1	Valorization of Bambara Groundnut Shell via Intermediate Pyrolysis : Products					
2	Distribution and Characterization					
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24 Abstract

This study provides first report on thermochemical conversion of residue from one of the 25 underutilized crops, Bambara groundnut. Shells from two Bambara groundnut landraces KARO 26 and EX-SOKOTO were used. Pyrolysis was conducted in a vertical fixed bed reactor at 500, 27 550, 600 and 650 °C; 50 °C/min heating rate and 5 L/min nitrogen flow rate. The report gives 28 experimental results on characteristic of the feedstock, impact of temperature on the pyrolysis 29 product distribution (bio-oil, bio-char and non-condensable gas). It evaluates the chemical and 30 physicochemical properties of bio-oil, characteristics of bio-char and composition of the non-31 condensable gas using standard analytical techniques. KARO shell produced more bio-oil and 32 was maximum at 600 °C (37.21 wt%) compared to EX-SOKOTO with the highest bio-oil yield 33 of 32.79 wt% under the same condition. Two-phase bio-oil (organic and aqueous) was collected 34

35 and analyzed. The organic phase from both feedstocks was made up of benzene derivatives which can be used as a precursor for quality biofuel production while the aqueous from KARO 36 consisted sugars and other valuable chemicals compared to the aqueous phase from EX-37 SOKOTO which comprised of acids, ketones, aldehydes and phenols. Characteristics of bio-char 38 and composition of the non-condensable were also determined. The results show that bio-char is 39 rich in carbon and some minerals which can be utilized either as a solid fuel or source of bio-40 fertilizer. The non-condensable gas was made up of methane, hydrogen, carbon monoxide and 41 carbon dioxide, which can be recycled to the reactor as a carrier gas. This study demonstrated 42 43 recovery of high quality fuel precursor and other valuable materials from Bambara groundnut shell. 44

Keywords: Bambara groundnut shell; Pyrolysis; Bio-oil; Bio-char; Non-condensable gas;
 Characterization

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Abbreviations

AAK	Acid, aldehyde, ketones
AMH	Atomic mass of hydrogen
AMO	Atomic mass of oxygen
С	Carbon
CFF	Crops for the future
DTG	Derivative thermogravimetric
EDX	Energy dispersive x-ray
ES	Esters
FTIR	Fourier transform infrared
GC	Gas chromatograph
GC-MS	Gas chromatograph-mass spectrometer
Н	Hydrogen
HC	Hydrocarbon
HHV	Higher heating value
KBr	Potassium bromide
LHV	Lower heating value
LH_w	Latent heat of vaporization of water

Ν	Nitrogen
0	Oxygen
PH	Phenolics
RKO	Karo Bambara shell
S	Sulfur
SGR	Sugars
SKT	EX-SOKOTO Bambara shell
TCD	Thermal conductivity detector
TG	Thermogravimetric
TGA	Thermogravimetric analyzer
VAC	Value added chemicals
XRD	X-ray diffraction

48 49

50 1. Introduction

Bambara groundnut (Vigna subterranea (L.) Verdc), is an underutilized pulse legume cultivated 51 primarily for its seeds, which are consumed in various forms of dishes, rich in protein (16-25 %), 52 carbohydrate (42-60 %), lipid (5-6 %) (Brough and Azam-Ali, 1992), fibre (4.8 %) and ash (3.4 53 %) (Brink et al., 2006). The fresh pods containing the seeds are boiled and eaten as snacks 54 55 whereas the haulm, is used as livestock feed in most of the drier parts of Africa (Heller et al., 1997). Legume such as Bambara groundnut with the ability to meet greater % of their N 56 requirement as a result of atmospheric N fixation have been identified as potential alternative 57 58 crops for mitigating climate change (Jensen et al., 2012) and offer an important alternative to the use of inorganic N fertilizers that are costly and unavailable to resource-poor farmers. The crop 59 is predominantly grown by subsistence farmers over much of semi-arid Africa and is reported to 60 spread into new production areas such as Indonesia, Malaysia and Thailand in the Asian 61 62 continent and is increasingly becoming popular due to its ability to thrive under conditions which 63 are too poor for other tropical legumes such as cowpea and groundnut (Mohale et al., 2014).

