohler A, Topfstedt E, Wuthiekanun V, Limmathurotsakul D, Peacock SJ, Steinmetz I. 2011. Highly sensitive direct detection and quantification of *Burkholderia pseudomallei* bacteria in environmental soil samples by using real-time PCR. *Appl Environ Microbiol* 77:6486–6494. <https://doi.org/10.1128/AEM.00735-11>.
- Parry CM, Wuthiekanun V, Hoa NT, Diep TS, Thao LT, Loc PV, Wills BA, Wain J, Hien TT, White NJ, Farrar JJ. 1999. Melioidosis in Southern Vietnam: clinical surveillance and environmental sampling. *Clin Infect Dis* 29:1323–1326. <https://doi.org/10.1086/313479>.
- Limmathurotsakul D, Wuthiekanun V, Chantratita N, Wongsuvan G, Amornchai P, Day NPJ, Peacock SJ. 2010. *Burkholderia pseudomallei* is spatially distributed in soil in northeast Thailand. *PLoS Negl Trop Dis* 4:e694. <https://doi.org/10.1371/journal.pntd.0000694>.
- Wuthiekanun V, Limmathurotsakul D, Chantratita N, Feil EJ, Day NP, Peacock SJ. 2009. *Burkholderia pseudomallei* is genetically diverse in agricultural land in northeast Thailand. *PLoS Negl Trop Dis* 3:e496. <https://doi.org/10.1371/journal.pntd.0000496>.
- Knappik M, Dance DA, Rattanavong S, Pierret A, Ribolzi O, Davong V, Silisouk J, Vongsouvanh M, Newton PN, Ditttrich S. 2015. Evaluation of molecular methods to improve the detection of *Burkholderia pseudomallei* in soil and water samples from Laos. *Appl Environ Microbiol* 81:3722–3727. <https://doi.org/10.1128/AEM.04204-14>.
- Price EP, Dale JL, Cook JM, Sarovich DS, Seymour ML, Ginther JL, Kaufman EL, Beckstrom-Sternberg SM, Mayo M, Kaestli M, Glass MB, Gee JE, Wuthiekanun V, Warner JM, Baker A, Foster JT, Tan P, Tuanyok A, Limmathurotsakul D, Peacock SJ, Currie BJ, Wagner DM, Keim P, Pearson T. 2012. Development and validation of *Burkholderia pseudomallei*-specific real-time PCR assays for clinical, environmental or forensic detection applications. *PLoS One* 7:e37723. <https://doi.org/10.1371/journal.pone.0037723>.
- Sermiswan RW, Royros P, Khakhum N, Wongratanacheewin S, Tuanyok A. 2015. Direct detection of *Burkholderia pseudomallei* and biological factors in soil. *Trans R Soc Trop Med Hyg* 109:462–468. <https://doi.org/10.1093/trstmh/trv040>.
- Kim Y, Liesack W. 2015. Differential assemblage of functional units in paddy soil microbiomes. *PLoS One* 10:e0122221. <https://doi.org/10.1371/journal.pone.0122221>.
- Liesack W, Schnell S, Revsbech NP. 2000. Microbiology of flooded rice paddies. *FEMS Microbiol Rev* 24:625–645. <https://doi.org/10.1111/j.1574-6976.2000.tb00563.x>.
- Song T, Martensson L, Eriksson T, Zheng W, Rasmussen U. 2005. Biodiversity and seasonal variation of the cyanobacterial assemblage in a rice paddy field in Fujian, China. *FEMS Microbiol Ecol* 54:131–140. <https://doi.org/10.1016/j.femsec.2005.03.008>.
- Edwards J, Johnson C, Santos-Medellin C, Lurie E, Podishetty NK, Bhatnagar S, Eisen JA, Sundaresan V. 2015. Structure, variation, and assembly of the root-associated microbiomes of rice. *Proc Natl Acad Sci U S A* 112:E911–E920. <https://doi.org/10.1073/pnas.1414592112>.
- Lu YH, Rosencrantz D, Liesack W, Conrad R. 2006. Structure and activity of bacterial community inhabiting rice roots and the rhizosphere. *Environ Microbiol* 8:1351–1360. <https://doi.org/10.1111/j.1462-2920.2006.01028.x>.
