University of Massachusetts Amherst ScholarWorks@UMass Amherst

Cranberry Station Resistance Management

Cranberry Station Outreach and Public Service Activities

2016

Pesticide Resistance Management

Katherine Ghantous UMass Amherst - Cranberry Station

Hilary Sandler University of Massachusetts Amherst

L. McDermott

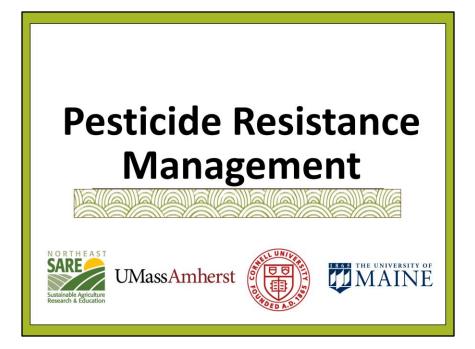
Follow this and additional works at: https://scholarworks.umass.edu/ cranberry_outreach_resistance

Part of the <u>Agriculture Commons</u>

Ghantous, Katherine; Sandler, Hilary; and McDermott, L., "Pesticide Resistance Management" (2016). Cranberry Station Resistance Management. 1.

Retrieved from https://scholarworks.umass.edu/cranberry_outreach_resistance/1

This Article is brought to you for free and open access by the Cranberry Station Outreach and Public Service Activities at ScholarWorks@UMass Amherst. It has been accepted for inclusion in Cranberry Station Resistance Management by an authorized administrator of ScholarWorks@UMass Amherst. For more information, please contact scholarworks@library.umass.edu.



What is Pesticide Resistance?

An <u>inheritable</u> characteristic of a pest that makes it less sensitive to a pesticide

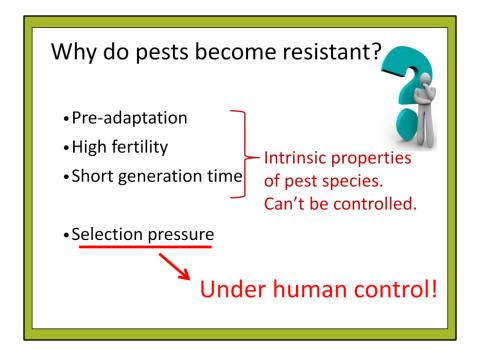
- Renders the pest able to survive exposure to that pesticide that would normally kill those without the genes
- Occurs in all pests weeds, insects, fungi, etc.
- Reflected by failure of a product to achieve expected level of control

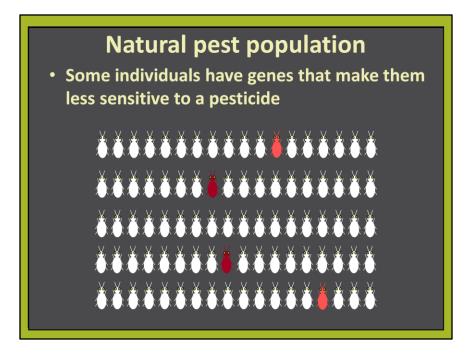
What is Pesticide Resistance?

- A mutation makes a pest less sensitive to a pesticide, can naturally occur in pest population before pesticide use
- Pesticide use kills susceptible individuals (those without the mutation/gene), and "selects" those with the mutation to survive
- Pests with the mutation live, reproduce, and pass on the genes, which made them less sensitive to the pesticide, to their offspring
- The pest population has increasing numbers of resistant individuals

What is tolerance? Tolerance - natural ability of a species or individual member of a species to survive and reproduce after a pesticide treatment. This implies no selection or genetic manipulation to make the organism this way. Tolerance is a natural tendency and is not a result of selection pressure. Example: Mature caterpillars are more tolerant to many insecticides than younger ones of the same species due to differences in body size, exoskeleton thickness, and the ability to metabolize a poison. These differences are identified as tolerance or natural resistance rather than true insecticide resistance.

In contrast to resistance, insecticide tolerance is a natural tendency and is not a result of selection pressure. Mature caterpillars are more tolerant to many insecticides than younger ones of the same species due to differences in body size, exoskeleton thickness, and the ability to metabolize a poison. These differences are identified as tolerance or natural resistance rather than true insecticide resistance.