Bambara groundnut shell constitutes the weight of the pod wall after the grain is removed and ishighly variable depending on the pod yield and shelling percentage and sometimes could reach

up to 1000 kg ha⁻¹. Musa et al. (2016) reported 374-896 kg ha⁻¹ of shell weight which 66 corresponds to about 28.6-34.2 % of the total pod weight. Kishinevsky et al. (1996) reported a 67 shelling percentage of 59-76 %, which translates to about 648-1107 kg ha⁻¹ (24-41 % of the pod 68 weight) considering the average pod yield of 2700 kg ha⁻¹ reported by <u>Collinson et al. (1999)</u>. 69 Mwale et al. (2007) reported pod yield of 2980 kg ha⁻¹ and a shelling percentage of 80 %, which 70 corresponds to a shell weight of 596 kg ha⁻¹ (20 % of the total pod weight). Despite the high 71 weight of the crop's shells, information on the utilization of this potential is limited and is 72 seldom reported as a yield component due to lack of a specific value (Mwale et al., 2007). There 73 74 is need to explore alternative ways to promote the utilization of this product which could address the problem of crop waste disposal and promote the integration of the crop into new production 75 areas. Crop residue disposal in the sub-humid tropical regions such as Malaysia and Indonesia 76 77 has been attracting a major global concern due to inappropriate methods adopted by the resource poor farmers, particularly in Indonesia, where farmers resort to the use of open burning as a 78 cheap way of clearing crop residues from the farm and thereby causing air pollution (haze) to 79 the neighboring countries and the entire region (Varkkey, 2013; Forsyth, 2014). 80

Pyrolysis is currently one of the most promising thermochemical processes for converting 81 biomass materials to products with high energy potentials. Typical pyrolysis products are bio-oil, 82 bio-char and non-condensable gas (Alburquerque et al., 2016; Gómez et al., 2016). Distribution 83 of pyrolysis products depends heavily on the process parameters such as pyrolysis temperature, 84 heating rate and vapor residence time in the reactor. There are different types of pyrolysis 85 namely; slow, intermediate and fast pyrolysis. Slow pyrolysis is also referred to as 86 carbonization. It is carried out at a temperature up to 400 °C, solid residence time of 1 h to 87 88 several hours, with product distribution of about 35 % bio-char, 30% bio-oil, and 35% non-

condensable gas. Fast pyrolysis can produce up to 80 % bio-oil, 12 % bio-char, and 13 % non-89 condensable gas at temperature around 500 °C, high heating rate, a short vapor residence time of 90 about 1 s, and rapid cooling of volatile (Bridgwater, 2012; Mohammed et al., 2015a). For 91 intermediate pyrolysis, 500-650 °C pyrolysis temperature and vapor residence time of 92 approximately 10 to 30 s are typical operating conditions. About 40-60% of the total product 93 yield is usually bio-oil, 15-25% bio-char and 20-30% non-condensable gas. Intermediate 94 pyrolysis produce bio-oil with less reactive tar which can be used directly as fuel in engines and 95 boilers, and dry char suitable for both agricultural and energy applications (Tripathi et al., 2016). 96 97 To the best of our knowledge, thermochemical conversion of Bambara groundnut shells has not been reported in the literature. In this study, intermediate pyrolysis of Bambara shell was 98 investigated in a fixed bed reactor and pyrolysis products were characterized. 99

100 **2. Materials and methods**

101 Shells from two Bambara groundnut landraces, Karo (RKO) and EX-Sokoto (SKT) were collected from Crops for the Future (CFF) field research Centre Semenyih, Selangor, Malaysia. 102 The material was dried at 105 °C in electrical oven. The dried materials were then shredded in a 103 Retsch[®] rotor beater mill (SM100, Retsch GmbH, Germany) to particle sizes between 0.2 mm 104 and 2.5 mm and stored in airtight plastic bags for further analysis. Volatile matter and ash 105 content on dry basis were determined. Fixed carbon was computed from the remaining bone dry 106 sample mass. The biomass ash elemental composition was analyzed with energy dispersive X-107 ray EDX (X-Max 20 mm², Oxford Instruments, UK). Higher heating value (HHV) was 108 determined using an oxygen bomb calorimeter (Parr 6100, Parr Instruments, Molin, USA). 109 Elemental compositions were determined using CHNS/O analyzer (2400 Series II CHNS/O 110 analyzer, Perkin Elmer Sdn Bhd, Selangor, Malaysia). The nature of chemical functional groups 111