- Inglis TJ, Garrow SC, Henderson M, Clair A, Sampson J, O'Reilly L, Cameron B. 2000. *Burkholderia pseudomallei* traced to water treatment plant in Australia. *Emerg Infect Dis* 6:56–59.
- Levy A, Chang BJ, Abbott LK, Kuo J, Harnett G, Inglis TJJ. 2003. Invasion of spores of the arbuscular mycorrhizal fungus *Gigaspora decipiens* by *Burkholderia* spp. *Appl Environ Microbiol* 69:6250–6256. <https://doi.org/10.1128/AEM.69.10.6250-6256.2003>.
- Kaestli M, Schmid M, Mayo M, Rothballer M, Harrington G, Richardson L, Hill A, Hill J, Tuanyok A, Keim P, Hartmann A, Currie BJ. 2011. Out of the ground: aerial and exotic habitats of the melioidosis bacterium *Burkholderia pseudomallei* in grasses in Australia. *Environ Microbiol* 14:2058–2070. <https://doi.org/10.1111/j.1462-2920.2011.02671.x>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Li L, Stoekert CJ, Jr, Roos DS. 2003. OrthoMCL: identification of ortholog groups for eukaryotic genomes. *Genome Res* 13:2178–2189. <https://doi.org/10.1101/gr.1224503>.
- Spring-Pearson SM, Stone JK, Doyle A, Allender CJ, Okinaka RT, Mayo M,

- Broomall SM, Hill JM, Karavis MA, Hubbard KS, Insalaco JM, McNew LA, Rosenzweig CN, Gibbons HS, Currie BJ, Wagner DM, Keim P, Tuanyok A. 2015. Pangenome analysis of *Burkholderia pseudomallei*: genome evolution preserves gene order despite high recombination rates. *PLoS One* 10:e0140274. <https://doi.org/10.1371/journal.pone.0140274>.
37. Gee JE, Glass MB, Novak RT, Gal D, Mayo MJ, Steigerwalt AG, Wilkins PP, Currie BJ. 2008. Recovery of a *Burkholderia thailandensis*-like isolate from an Australian water source. *BMC Microbiol* 8:54. <https://doi.org/10.1186/1471-2180-8-54>.
38. Wuthiekanun V, Smith MD, Dance DA, White NJ. 1995. Isolation of *Pseudomonas pseudomallei* from soil in north-eastern Thailand. *Trans R Soc Trop Med Hyg* 89:41–43. [https://doi.org/10.1016/0035-9203\(95\)90651-7](https://doi.org/10.1016/0035-9203(95)90651-7).
39. Ashdown LR. 1979. An improved screening technique for isolation of *Pseudomonas pseudomallei* from clinical specimens. *Pathology* 11: 293–297. <https://doi.org/10.3109/00313027909061954>.
40. Galimand M, Dodin A. 1983. Diagnosis of the Whitmore bacillus in the laboratory. *Bull Soc Pathol Exot Filiales* 76:652–656. (In French.)
41. Gohler A, Hetzer A, Holtfreter B, Geisel MH, Schmidt CO, Steinmetz I, Kocher T. 2014. Quantitative molecular detection of putative periodontal pathogens in clinically healthy and periodontally diseased subjects. *PLoS One* 9:e99244. <https://doi.org/10.1371/journal.pone.0099244>.
42. Novak RT, Glass MB, Gee JE, Gal D, Mayo MJ, Currie BJ, Wilkins PP. 2006. Development and evaluation of a real-time PCR assay targeting the type III secretion system of *Burkholderia pseudomallei*. *J Clin Microbiol* 44: 85–90. <https://doi.org/10.1128/JCM.44.1.85-90.2006>.
43. Supaprom C, Wang D, Leelayuwat C, Thaewpia W, Sussaengrat W, Koh V, Ooi EE, Lertmemongkolchai G, Liu Y. 2007. Development of real-time PCR assays and evaluation of their potential use for rapid detection of *Burkholderia pseudomallei* in clinical blood specimens. *J Clin Microbiol* 45:2894–2901. <https://doi.org/10.1128/JCM.00291-07>.