Pesticide application

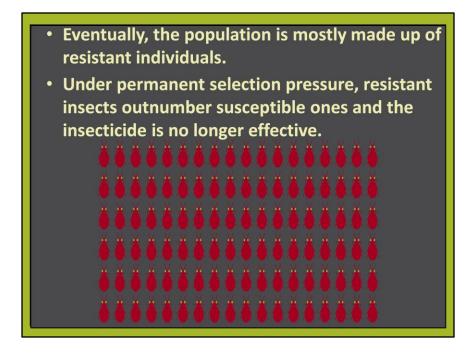
 Individuals with naturally occurring genes that make them less sensitive to a pesticide survive...

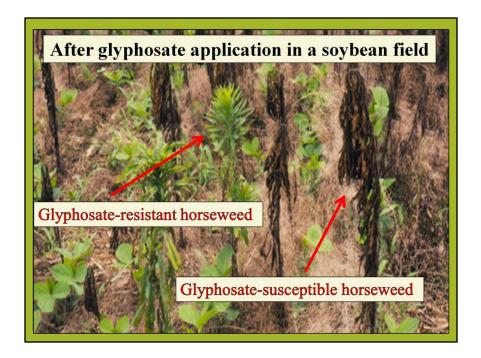


After pesticide application

- Humans have applied selection pressure.
- Individuals with genes that make them less sensitive to a pesticide reproduce.
- The offspring have the genes that make them less sensitive to the pesticide.
- The new population is more resistant than a natural population



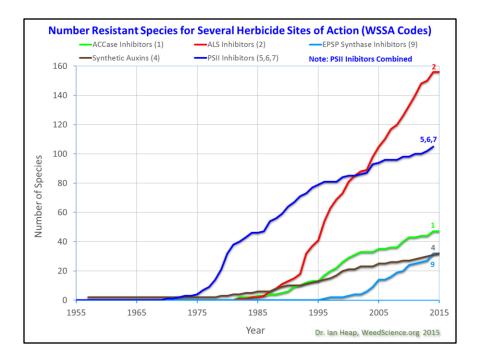




Weed pest resistance occurs in a similar fashion as with insect pest resistance. This slide shows what happens as resistance is developed – the resistant individuals will eventually make up the entire population.

- Currently:
 - $_{\odot}~$ 520 insect and mite species
 - At least 17 insect species are resistant to <u>all</u> major classes of insecticides
 - 273 weed species
 - 150 plant diseases
 - 10 rodent species

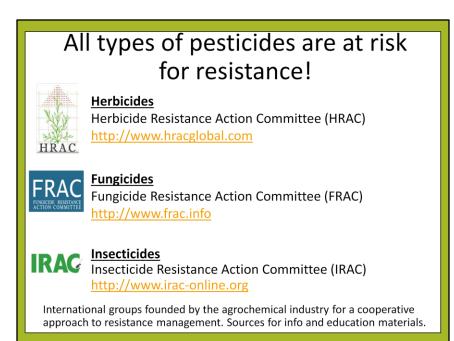




This graph presents the chronological increase in resistance to 5 herbicide sites of action. The letters refer to the Weed Science Society of America code (WSSA) to identify herbicide sites of action. Different herbicide sites of action have different propensities to select resistance. PSII inhibitor (Group 5, 6, 7) herbicides, primarily atrazine resistant weeds in corn, dominated in the USA and Europe in the 1970's and 80's. ALS inhibitor (Group 2) herbicides are the most prone to resistance. Note that the Y axis is the number of species (because a species is only plotted once per site of action).

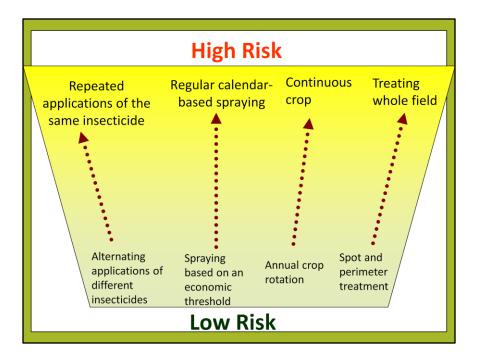
Why is Managing Resistance Important?

- We want pesticides to provide effective control of pests
- It's hard to develop new products for pest control
 - If we exhaust current technology, it might be a long time until something is developed to replace it!
- Environmental stewardship
 - Pest are mobile, resistance that develops in one crop can spread
 - Minimize ineffective use?? of pesticides



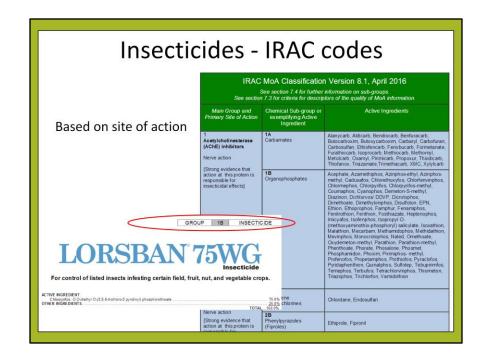
What Products are Resistant Prone?