were evaluated with Fourier transform infrared spectroscopy (Spectrum RXI, PerkinElmer, 112 Selangor, Malaysia) using the potassium bromide (KBr) method. Potassium bromide (KBr) disc 113 (13 mm diameter translucent) was made from a homogenized 2 mg sample in 100 mg KBr using 114 a bench press (Carver 43500, Carver Inc., USA) by applying 5.5 tonnes load for 5 min. Spectra 115 were recorded with Spectrum V5.3.1 software within wave number range of 400 to 4000 cm⁻¹ at 116 32 scans and a resolution of 4 cm⁻¹. Thermogravimetric study was carried out using 117 thermogravimetric simultaneous thermal analyzer (TGA) (STA 6000, Perkin Elmer Sdn Bhd, 118 Selangor, Malaysia) in nitrogen atmosphere, flow rate 20 mL/min at temperatures between 30-119 950 °C and heating rate of 20 °C/min. About 10 mg (particle size of 0.2 mm) of sample was used. 120

Intermediate pyrolysis study was conducted at 500, 550, 600 and 650 °C in a vertical fixed bed 121 122 pyrolysis system as shown in Figure 1. The system consists of a fixed bed reactor made of 123 stainless steel (115 cm long, 6 cm inner diameter), a distribution plate with 1.5 mm hole diameter which sit at 25 cm from the bottom of the tube, two nitrogen preheating sections, a cyclone, oil 124 collector, gas scrubbers and water chiller operating at 3 °C attached to a coil condenser. 200 g of 125 shell (bone dried, 2.5mm particle size) was placed on a gas distribution plate inside the reactor 126 127 tube. The setup was heated in a vertical furnance at 50 °C/min under 5 L/min nitrogen flow. The 128 reaction temperature was monitored with a thermocouple (K-type, NTT Heating, Sdn Bhd, 129 Selangor, Malaysia). The reaction time was kept at 60 min after the temperature attained the setvalue. Pyrolysis vapor was condensed by passing through a condenser attached to the water 130 131 chiller and condensate (bio-oil) was collected in a container. Bio-oil, bio-char and noncondensable gas yields were calculated using Eq (1), (2) and (3). The experiment was repeated in 132 triplicates and stanadard deviations were computed. 133

134 Yield
$$(wt\%)_{bio-oil} = \left[\frac{weight of the bio-oil collected}{weight of biomass feed} \times 100\right] - \dots (1)$$

135 Yield $(wt\%)_{bio-char} = \left[\frac{weight of the bio-char collected}{weight of biomass feed} \times 100\right] - \dots (2)$
136 Yield $(wt\%)_{non-condensable gas} = 100 - (Eq(1) + Eq(2)) - \dots (3)$

137

Characterization of bio-oil, bio-char and non-condensable gas were carried out accordingly using 138 139 analytical instruments as summerized in Table 1. FTIR of bio-oil was determined using a pair of circular demountable KBr cell windows (25 mm diameter and 4 mm thickness). Detail chemical 140 composition of the bio-oil was determined using a gas chromatograph-mass spectrometer (GC-141 MS) system equipped with a quadruple detector and column (30m x 0.25mm x 0.25mm). The 142 oven was programmed at an initial temperature of 40 °C, ramp at 5 °C /min to 280 °C and held 143 there for 20 min. The injection temperature, volume, and split ratio were 250 °C, 1 µL, and 50:1 144 respectively. Helium was used as carrier gas at a flow rate of 1 mL/min. The peaks of the 145 chromatogram were identified by comparing with standard spectra of compounds in the National 146 147 Institute of Standards and Technology library (NIST, Gaithersburg, USA).

148 The composition of non-condensable pyrolysis product was monitored offline. The gas sample was collected in a gas sample bag (Tedlar, SKC Inc., USA) and its composition analyzed using a 149 150 gas chromatography equipped with stainless steel column (Porapak R 80/100) and thermal conductivity detector (TCD). Helium was used as a carrier gas and the GC was programed at 60 151 °C, 80 °C and 200 °C for oven, injector and TCD temperature respectively. Bio-char ultimate 152 analysis and inorganic composition were carried out following the procedure outlined for the 153 feedstock characterization. Crystallographic structure in the produced bio-char was examined 154 between 2O angle of 10°-70° at 25 mA, 45 kV, step size of 0.025° and 1.0 s scan rate. Surface 155

and structural characteristics of the bio-char were also determined .

157 **3. Results and Discussion**

158 3.1 Characterization of feedstock

159 Characteristic of the feedstock is summarized in Table 2. Moisture content of SKT and RKO (as received) was 4.67 and 7.12 wt%. Higher volatile matter (76.24 wt%) and lower ash content 160 161 (8.98 wt%) was recorded for the RKO compared to 73.83 wt% volatile matter and 10.10 wt% ash content in the SKT. Similarly, the ultimate analysis revealed that RKO has carbon and 162 oxygen content of 37.36 wt% and 49.11 wt%, while 34.63 wt% carbon and 51.93 wt% oxygen 163 was recorded for the SKT. RKO has higher heating value (HHV) of 20.46 MJ/kg relative to 164 19.19 MJ/kg recorded for SKT. The higher energy content recorded for the RKO could be 165 attributed to its lower ash and oxygen contents. These physiocchemical characteristics are similar 166 to properties of peanut shell reported by Zhang et al. (2011). The authors recorded high volatile 167 matter and heating value of 71.5 wt% and 18.1 MJ/kg respectively. Their ultimate analysis result 168 169 showed higher carbon (47 wt%), and lower oxygen (34 wt%) and hydrogen (6 wt%) contents compared to that of the Bamabara shells. EDX analysis of the ash showed that the inorganic 170 element in the RKO and SKT is in the following order potassium (K)>silica (Si)>Aluminum 171 172 (Al)>phosphorus (P)>Chlorine (Cl)>iron (Fe)>calcium (Ca)>magnesium (Mg)and K>Cl>Si>Al>P>Mg respectively. The proportion of metallic elements in the RKO is similar to 173 that of peanut reported by Kuprianov and Arromdee (2013). The authors also reported 174 distribution of minerals in tamarin shell which is comparable to that of SKT except for the Si, 175 176 which is higher in SKT. The result of thermogravimetric analysis (TGA) analysis is shown in Figure 2. The thermogravimetric (TG) and derivative thermogravimetric (DTG) profiles of both 177 samples exhibited similar decomposition characteristics. The residual weight at the end of the 178

179 decomposition process was found to be 23 and 29 wt% for RKO and SKT respectively. From the DTG curves, two main peaks from ambient to 100 °C and from 240 to 420 °C, which is attributed 180 to evaporation of moisture and decomposition of holocellulose (hemicellulose and cellulose) 181 component of the biomass. Hemicellulose which normally decomposes around 250 °C in a 182 lignocellulosic material was not observed in this study. It means that Bambara shells have less or 183 infinitesimal amount of hemicellulose. A noticeable peak around 200 °C was also observed, 184 which is ascribed to the decomposition of extractives. The pattern observed at the temperature 185 above 420 °C is attributed to the decomposition of lignin. This region exhibited unsteady thermal 186 187 degradation, which is ascribed to the presence of different oxygen functional groups in the lignin (Mohammed et al., 2015b). TGA analysis of almond shell by Grioui et al. (2014) showed a 188 residual weight fraction of 27.5 wt%. The DTG curve exhibited main peak at temperature 189 between 220 and 400 °C, which was attributed to the decomposition of cellulose and 190 hemicellulose. The decomposition temperature values observed in this study are aslo similar to 191 those of other lignocellulosic biomass in the literature (Mohammed et al., 2015c). 192

193 3.2 Pyrolysis product distribution and physicochemical properties of the bio-oil

Product distribution from the pyrolysis of RKO and SKT at temperature between 500 and 650 194 ^oC, 5 L/min nitrogen flow and heating rate 50 ^oC/min is shown in Fig. 3. From Figure 3(a), total 195 bio-oil yield from RKO increased from 33.26 to 37.21 wt% as the temperature increased from 196 500 to 600 °C. This observation could be attributed to the decomposition of more lignin at such a 197 high temperature. At 650 °C, the oil yield dropped to 27.69 wt%, which is due to secondary 198 199 reactions of pyrolysis vapor at the elevated temperature (Mohammed et al., 2015a). The bio-oil collected consist of organic phase and an aqueous phase. The yield of organic phase increased 200 from 9.8 wt% to 15.0 wt% at the temperature between 500 and 600 °C and latter declined to 9.3 201