- All pesticides have some risk, BUT not to the same extent
- Those with single-site mode of action, targeted and narrow-spectrum more at risk

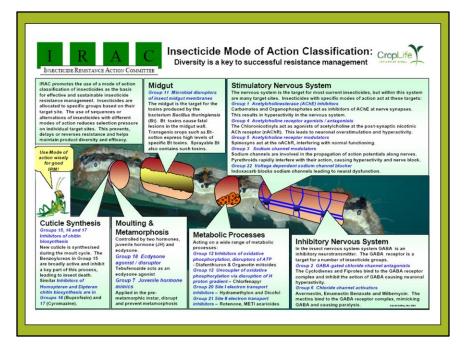


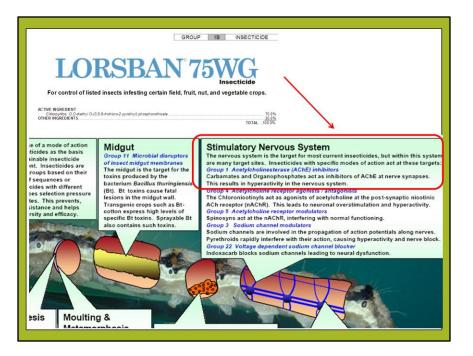
Mode of action (MoA)

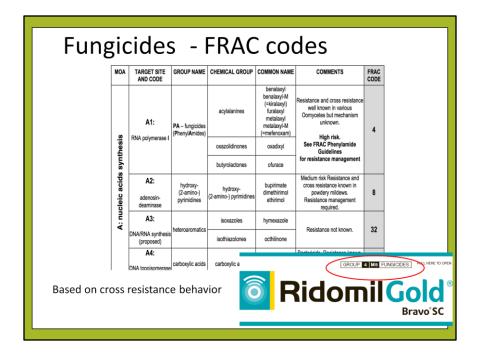
- The chemical structure of a pesticide generally defines its target site and its mode of action at that target site.
 - <u>Target site</u> the physical location within an organism where the pesticide acts
 - <u>Mode of action</u> the action of a pesticide at its target site. The way in which it causes physiological disruption at the target site.
- Each pesticide has a **Group Number** to help growers make resistance management decisions
- Group number is clearly marked on all labels

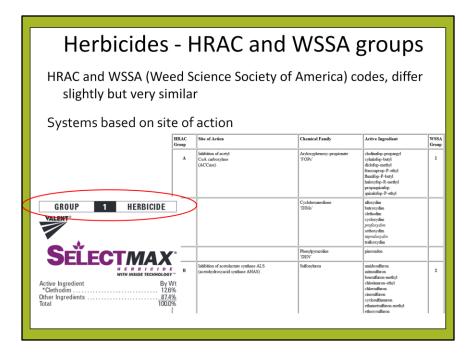


Educators are encouraged to show farmers how to find IRAC group codes in the pesticide guideline that they use routinely.

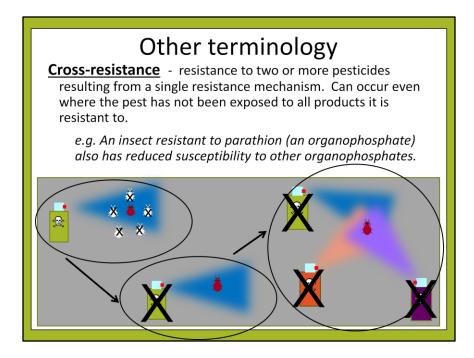








Just as a note to educators: WSSA is currently in discussion with EPA to see if these 2 systems can be unified somehow.



Other terminology

<u>**Multiple resistance**</u> - resistance to several pesticides from two or more distinct resistance mechanisms in the same organism.

e.g. Palmer amaranth is resistant to atrazine (a triazine herbicide), and also mesotrione (an HPPD inhibitor herbicide).



Other terminology

Quantitative Resistance

- Either the pesticide works.... or it does not.
- Complete loss of control.

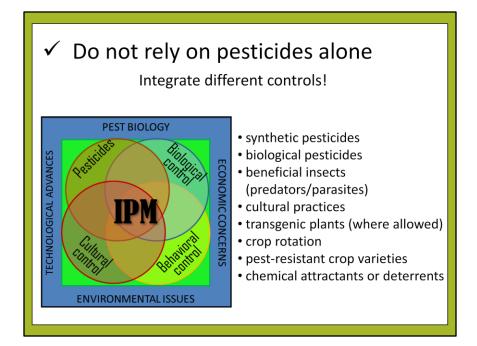
Qualitative Resistance

- Pest exhibits range in sensitivity.
- Gradual loss of control.
- Can be regained with higher rate or more active pesticide.