wt% at 650 $^{\circ}$ C. On the other hand, a continuous decrease in the aqueous phase from 24.3 to 18.4 202 wt% was recorded. Generally, the aqueous phase bio-oil is a product of holocellulose 203 decomposition whose yield and composition is greatly affected by the inorganic minerals in the 204 source biomass. The significant reduction in the bio-oil aqueous phase and increased non-205 condensable yield observed could be attributed to the synergistic effect of the most abundant 206 207 minerals in the biomass ash, which are Al and K (Table 2) since Si is an inert material and may not possess any catalytic activity (Mohammed et al., 2016a). The yield of non-condensable gas 208 was 32.43 and 32.57 wt% at 500 and 550 °C but increased rapidity to 33.28 and 43.11 wt% at 209 210 600 and 650 °C. The corresponding bio-char yield was 32.96, 32.26, 29.51 and 29.20 wt%. The high yield of non-condensable gas observed above could be mainly due to the secondary 211 reactions of pyrolysis vapor at the elevated temperature (Mohammed et al., 2015a) since there 212 was no substantial reduction in the biochar yield. Figure 3b shows pyrolysis product distribution 213 from the SKT. The total bio-oil yield was 26.17, 32.18, 32.78 and 27.58 wt% at 500, 550, 600 214 and 650 °C respectively. Similar to RKO, two phase bio-oil was collected from the pyrolysis of 215 SKT and the corresponding organic and aqueous phases recorded were 5.90, 10.84, 11.17, 5.77 216 wt% and 20.27, 21.34, 21.61, 21.81 wt%. The bio-char yield decreased from 39.91 wt% at 500 217 °C to 28.56 wt% at 650 °C while the corresponding yield of non-condensable gas increased from 218 33.92 to 43.86 wt%. The high yield of non-condensable gas could be a resultant effect of a 219 secondary reaction of pyrolysis vapor and devolatilization of the bio-char at high temperature. 220 221 Comparing the product distribution between RKO and SKT, the highest total bio-oil yield of 37.21 wt% was recorded at 600 °C from RKO compared to 32.78 wt% from SKT. A substantial 222 decrease in the biochar yield (11.35 %) with increasing temperature (500-650 °C) was observed 223 224 from SKT compared to 4.06 % recorded from the RKO. Higher non-condensable gas yield was

also recorded with SKT at all temperatures relative to that obtained from RKO. These 225 observations can be attributed to the differences in the characteristics of the 226 feedstocks(Mohammed et al., 2016b), as summerized in Table 2. Comparing the pyrolysis 227 product distribution with products yield from other shells, Zhang et al. (2011) reported a total 228 pyrolysis oil of 60 wt% from peanut shell at 500 °C with corresponding 19 wt% char and 15 229 230 wt% non-condensable gas. Grioui et al. (2014) reported pyrolysis products distribution of 40 wt% bio-oil, 47 wt% gas and 13 wt% solid. Pyrolysis oil yield from other biomass species have 231 been reported by Lim et al. (2015) and (2016). They recorded bio-oil yield of 41.91-57.21 wt% 232 and 37.98-43.66 wt% from Napier grass and sago waste respectively at 600 °C pyrolysis 233 temperature. The variation in the products distribution can be linked to physicochemical and 234 structural characteristics of the respective feedstock. Physicochemical properties of Bamabara 235 shell bio-oil are summarized in Table 3. The pH value of organic phase from RKO and SKT was 236 4.4 and 3.3, which indicate that the oils are less acidic relative to the almond bio-oil (pH = 3.1) 237 reported by Grioui et al. (2014) (Table 3). Similarly, density of the bio-oils are relatively lower 238 than the values reported in the literature for almond and peanut bio-oil. Ultimate analysis result 239 (dry basis) calculated from the wet basis results using Eq (4), (5), (6) (de Miguel Mercader et al., 240 241 2010; Mohammed et al., 2016c) and (7) (Mohammed et al., 2015d) indicated that C, H, O and HHV of the organic phase bio-oil from both Bambara shells are comparable to the elemental 242 composition of peanut and almond shell bio-oil. 243

244
$$C_{dry} = \frac{C_{wet}}{\left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right)} - \dots - \dots - \dots - (4)$$

245
$$H_{dry} = \frac{\left(H_{wet}\right) - \left[\eta_{H_2O} \times \left(\frac{2 \times AMH}{(2 \times AMH + AMO)}\right)\right]}{\left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right)}$$
246
$$O_{dry} = \frac{\left(O_{wet}\right) - \left[\eta_{H_2O} \times \left(\frac{AMO}{(2 \times AMH + AMO)}\right)\right]}{\left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right)}$$
247
$$HHV_{dry} = \frac{\left(LHV_{wet}\right) + \left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right) \times LH_w}{\left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right)}$$
247
$$(LHV_{dry} = \frac{\left(LHV_{wet}\right) + \left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right) \times LH_w}{\left(1 - \left(\frac{\eta_{H_2O}}{100}\right)\right)}$$

C, H and O is carbon, hydrogen and oxygen content in wt %; η_{H2O} is the amount of water in the oil (wt %); AMH and AMO is atomic mass of hydrogen and oxygen; LH_w is the latent heat of vaporization of water (2.2MJ/kg)

251

252 3.3 Fourier-Transform Infra-Red (FTIR)

FTIR spectra of chemical species in the bio-oil are shown in Figure 4. The common broad peak around 3439 cm⁻¹ implies that the samples contain chemical compounds with hydroxyl group (O-H) such as water, alcohols and phenol (Bordoloi et al., 2015). The peaks between 2838 and 2948 cm⁻¹ common to the organic phase bio-oils (Figure 4a and 4b) is C–H stretching vibration, which indicate the presence of saturated hydrocarbon in the organic phase while the

peak between 2041 and 2132 cm⁻¹ common to all the samples is ascribed to the C = N258 functional group (Guo et al., 2015). Vibrations observed between 1625 cm⁻¹ and 1707 cm⁻¹ in all 259 the oil phases are attributed to C = O, which signifies the presence of aldehydes, ketones or 260 carboxylic acids. The vibration at a frequency of 1515 cm⁻¹ in both organic phases is ascribed to 261 C = C bond while the band around 1454 cm⁻¹ in all the samples is ascribed to C - H bending, 262 suggesting the presence of alkenes/aromatic hydrocarbons. The vibrations observed at 1364, 263 1230 and 1273 cm⁻¹ signify C - N stretching (Yorgun and Yıldız, 2015). The sharp band 264 around 1112, 1093, 1050 and 1025 cm^{-1} are due to C-O vibration indicating the presence of 265 alcohol and esters. The fingerprint at 755 cm^{-1} is ascribed to aromatic C – H bending vibrations 266 while the fingerprint between 655 and 531 cm⁻¹ is ascribed to alkyl halide (Yorgun and Yıldız, 267 2015). 268

269 3.4 GC-MS analysis of the bio-oil samples

Identification of detail chemical compounds in the bio-oil samples collected at 600°C was carried 270 271 out by GC-MS. The chromatograms (Figure 5) show twenty (20) most abundant compounds in the bio-oils. The analysis revealed that the organic phase from both RKO and SKT consist of 272 273 similar compounds such as alkenes, benzene derivatives, nitrogenated compounds, esters, aldehydes, ketones and acids in different proportions. A trace of sugar (L-Galactose) was also 274 detected in the oil from RKO while none was observed in the oil from SKT. The aqueous phase 275 from RKO consists predominant sugars and fine chemicals while that from SKT consist mainly 276 acids, ketones, aldehyde and phenolics. In order to elucidate the variation of these compounds in 277 the oil samples, the compounds were further grouped to hydrocarbon(HC), esters (ES), phenolics 278 (PH), sugars (SGR), value added chemicals (VAC) and acid, aldehyde, ketones (AAK). From 279 Figure 6, based on the GC-MS peak area (%), 7.09 % HC and 52.17 % PH was recorded in the 280