Key Points About Managing Resistance

- Goal is **delaying** development of resistance, <u>not</u> managing resistant pest biotypes once detected
- Use Integrated Pest Management (IPM) program
- Minimize use of at-risk products





Alternate, rotate, or sequence different pesticide MoA classes				
Use FRAC, IRAC, and HRAC when choosing chemicals!Do not rely on product names				
 Do not rely on active ingredients Many different products and active ingredients can be in the same group! 				
Alternaria leaf spot – Cantaloupe				
Fungicide FRA	FRAC Code			
Cabrio (pyraclostrobin)	11			
Pristine (pyraclostrobin + boscalid)	7 + 11			
Quadris (azoxystrobin) 11				
Quadris Top (azoxystrobin + difenoconazol 3 + 11				
Reason (fenamidone)	11			

The individual educators should change slide to represent a spray program that they would routinely use for their crop. Then the educator could pick and choose the ones that they focused on during the presentation.

Сгор	Target Diseases	Use Rate fl oz product/A (lb ai/A)	Remarks	
Cucurbits Cantaloupe Chayote Chinese-waxgourd Cucumber Gourds Honeydew Melons Momordica spp. (bitter melon, balsam apple) Muskmelon Watermelon Pumpkin Squash Zucchini Including cultivars and/or hybrids of these	Alternaria blight (Alternaria cucumerina) Anthracnose (Colletotrichum lagenarium) Belly rot (Rhizoctonia solani) Cercospora laf spot (Cercospora citrulina) Downy mildew (Pseudoperonospora cubensis) Gummy stem blight (Didymella bryoniae) Leaf spot (Alternaria spp., Cercospora spp.) Myrothecium canker (Myrothecium canker (Myrothecium maker (Myrothecium blight (Plectosporium blight (Sphaerotheca fuliginea, Ensiphe cichoracearum) Target leaf spot (Corynespora cassicola)	11.0-15.5 (0.18-0.25)	For both downy and powdery mildew, make preventative applications on a 5- to 7-day schedule. For belly rot control, the first application should be made at the 1-3 leaf crop stage with a second application just prior to vine tip over or 10-14 days later whichever occurs first. For all other diseases, Quadris applica- tions should begin prior to disease development and continue throughout the season every 7-14 days following the resistance management guidelines. Applications may be made by ground, air or chemigation. An adjuvant may be added at specified rates. Do not tank mix Quadris with crop oil concentrates (COC), methylated spray oil (MSO) or silicon adjuvants. Do not tank mix Quadris with Malathion, Kelthane®, Thiodan®, Phaser®, Lannate®, Lorsban®, M-Pede® or Botran®. Do not apply more than one applications of Quadris or other Group 11 fungicides before alternation with a fungicide that is not in Group 11. Do not make more than four (4) foliar applications of Quadris or other Group 11 fungicides per crop per acre per year.	Resistance management guidelines on label MUST be followed.

Applications must be timed correctly

- Target the most vulnerable life stage of the pest
- Use spray rates and application intervals recommended by the manufacturer and in compliance with local agricultural extension regulations.
 - A high rate can take out pests that might be somewhat resistant, but using too low a rate may allow them to survive

Challenges to Managing Resistance

- Do not have adequate tools
 - Not enough registered products to permit rotation
- The program may not be as effective
- The program may be more expensive
- Products with resistance risk for one pest are also used for others

Preserve susceptible genes

Preserve susceptible individuals within the target population by providing a **refuge** or haven for susceptible insects

- Unsprayed areas within treated fields
- Adjacent refuge fields

These susceptible individuals may out-compete and interbreed with resistant individuals, diluting the impact of any resistance that may have developed in the population.

Do genetically engineered crops (GMOs) contribute to resistance development?

Not directly!

Growers may manage those crops differently, encourage resistance development inadvertently

• Example: "RoundUp" ready crops, over reliance on single herbicide

Supporting funds were provided by NE-SARE Professional Development Program Grant, ENE15-140-29994



Presentation developed by K. Ghantous, H. Sandler, and L. McDermott.

With special thanks to:

- Dr. Margaret McGrath, Cornell University
- Dr. Andrei Alyokhin, University of Maine
- Dr. Richard Bonanno, University of Massachusetts

for their expertise and contribution of materials for presentation