organic phase RKO while 66.06 %PH was observed in the SKT with 2.31 % HC. The high 281 content of SGR (30.69 %) and low content of AAK (7.04 %) in the aqueous phase from the RKO 282 compared to low SGR (4.42 %) and high AAK (36.95 %) in the SKT is attributed to the 283 catalytic effect of the respective biomass ash. Studies have shown that alkaline metals tend to 284 catalyze pyrolysis reactions which usually lead to degradation and polymerization of pyrolysis 285 286 product intermediate (Deshmukh et al., 2015; Bordoloi et al., 2015). Potassium (K) has been identified to have impacts on both the pyrolysis product distribution and the composition of bio-287 oil. Studies by Shimada et al. (2008); Nowakowski et al. (2008); Mohammed et al. (2016a) have 288 demonstrated that potassium strongly promoted the formation of low molecular weight 289 compounds and inhibited the formation of sugar during biomass pyrolysis. Similar observations 290 have also been reported by Wang et al. (2010) and Trendewicz et al. (2015) during the pyrolysis 291 of pine wood and cellulose respectively. They also reported greater phenol yield with K as a 292 catalyst during the pyrolysis. Therefore, the high proportion of PH, AAK and lower content of 293 SGR in the oil from the SKT relative to RKO is ascribed to the higher K content in the SKT 294 (72.57 wt%) relative to 44.96 wt% in the RKO (Table 2). Compositions of bio-oil from almond 295 shell bio-oil were mainly HC (alkanes and oleifins) as reported by Grioui et al. (2014) while 296 297 peanut shell bio-oil constitutes about 59 % PH and 31 % AAK (Zhang et al., 2011). Lim et al. (2016) reported that bio-oil from Napier grass and sago waste consititute mainly PH and AAK 298 and PH, AAK and trace of HC respectively. The authors linked the difference in the chemical 299 300 composition to the presence of different mineral elements in the respective biomass.

301 *3.5 GC* analysis of the non-condensable gas sample

Composition the non-condensable gas collected at 600 $^{\circ}$ C is summarized in Table 4. The gas constitutes a high percentage of methane (CH₄), hydrogen (H₂), carbon monoxide (CO) and

carbon dioxide (CO_2). The high composition of CH_4 signifies thermal cracking mechanisms that 304 produce small organic molecules during the pyrolysis. The proportion of CH₄ recorded in the 305 gas from RKO (9.43 vol%) was higher than that obtained from the SKT (7.42 vol%). This 306 suggests that there is interaction between the minerals Al (19.61 wt%) and K (44.96 wt%) since 307 Si is an inert material, (Table 2) in the biomass ash promoted the conversion of CH₄ precursors 308 309 to form hydrocarbons. This observation is in good agreement with the higher percentage of total hydrocarbons recorded in the bio-oil from RKO (18.28 % in both phases) relative to SKT (2.31 310 %). On the other hand, higher composition of H₂, CO and CO₂ in the gas from SKT was recorded 311 312 compared to RKO and could be linked to the higher percentage of K in the SKT ash. Study by Wang et al. (2010) has shown that K promotes production of H_2 , CO and CO₂ during pyrolysis 313 of lignocellulosic biomass. 314

315 3.5 Characteristics of produced bio-char

The properties of bio-char collected at 600 °C (Table 5) shows that both RKO and SKT produced 316 bio-char rich in carbon. The atomic ratio, H/C and O/C were 0.03 and 0.14 in RKO-char while 317 0.01 and 0.13 were recorded in SKT-char. The relative lower ratios observed in the SKT-char 318 indicates that SKT produced higher carbonized char which connotes a greater degree of 319 aromaticity. Comparing this characteristic with the individual biomass feedstock, H/C, O/C of 320 RKO and SKT biomass was 0.3, 1.3 and 0.33, 1.5 (Table 2). The decrease in the ratio, 321 particularly, the O/C in the produced bio-char was due to dehydration reaction during the 322 pyrolysis (Al-Wabel et al., 2013). 323

FTIR of the produced bio-char was compared with the source biomass as shown in Figure 7. The averaged spectra showed similar characteristic peaks for both RKO and SKT biomass. Similarly, common characteristic peaks were observed for the produced biochars. The peaks detected

between 3723 and 3505 cm^{-1} in the feedstocks are assigned to different hydroxyl groups (O-H), 327 principally from the cellulose component of the biomass. The peak at 3334 cm⁻¹ is ascribed to 328 N-H functional group, indicating the presence of nitrogenous group in the feedstock while the 329 peak observed at 3165, 3111 and 2960 cm⁻¹ are due to C - H stretching vibrations of cellulose, 330 hemicellulose and lignin in the biomass samples (Usman et al., 2015). These peaks disappeared 331 in the produced bio-chars, confirming the decomposition of structural component of the biomass 332 feedstock (Mimmo et al., 2014) while that observed around 3700 cm⁻¹ due to O-H could be 333 334 attributed to moisture adsorbed during sample preparations (Chen et al., 2016). The vibration at 1658 and 1588 cm⁻¹ in the feedstock and produced bio-char is due to C = C stretching while the 335 band at around 1417 cm^{-1} in all the samples is ascribed to C-H bending. The band observed in 336 the feedstock at 1246 and 1033 is ascribed to C-O-C and C-O stretching vibrations of 337 polysaccharides while the fingerprint at 545 and 462cm⁻¹ is due to alkyl halides (Mohammed et 338 al., 2015b). These peaks completely disappeared in the produced bio-char confirming the 339 degradation of polysaccharides and alkyl halides during pyrolysis. 340

Physisorption analysis (Table 5) revealed that the produced bio-char is a porous material with a pore volume of 0.01 cm³/g (RKO-char), 0.2 cm³/g (SKT-char) with corresponding specific surface area (BET) of 1.2 and 1.72 m²/g. This observation is also evident in the SEM micrograph (Figure 8) which showed cracked, uneven and visible porous structure in the bio-char due to the high degree of devolatilization at high temperature (600 °C) during the pyrolysis.

346 XRD analysis of the feedstocks and the produced bio-char is presented in Figure 9. The 347 diffractogram showed main characteristic peak for both RKO and SKT biomass at 22.23° and 348 22.09° respectively, which represent the crystalline component in the biomass, mainly cellulose 349 (<u>Timpano et al., 2015</u>). This peak completely disappeared in the produced bio-char,

350 confirming complete decomposition of cellulose during the pyrolysis. New sets of peaks were identified in the produced bio-chars around 26, 28, 30, 31, 40 and 50°. XRD search and 351 match reveal that the bio-char constituted predominantly sylvite mineral represented by the 352 peaks in both chars around 28.63°, 40.65° and 50.43° (Usman et al., 2015). The peak observed 353 around 26.83° and 30°, 31° indicate the presence of quartz and calcite (Yuan et al., 2011). 354 Thermogravimetic analysis (Figure 10a) of the produced bio-char showed a total residual weight 355 of 85.38 wt% and 86.49 wt% at 600 °C for SKT-bio-char and RKO-bio-char respectively 356 compared to the individual feedstock with 37.67 wt% and 32.47 wt%. This result indicates that 357 almost all the volatile matters in the biomass were completely volatilized during the pyrolysis. 358 This observation is also confirmed in the DTG plot (Figure 10b) as all the peaks in the feedstock 359 completely disappeared at 600 °C and above. The weight loss recorded at 600 °C in the bio-char 360 (13.51 wt%-RKO-char, 14.62 wt%-SKT-char) is due to combustion of the remaining organic 361 material in the bio-char. At higher temperature (900 °C), the total weight loss was 78.77 wt% in 362 RKO-char and 76.76 wt% in SKT-char, which correspond to about 7.72 wt% and 8.62 wt% 363 weigh loss between 600 and 900 °C. This observation could be attributed to the decomposition 364 of inorganic materials in the biochar at higher temperature (Azargohar et al., 2014). 365

366 **4. Conclusion**

Pyrolysis of Bambara groundnut shell was carried out in a fixed bed reactor. This study evaluates impact of temperature on the pyrolysis product distribution (bio-oil, bio-char and noncondensable gas). It assesses the chemical and physicochemical properties of bio-oil, characteristics of bio-char and composition of the non-condensable gas. The bio-oil collected was a two-phase liquid, organic and aqueous phase. The organic phase from both shells have high energy content (25-29 MJ/kg) and could be used directly as a fuel in boilers or a precursor

373 for quality biofuel production. The aqueous phase particularly from RKO was predominantly made up of sugars and other value-added chemicals, which can be used as a raw material for 374 production of consumer products and fine chemicals. Bio-char was a porous material rich in 375 carbon, which may well be used as a solid fuel. The bio-char also consist of macronutrient and 376 may be applied as a source of nutrient for plant growth. Non-condensable gas consists of 377 methane, hydrogen, carbon monoxide and carbon dioxide which can be recycled to the reactor as 378 an energy supplement for the endothermic pyrolysis reaction or as a carrier gas. Using Bambara 379 shells as feedstock for bioenergy production will encourage more production of the groundnut, 380 which will bring about food security, particularly in challenging environments where major 381 crops are severely limited by adverse climatic conditions in addition to income generation. 382

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